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# 干细胞治疗终末期肝病的进展与挑战

施明, 刘振文, 张政, 王福生

[摘要] 干细胞包括胚胎干细胞和成体干细胞,具有无限自我更新能力,可以分化成特定组织,修复各种损伤。骨髓是成体干细胞的重要来源,主要含骨髓造血干细胞和间充质干细胞。目前利用干细胞治疗终末期肝病越来越受到关注。本文从体外、动物实验和临床研究三个方面总结了干细胞特别是成体干细胞治疗终末期肝病的研究进展,重点阐述各种来源干细胞在临床应用中取得的进展,探讨干细胞治疗终末期肝病的可能机制,提出目前研究存在的问题,展望未来的应用前景。

[关键词] 干细胞; 肝移植; 肝疾病

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## Progress and challenge in the treatment of end-stage liver disease with stem cells

SHI Ming<sup>1</sup>, LIU Zhen-wen<sup>1</sup>, ZHANG Zheng<sup>2</sup>, WANG Fu-sheng<sup>2\*</sup>

<sup>1</sup>Research Center for Liver Transplantation, <sup>2</sup>Research Center for Biological Therapy, 302 Hospital of PLA, Beijing 100039, China <sup>\*</sup>Corresponding author, E-mail: fswang302@163.com

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[Abstract] Stem cells, including embryonic stem cells and adult stem cells, are multipotent cells that have self-renewing abilities and the potential to differentiate into various types of cells, and to repair various tissue damages. Bone marrow is the major source of adult stem cells, including mainly the bone marrow hematopoietic stem cells and mesenchymal stem cells. End-stage liver disease is a serious clinical systemic syndrome, and the use of stem cells in the treatment of end-stage liver disease has received more and more attention. This review summarizes the research progresses of stem cells, especially adult stem cells, in respect of the treatment of end-stage liver disease in the form of *in vitro* study, animal experimentation and clinical research, with emphasis on the research progress in the use of stem cells of various sources in clinical application, discussion of possible mechanisms of stem cell therapy for end-stage liver disease, to point out the problems existing in current research, and to look forward to the prospect of future application.

[Key words] stem cells; liver transplantation; liver diseases

肝移植是治疗终末期肝病最有效的方法,但供肝来源短缺与肝移植需求之间的巨大差距促使人们努力寻求其他的替代疗法。近来,干细胞治疗终末期肝病越来越受到关注,特别是骨髓来源的干细胞[主要含造血干细胞(HSC)和间质干细胞(MSC)]治疗已有体外和动物实验支持,自体回输骨髓干细胞治疗亦有相关临床报道。此外,其他来源的MSC治疗肝病亦取得了较好进展。本文就此方面的研究进展综述如下。

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[作者单位] 100039 北京 解放军302医院肝移植研究中心(施明、刘振文), 肝病生物治疗研究中心(张政、王福生)

[通讯作者] 王福生, E-mail: fswang302@163.com

### 1 研究背景

干细胞根据其发育阶段可分为胚胎干细胞(ES)和成体干细胞,其重要特征是具有无限自我更新能力,可以分化成特定组织,在细胞发育过程中处于较原始阶段。骨髓是成体干细胞的重要来源,主要含HSC和MSC。MSC在骨髓中含量最多,在其他组织器官如脐带组织、脐带血、胎盘、外周血、脂肪组织中的含量也较丰富。基于干细胞强大的分化潜能,人们开始尝试应用干细胞治疗各种疾病。自1999年Petersen等门发现骨髓中某些干细胞具有向肝细胞分化的潜能以来,人们开始研究应用干细胞治疗肝病的可行性,后来发现不同来源的干细胞的疗肝病的可行性,后来发现不同来源的干细胞如骨髓HSC、骨髓MSC等均可定向诱导分化为肝细胞样细胞,具有正常肝细胞功能,如分泌尿素和白蛋白

等<sup>[2]</sup>,且得到动物实验的验证,临床亦有自体回输骨髓干细胞治疗肝病的报道。目前开展的肝病治疗研究的干细胞类型如图1所示。

Has been in clinical application Not for clinical application

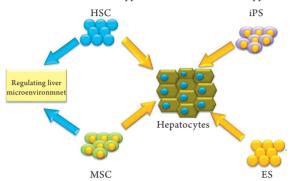


图1 目前已经开展的治疗肝病的干细胞类型及其治疗机制 Fig.1 Types of stem cells on current liver disease therapy and its mechanism

HSC. Hematopoietic stem cell; iPS. Induced pluripotent stem cell; MSC. Mesenchymal stem cells; ES. Embryonic stem cell

## 2 临床前研究

骨髓干细胞治疗肝病 以前认为只有具 备HSC特性的细胞才能转化为肝细胞, 1999年 Petersen等[1]发现骨髓干细胞或HSC能够在鼠肝内转 化为肝卵圆细胞甚至成熟的肝细胞和胆管细胞。 2000年Theise等[3]在异性间骨髓移植或肝移植的 受体中也发现了源于供体的肝细胞和胆管细胞。 Lagasse等[4]用正常小鼠骨髓细胞移植治疗延胡索酰 乙酰乙酸盐水解酶缺乏(FAH-/)导致的遗传性 I 型 酪氨酸血症小鼠,发现骨髓中只有HSC能在体内转 分化为肝细胞,并在受体内形成供体源造血和肝细 胞再生,受体小鼠血液生化指标明显改善。之后, 人们尝试应用粒细胞集落刺激因子(G-CSF)直接动 员骨髓释放干细胞来治疗肝病。动物部分肝切除后 应用G-CSF可以促进肝再生,其机制可能与血液中 CD133<sup>+</sup>或CD34<sup>+</sup>细胞动员有关,且体外培养G-CSF 动员后CD133<sup>+</sup>细胞可表达肝细胞特异性标志物<sup>[5]</sup>。 为进一步验证CD133<sup>+</sup>细胞对肝再生的促进作用, am Esch等[6]将分选出的自体骨髓CD133<sup>+</sup>细胞回输至 选择性门静脉栓塞的中央巨型肝脏肿瘤患者中,发 现CD133<sup>+</sup>细胞移植组非栓塞部分肝脏体积明显增 大,有利于患者接受进一步手术。

Sakaida等<sup>[7]</sup>应用四氯化碳(CCl<sub>4</sub>)建立的小鼠肝纤维化模型进行研究发现,异体骨髓细胞移植能减轻肝纤维化程度,提高小鼠生存率。大鼠未受损的肝血窦内皮细胞上有HSC表面标志(CD45<sup>†</sup>和CD33<sup>†</sup>)及内皮细胞的表面标志(CD31<sup>†</sup>),肝血窦内皮细胞受损后,回输骨髓来源的CD133<sup>†</sup>干细胞,在受体

肝脏中可发现供体来源的肝血窦内皮细胞,替代了原来的受损细胞<sup>[8]</sup>。

也有部分学者对HSC向肝细胞分化的潜能及治疗意义提出了质疑,他们应用动物的骨髓造血祖细胞在体外并不能诱导成功能性的肝细胞<sup>[9]</sup>,应用转基因的HSC研究发现,在体内只有极少部分HSC来源的肝细胞与受体的肝细胞形成融合细胞,几乎没有任何治疗意义<sup>[10]</sup>。

2.2 MSC治疗肝病 由于骨髓细胞成分复杂,何种细胞更适合用于肝细胞再生治疗尚不清楚,但越来越多的研究倾向于骨髓MSC。Sato等[11]比较了人骨髓中的MSC、CD34<sup>+</sup>、非MSC/CD34<sup>-</sup>等3种细胞直接注射到大鼠肝内后转分化成肝细胞的能力,结果骨髓MSC显示出最强的向肝细胞分化的潜能。

在适当条件尤其是肝脏损伤的情况下, 骨髓 MSC可以分化为肝细胞样细胞,这些分化细胞表现 出成熟肝细胞的形态和特征, 如表达肝细胞特异性 基因, 具有合成和分泌白蛋白、储存糖原、代谢尿 素及解毒功能等。Kuo等[12]应用人骨髓来源的MSC 进行移植治疗, 使小鼠暴发型肝衰竭得到有效恢 复,并促进了肝脏的再生。脂肪组织来源的MSC 与骨髓来源的MSC相似,在体外可诱导成肝细胞 样细胞,将这些肝细胞样细胞移植到CCl4肝损伤模 型的小鼠体内, 可整合入宿主的肝脏中, 使其肝脏 功能得到改善[13-14]。Van Poll等[15]应用D-半乳糖胺 建立小鼠急性肝损伤模型, 再输入MSC条件培养 基(MSC-CM),结果有效地减少了肝细胞坏死,促 进了肝细胞增殖, 并抑制了肝损伤标志物的释放。 Chamberlain等[16]将人MSC分别通过腹腔和肝内注射 至胎羊中,发现经肝内途径注射MSC后,人来源的 肝细胞广泛分布于肝实质中, 而通过腹腔途径注射 MSC后,人来源的肝细胞主要分布于门静脉周围。 用人羊水来源的MSC, 以及由MSC诱导分化的肝 祖细胞样细胞和肝细胞样细胞治疗CCl。损伤的急性 肝衰竭动物模型,结果羊水来源的MSC和肝祖细胞 样细胞取得了很好的治疗效果,且回输肝祖细胞样 细胞条件培养基也能取得较好效果[17]。人肝干细胞 及其条件培养基对暴发性肝衰竭也有很好的保护作 用,可通过分泌细胞因子抑制肝细胞凋亡,促进肝 细胞再生[18]。最近国内有学者研究发现,通过门静 脉输注人骨髓来源的MSC可提高急性肝衰竭猪的生 存率,促进肝细胞再生[19]。

但同时也有研究表明,MSC不能在体外诱导成成熟的肝细胞,但可在体内分化为肝细胞,且免疫原性很低,是MSC治疗肝病的良好细胞来源<sup>[20]</sup>。将人骨髓来源的MSC直接注入免疫抑制的肝脏损伤大鼠中,其分布只限于注射部位周围,虽然可以分化

成肝细胞,但分化效率很低,且供体的MSC并不与受体的肝细胞发生融合,MSC分化的肝细胞单纯来源于供体MSC<sup>[11]</sup>。Arikura等<sup>[21]</sup>将先天性白蛋白缺乏症大鼠行70%肝切除后植入正常大鼠骨髓MSC,4周后在受体肝脏中可检测到表达白蛋白mRNA的肝细胞。

- 2.3 胚胎干细胞(ES)治疗肝病 ES是指从囊胚期的内细胞团中分离出来的尚未分化的胚胎细胞,可分化形成各种类型的组织<sup>[22]</sup>。ES具有在体外无限增殖并保持分化成所有细胞类型的特性,理论上可以向3个胚层的任何类型细胞分化,其中也包括肝细胞<sup>[23-25]</sup>。已有研究表明ES对大鼠肝衰竭<sup>[26]</sup>和小鼠肝硬化<sup>[27]</sup>均有显著疗效。目前,有关ES研究的限制主要集中在伦理学、组织相容性以及移植后畸胎瘤发生等方面。
- 2.4 诱导多能干细胞(iPS)治疗肝病 已有研究显 示iPS细胞可以诱导分化为肝细胞<sup>[28]</sup>,也可将人原 代肝细胞重编程为肝细胞来源的iPS细胞,此iPS细 胞可定向诱导分化为内胚层细胞、肝祖细胞和成熟 的肝细胞<sup>[29]</sup>。有研究发现小鼠iPS细胞可以在四倍体 囊胚中发育成完整的胎肝,同时人iPS细胞可在体外 诱导分化为具备相关功能的肝细胞样细胞,这些细 胞可以在小鼠的肝脏内增殖并整合到肝实质中[30]。 最近发现应用三种转录因子(缺少转录因子c-Myc) 的iPS细胞及其诱导的具肝细胞表型和功能的肝细 胞样细胞,均能减轻CCI\_所致的急性肝损伤,提高 动物生存率[31]。有研究者建立了以代谢性疾病患者 真皮成纤维细胞制备的人iPS细胞系平台,此iPS细 胞系能诱导分化成具有肝细胞表型、基因型和功能 的肝细胞样细胞,显示出较强的治疗潜力[32]。利用 iPS细胞治疗肝病的具体疗效及其安全性尚需在动 物模型中进一步验证。

#### 3 临床研究

- 3.1 HSC治疗肝病的临床研究 2000年Alison等<sup>[33]</sup> 在移植了男性骨髓细胞的女性患者肝脏内发现了Y 染色体阳性的肝细胞,2002年Korbling等<sup>[34]</sup>在移植了男性外周血干细胞的女性患者肝脏内也发现了来自男性的肝细胞,证明骨髓和外周血干细胞在人体内可以分化为肝细胞。骨髓干细胞治疗可以通过 G-CSF动员或直接抽取骨髓的方式进行。
- 3.1.1 G-CSF动员的骨髓干细胞 Gaia等<sup>[35]</sup>应用 G-CSF动员骨髓干细胞治疗终末期肝病,患者有较好的耐受性,部分患者Child和终末期肝病模型 (MELD)评分改善,临床症状好转。Garg等<sup>[36]</sup>应用G-CSF动员骨髓CD34<sup>+</sup>干细胞治疗慢加急性肝衰竭,结果患者耐受性良好,Child、MELD和序贯器

官衰竭估计(SOFA)评分均显著改善,生存率显著提高。Levicar等<sup>[37]</sup>应用G-CSF动员骨髓,通过免疫磁珠分选外周血中的CD34<sup>+</sup>干细胞,并通过门静脉或肝动脉回输治疗慢性肝衰竭,结果未发现短期和长期不良反应,患者的肝脏功能有一定程度改善。Salama等<sup>[38]</sup>应用G-CSF动员骨髓,而后通过收集外周血中的骨髓HSC治疗终末期肝病,结果患者白蛋白升高,胆红素和丙氨酸转氨酶(ALT)下降,国际标准化比值(INR)恢复正常,所有患者均耐受。但也有报道通过门静脉回输自体骨髓CD133<sup>+</sup>细胞和单个核细胞治疗失代偿性肝硬化效果均不十分理想<sup>[39]</sup>。

- 3.1.2 未经动员的骨髓干细胞 Terai等[40]通过骨 髓穿刺获取骨髓干细胞后, 经外周静脉回输治疗肝 硬化(包括失代偿),在24周随访期内,患者血浆白 蛋白水平明显升高, Child评分明显改善, 肝脏活 检组织中甲胎蛋白(AFP)及增殖细胞核抗原(PCNA) 水平明显升高。Lyra等[41-42]分离终末期肝病患者骨 髓单个核细胞后经肝动脉回输,结果无明显不良 反应,患者Child、MELD评分明显改善,血清总 胆红素下降,白蛋白升高,INR明显下降,通过肝 动脉回输骨髓干细胞治疗终末期肝病安全可行。 Mohamadnejad等[43]采用抽取自体骨髓体外培养的 方法,将富含干细胞标记的细胞亚群回输至等待 肝移植的患者,4例患者在等待期内均存活,1年内 MELD评分有所改善。Kim等[44]应用自体骨髓干细 胞(包括造血干细胞和上皮细胞等单核细胞)治疗慢 性乙肝相关的肝硬化患者,结果患者生活质量明显 改善、80%患者肝脏体积增大,腹水减少,Child评 分改善, 肝脏祖细胞(HPC)活性增强, 可在6个月 继续分化成肝细胞,但肝硬化的临床分级无明显改 善(表1)。
- 3.2 MSC治疗肝病的临床研究 Mohamadnejad等[43] 最早报道应用自体骨髓MSC治疗终末期肝病患者, 经穿刺获取骨髓后通过密度梯度离心获得骨髓单个 核细胞, 在体外诱导成MSC后, 经外周静脉回输至 患者,经过12个月的随访,该方法安全无副作用, 患者症状明显改善,血浆白蛋白升高,腹水减少, 肝体积增大, MELD评分改善。Khayaziha等[45]报道 应用自体骨髓MSC经外周或门静脉回输治疗由乙 肝、丙肝、酒精性肝硬化及不明原因导致的终末期 肝病患者,结果表明该方法安全无副作用,能提升 血浆白蛋白,降低胆红素水平,促进凝血酶原时间 (PT)复常,改善MELD评分。近两年Amer等[46]用骨 髓MSC体外诱导成肝细胞样细胞后进行自体回输 治疗丙肝相关的终末期肝病,结果患者症状明显改 善,腹水减少,白蛋白显著增加,Child和MELD评 分改善。

#### 表1 干细胞治疗终末期肝病的临床应用情况

**Tab. 1** Clinical application of stem cells on treatment of end-stage liver disease

Enrolled patients	Source and type of stem cells	Number of infused cells	Infused approach	Efficacy
Child score ≥9, MELD score >10 <sup>[35]</sup>	CD34 <sup>+</sup> cells from G-CSF mobilized peripheral blood	_	_	Improved Child score, MELD score, and clinical symptoms
Acute-on-chronic liver failure [36]	CD34 <sup>+</sup> cells from G-CSF mobilized peripheral blood	_	_	Improved Child score, MELD score, SOFA score, and survival rate
Chronic liver failure <sup>[37]</sup>	CD34 <sup>+</sup> cells from G-CSF mobilized peripheral blood	$1\times10^6-2\times10^8$	Portal vein or hepatic artery	Improved serum bilirubin
End-stage liver disease associated with hepatitis $C$ and autoimmune liver disease <sup>[38]</sup>	Autologous bone marrow $\mathrm{CD34}^{^{+}}$ cells	-	Hepatic artery or portal vein	Improved serum albumin, serum bilirubin ALT, and INR $$
Decompensated liver cirrhosis [39]	Autologous bone marrow-derived $\mathrm{CD}133^+$ and mononuclear cells	_	Portal vein	Changes in clinical parameters is not obvious
Liver cirrhosis (including decompensated) $^{[40]}$	Autologous bone marrow-derived mononuclear cells	$(5.20 \pm 0.63) \times 10^9$	Peripheral vein	Improved serum albumin, and Child score
End-sage liver disease cause by various pathogenesis [41]	Autologous bone marrow-derived CD34 <sup>+</sup> and CD45 <sup>+</sup> cells	$(3.78 \pm 2.69) \times 10^8$	Hepatic artery	Imrpoved Child score, MELD score, serum albumin, and serum bilirubin
Decompensated liver cirrhosis <sup>[43]</sup>	Autologous bone marrow-derived MSCs	$(1.02 - 6.00) \times 10^7$	Peripheral vein	Improved MELD score, liver function, serum albumin; decreased ascites; increased liver volume
Liver cirrhosis associated with chronic hepatitis $B^{[44]}$	Autologous bone marrow-derived stem cells (mononuclear cells)	$0.995\times10^8/kg$	Peripheral vein	Liver volume increased in 80% of the patients; decreased ascites; improved Child score; increased activation of the hepatic progenitor cell compartment
End-stage liver disease caused by hepatitis B, hepatitis C, alcoholic, and cryptogenic [45]	Autologous bone marrow-derived MSCs	$(3-5)\times10^7$	Peripheral vein or portal vein	Improved MELD score, serum albumin, serum bilirubin, serum creatinine, and PT
End-stage liver disease associated with hepatitis $C^{[46]}$	Hepatocyte-like cells induced by autologous bone marrow-derived MSCs	$2 \times 10^7$	Intrasplenic or intrahepatic	Improved serum albumin, Child score, MELD score, fatigue scale and performance status; decreased ascites
Liver failure associated with hepatitis $\boldsymbol{B}^{[47]}$	autologous bone marrow-derived MSCs	$(3.4 \pm 3.8) \times 10^8$	Hepatic artery	Improved serum albumin, serum bilirubin, MELD score and PT $$
Decompensated liver cirrhosis associated with hepatitis $B^{[48]}$	Umbilical cord- derived MSCs	$0.5\times10^6/kg$	Peripheral vein	Improved liver function, serum albumin, serum bilirubin, sodium MELD score; decreased ascites
Acute-on-chronic liver failure associated with hepatitis $B^{[49]}$	Umbilical cord- derived MSCs	$0.5 \times 10^6/\text{kg}$	Peripheral vein	Improved survival rate, MELD score, serum albumin, serum bilirubin and PTA
Primary biliary cirrhosis <sup>[50]</sup>	Umbilical cord- derived MSCs	$0.5 \times 10^6/\text{kg}$	Peripheral vein	Decreased serum alkaline phosphatase and g-glutamyltransferase

国内学者最近应用MSC治疗终末期肝病取得了较好进展。Peng等<sup>[47]</sup>应用自体骨髓MSC治疗乙肝相关的肝衰竭患者,结果短期(4周)内患者白蛋白、PT水平明显增加,而总胆红素水平和MELD评分明显下降,无明显不良反应,但长期效果不明显。Zhang等<sup>[48]</sup>应用脐带MSC移植治疗慢性乙肝相关失代偿性肝硬化,结果患者腹水明显减少,肝脏功能明显改善,白蛋白增加,血清胆红素水平和MELD评分明显下降。Shi等<sup>[49]</sup>应用脐带MSC移植治疗慢性乙肝相关的慢加急性肝衰竭患者,结果发现可显著提高患者生存率,改善MELD评分,增加白蛋白、胆碱酯酶、凝血酶原活动度水平,而总胆红素和转氨酶水平明显下降。Wang等<sup>[50]</sup>采用脐带MSC治疗原发性胆汁性肝硬化患者,结果血清胆碱磷酸和 γ-谷氨酰转移酶水平明显下降(表1)。

3.3 干细胞回输与移植途径 目前,干细胞治疗 肝病的主要移植途径有外周循环移植、门静脉移 植、肝动脉移植等,现有的结果显示安全性好,患者可耐受,无明显不良反应。但也有报道经穿刺获得骨髓后应用免疫磁珠分选出CD34<sup>+</sup>干细胞,经过肝动脉回输,结果发生显影剂性肾病,最后发展成1型肝肾综合征<sup>[51]</sup>。因此,通过肝动脉回输干细胞的安全性尚需进一步观察。

3.4 疗效评价及影响疗效的因素 移植前的肝功能状况直接影响干细胞移植的结局。Barba等<sup>[52]</sup>报道HSC移植前高胆红素和高谷酰转肽酶(GGT)水平可影响移植后病死率和生存率。现有肝功能改善的评价方法主要包括Child和MELD评分、血液生化、临床表现、生活质量、生存时间(率),以及应用甲胎蛋白和PCNA评价肝细胞再生,利用TNF、IL-6和IL-10等评价肝脏炎症环境的改善等<sup>[53]</sup>。利用磁性标记的大鼠MSC进行肝脏移植,再行磁共振成像活体示踪,可在肝实质中发现标记的移植细胞<sup>[54]</sup>。肝组织活检对评价肝再生及移植细胞状态有重要意

义,但有创性使其对终末期肝病患者的应用受到限制。一些无创或微创的活体内移植细胞示踪方法如Y染色体探查等,可应用于不同性别间的细胞移植,放射性同位素标记移植细胞后应用核素显像或PET-CT等也已有临床报道<sup>[55]</sup>。这些方法均可在不同程度上说明移植细胞的存活和增殖情况,为评价于细胞移植的作用提供依据。

## 4 干细胞治疗肝病的机制

- 4.1 干细胞归巢功能 肝脏微环境改变是MSC归巢的始动因素,肝组织损伤时存在炎症反应,局部可表达及分泌多种趋化因子、黏附因子、生长因子、基质金属蛋白酶9(MMP-9)等,这些因子与其受体的相互作用可引导干细胞特异性迁移至病损部位<sup>[56-58]</sup>。基质细胞衍生因子SDF-1及其受体CXCR4构成的SDF-1/CXCR4轴是引导MSC向损伤组织迁移的重要生物轴<sup>[59]</sup>。此外,肝细胞生长因子(HGF)及其受体c-met构成的HGF/c-met轴、G-CSF、血管内皮生长因子(VEGF)、MMP等均与MSC的归巢有关<sup>[58,60-61]</sup>。还有研究认为,肝脏受损后肝脏环境中的鞘脂代谢物——鞘氨醇1-磷酸盐(S1P)水平增高,且其与骨髓之间的浓度梯度差通过S1P3受体介导是骨髓MSC向肝脏归巢的重要因素<sup>[62]</sup>。
- 4.2 干细胞分化功能 Petersen等[1]于1999年首先报道,在性别交叉骨髓细胞移植或全肝移植受体肝脏中发现来源于供体骨髓的肝细胞,随后Alison等[33]和Theise等[3]相继发现骨髓移植和肝移植患者的肝脏中也有来源于供体骨髓的肝细胞。Lagasse等[4]在延胡索酰乙酰乙酸水解酶(FAH)缺陷大鼠模型中发现骨髓HSC可在肝脏内分化为具有功能的肝细胞,改善FAH缺陷大鼠的症状。Schwartz等[63]也报道,从大鼠、小鼠和人的骨髓中分离得到多能成体祖细胞,体外经成纤维细胞生长因子(FGF)和HGF诱导可分化为功能肝细胞。骨髓中存在可分化为肝细胞的干细胞,直接将其移植到肝脏,在肝脏微环境下可分化为肝细胞。因此骨髓干细胞移植为多种严重肝病的治疗提供了新的策略(图1)。

MSC具有跨胚层多向分化潜能,在合适的条件下,如HGF、FGF、表皮生长因子(EGF)或制瘤素M(OSM)等诱导下,通过特定的细胞信号传导途径,可以跨胚层向内胚层的肝细胞样细胞、胆管细胞和血管内皮样细胞分化<sup>[64-66]</sup>,而Notch/Jagged信号通路在此过程中可能起重要作用<sup>[67-68]</sup>。在此分化系统中,肝脏局部微环境,如细胞因子、细胞外基质(ECM)、激素、基质细胞等,是诱导MSC定向分化的决定因素,其中细胞因子的类型、浓度和添加次序是影响MSC分化的主要因素<sup>[69]</sup>。

4.3 干细胞旁分泌功能 以往认为,干细胞移植可 提供大量肝细胞样细胞替代受损的肝细胞功能,然 而急性肝损伤所造成的疾病进展快, 肝功能恢复不 能依赖于干细胞分化为肝细胞过程, 而更大程度上 依赖其他机制,目前认为干细胞旁分泌机制改变组 织微环境比向肝细胞的转分化更重要[70]。如MSC可 以分泌多种细胞因子、生长因子,发挥局部效应, 促进受损肝脏增生及肝脏血管再生,抑制免疫细胞 增殖及向肝脏迁移,调节肝脏及全身免疫炎症反 应,从而减轻肝脏的急性损伤,提高生存率[12,71-72]。 MSC的条件培养基能抑制肝细胞的抗凋亡,刺激肝 脏再生[15], 机制可能与MSC的旁分泌功能一致, 即为损伤的肝脏提供营养和有利的生存环境, 因为 蛋白质组分析显示条件性培养基中含有许多抗炎症 因子,如IL-10、IL-1ra、IL-13和IL-27等[17],以及 促进肝细胞再生、抑制肝细胞凋亡的细胞因子,如 HGF、IL-10、VEGF、IL-6和IL-8等[18]。此外,研 究还发现移植的骨髓MSC可通过释放促细胞增生因 子和MMP-9刺激内源性肝细胞再生[73-74]。

#### 5 存在的问题与展望

基于干细胞具有自我更新、无限增殖的能力以及多向分化潜能的特性,将其应用于终末期肝病的临床研究取得了令人鼓舞的进展,但干细胞治疗终末期肝病的长期安全性(特别是致畸或癌变的风险)和远期疗效仍有待观察,在疗效评价方面如何检测受体肝脏内移植细胞的状态和功能,如何减少和避免排斥反应及其他不良反应,特别是MSC具有形成肝细胞或促进肝纤维化的双刃剑作用<sup>[75]</sup>,仍需进一步探讨。此外,干细胞的来源、诱导培养条件、细胞质量控制(包括表型、功能、微生物安全)、使用时机、途径和剂量、临床适应证等都可能对治疗的后果产生影响,这一系列与安全和疗效密切相关的问题还须进一步深入研究。因此,要在阐明其机制的基础上,不断积累各方面的资料,明确其临床应用的安全性和有效性,保证患者利益。

总之,干细胞强大的治疗潜力有可能成为终末 期肝病患者的有效治疗手段,从而给患者带来新的 希望。

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