Biomechanical constitutive modeling of the gastrointestinal tissues: where are we? SUPPLEMENTAL MATERIAL

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S1. Constitutive modeling of cell electrophysiology

In view of active models of GI tissues provided in Section 6, we briefly introduce GI electrophysiology for the sake of clarity. The active component of GI constitutive models depends on cellular electrophysiology. Interstitial cells of Cajal (ICC) and smooth muscle cells (SMCs) electrical activity dictates GI motility [1]. Here, plasma membrane proteins, acting as selective voltage-dependent ion channels (gates), rule inward and outward ionic fluxes and allow large excursions of the transmembrane voltage, known as action potential (AP). Unlike the heart, the generation and propagation of slow excitation waves in GI are realized by ICC. SMCs, then, get electrically excited, transforming these stimuli into mechanical responses upon threshold activation. Accordingly, the multiple lengths and time scales involved rely on simplified phenomenological approaches or detailed biophysical models [2]. We provide a concise basis of cell electrophysiology constitutive modeling within deformable substrate.

At the cell level, electrophysiological constitutive models are based on Kirchhoff's balance low (principle of conservation of total current), assuming the cell membrane composed by the superposition of capacitive and resistive contributions. When the tissue level is considered, such local representation is extended into a space-dependent constitutive description in which ion flux balances are homogenized into the so-called *cable equa-*

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tion (or monodomain model): this is a diffusive Fourier-like PDE accounting for resistive inter-cellular connections included in the conductivity tensor Σ . In general, the cell-cell conductivity incorporates heterogeneous and anisotropic features, e.g., a function of local spatial coordinate \mathbf{x} and material direction \mathbf{n} , respectively [3–11]. Recently, the active strain approach described in Section 6.2 has been coupled to gastric electrophysiology and successfully implemented in large-scale finite element models [12]. Such a multiphysics framework requires the pull-back operation applied on the monodomain model framing the current (deformed) configuration, in which constitutive laws are assigned, into a material (undeformed) reference configuration, amenable of numerical implementation frame invariance.

The cable equation in the material framework reads:

$$C_m \frac{\partial V}{\partial t} = \frac{1}{J} \nabla \cdot J \mathbf{C}^{-1} \mathbf{\Sigma}(\mathbf{x}, \mathbf{n}) \, \nabla V + I_{\text{ion}}(V, \mathbf{y}) \tag{1}$$

with $J = \det(\mathbf{F})$ and $\mathbf{C} = \mathbf{F}^T \mathbf{F}$ the right Cauchy-Green deformation tensor, C_m the membrane capacitance, I_{ion} the sum of the ionic currents contributing to the membrane potential V, \mathbf{y} a list of local kinetics associated with voltage- or ion-dependent channels regulated by specific reaction rates, and $\mathbf{g}(V, \mathbf{y})$ is a vector of coupled nonlinear functions, readily:

$$\frac{\mathrm{d}\mathbf{y}}{\mathrm{d}t} = \mathbf{g}(V, \mathbf{y}) \tag{2}$$

Other general models proposed to reproduce GI electrical behavior comprise the *bidomain*, and *extended bidomain* (also named tridomain) models [13], further incorporating the inter-cellular space among two different cell types. These generalized constitutive descriptions allow investigating advanced phenomena, e.g., external electric stimulation of the tissue, though poorly applied in the GI context [14].

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Table S1: Search terms and combinations used on PubMed to identify relevant studies for each GI organ.

Organ	Search terms and combinations
Esophagus	(constitutive AND (model OR modeling OR modelling)) OR (mechanical AND (model OR modeling OR modelling)) OR (mechanical AND (active OR passive)) OR (mechanical AND stress AND strain) OR (strain AND energy) OR (finite AND element) OR ((viscoelastic OR viscoelasticity) AND (model OR modeling OR modelling)) AND (esophagus OR esophageal)
Stomach	(constitutive AND (model OR modeling OR modelling)) OR (mechanical AND (model OR modeling OR modelling)) OR (mechanical AND (active OR passive)) OR (mechanical AND stress AND strain) OR (strain AND energy) OR (finite AND element) OR ((viscoelastic OR viscoelasticity) AND (model OR modeling OR modelling)) AND (stomach OR (gastric AND tissue))
Small intestine	(constitutive AND (model OR modeling OR modelling)) OR (mechanical AND (model OR modeling OR modelling)) OR (mechanical AND (active OR passive)) OR (mechanical AND stress AND strain) OR (strain AND energy) OR (finite AND element) OR ((viscoelastic OR viscoelasticity) AND (model OR modeling OR modelling)) AND (small intestine OR duodenum OR jejunum OR ileum)
Large intestine	(constitutive AND (model OR modeling OR modelling)) OR (mechanical AND (model OR modeling OR modelling)) OR (mechanical AND (active OR passive)) OR (mechanical AND stress AND strain) OR (strain AND energy) OR (finite AND element) OR ((viscoelastic OR viscoelasticity) AND (model OR modeling OR modelling)) AND (large intestine OR colon OR sigmoid OR cecum)
Rectum	(constitutive AND (model OR modeling OR modelling)) OR (mechanical AND (model OR modeling OR modelling)) OR (mechanical AND (active OR passive)) OR (mechanical AND stress AND strain) OR (strain AND energy) OR (finite AND element) OR ((viscoelastic OR viscoelasticity) AND (model OR modeling OR modelling)) AND (rectum OR rectal)

Table S2: List of studies where a hyperelastic Fung-type model is presented. We refer to Equation (8) and Equation (10) in the manuscript for the definitions of the parameters a and b, respectively. The numbers of parameters a and b correspond to the best performing model in case multiple configurations were tested.

Study	Organ	Test protocol	Modeled entity	Number of parameters a (and b when applicable)
Miftakhov & al. (1994) [15]	Small intestine	Numerical adjustment	Intact tissue	3 (3)
Dou & al. (2003) [16]	Small intestine	Tubular inflation	Intact tissue	3
Liao & al. (2003) [17]	Esophagus	Tubular inflation	Intact tissue - Individual layer	6
Zhao & al. (2003) [18]	Small intestine	Tubular inflation	Intact tissue	3
Yang & al. (2004) [19]	Esophagus	Tubular extension - Tubular inflation - Tubular torsion	Intact tissue	3
Zeng & al. (2004) [20]	Esophagus	Tubular extension - Tubular inflation - Tubular torsion	Intact tissue	3
Stavropoulou & al. (2009) [21]	Esophagus	Tubular extension - Tubular inflation	Individual layer	3

Liao & al. (2010) [22]	Small intestine	Tubular extension - Tubular inflation	Intact tissue	6
Sokolis & al. (2010) [23]	Esophagus	Tubular extension - Tubular inflation	Intact tissue - Individual layer	3
Bellini & al. (2011) [24]	Small intestine	Planar biaxial	Intact tissue	3
Sokolis & al. (2011) [25]	Large intestine & Rectum	Tubular extension - Tubular inflation	Intact tissue	3
Stavropoulou & al. (2012) [26]	Esophagus	Planar uniaxial	Individual layer	3
Aydin & al. (2017) [27]	Stomach	Planar biaxial	Intact tissue	3
Sokolis (2017) [28]	Small intestine	Tubular extension - Tubular inflation	Intact tissue	3 (3)
Sun & al. (2017) [29]	Small intestine	Tubular extension - Tubular inflation - Tubular torsion	Intact tissue	7

Table S3: List of studies where an isotropic hyperelastic phenomenological model is proposed.

Study	Organ	Test protocol	Modeled entity	Hyperelastic model type
Gao & al. (2008) [30]	Stomach	Planar uniaxial	Intact tissue	Ogden
Boubaker & al. (2009) [31]	Rectum	Planar uniaxial	Intact tissue	Ogden
Tran & al. (2011) [32]	Large intestine	Compression - Planar uniaxial	Intact tissue	Ogden
Rubod & al. (2012) [33]	Rectum	Planar uniaxial	Intact tissue	Mooney-Rivlin
Puértolas & al. (2013) [34]	Large intestine	Planar uniaxial	Intact tissue	Mooney-Rivlin
Boubaker & al. (2015) [35]	Rectum	Planar uniaxial	Intact tissue	Ogden
Dargar & al. (2016) [36]	Stomach	Compression	Individual layer (mucosa, submucosa, and muscle)	2nd order reduced polynomial
Zhou & al. (2018) [37]	Large intestine	Planar uniaxial	Intact tissue	Mooney-Rivlin extended

Table S4: List of studies where a Holzapfel-type hyperelastic model is proposed.

Study	Organ	Test protocol	Modeled entity	Number of fiber families (details about directions)
Ciarletta & al. (2009) [38]	Large intestine	Planar uniaxial - Planar shear	Intact tissue	4 (longitudinal, circumferential, and two symmetrical directions)
Sokolis & al. (2013) [39]	Esophagus	Tubular extension - Tubular inflation	Intact tissue - Individual layer (mucosa and mucosa-submucosa)	3 (longitudinal and two symmetrical directions)
			Individual layer (muscle)	4 (longitudinal, circumferential, and two symmetrical directions)
Sokolis & al. (2013) [40]	Large intestine & Rectum	Tubular extension - Tubular inflation	Intact tissue	3 (longitudinal and two symmetrical directions)
Sommer & al. (2013) [41]	Esophagus	Planar uniaxial - Planar biaxial - Tubular extension - Tubular inflation	Individual layer (mucosa-submucosa and muscle)	1
Patel & al. (2018) [42]	Large intestine	Tubular extension - Tubular inflation	Intact tissue	3 (longitudinal and two symmetrical directions)

Puertolas & al. (2019) [43]	Large intestine	Planar biaxial	Intact tissue	4 (longitudinal, circumferential, and two symmetrical directions)
Zhao & al. (2020) [44]	Large intestine & Rectum	Planar biaxial - Tubular inflation	Individual layer (mucosa-submucosa and muscle)	1
Sokolis (2021) [45]	Small intestine	Tubular inflation - Tubular extension	Intact tissue	2 (longitudinal and circumferential)

Table S5: List of studies where Natali-type hyperelastic model is presented.

Study	Organ	Test protocol	Modeled entity
Natali & al. (2009) [46]	Esophagus	Planar uniaxial	Individual layer (mucosa-submucosa and muscle)
Carniel & al. (2013) [47]	Large intestine	Planar uniaxial	Intact tissue
Carniel & al. (2014) [48]	Large intestine	Planar uniaxial	Intact tissue

Table S6: List of studies where a visco-hyperelastic model is presented.

Study	Organ	Test protocol	Modeled entity	Hyperelastic model type	Viscous model type
Higa & al. (2007) [49]	Large intestine	Tubular inflation	Intact tissue	Neo-Hookean	Convolution integral
Fontanella & al. (2019) [50]	Stomach	Tubular inflation	Intact tissue	Natali	Viscous variables
Panda & al. (2019) [51]	Small intestine	Tubular inflation	Intact tissue	Neo-Hookean	Panda type linear and non linear
	Large intestine	Tubular inflation	Intact tissue	Fung	Panda type linear
	Rectum	Tubular inflation	Intact tissue	Neo-Hookean	Panda type linear

Table S7: List of studies where an active model is presented.

Study	Organ	Test protocol	Modeled entity	Hyperelastic model n category	Viscous nodel type	Active model category
Yang & al. (2007) [52]	Esophagus	Planar uniaxial - Tubular inflation - Other	Intact tissue - Individual layer (mucosa- submucosa and muscle)	Structure- based	NA	Active stress
Yassi & al. (2009) [53]	Esophagus	NP	Individual layer (muscle)	Phenomenologica	l NA	Active stress
Gao & al. (2009) [54]	Small intestine	Impedance planimetry	Intact tissue	NA	NA	Active stress
Altomare & al. (2014) [55]	Large intestine	Planar uniaxial - Other	Individual layer (muscle)	Structure- based		Active strain
Kou & al. (2015) [56]	Esophagus	NA	Individual layer	NA	NA	Active stress
Gizzi & al. (2016) [57]	Small intestine	Planar biaxial	Intact tissue (muscosa- submucosa and muscle)	Structure- based	Neo- Hookean	Active electromechanics

Peirlinck & al. (2018) [58]	Esophagus	Planar uniaxial - Planar biaxial - Tubular extension - Tubular inflation	Individual layer (mucosa, submucosa, and muscle)	Structure- based	NA	Active strain
Brandstaeter & al. (2018) [14]	Stomach	NA	Intact tissue	Phenomenological	NA	Active strain
Klemm & al. (2020) [59]	Stomach	Planar uniaxial	Individual layer (muscosasubmucosa and muscle)	Structure- based	NA	Active stress

Table S8: List of studies where a model is developed based on human tissue testing.

Study	Organ	Test protocol	Modeled entity	Hyperelastic model category	Viscous model type	Active model category
Boubaker & al. (2009) [31]	Rectum	Intact tissue	Ex vivo	Ogden	NA	NA
Gao & al. (2009) [54]	Small intestine	Intact tissue	In vivo	NA	NA	Active stress
Rubod & al. (2012) [33]	Rectum	Intact tissue	Ex vivo	Mooney- Rivlin	NA	NA
Puértolas & al. (2013) [34]	Large intestine	Intact tissue	Ex vivo	Mooney- Rivlin	NA	NA
Altomare & al. (2014) [55]	Large intestine	Individual layer (muscle)	Ex vivo	Holzapfel	NA	Active strain
Panda & al. (2019) [51]	Small intestine	Intact tissue	Ex vivo	Neo- Hookean	Panda-type linear and nonlinear	NA