

Adding a new model to NPSGD

Thomas Dimson

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1 Introduction

NPSGD was created with the design goal of making the addition of new models easy. All models are actually just python classes that inherit from a specific base class. By adding names and specific parameters, NPSGD will do the work of shipping the parameters from the web interface into the module. It is up to the module to perform the work of running the model and giving meaningful output to the user.

2 Quick Start

All models are added to the `models/` subdirectory and are really just python modules in disguise. The quickest start possible is to copy `example.py` to a new filename called `my_model.py` and change `short_name` to `my_model`. Visit http://npsgdserver:8000/models/my_model and your model will be up and running.

Please note that you will have to make sure that the model is available on all servers running NPSGD daemons (queue, web, and workers).

3 Models

In NPSGD, a “model” is actually a Python class that inherits, however distantly, from the base class of `ModelTask`. This gives the user the flexibility to be language-agnostic in terms of model implementation, with a quick Python wrapper as a model runner.

All models (with a `.py` extension) are placed in the `models/` subdirectory. Periodically, the **queue**, **web**, and **worker** daemons will scan this directory for conforming (or newly timestamped) classes and load them into memory. From there, the model will be immediately accessible from the web, and therefore from the worker and queue. By default, the scanner propagation time is 120 seconds.

4 Helper classes

Rather than inheriting from `ModelTask` directly, NPSGD provides two helper classes that can greatly speedup creation of models. These are `MatlabTask` and `StandaloneTask`.

4.1 StandaloneTask

Models inheriting from `StandaloneTask` are wrappers for standalone executables that communicate using command line arguments (e.g. specifying the path to specific data input). When a model executes the pre-requisites are prepared in the working directory, and a subprocess is spawned using the executable and command line parameters.

4.2 MatlabTask

`MatlabTask` aids in connecting models that were programmed in Matlab. Essentially, this allows the user to access web parameters *directly* in Matlab code, bypassing the need worry about technical details such as spawning subprocesses to run the model.

5 NPSGD Pipeline

The purpose of NPSGD is to perform all the administrative work needed to deliver data to your model implementation. It is useful to understand exactly the processes that occurs in order for this to happen, though the model implementor will not have to touch this pipeline at all.

1. Model, implemented in `models/example.py` is noticed and loaded by all NPSGD daemons
2. Online web user visits the web interface for the model, typically at `http://npsgdserver:8000/models/example`. The user specifies all the parameters present in the model and clicks submit
3. Web daemon `npsgd_web` submits the request to the queue
4. Queue daemon, `npsgd_queue` sends an email to the user with a confirmation code for the request.
5. User receives email, clicks the confirmation code link which is typically `http://npsgdserver:8000/confirm_submission/code`. The web daemon submits the confirmation code to the queue daemon at this time.
6. Queue daemon waits for a worker to poll. When a worker polls with this model available, queue will hand off the request.
7. Worker spawns the model class with the parameters that were specified in the web interface. Model proceeds with model pipeline documented in Section 6
8. Model pipeline completes, sends an e-mail to the user and then tells the queue that everything has completed successfully

6 Model Pipeline

The model pipeline is the section that a model implementor will actually have to create in the model. Typically, these follow a strict order that is specified in `ModelTask` class under the method `run`:

1. A working directory is created for the model (usually `/var/tmp/npsgd/unique.id`).
2. Execution is setup in the `prepareExecution` method. This is generally done by converting parameters into a form (e.g. a file in the working directory) that the model can work.
3. The model is run under the `runModel` method. A typical model run includes calling the subprocess with specified parameters and waiting for the result.
4. Graphs are prepared under the `prepareGraphs` method using the data that was outputted from `runModel`.
5. PDF document is created in `generatePDF`, through `getAttachments` using PDFLatex.
6. E-mail is sent using `sendResultsEmail`

Note that the pipeline can be greatly simplified by using the helper methods outlined in Section 4.

7 Model Implementation

This section is a basic guide about specifying a model to run.

7.1 Defining a class

In order to create a model, you must inherit from `ModelTask`, or one of the helpers specified in Section 4. After doing so, you have to save your model under `models/my_model.py`. For example, a skeleton model could take the form of

```
from npsgd.model_task import ModelTask
from npsgd.model_parameters import *

class MyModel(ModelTask):
    short_name = "my_model"
    full_name = "My Model"
    subtitle = "The best model money can buy"
```

7.2 Parameters

Models are pretty useless without parameters which the user can play around with in order to get different result. All parameters are specified in the class structure itself, under a list with the name `parameters`. As an example to start, we could specify an integer representing the number of samples to take into account during a simulation, with a default of 1000:

```
...
class MyModel(ModelTask):
    ...
    parameters = [
        ...
        IntegerParameter('nSamples', description="Number of
            samples to specify",
            default=1000)
        ...
    ]
```

7.2.1 Parameter Options

All parameters support a variety of different options in order to modify behaviour. The first value passed into the parameter is always the **name**, which is used as a reference through NPSGD. The rest of the options are specified using keyword arguments. Each parameter type outlined in Section 7.2.2 has a superset of these options:

Option	Description
name	Unique identifier used to reference the parameter through NPSGD. These generally should not have spaces, and by convention are camel cased.
description	As it says, used to describe the parameter. The string specified here will appear in a number of places, including the output L ^A T _E X document, and the html page.
default	Used to specify the default value of a parameter.
hidden	Boolean used to toggle whether the parameter is hidden or not. Currently, this only affects the output html. Hidden parameters will simply be passed along with their default value. This is useful for subclassing models
helpText	Used to give “helpful hints” when a user is specifying the model. This currently appears as balloon text on the html output page.

7.2.2 Parameter Types

Currently, NPSGD supports the following parameter types:

- **StringParameter**: Basic string input. The only extra option is **units**, which represents the units of input.
- **FloatParameter**: Basic float input. Float parameters can be used to specify a specific range of inputs by specifying the **rangeStart**, **rangeEnd** and **step** inputs. If both **rangeStart** and **rangeEnd** are specified, the float parameter will function as a slider. If only **rangeStart** or **rangeEnd** are specified, the input will be clamped to values accordingly. Additionally, floats take a **units** parameter that can be used to specify the units of the float (e.g. nm, cm)
- **IntegerParameter**: Basic integer input. Integers have exactly the same options as **FloatParameter**, but will be verified to ensure integrality.
- **RangeParameter**: Range parameters are used for specifying a range of floating point inputs (e.g. 400-2500nm). Options for **RangeParameter** match **FloatParameter**, but must have all of **rangeEnd**, **rangeEnd** and **step** specified. The output and default values of a range must be specified as a pair of floats, e.g. **default**=(1,5), representing the range of choices.
- **SelectParameter**: Select parameters are used to clamp input to a specific set of options, something like a combo box. Select parameters have an option called **options**, which is a list of valid inputs that the select box can take (e.g. “Strong”, “Weak”).
- **BooleanParameter**: Boolean parameters are similar to the **SelectParameter** type, but now are clamped to true/false. These parameters will display as text boxes in html and have no additional options.

7.2.3 Attachments

By default, NPSGD includes only the PDF created via L^AT_EX, outlined in Section 7.2.5 as an attachment. If your model creates more output (such as data files, graphs, pictures, etc.) then you may want to add additional attachments. In NPSGD, this is performed by adding a class variable by the name of **attachments** consisting of a list of all additional attachments within the working directory to include. For example,

```

...
class MyModel(ModelTask):
    ...
    attachments = [picture.jpg, data.txt]
    ...

```

The above code listing would include two e-mail attachments (`picture.jpg` and `data.txt`) along with the usual `results.pdf`.

If you want more flexibility in specifying attachments, consider overriding the `getAttachments` method on `ModelTask`.

7.2.4 Graphical Output

After an executable has run, usually a task will create graphs out of the output. In NPSGD, this can be accomplished by saving files in the working directory by overriding the `prepareGraphs` method in a model. Python has an excellent library called `matplotlib` (<http://matplotlib.sourceforge.net/>) which creates graphs that are on-par with Matlab's using commands that are almost identical to Matlab plot syntax. For example,

```

...
import os
import matplotlib
matplotlib.use("Agg") #suppress graphical user interface
import matplotlib.pyplot as plt
...
class MyModel(ModelTask):
    ...
    attachments = [plot.png]
    ...
    def prepareGraphs(self):
        x = [1,2,3,4,5]
        y = [10,5,2,6,7]
        plt.clf() #clear previous plot
        plt.plot(x,y)
        plt.xlabel("X")
        plt.ylabel("Y")
        plt.title("Demo Plot")
        plt.savefig(os.path.join(self.workingDirectory, "plot.
                               png"))
    ...

```

Note that the `prepareGraphs` method is completely optional - if your model does not have graphical output, or creates the output inside the executable then you may include the relevant files using the attachment mechanism outlined in Section 7.2.3.

7.2.5 Latex Output

Output for all models is generally routed through PDFLatex. Each model will **definitely** want to override the `latexBody` method. This method returns a string that is then run through PDFLatex

in order to generate a file called `result.pdf`. This file, by default, is included in every e-mail that is sent out.

`ModelTask` has a useful method for creating a `LATEX`table containing all the parameters that the user has specified. By including `self.latexParameterTable()` somewhere in the `LATEX`output the output pdf will contain a very nicely formatted parameter table.

A complete example:

```
...
class MyModel(ModelTask):
    ...
    def latexBody(self):
        return r"""
        Hello there!

        \section{Main Section}
        This is the result of an example model run for NPSGD.
        Your
        parameters were:

        %s
""" % self.latexParameterTable()
```

It is highly recommended that you use python docstrings (triple quoted strings) in order to specify output, as well as using the `r` prefix to the string (raw string mode, so you don't have to escape slashes).

7.2.6 Reading Parameter Values

When executing your script, preparing graphs and outputting `LATEX` it is often necessary to have access to the parameters that the user has specified at the web interface. These are *automatically* delivered to the script using the names that you specified in for your parameters. By accessing `self.parametername.value` in any instance method, you will get access to the value that the user specifies. This is best illustrated by example:

```
...
class MyModel(ModelTask):
    ...
    parameters = [
        ...
        IntegerParameter('nSamples', description="Number of
        samples to specify",
        default=1000)
        ...
    ]

    def prepareGraphs(self):
        print self.nSamples.value #Will output the number of
        samples the
        #user specified
```

7.2.7 Helpers: StandaloneTask

Subclassing `StandaloneTask` automates the process of running a subprocess in order to execute a model on the command line. This is the most technical part of the process.

`StandaloneTask` specifies a method of `runModel` that simply executes a command as a python subprocess, and stores the stderr/stdout of the subprocess in instance parameters `self.stdout` and `self.stderr`. The subprocess is executed within the model's working directory.

The model creator must specify one additional parameter, and one additional method for running. The class variable `executable` specifies the path to the executable we wish to run (typically the model executable, or something like java for a java task). The instance method `executableParameters` returns a list of parameters for the executable along with values. The parameters are specified as a python list. This is best shown via example:

```
from npsgd.model_task import StandaloneTask
from npsgd.model_parameters import *

class LsModel(StandaloneTask):
    ...
    executable = "ls"
    ...
    def executableParameters(self):
        return [
            "-al",
            "/var/tmp"
        ]
```

Such a model will execute `ls` in a subprocess and return the results in `self.stdout`. Many more examples ship along with NPSGD.

7.2.8 Helpers: MatlabTask

Subclassing `MatlabTask` automates the process of spawning a matlab subprocess, which can be a tricky and time consuming process. It also delivers the parameter values **directly** into the matlab environment. The script will just “magically” have access to all the values of user input in variables that match the names specified in the model class.

A user of the `MatlabTask` helper need only specify one additional class parameter, namely `matlabScript` which gives the location of the script Matlab should execute. A full example of using `MatlabTask` is specified in Appendix A.

A Complete Example of a Matlab Task

A.1 NPSGD Code

```
from npsgd.matlab_task import MatlabTask
from npsgd.model_parameters import StringParameter, IntegerParameter
    , RangeParameter, FloatParameter

class ExampleModel(MatlabTask):
    short_name = 'example'
```



```

subtitle    = 'A demo model'

parameters = [
    StringParameter('test',      description="This a test string
    "),
    IntegerParameter('graphEnd', description="Graph end point"),
    RangeParameter('ranger',     description="Sample range
    parameter",\
        rangeStart=400, rangeEnd=700, step=1),
    FloatParameter('floater',    description="Sample Float
    Parameter", rangeStart=10, rangeEnd=1000, step=1)
]

matlabScript = '/home/tdimson/public_html/npsg/npsgd/models/
example/example.m'

def latexBody(self):
    return r"""
        This is a test of including a figure.
        \begin{figure}
        \caption{A nice looking function}
        \includegraphics[width=5in]{test_figure}
        \end{figure}

        \newpage\appendix\section{Parameter List}
        %s
    """ % self.latexParameterTable()

```

A.2 Matlab Code

```

x = rangerStart:1:rangerEnd;
y = x.^2;

plot(x,y)
title('Plot of y = x^2')
print -dpng test_figure

```

B Complete Example of ABM-U

ABM-U is a more sophisticated example of a model task. An example of this code running is available at <http://www.npsg.uwaterloo.ca/models/ABMU.php>.

```

import os
import sys
import csv
import json
import matplotlib
matplotlib.use("Agg")
import matplotlib.pyplot as plt

```

```

from npsgd.standalone_task import StandaloneTask
from npsgd.model_parameters import *

class ABMU(StandaloneTask):
    short_name = 'abmu_c'
    full_name = 'ABM-U'

    subtitle='Algorithmic BDF Model Unifacial'
    parameters = [
        IntegerParameter('nSamples', description="Number of
            samples",
            rangeStart=1000, rangeEnd=100000, step=1, default
            =10000),
        RangeParameter('wavelengths', description="Wavelengths",
            rangeStart=400, rangeEnd=2500, step=5, units="nm",
            helpText="Output will generated in steps of 5nm."
            ),
        FloatParameter('angleOfIncidence', description="Incident
            angle",
            default=8, rangeStart=0, rangeEnd=90, step=0.1,
            units="degrees"),
        SelectParameter('surfaceOfIncidence', description="
            Surface of incidence",
            options=["Adaxial", "Abaxial"], default="Abaxial",
            helpText="Adaxial is the top epidermal layer of the
            leaf, abaxial is the bottom epidermal layer of
            the leaf."),
        FloatParameter('wholeLeafThickness', description="Leaf
            thickness",
            default=2.04e-4, units="m"),
        FloatParameter('mesophyllPercentage', description="
            Mesophyll percentage",
            default=80, units="%", rangeStart=0, rangeEnd=100,
            step=0.1,
            helpText="Percentage of the total leaf thickness
            occupied by the mesophyll tissue."),
        FloatParameter('proteinConcentration', description="
            Protein concentration",
            default=0.05308714, units="g/cm^3", rangeStart=0.0),
        FloatParameter('celluloseConcentration', description="
            Cellulose concentration",
            default=0.05318708961, units="g/cm^3", rangeStart
            =0.0),
        FloatParameter('linginConcentration', description="
            Lingin concentration",
            default=0.006058529380, units="g/cm^3", rangeStart
            =0.0),
    ]

```

```

FloatParameter('chlorophyllAConcentration', description=
    "Chlorophyll A concentration",
    default=0.002895146, units="g/cm^3", rangeStart=0.0)
,
FloatParameter('chlorophyllBConcentration', description=
    "Chlorophyll B concentration",
    default=0.00079866, units="g/cm^3", rangeStart=0.0),
FloatParameter('carotenoidConcentration', description="
    Carotenoid concentration",
    default=0.000658895, units="g/cm^3", rangeStart=0.0)
,
FloatParameter('cuticleUndulationsAspectRatio',
    description="Cuticle undulations aspect ratio",
    default=10.0, rangeStart=1.0, rangeEnd=50.0, step
        =0.5,
    helpText="Lower values result in more roughness and
        a more diffuse behaviour."),
FloatParameter('epidermisCellCapsAspectRatio',
    description="Epidermis cell caps aspect ratio",
    default=5.0, rangeStart=1.0, rangeEnd=50.0, step
        =0.5,
    helpText="Lower values correspond to more prolate (
        or rough) cell caps. This results in more
        diffusion of the propegated light."),
FloatParameter('spongyCellCapsAspectRatio', description=
    "Spongy cell caps aspect ratio",
    default=5.0, rangeStart=1.0, rangeEnd=50.0, step
        =0.5,
    helpText="Lower values correspond to more prolate (
        or rough) cell caps. This results in more
        diffusion of the propegated light."),
BooleanParameter('sieveDetourEffects', description="
    Simulate sieve and detour effects",
    default=True, helpText="To account for the non-
        homogeneous distribution of pigments (for details
        , please refer to our related publications).")
]

attachments = ['spectral_distribution.csv', 'reflectance.png',
    'transmittance.png', 'absorptance.png']

executable = "/home/tdimson/public_html/npsg/abmb-abmu_cpp/abmu"

def executableParameters(self):
    if self.surfaceOfIncidence.value == "Adaxial":
        angleIn = 180 - self.angleOfIncidence.value
    else:
        angleIn = self.angleOfIncidence.value

```

```

params = [
    "-d", os.path.join(os.path.dirname(self.executable), "
        data"),
    "-n", str(self.nSamples.value),
    "-p", str(angleIn),
    "-s", str(5), #step
    "-w", str(self.wavelengths.value[0]),
    "-e", str(self.wavelengths.value[1]),
]

if not self.sieveDetourEffects.value:
    params.append("-q")

params += ["sample.json",
           "spectral_distribution.csv"]

return params

def readDataTable(self):
    wavelengths, reflectance, transmittance, absorptance = ([],
        [], [], [])
    with open(os.path.join(self.workingDirectory, "
        spectral_distribution.csv"), 'r') as f:
        spectralReader = csv.reader(f)
        headers = [e.strip() for e in spectralReader.next()]
        wIndex = headers.index("wavelength")
        rIndex = headers.index("reflectance")
        tIndex = headers.index("transmittance")
        aIndex = headers.index("absorptance")

        for row in spectralReader:
            wavelengths.append(float(row[wIndex]))
            reflectance.append(float(row[rIndex]))
            transmittance.append(float(row[tIndex]))
            absorptance.append(float(row[aIndex]))

    return wavelengths, reflectance, transmittance, absorptance

def latexDataTable(self):
    wavelengths, reflectance, transmittance, absorptance = self.
        readDataTable()
    latex = r"""
    \begin{centering}
    \begin{longtable}{l l l l}

```

```

\textbf{Wavelength} & \textbf{Reflectance} & \textbf{Transmittance} & \textbf{Absorptance} \\
\hline
\endhead
%s
\end{longtable}
\end{centering}
""" % "\n".join("%snm & %s & %s & %s\\\\" % (w,r,t,a) for (w
,r,t,a) in zip(wavelengths,reflectance, transmittance,
absorptance))

return latex

def prepareExecution(self):
    with open(os.path.join(self.workingDirectory, "sample.json")
, 'w') as f:
        f.write(json.dumps({
            "wholeLeafThickness": self.wholeLeafThickness.value,
            "cuticleUndulationsAspectRatio": self.
                cuticleUndulationsAspectRatio.value,
            "epidermisCellCapsAspectRatio": self.
                epidermisCellCapsAspectRatio.value,
            "spongyCellCapsAspectRatio": self.
                spongyCellCapsAspectRatio.value,
            "palisadeCellCapsAspectRatio": 0.0,
            "linginConcentration": self.linginConcentration.
                value,
            "proteinConcentration": self.proteinConcentration.
                value,
            "celluloseConcentration": self.
                celluloseConcentration.value,
            "chlorophyllAConcentration": self.
                chlorophyllAConcentration.value,
            "chlorophyllBConcentration": self.
                chlorophyllBConcentration.value,
            "carotenoidConcentration": self.
                carotenoidConcentration.value,
            "mesophyllFraction": self.mesophyllPercentage.value
                / 100
        })))

def prepareGraphs(self):
    wavelengths, reflectance, transmittance, absorptance = self.
        readDataTable()
    axisWavelengthStart = wavelengths[0]
    axisWavelengthEnd = wavelengths[-1]
    plotCommand = plt.plot
    if len(wavelengths) == 1:

```

```

        axisWavelengthStart = wavelengths[0] - 100
        axisWavelengthEnd   = wavelengths[0] + 100
        plotCommand = plt.scatter

plt.clf()
plotCommand(wavelengths, [e*100 for e in reflectance])
plt.xlabel("Wavelength (nm)")
plt.ylabel("Reflectance (%)")
plt.title(self.full_name)
plt.axis([axisWavelengthStart, axisWavelengthEnd, 0, max(
    reflectance) * 100 + 5])
plt.savefig(os.path.join(self.workingDirectory, "reflectance
.pdf"))
plt.savefig(os.path.join(self.workingDirectory, "reflectance
.png"))
plt.clf()

plotCommand(wavelengths, [e*100 for e in transmittance])
plt.xlabel("Wavelength (nm)")
plt.ylabel("Transmittance (%)")
plt.title(self.full_name)
plt.axis([axisWavelengthStart, axisWavelengthEnd, 0, max(
    transmittance) * 100 + 5])
plt.savefig(os.path.join(self.workingDirectory, "
transmittance.pdf"))
plt.savefig(os.path.join(self.workingDirectory, "
transmittance.png"))
plt.clf()

plotCommand(wavelengths, [e*100 for e in absorptance])
plt.xlabel("Wavelength (nm)")
plt.ylabel("Absorptance (%)")
plt.title(self.full_name)
plt.axis([axisWavelengthStart, axisWavelengthEnd, 0, max(
    absorptance) * 100 + 5])
plt.savefig(os.path.join(self.workingDirectory, "absorptance
.pdf"))
plt.savefig(os.path.join(self.workingDirectory, "absorptance
.png"))
plt.clf()

def latexBody(self):
    return r"""
        These are the results of your model run of \textbf{ABM-U}
        } for the

```

*Natural Phenomenon Simulation Group (NPSG) at the
University of Waterloo.*

*The ABM-U employs an algorithmic Monte Carlo formulation
to simulate light interactions with unifacial plant
leaves
(e.g., corn and sugar cane). More specifically,
radiation propagation
is treated as a random walk process whose states
correspond
to the main tissue interfaces found in these leaves. For
more
details about this model, please refer to our related
publications~\cite{Ba06,Ba07}.
Although the ABM-U provides bidirectional readings,
directional-hemispherical quantities (provided by our
online system)
can be obtained by integrating the outgoing light (rays)
with respect
to the outgoing (collection)
hemisphere. Similarly, bihemispherical quantities can be
calculated
by integrating the BDF (bidirectional scattering
distribution function)
values with respect to incident and collection
hemispheres.*

*The provided spectral curves (directional-hemispherical,
reflectance,
transmittance and absorptance) were obtained considering
an angle of incidence
measured with respect to the specimen's normal (zenith).
The curves
were obtained using a virtual spectrophotometer~\cite{Ba01}.*

*The researcher interested in BDF
(bidirectional scattering distribution function)
plots is referred to a publication describing the
implementation of virtual
goniophotometers~\cite{Kr04}. These publications can be
found at:
\url{http://www.npsg.uwaterloo.ca/pubs/measurement.php}*

*\begin{figure}
\begin{centering}
\includegraphics[width=5in]{reflectance}
\caption{Directional-hemispherical reflectance}
\end{centering}*

```

\end{figure}

\begin{figure}
\begin{centering}
\includegraphics[width=5in]{transmittance}
\caption{Directional-hemispherical transmittance}
\end{centering}
\end{figure}

\begin{figure}
\begin{centering}
\includegraphics[width=5in]{absorptance}
\caption{Directional-hemispherical absorptance}
\end{centering}
\end{figure}

\newpage
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Virtual spectrophotometric Measurements for biologically
and physically-based rendering.
\textit{The Visual Computer}, Volume 17, Issue 8, pp.
506-518, 2001.

\bibitem{Ba06}
Baranoski G.V.G.
Modeling the interaction of infrared radiation (750 to
2500 nm) with bifacial and unifacial plant leaves.
\textit{Remote Sensing of Environment}, 100(3):335-347,
2006.

\bibitem{Ba07}
Baranoski G.V.G.; Eng D.
An investigation on sieve and detour effects affecting
the interaction of collimated and diffuse infrared
radiation (750 to 2500 nm) with plant leaves.
\textit{IEEE Transactions on Geoscience and Remote
Sensing}, 45 (8):2593-2599, 2007.

\bibitem{Kr04}
Krishnaswamy, A.; Baranoski, G.V.G.; Rokne, J.G.
Improving the reliability/cost ratio of goniophotometric
comparisons.
\textit{Journal of Graphics Tools}, Volume 9, Number 3,
pp. 1-20, 2004.
\end{thebibliography}

```



```

        \appendix
        \section{Parameter List}
        %s
        \section{Data List}
        %s
    """ % (self.latexParameterTable(), self.latexDataTable())

```