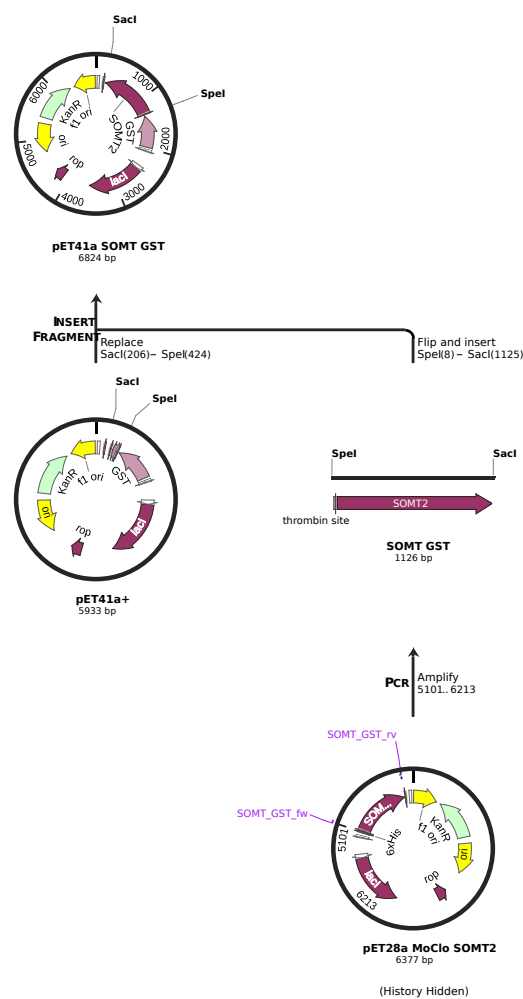


# WEB324 - Cloning SOMT into GST-tag containing pET41a(+)

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**Figure 1:** Cloning strategy towards GST-tagged SOMT2. pET41a(+)-SOMT.

# 1 Cloning to pET41a(+)

**ATTENTION:** SOMT gene has an endogenous BsaI cleavage site → site needs to be removed

**Table 1:** The primers used for cloning.

name	sequence [5'→3']
somt_gst_fw	cat cat act agt AGC AGC GGC CTG GTG
somt_gst_rv	tt gaagac aa gag ctc TCA AGG ATA GAT CTC AAT

## 1.1 PCR SOMT2

- cloning from pET28a MoClo SOMT2 (2) [51  $\frac{ng}{\mu l}$ ]

### Reaction mix:

substance	volume	final concentration
10x KOD Buffer	5 $\mu l$	1x
25 mM MgSO <sub>4</sub>	3 $\mu l$	1.5 mM
2 mM dNTPs	5 $\mu l$	0.2 mM
51 ng/ $\mu l$ template	0.25 $\mu l$	12.5 ng
10 pmol/ $\mu l$ SOMT_GST_fw	1.5 $\mu l$	
10 pmol/ $\mu l$ SOMT_GST_rv	1.5 $\mu l$	
KOD HS Polymerase	1 $\mu l$	1 $\mu l$
water	32.75 $\mu l$	

**PCR Program:** theoretical size = 1100 bp

Step	Description	Temperature	Time
1	Denaturation	95°C	2 min
2	Denaturation	95°C	20 sec
3	Annealing	55°C	10 sec
4	Extension	70°C	21 sec
5	Final Extention	70°C	2 min
	Pause	4°C	∞

repeat 2-3 24x



**Figure 2:** Agarose gel of PCR to amplify SOMT fragment

## 2 Digestion of pET41 and fragment by *SpeI* (*BcuI*) and *SacI*

### Plasmid digestion:

component	amount	volume
FD Buffer Green		1 $\mu$ l
pET41a(+)	1 $\mu$ g	2 $\mu$ l
<i>SacI</i> FD		0.5 $\mu$ l
<i>BcuI</i> FD		0.5 $\mu$ l
ad to 10 $\mu$ l MQ-H <sub>2</sub> O (6 $\mu$ l )		

### fragment digestion:

component	amount	volume
FD Buffer Green		1 $\mu$ l
SOMT fragment	1 $\mu$ g	5 $\mu$ l
<i>SacI</i> FD		0.5 $\mu$ l
<i>BcuI</i> FD		0.5 $\mu$ l
ad to 10 $\mu$ l MQ-H <sub>2</sub> O (3 $\mu$ l )		

- digested plasmid was cleaned by agarose gel > cut band > MN gel cleanup (eluted with H<sub>2</sub>O: 40 ng/ $\mu$ l )
- digested fragment was only cleaned by MN PCR cleanup (eluted with H<sub>2</sub>O: 46 ng/ $\mu$ l )

### 2.1 Second Ligation

#### Ligation:

component	amount	volume
T4 ligase buffer	1x	1 $\mu$ l
cut pET41a(+)	~20 ng	0.5 $\mu$ l
cut fragment	?	0.5 $\mu$ l
T4 ligase		0.25 $\mu$ l
ad to 10 $\mu$ l MQ-H <sub>2</sub> O (7.5 $\mu$ l )		

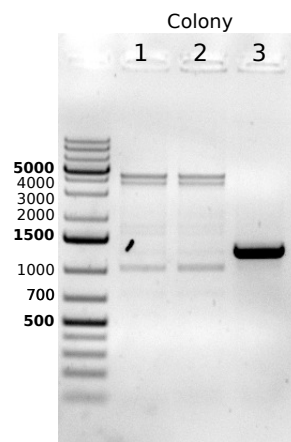
- ligated at 22°C for 1 h and 4°C over the weekend

**Colony PCR:** Several small colonies → colony PCR (SOMT1, T7\_term as primers) → mini-prep of clone 3 and sequencing → Sequencing results were screwed up → Ligation didn't work. → **ligate again**

#### Colony PCR:

component	amount	volume (* samples)
DreamTaq Buffer	1x	2 $\mu$ l
SOMT1 primer		0.4 $\mu$ l
T7_term primer		0.4 $\mu$ l
dNTP mix		0.4 $\mu$ l
DreamTaq Polymerase	0.5 U	0.1 $\mu$ l
ad to 20 $\mu$ l MQ-H <sub>2</sub> O (16.7 $\mu$ l )		

For more samples the volumes need to be multiplied by the number of samples.



**Figure 3:** Agarose gel of colony-PCR. The theoretical size of the fragment, if the ligation was successful is 1216 bp. Only clone 3 gave a fragment with a comparable size. This clone was sequenced.

## 2.2 Second Ligation

#### Ligation:

component	amount	volume
T4 ligase buffer	1x	1 $\mu$ l
cut pET41a(+)	~20 ng	0.5 $\mu$ l
cut fragment	18.98 ng	0.41 $\mu$ l
T4 ligase		0.25 $\mu$ l
ad to 10 $\mu$ l MQ-H <sub>2</sub> O (7.84 $\mu$ l )		

- ligated at 22°C for 1 h and 4°C over night

No colonies grew on the plate.