

SIGNIFICANCE

RNA lies at the center of cellular function. Its most appreciated function is to carry protein blueprints to the ribosome for manufacture. Only recently have researchers begun to appreciate its myriad of other functions, many of which also lie at the center of important cellular processes and are implicated in a variety of disease states[1–3]. Long noncoding RNA (lncRNA) comprise one such important class of RNA that does not participate in the central dogma[4]. Alarming, the human genome encodes for as many lncRNA as proteins[5]. These transcripts are typically greater than 200 bases, and participate in binding both proteins and nucleic acids, often both at the same time [cite]. In a striking example, *SNHG5* is a long noncoding RNA (lncRNA) of largely unknown function, that has been linked to cancer cell survival and metastasis in a diversity of different cancers from leukemias to melanoma[6–13]. This 2 kilobase cytoplasmic transcript is upregulated in these disease states, and has been implicated to interact with a variety of mRNAs in the cytoplasm to control expression. Despite such burning questions in the field of lncRNA, a tool to directly image these species and their interacting partners in living cells has yet to be developed.

BACKGROUND

Protein studies benefit from a large imaging toolkit that has been developed over the last 25 years. Genetically encodable fluorescent proteins are now ubiquitous for the study of the localization of any translated target in the cell. While fluorescent proteins are a mature, well-understood technology, tools for imaging the localization of individual RNA transcripts in living cells remain limited. The most popular systems to date include dye-binding aptamers (Spinach[14], Broccoli[15], and Mango[16, 17]), and RNA-binding protein fusions (MS2-FPs)[18].

The most analogous to fluorescent proteins, dye-binding aptamers utilize exogenously administered dyes that give fluorescence induction upon binding their RNA partner[14–17]. This sequence is encoded downstream of an RNA of interest to track its location in the cell. Though excellent binders for their dyes, the aptamers utilized by this technology are unstable in mammalian cells due to their nonnative structure[19]. Additionally, though fluorescence turn-on is excellent *in vitro*, often reaching 1,000 fold, signal induction in cells is typically two to four fold, most likely due to nonspecific binding of the dye.

RNA-binding protein fusions are a much more robust technique for imaging RNA transcripts[18]. This method often utilizes the MS2 bacteriophage coat protein that binds an encoded stem loop of RNA. When MS2 is fused to a fluorescent protein, transcripts can be visualized in the cell. In order to concentrate the fluorescence signal above background, multiple stem loops are placed in series (up to 24 in a row). Though this technique has enabled imaging of single transcripts in the cell[18, 20], the resulting protein-RNA complex is prohibitively large for many studies.

Riboglow is a recently developed platform in the Palmer lab that solves many of the drawbacks of current RNA imaging techniques (Fig. 1)[21]. The two-component system utilizes a synthetic fluorophore-quencher pair, and a genetically-encoded riboswitch. A transcript of interest is first tagged within the cell, and the probe construct is administered via bead loading[20, 22, 23]. When the construct binds the transcribed riboswitch, the fluorophore is dequenched, giving fluorescent signal. Like the dye-binding aptamers, Riboglow utilizes a riboswitch receptor domain that natively binds vitamin B₁₂ (cobalamin, Cbl, Fig. 1B)[24]. Conveniently, cobalamin is known to act as a fluorescence quencher for a variety of fluorophores[25–27]. The cobalamin center is conjugated to the fluorophore through a flexible linker that promotes quenching in the unbound state, but enables the fluorophore to reside at a distance in the bound state (Fig. 1B).

The Palmer lab found that this initial generation of Riboglow has the potential to outperform the existing tools in the field. Its use of a naturally occurring riboswitch imparts stability to the construct that is not present in

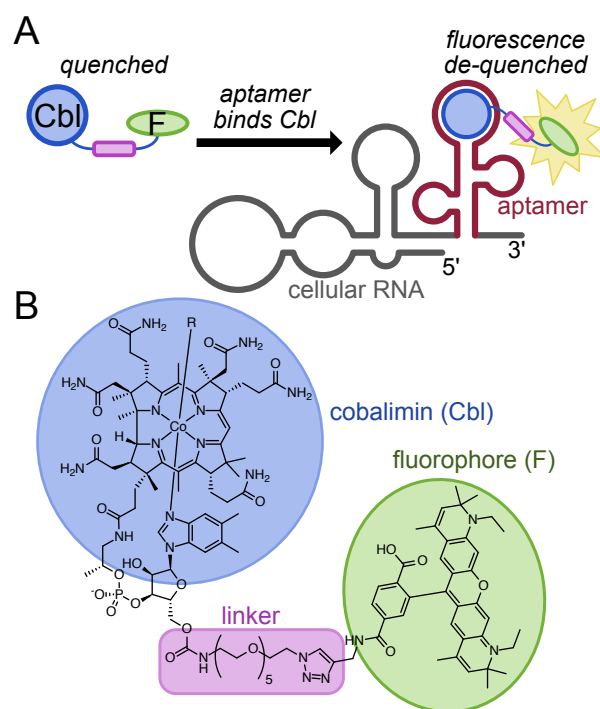


Figure 1. A) Cobalamin acts as a quenching and localization moiety to guide a fluorescent probe to an RNA transcript of interest. When unbound, fluorescence is quenched. In the presence of RNA tagged with the cobalamin riboswitch, fluorescence is restored. B) Structure of the generation 1 Riboglow probe. A polyethylene glycol linker of five units (5xPEG) connected to the 5' hydroxyl of the cobalamin ribose was used to tether an ATTO 590 fluorophore to the construct.

aptamer-based techniques. Additionally, the use of a donor-quencher pair reduces nonspecific fluorescence in the unbound state. Such signal induction is low in the aptamer-binding dyes, and cannot be obtained at all with the MS2-fluorescent protein fusions. Due to these advantages, Riboglow outperformed Broccoli and the MS2 system in initial studies of stress granule (SG) detection in mammalian cells.

Though promising, the initial iteration of Riboglow has significant room for improvement. With initial constructs, signal induction upon riboswitch binding never exceeded seven fold *in vitro*, and the pendant dye was never fully quenched (with 10-20% residual fluorescence). Additionally, to effectively detect stress granules, four riboswitches had to be placed in series to concentrate fluorescence signal. This high background precluded single-molecule imaging. Finally, the strategy cannot currently be used to monitor the location of multiple transcripts in tandem. Herein, I will propose strategies to overcome these limitations through simultaneous modulation of the small molecule construct and riboswitch sequence. This is an undertaking for which I am uniquely suited, due to my experience of tandem modification of small molecule-protein interactions during my graduate studies. First, I will synthesize a variety of new Riboglow probes to optimize fluorescence quenching (Aim 1). Next, I will turn to the sequence of the riboswitch itself. I'll design a screen for brightness that will select the best riboswitch sequence to match the optimized probe structure (Aim 2). This new, brighter pair will be tested for single-molecule detection. Finally, I will use SELEX to identify mutually orthogonal probe-riboswitch pairs for multicomponent RNA imaging (Aim 3).

APPROACH Aim 1. Synthesize improved Riboglow probes.

The main drawback of Riboglow is the poor turn-on that is observed upon probe binding. In this aim, I intend to leverage my background in synthetic chemistry to produce a panel of diverse probe structures that improve fluorescence quenching (and thus signal induction). In previous studies in collaboration with Professor Dorota Gryko (see Gryko letter of support) a small number of linkers and fluorophores were evaluated for quenching and fluorescence turn-on. Linker length and fluorophore wavelength were varied to gauge the quenching ability of cobalamin. Remarkably, some degree of quenching occurred in all of the constructs synthesized, regardless of the spectral overlap of the fluorophore and the cobalamin. However, most retained 10-20% residual fluorescence, which increased background signal significantly. *To optimize probe function, I will vary linker composition, attachment point, and pendant fluorophore.*

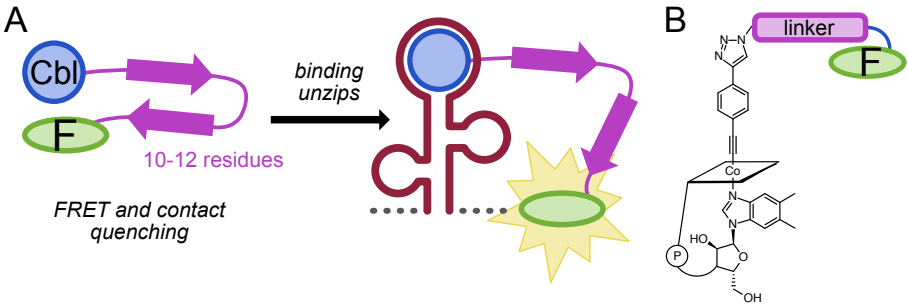


Figure 2. A) Tryptophan zipper peptides will be used to maximize FRET and contact quenching in the unbound state. B) Conjugation via the axial site of the cobalt metal center will provide additional probe structural diversity.

Sequence	Tm (°C)	% folded (25 °C)
KKWTW-NPATGK-WTWQE	85	>96
KKYTW-NPATGK-WTVQE	66	92
GEWTY-NPATGK-FTVTE	47	74
GGGGG-GGGGGG-GGGGG	N/A	N/A

Table 1. Tryptophan zippers of varying stability will be tested to maximize fluorescence quenching. Values were obtained from Ref. [28].

motifs are as small as twelve amino acids and many are stable to denaturation up to 85 °C (Table 1). In the unbound state, such a linker would hold the quencher and fluorophore in close proximity (due to the short distance between the N and C termini of the peptide, Fig. 2A). When the cobalamin is bound by the riboswitch, steric occlusion would force the β -hairpin turn to unfold to place the fluorophore-quencher pair at a larger distance. A wide range of tryptophan zippers have been developed with a variety of structures and stabilities, enabling easy synthesis of a range of different linkers via solid phase peptide synthesis[28, 31–33]. Cobalamin and a

Ideally, in the unbound state, the cobalamin and fluorophore would be closely associated to maximize FRET and contact quenching[25, 29, 30]. In the RNA-bound state, the molecules would reside at their maximal distance to promote fluorescence[26]. To strike this balance, I will use a synthetic β -hairpin as the linker between cobalamin and the fluorophore (Fig. 2A, Table 1). Such a linker would hold the molecules close in solution, but would be linearized upon binding to the riboswitch. A number of such β -hairpin have been developed, and are referred to as tryptophan zippers (trzip)[31]. These

fluorophore will then be coupled to each terminus of the linker via additional amide couplings[34]. A starting set of linkers are listed in Table 1. The polyglycine linker will be used as a negative control for a linker with little secondary structure, but identical length. If the fold proves to be too stable (dequenching is not observed), the amino acids of the peptide linker will be varied to tune the melting temperature. Or, if adequate quenching is not observed, the relative positions of the N and C termini will be adjusted based upon structural data available on these β -hairpin, available in the Protein Data Bank (PDB). Each new construct will be tested for their ability to quench fluorescence in vitro by comparing the fluorescence of the construct with that of the free fluorophore in solution. Fluorescence signal in the presence of the Riboglow riboswitch will also be measured to calculate fluorescence turn-on.

Another underexplored variable of the original Riboglow probes is the linker attachment point. Though the 5' hydroxyl of the cobalamin ribose is the most accessible nucleophile on the structure, there exist several other possible sites of conjugation. Perhaps the second most common site is the axial ligand of the cobalt metal itself. Though many studies have taken advantage of the labile nature of certain alkyl modifications at this position[29], others have found alkynyl modifications to be stable to air and light[35, 36]. Following this precedent, I will synthesize a cobalamin with an alkyne handle attached to the cobalt metal center (Fig. 2B). Such a molecule has already been synthesized in the lab of Dorota Gryko (see Gryko letter of support), and shown to be amenable to functionalization via dipolar azide-alkyne cycloaddition (AAC)[37]. The alkyne handle will be readily conjugated to a variety of linkers with terminal azides[38, 39].

These constructs will also be evaluated for degree of fluorescence quenching, and brightness in the presence and absence of the cobalamin riboswitch. It is possible that incorporation of a ligand at this site on the metal center will abolish binding to the native riboswitch due to its positioning in the binding pocket[24]. If this is the case, these probes could serve as excellent starting points for directed evolution of orthogonal aptamer-probe pairs (see Aim 3). Additionally, perturbations to the native binding mode of cobalamin could reduce off-target association with native B₁₂ machinery[40], further reducing fluorescence background.

The final variable in the molecular structure of the Riboglow probe is the fluorophore itself. The ideal probe would minimize cellular autofluorescence through the use of a red fluorophore with a high extinction coefficient. The Janelia Fluor series of dyes is well suited for our application (see Lavis support letter)[41, 42]. Through minor modifications to the rhodamine scaffold, the Lavis lab has developed a range of fluorophores with varying excitation and emission spectra, and excellent photophysical properties. Table 2 includes a subset of the probes that will be explored. Emphasis will be placed on fluorophores with high extinction coefficients and quantum yields, with the intent for these probes to be used in single-molecule imaging (Aim 2). Additionally, JF₅₈₅ and JF₆₃₅ were shown to be highly fluorogenic upon conjugation to a protein tag, with absorbance increases of 80 and 113-fold respectively[41]. The Lavis lab makes these probes freely available, and has offered to provide us with alkynylated versions for easy conjugation via CuAAC (see support letter). Probes containing these new fluorophores will be evaluated as previously described, with the addition of a test to fluorophore photostability. Stability under extended illumination will be important for single-molecule experiments (Aim 2), thus these experiments will give an additional point of comparison for probe quality.

Dye	λ_{ex} (nm)	λ_{em} (nm)	ϵ (M ⁻¹ cm ⁻¹)	ϕ
Cy5	646	662	271,000	0.2
ATTO 488	501	523	90,000	0.8
ATTO 590	594	624	120,000	0.8
ATTO 633	629	657	130,000	0.6
JF ₅₄₉	549	571	101,000	0.88
JF ₅₈₅	585	609	156,000	0.78
JF ₆₃₅	635	652	167,000	0.56

Table 2. A range of fluorophores will be explored for optimal quenching and signal induction. Cy5 and the ATTO series were utilized in the first generation Riboglow system, and will also be used here with new linkers and Cbl attachment points. Photophysical values were obtained from Atto tec and refs. [21] and [41]

Summary and alternate approaches:

To identify probes with improved quenching and fluorescence turn-on, linker, attachment point, and fluorophore will be varied. With four linkers, two attachment points, and seven fluorophores, 56 constructs are possible. Though not all of these will be evaluated, simple chemistries involving amide bond couplings and click chemistry will enable us to sample a large amount of this structure space in a rapid fashion. If β -hairpin zippers do not yield improvements in quenching, other linkers with varying rigidity and lengths will be explored[26], specifically those that promote direct contact between the fluorophore and the cobalamin. Also, it should be noted that new cobalamin constructs are not necessary for the success of aims 2 or 3. It is likely that all three aims will be pursued in parallel, with new discoveries informing the direction of each.

Aim 2. Adapt Riboglow for single-molecule imaging.

Visualization of the lifecycle of single RNA transcripts as they move throughout the cell remains a holy grail of RNA imaging.[43] Though several recent studies have demonstrated visualization of individual mRNA transcripts, all have required the use of at least 24 repeats of the MS2-binding stem loop[18, 44–46]. When these loops (which each bind two MS2 proteins) are fully populated, a complex of more than 2,000 kilodaltons is produced. Such a structure is several times larger than most mRNA, giving it the potential to be highly perturbing to normal mRNA behavior. Riboglow has the potential to reduce the size of the fluorescent probe by 10–100-fold (relative to the MS2 system), however, high fluorescence background and poor signal induction precluded these studies[21].

With the wide variety of new probe constructs developed to optimize quenching (Aim 1), changes will be made to the sequence of the cobalamin riboswitch with the goal of increasing fluorescence turn-on. *A directed evolution screen for fluorescence brightness will identify optimal riboswitch sequences that promote fluorescence signal induction.*

First, a library will be designed that varies the environment surrounding the binding pocket of the cobalamin riboswitch (Fig. 3A). Through the guidance of Robert Batey (see Batey support letter), sites will be chosen to retain binding of the cobalamin, yet maximize the distance between the quencher and the fluorophore upon binding[24]. The Palmer lab is a leader in technologies for tool development in mammalian cells[47, 48]. This expertise will be leveraged for screening libraries of cobalamin riboswitches (Fig. 3B). Libraries of transcripts will be transduced into mammalian cells, the probe of interest will be administered, and cells will be sorted via flow cytometry. Cells that show elevated brightness relative to a fluorescent protein expression control will be sorted. In this way, libraries of up to one million members will be screened. Bright variants will be collected, cultured, and resubjected to sorting until only bright variants remain. Sequences will be evaluated through deep sequencing.

New sequences identified through this screen will undergo rigorous characterization of their biophysical and photophysical properties *in vitro*. It will be important to characterize the extinction coefficient of each complex upon binding the RNA tag, as well as their quantum yields and photostability. The binding affinity for each probe-RNA combination will also be measured via isothermal titration calorimetry. Such a value is important because it informs the probe dosage that will be necessary for imaging (and lower probe dosages will contribute less fluorescence background).

Candidate probes that exhibit low background fluorescence, high signal induction, and high RNA affinity will be tested to determine the absolute level of turn-on fluorescence, and the minimum number of fluorophores that can be localized. To do this, beta-actin mRNA will be tagged with one, two, or four repeats of the mutant riboswitch of interest in U2-OS cells through transient transfection. The cells will be imaged under conditions analogous to previous studies[44] using a TIRF microscope (see Equipment document). If puncta that appear to be single molecules are observed under these conditions, probe localization will be verified via fluorescence in situ hybridization (FISH) following cell fixation. If puncta are not visible in the initial TIRF images, cells will be treated with arsenite to induce the formation of stress granules (SGs) that also contain the protein G3BP1[49–51]. G3BP1 can be tagged with the Halo-tag, and subsequently labeled with an orthogonal fluorophore to evaluate co-localization. A check of direct mRNA location can then be carried out with FISH. This is a technique that the Palmer lab has previously used successfully to test Riboglow probes[21].

To quantify our ability to resolve single mRNA molecules, we will utilize single-molecule FISH (smFISH) imaging to correlate RNA count with Riboglow signal[52]. This same method was recently used to quantify the sensitivity of an improved MS2 system[53].

If the new Riboglow probes demonstrate single-molecule labeling in the β -actin model system, we will attempt to image migration of transcripts of long noncoding RNA (lncRNA) from the nucleus to the cytoplasm. Recently, a surprisingly large number of lncRNA were identified in the cytoplasmic compartment via APEX-RIP[54]. Most known functions of lncRNA involve networking of chromatin... One, a transcript called SNHG5 has been impli-

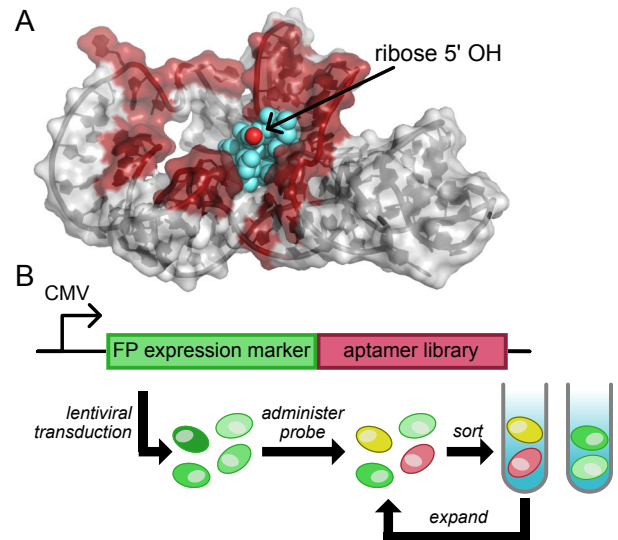


Figure 3. A) Riboswitch sites for mutation will be targeted based on potential contact with the linker and fluorophore. PDB: 4FRN[24] B) RNA sequences will be screened for brightness relative to a fluorescent protein control.

cated in colorectal cancer cell survival[7]. Little is known about cytosolic lncRNAs with respect to function, and less is known about localization.

Summary and alternate approaches:

To attain single-molecule resolution with Riboglow, the probes synthesized in Aim 1 will be screened with libraries of riboswitch mutants *in cellulo*. Cells containing bright mutants will be sorted via FACS and analyzed. Library hits will be characterized *in vitro*, and tested via established assays for single-molecule localization *in cellulo*. Probes that are successful in our model systems will be used to image long noncoding RNA under the supervision of Professor John Rinn. If single-molecule resolution is not observed in the library hits, additional riboswitches (over the four repeats already planned) will be added to concentrate fluorescent signal. Even if 24 repeats are required, the construct will still be less than half the size of the MS2 system.

Aim 3. Develop mutually orthogonal Riboglow probes for multicomponent imaging.

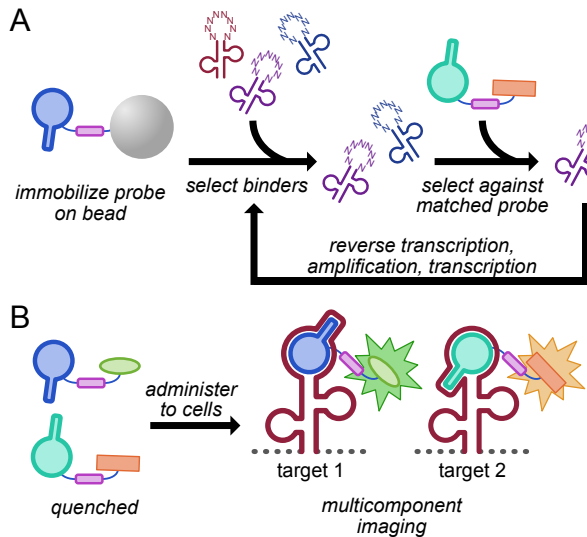


Figure 4. A) SELEX will be used to screen for riboswitches that bind each cobalamin in a mutually-exclusive manner. B) Mutually orthogonal cobalamin analogs will enable multicomponent RNA imaging. Signal turn-on will only be observed in the presence of the matched pair.

The ability to track multiple RNA transcripts simultaneously would enable study of the interactions of RNA as they move through the cell. Upregulation of *SNHG5* in colorectal cancer has been implicated in stabilization of a variety of mRNA in the cytoplasm[7]. Direct hybridization of the lncRNA with mRNA protects transcripts from degradation, and enables translation, promoting disease states. Despite these implications, these interactions have never been visualized in living cells, and thus the functions of *SNHG5* and other lncRNA remain unknown. Localization of these lncRNA and their mRNA targets in living cells would shed light on these mysterious processes.

The modularity of the Riboglow platform is well suited to enable such an experiment. Probes will be distinguished through the use of different fluorophores, and different riboswitch sequences will be used to bind each structurally different probe in a mutually exclusive manner (Fig. 4B).

Fortunately, screening for substrate selectivity is well-precedented in RNA engineering through systematic evolution of ligands by exponential enrichment (SELEX)[55, 56]. In fact, *de novo* riboswitches have already been developed for cobalamin and a few of its analogs via SELEX[57]. This important precedent shows that even small changes to the structure of the cobalamin can be distinguished by engineered RNA, and implies that no additional changes will need to be made to the cobalamin structure apart from those constructs already pro-

posed above. However, to improve our chances at identifying mutually-orthogonal probes, we will begin by screening probes that contain each of the two linker attachment points (Aim 1) to maximize structural differences at the cobalamin center.

Importantly, for our selection experiments, we will not be starting from a completely randomized sequence of nucleic acids. It will be crucial to retain the original fold (and thus *in cellulo* stability) of the native riboswitch. RNA biosensors developed in this way are known to have increased stability relative to aptamers selected *de novo*[58]. The design of the starting RNA libraries, and subsequent selection experiments will be carried out under the supervision of Professor Robert Batey (see Batey letter of support), an expert in SELEX and RNA engineering[58, 59]. Libraries will target bases known to participate in contacts with the cobalamin small molecule[24, 58]. The selection will be carried out as shown in Figure 4A. Briefly, the cobalamin probe of interest will be immobilized on a bead via the same linker that will be used in the final construct. The synthetic library of riboswitches will be incubated with the beads, and unbound sequences will be rinsed away. Riboswitches that bind selectively will be collected off the beads and amplified (via reverse transcription, PCR amplification, and transcription) to undergo another round of selection (for a total of 7-10 rounds). In subsequent rounds, the stringency of the screen will be increased via additional washes and reduction of RNA concentration. In the final 2-3 rounds of selection, a counterselection will be conducted to ensure orthogonality with other synthetic probes (Fig. 4A). In this way, the probe (or probes) intended to be orthogonal to the construct under selection will wash away any riboswitches that do not bind selectively. Riboswitches will be deep sequenced to identify library convergence and optimal clones[58]. This process will be repeated for each probe of interest, with negative screens against each other

probe that is intended for multiplexed use.

Mutually-orthogonal probes identified through the *in vitro* selection will be verified in a similar fashion as described in Aim 2. Probe extinction coefficient, quantum yield, photostability, and binding affinity will be characterized *in vitro*. Probes will also be independently verified *in cellulo* via the β -actin fusion assay described in Aim 2.

Once mutually-selective Riboglow probes are identified, they will be tested in mammalian cells to probe the localization

Maybe look at two differently localized RNAs that both migrate to SGs upon arsenite addition?

Summary and alternate approaches:

Bibliography and References Cited

- [1] Cech, T. R. and Steitz, J. A.: The Noncoding RNA Revolution—Trashing Old Rules to Forge New Ones. Cell. 157(1): 77–94, March 2014.
- [2] Della Ragione, F., Gagliardi, M., D'Esposito, M., and Matarazzo, M. R.: Non-coding RNAs in chromatin disease involving neurological defects. Front. Cell. Neurosci. 8, 2014.
- [3] Anastasiadou, E., Jacob, L. S., and Slack, F. J.: Non-coding RNA networks in cancer. Nature Reviews Cancer. 18(1): 5–18, January 2018.
- [4] Rinn, J. L. and Chang, H. Y.: Genome Regulation by Long Noncoding RNAs. Annual Review of Biochemistry. 81(1): 145–166, 2012.
- [5] Rinn, J. L., Euskirchen, G., Bertone, P., Martone, R., Luscombe, N. M., Hartman, S., Harrison, P. M., Nelson, F. K., Miller, P., Gerstein, M., Weissman, S., and Snyder, M.: The transcriptional activity of human Chromosome 22. Genes Dev. 17(4): 529–540, February 2003.
- [6] Tanaka Ritsuko, , Satoh Hitoshi, , Moriyama Masatsugu, , Satoh Kasumi, , Morishita Yasuyuki, , Yoshida Syouko, , Watanabe Toshiki, , Nakamura Yoshikazu, , and Mori Shigeo, : Intronic U50 small-nucleolar-RNA (snoRNA) host gene of no protein-coding potential is mapped at the chromosome breakpoint t(3;6)(q27;q15) of human B-cell lymphoma. Genes to Cells. 5(4): 277–287, December 2001.
- [7] Damas, N. D., Marcatti, M., Côme, C., Christensen, L. L., Nielsen, M. M., Baumgartner, R., Gylling, H. M., Maglieri, G., Rundsten, C. F., Seemann, S. E., Rapin, N., Thézenas, S., Vang, S., Ørntoft, T., Andersen, C. L., Pedersen, J. S., and Lund, A. H.: *SNHG5* promotes colorectal cancer cell survival by counteracting STAU1-mediated mRNA destabilization. Nature Communications. 7: 13875, December 2016.
- [8] Zhao, L., Guo, H., Zhou, B., Feng, J., Li, Y., Han, T., Liu, L., Li, L., Zhang, S., Liu, Y., Shi, J., and Zheng, D.: Long non-coding RNA SNHG5 suppresses gastric cancer progression by trapping MTA2 in the cytosol. Oncogene. 35(44): 5770–5780, November 2016.
- [9] He, B., Bai, Y., Kang, W., Zhang, X., and Jiang, X.: LncRNA SNHG5 regulates imatinib resistance in chronic myeloid leukemia via acting as a CeRNA against MiR-205-5p. Am J Cancer Res. 7(8): 1704–1713, PMC5574942, August 2017.
- [10] Yan, L., Wang, S., Li, Y., Tognetti, L., Tan, R., Zeng, K., Pianigiani, E., Mi, X., Li, H., Fimiani, M., and Rubegni, P.: SNHG5 promotes proliferation and induces apoptosis in melanoma by sponging miR-155. RSC Advances. 8(11): 6160–6168, 2018.
- [11] Ma, Z., Xue, S., Zeng, B., and Qiu, D.: LncRNA SNHG5 is associated with poor prognosis of bladder cancer and promotes bladder cancer cell proliferation through targeting p27. Oncology Letters. 15(2): 1924–1930, February 2018.
- [12] Shen, H., Wang, Y., Shi, W., Sun, G., Hong, L., and Zhang, Y.: LncRNA SNHG5/miR-26a/SOX2 signal axis enhances proliferation of chondrocyte in osteoarthritis. Acta Biochim Biophys Sin (Shanghai). 50(2): 191–198, February 2018.
- [13] Zheng, L., Hu, N., Guan, G., Chen, J., Zhou, X., and Li, M.: Long noncoding RNA SNHG6 promotes osteosarcoma cell proliferation through regulating p21 and KLF2. Archives of Biochemistry and Biophysics.
- [14] Paige, J. S., Wu, K. Y., and Jaffrey, S. R.: RNA Mimics of Green Fluorescent Protein. Science. 333(6042): 642–646, July 2011.
- [15] Filonov, G. S., Moon, J. D., Svensen, N., and Jaffrey, S. R.: Broccoli: Rapid Selection of an RNA Mimic of Green Fluorescent Protein by Fluorescence-Based Selection and Directed Evolution. J. Am. Chem. Soc. 136(46): 16299–16308, November 2014.
- [16] Autour, A., Jeng, S., Cawte, A., Abdolazadeh, A., Galli, A., Panchapakesan, S. S. S., Rueda, D., Ryckelynck, M., and Unrau, P. J.: Fluorogenic RNA Mango aptamers for imaging small non-coding RNAs in mammalian cells. Nature Communications. 9(1): 656, February 2018.

- [17] Dolgosheina, E. V., Jeng, S. C. Y., Panchapakesan, S. S. S., Cojocaru, R., Chen, P. S. K., Wilson, P. D., Hawkins, N., Wiggins, P. A., and Unrau, P. J.: RNA Mango Aptamer-Fluorophore: A Bright, High-Affinity Complex for RNA Labeling and Tracking. ACS Chem. Biol. 9(10): 2412–2420, October 2014.
- [18] Fusco, D., Accornero, N., Lavoie, B., Shenoy, S. M., Blanchard, J.-M., Singer, R. H., and Bertrand, E.: Single mRNA Molecules Demonstrate Probabilistic Movement in Living Mammalian Cells. Current Biology. 13(2): 161–167, January 2003.
- [19] Etzel, M. and Mörl, M.: Synthetic Riboswitches: From Plug and Pray toward Plug and Play. Biochemistry. 56(9): 1181–1198, March 2017.
- [20] Morisaki, T., Lyon, K., DeLuca, K. F., DeLuca, J. G., English, B. P., Zhang, Z., Lavis, L. D., Grimm, J. B., Viswanathan, S., Looger, L. L., Lionnet, T., and Stasevich, T. J.: Real-time quantification of single RNA translation dynamics in living cells. Science. 352(6292): 1425–1429, June 2016.
- [21] Braselmann, E., Wierzb, A. J., Polaski, J. T., Chromiński, M., Batan, D., Gryko, D., Batey, R. T., and Palmer, A. E.: Development of a riboswitch-based platform for live cell imaging of RNAs in mammalian cells. bioRxiv. 199240 pp, October 2017.
- [22] McNeil, P. L. and Warder, E.: Glass beads load macromolecules into living cells. Journal of Cell Science. 88(5): 669–678, December 1987.
- [23] Hayashi-Takanaka, Y., Yamagata, K., Wakayama, T., Stasevich, T. J., Kainuma, T., Tsurimoto, T., Tachibana, M., Shinkai, Y., Kurumizaka, H., Nozaki, N., and Kimura, H.: Tracking epigenetic histone modifications in single cells using Fab-based live endogenous modification labeling. Nucleic Acids Res. 39(15): 6475–6488, August 2011.
- [24] Johnson Jr, J. E., Reyes, F. E., Polaski, J. T., and Batey, R. T.: B₁₂ cofactors directly stabilize an mRNA regulatory switch. Nature. 492(7427): 133–137, December 2012.
- [25] Rosendahl, M. S., Omann, G. M., and Leonard, N. J.: Synthesis and biological activity of a profluorescent analogue of coenzyme B12. PNAS. 79(11): 3480–3484, June 1982.
- [26] Lee, M. and Grissom, C. B.: Design, Synthesis, and Characterization of Fluorescent Cobalamin Analogues with High Quantum Efficiencies. Org. Lett. 11(12): 2499–2502, June 2009.
- [27] Smeltzer, C. C., Cannon, M. J., Pinson, P. R., Munger, J. D., West, F. G., and Grissom, C. B.: Synthesis and Characterization of Fluorescent Cobalamin (CobalaFluor) Derivatives for Imaging. Org. Lett. 3(6): 799–801, March 2001.
- [28] Fesinmeyer, R. M., Hudson, F. M., and Andersen, N. H.: Enhanced Hairpin Stability through Loop Design: The Case of the Protein G B1 Domain Hairpin. J. Am. Chem. Soc. 126(23): 7238–7243, June 2004.
- [29] Shell, T. A. and Lawrence, D. S.: Vitamin B12: A Tunable, Long Wavelength, Light-Responsive Platform for Launching Therapeutic Agents. Acc. Chem. Res. 48(11): 2866–2874, November 2015.
- [30] Shell, J. R., Rodgers, Z. L., and Lawrence, D. S.: Tunable Visible and Near-IR Photoactivation of Light-Responsive Compounds by Using Fluorophores as Light-Capturing Antennas. Angew. Chem. Int. Ed. 53(3): 875–878, January 2014.
- [31] Cochran, A. G., Skelton, N. J., and Starovasnik, M. A.: Tryptophan zippers: Stable, monomeric β -hairpins. PNAS. 98(10): 5578–5583, May 2001.
- [32] Kier, B. L. and Andersen, N. H.: Probing the Lower Size Limit for Protein-Like Fold Stability: Ten-Residue Microproteins With Specific, Rigid Structures in Water. J. Am. Chem. Soc. 130(44): 14675–14683, November 2008.
- [33] Andersen, N. H., Olsen, K. A., Fesinmeyer, R. M., Tan, X., Hudson, F. M., Eidenschink, L. A., and Farazi, S. R.: Minimization and Optimization of Designed β -Hairpin Folds. J. Am. Chem. Soc. 128(18): 6101–6110, May 2006.
- [34] Jackowska, A., Chromiński, M., Giedyk, M., and Gryko, D.: 5'-Vitamin B12 derivatives suitable for bioconjugation via the amide bond. Org. Biomol. Chem. 16(6): 936–943, February 2018.

- [35] Chromiński, M., Lewalska, A., and Gryko, D.: Reduction-free synthesis of stable acetylide cobalamins. Chemical Communications. 49(97): 11406–11408, 2013.
- [36] Ruetz Markus, , Salchner Robert, , Wurst Klaus, , Fedosov Sergey, , and Kräutler Bernhard, : Phenylethynyl-cobalamin: A Light-Stable and Thermolysis-Resistant Organometallic Vitamin B12 Derivative Prepared by Radical Synthesis. Angewandte Chemie International Edition. 52(43): 11406–11409, September 2013.
- [37] Chromiński, M., Lewalska, A., Karczewski, M., and Gryko, D.: Vitamin B12 Derivatives for Orthogonal Functionalization. J. Org. Chem. 79(16): 7532–7542, August 2014.
- [38] Kolb Hartmuth C., , Finn M. G., , and Sharpless K. Barry, : Click Chemistry: Diverse Chemical Function from a Few Good Reactions. Angewandte Chemie International Edition. 40(11): 2004–2021, May 2001.
- [39] Patterson, D. M., Nazarova, L. A., and Prescher, J. A.: Finding the Right (Bioorthogonal) Chemistry. ACS Chem. Biol. 9(3): 592–605, March 2014.
- [40] Pathare, P. M., Wilbur, D. S., Heusser, S., Quadros, E. V., McLoughlin, P., and Morgan, A. C.: Synthesis of Cobalamin-Biotin Conjugates That Vary in the Position of Cobalamin Coupling. Evaluation of Cobalamin Derivative Binding to Transcobalamin II. Bioconjugate Chem. 7(2): 217–232, January 1996.
- [41] Grimm, J. B., Muthusamy, A. K., Liang, Y., Brown, T. A., Lemon, W. C., Patel, R., Lu, R., Macklin, J. J., Keller, P. J., Ji, N., and Lavis, L. D.: A general method to fine-tune fluorophores for live-cell and *in vivo* imaging. Nature Methods. 14(10): 987–994, October 2017.
- [42] Grimm, J. B., English, B. P., Chen, J., Slaughter, J. P., Zhang, Z., Revyakin, A., Patel, R., Macklin, J. J., Normanno, D., Singer, R. H., Lionnet, T., and Lavis, L. D.: A general method to improve fluorophores for live-cell and single-molecule microscopy. Nature Methods. 12(3): 244–250, March 2015.
- [43] Li, G.-W. and Xie, X. S.: Central dogma at the single-molecule level in living cells. Nature. 475(7356): 308–315, July 2011.
- [44] Katz, Z. B., English, B. P., Lionnet, T., Yoon, Y. J., Monnier, N., Ovrzyn, B., Bathe, M., and Singer, R. H.: Mapping translation 'hot-spots' in live cells by tracking single molecules of mRNA and ribosomes. eLife Sciences. 5: e10415, January 2016.
- [45] Yamagishi, M., Ishihama, Y., Shirasaki, Y., Kurama, H., and Funatsu, T.: Single-molecule imaging of β -actin mRNAs in the cytoplasm of a living cell. Experimental Cell Research. 315(7): 1142–1147, April 2009.
- [46] Halstead, J. M., Lionnet, T., Wilbertz, J. H., Wippich, F., Ephrussi, A., Singer, R. H., and Chao, J. A.: An RNA biosensor for imaging the first round of translation from single cells to living animals. Science. 347(6228): 1367–1671, March 2015.
- [47] Fiedler, B. L., Van Buskirk, S., Carter, K. P., Qin, Y., Carpenter, M. C., Palmer, A. E., and Jimenez, R.: Droplet Microfluidic Flow Cytometer For Sorting On Transient Cellular Responses Of Genetically-Encoded Sensors. Anal. Chem. 89(1): 711–719, January 2017.
- [48] Dean, K. M., Davis, L. M., Lubbeck, J. L., Manna, P., Friis, P., Palmer, A. E., and Jimenez, R.: High-Speed Multiparameter Photophysical Analyses of Fluorophore Libraries. Anal. Chem. 87(10): 5026–5030, May 2015.
- [49] Zurla, C., Lifland, A. W., and Santangelo, P. J.: Characterizing mRNA Interactions with RNA Granules during Translation Initiation Inhibition. PLOS ONE. 6(5): e19727, May 2011.
- [50] Jain, S., Wheeler, J. R., Walters, R. W., Agrawal, A., Barsic, A., and Parker, R.: ATPase-Modulated Stress Granules Contain a Diverse Proteome and Substructure. Cell. 164(3): 487–498, January 2016.
- [51] Nelles, D. A., Fang, M. Y., O'Connell, M. R., Xu, J. L., Markmiller, S. J., Doudna, J. A., and Yeo, G. W.: Programmable RNA Tracking in Live Cells with CRISPR/Cas9. Cell. 165(2): 488–496, April 2016.
- [52] Mueller, F., Senecal, A., Tantale, K., Marie-Nelly, H., Ly, N., Collin, O., Basyuk, E., Bertrand, E., Darzacq, X., and Zimmer, C.: FISH-quant: Automatic counting of transcripts in 3D FISH images. Nature Methods. 10(4): 277–278, April 2013.

- [53] Tutucci, E., Vera, M., Biswas, J., Garcia, J., Parker, R., and Singer, R. H.: An improved MS2 system for accurate reporting of the mRNA life cycle. Nature Methods. 15(1): 81–89, January 2018.
- [54] Kaewsapsak, P., Shechner, D. M., Mallard, W., Rinn, J. L., and Ting, A. Y.: Live-cell mapping of organelle-associated RNAs via proximity biotinylation combined with protein-RNA crosslinking. eLife Sciences. 6: e29224, December 2017.
- [55] Mairal, T., Özalp, V. C., Sánchez, P. L., Mir, M., Katakis, I., and O'Sullivan, C. K.: Aptamers: Molecular tools for analytical applications. Anal Bioanal Chem. 390(4): 989–1007, February 2008.
- [56] Cho, E. J., Lee, J.-W., and Ellington, A. D.: Applications of Aptamers as Sensors. Annual Review of Analytical Chemistry. 2(1): 241–264, 2009.
- [57] Lorsch, J. R. and Szostak, J. W.: In vitro selection of RNA aptamers specific for cyanocobalamin. Biochemistry. 33(4): 973–982, February 1994.
- [58] Porter, E. B., Polaski, J. T., Morck, M. M., and Batey, R. T.: Recurrent RNA motifs as scaffolds for genetically encodable small-molecule biosensors. Nature Chemical Biology. 13(3): 295–301, March 2017.
- [59] Trausch, J. J. and Batey, R. T. Chapter Three - Design of Modular “Plug-and-Play” Expression Platforms Derived from Natural Riboswitches for Engineering Novel Genetically Encodable RNA Regulatory Devices. In Burke-Aguero, D. H. (Ed.): Methods in Enzymology, volume 550 of *Riboswitches as Targets and Tools*, pp 41–71. Academic Press, January 2015.