

# Analysis of visual cortical neurons of mice

Cristian Bargiacchi, Christian Mancini

May 10, 2024

## 1 What this graph is about

Data of this graph was collected from the study of the paper [\[3\]](#).

The aim of the graph is to understand better the connections and structure of neurons in the primary visual cortex of a mouse.

Researchers have simulated visual stimulus on some neurons and tracked where the synapses were propagated or not.

### 1.1 Data components

Three types of components were tracked:

- Characterized pyramidal neuron
- Cell body in EM volume
- Dendritic fragment

We will try to get a brief explanation of these components.

#### 1.1.1 Characterized pyramidal neuron

These are the neurons that were stimulated with two-photon calcium imaging. A specific visual stimulus is created with this technique.

Every synapse starts from one of them.

#### 1.1.2 Cell body in EM volume

These are the neurons that are targeted from the synapse. With the large-scale electron microscopy (EM), like we described in the [README](#), it is possible to trace a portion of the local network of these neurons.

#### 1.1.3 Dendritic fragment

Part of the neuron specialized in retrieving signals from other neurons. It is different from the axon which is specialized in sending signals to other neurons.

We have *fragment* because the Dendritic is highly branched, so only a fraction of it was tracked.

## 1.2 Further division of the components

We said that a **pyramidal neuron** “starts” the synapse targeting a specific neuron.

Post-Synaptic data was also collected, so the components: *Cell body in EM volume* and *Dendritic fragment* can be classified to:

- Postsynaptic excitatory target,
- Postsynaptic inhibitory target

**excitatory** and **inhibitory** means: “A target that *propagates* the signal and a target that *stops* the signal respectively.”

Since all the synapses start from the *characterized pyramidal neurons*, they do not have this further division and that attribute is marked as *NA*.

## 2 Descriptive analysis

We now start from the descriptive analysis since is easier and let’s us take confidence with these dataset.

```
[1]: import igraph as ig
import matplotlib.pyplot as plt
import random
```

First step is to importing the graph downloaded from the site [2].

Unfortunately with this version we are using of iGraph, we can’t read from an URL.

The data was downloaded and put in the [Data](#) folder.

```
[2]: neurons : ig.Graph
neurons = ig.Graph.Read_GraphML("../Data/mouse_visual.cortex_2.graphml")
neurons.to_directed()
```

Since the return type of `Read_GraphML` is erroneously hinted as `None` (which is not because is a `Graph`) we hint the interpreter that `neurons` is a `Graph`, so we can use autocompletion and suggestions.

As we saw before, we have three types of nodes in this network and are described as nodal attributes of name `type1`

```
[3]: print(f"The three node possible types are : {set(neurons.vs['type1'])}")
```

The three node possible types are : {'Cell body in EM volume', 'Characterized pyramidal neuron', 'Dendritic fragment'}

Furthermore if a node is a Postsynaptic target, i.e. if a node is not a Characterized pyramidal neuron, we have two others classifications:

```
[4]: print(f"We can further classify as: {set(neurons.vs['type2'])}")
```

We can further classify as: {'Postsynaptic inhibitory target', 'Postsynaptic excitatory target', 'NA'}

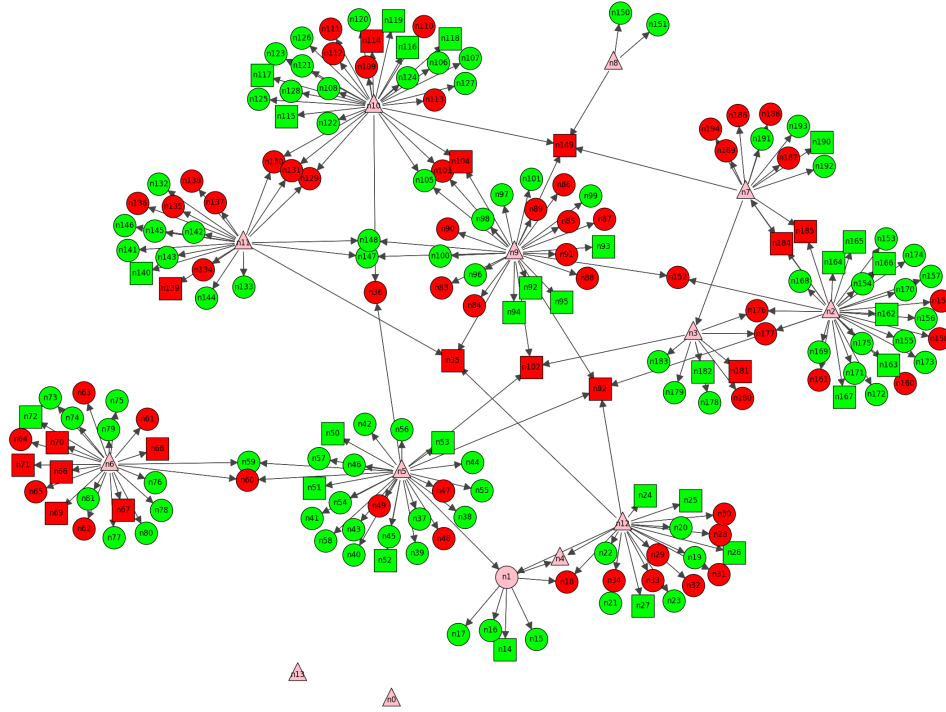
## 2.1 Plotting the network

Let's now plot the data.

We will use `matplotlib` for backend.

```
[5]: random.seed(42)

fig, ax = plt.subplots(figsize=(32,24))
ig.plot(
    neurons,
    target=ax,
    layout="fruchterman_reingold",
    vertex_shape = ["circle" if type == "Dendritic fragment" else "square" if
↪type == "Cell body in EM volume" else "triangle" for type in neurons.
↪vs["type1"]],
    vertex_size=[40 if type == "Characterized pyramidal neuron" else 50 for
↪type in neurons.vs["type1"]],
    vertex_label=neurons.vs["id"],
    vertex_color=["green" if type == "Postsynaptic excitatory target" else
↪"red" if type == "Postsynaptic inhibitory target" else "pink" for type in
↪neurons.vs["type2"]],
    edge_width = 1
)
plt.gcf().set_facecolor('none')
plt.savefig("../Plots/neurons.png",transparent=True)
plt.show()
```



## 2.2 Network Statistics

We now start to deep dive in to the graph structure, calculating and interpreting the major networks statistics.

We will use the following statistics:

- Density
- Reciprocity
- Transitivity (undirected case)
- Modularity/Assortative Mixing

All the network statistics are linked to the corresponding section.

Each of these statistics answer a different and a specific question which we will analyze one by one.

A recap of all these statistics are in [this](#) section.

### 2.2.1 Density

#### QUESTION

*How connected is the graph?*

The density  $\rho$  strictly lies between  $0 \leq \rho \leq 1$ .

If the density approaches 0 it means that the graph is *sparse*, if it approaches 1 it is said to be *dense*.

Density is also an estimate of the probability of observing a tie between randomly sampled nodes.

Documentation of the method can be found [here](#).

```
[6]: density = neurons.density(loops = False)
print(f"The density of the graph is: {density:.4f}")
print(f"The probability of observing a random tie is {density*100:.2f}%")
```

The density of the graph is: 0.0057

The probability of observing a random tie is 0.57%

As we can see the network is sparse. If we go back to the [plot](#) we can observe that indeed the graph is sparse and the majority of ties are the ones that forms stars with *Characterized pyramidal neurons* as the centers.

### 2.2.2 Reciprocity

#### Question

*How strong is the tendency to return a tie in the network?*

The reciprocity coefficient  $R$  lies in the range  $0 \leq R \leq 1$ .

The more this value is 0, the more relations observed between nodes in the network are **not** reciprocated.

Value close to 1 means relations observed between nodes in the network are reciprocated.

For what we know from our assumptions (i.e.) *All the synapses starts from a Characterized pyramidal neuron and the path have length 1*, we expect that this statistic approaches 0.

Documentation of the method can be found [here](#).

```
[7]: reciprocity = neurons.reciprocity()
print(f"The reciprocity of the graph is: {reciprocity:.1f}")
```

The reciprocity of the graph is: 0.0

Indeed the reciprocity is 0, in fact no node returns a tie in the network.

In our case we do not have enough information to state if in the reality the reciprocity is higher, because the data collected from the experiment contains only one step of the path of the propagation of the stimulation.

### 2.2.3 Transitivity

#### Question

*What proportion of triangle do we observe in the network?*

Transitivity is the ratio of the triangles and connected triplets in the graph.

In a social graph the question could be: *Are friends of friends also friends?*

In our case, finding a triangle is an interesting issue for understanding the structure of the visual cortex. However, as we stated before, paths collected are of length one, so we expect that even this statistic approaches to 0.

Documentation of the method can be found [here](#).

```
[8]: transitivity = neurons.transitivity_undirected()
print(f"Transitivity of the graph is: {transitivity:.4f}")
```

Transitivity of the graph is: 0.0047

The transitivity is not 0, this means that there is at least 1 triangle. Still the statistic approaches 0, this means that in general we can expect no triangle.

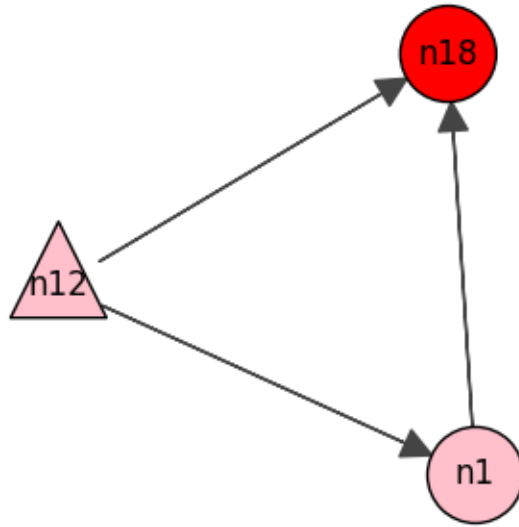
Going back to the `plot` we can see a triangle on the bottom-right of the plot, composed by node 1, 12 and 18.

For simplicity a subgraph of that triangle is reported below.

```
[9]: ids = ["n1", "n12", "n18"]
vertex_idx = [v.index for v in neurons.vs if v["id"] in ids]
triangle = neurons.subgraph(vertex_idx)

random.seed(42)
fig, ax = plt.subplots(figsize=(4,4))
ig.plot(
    triangle,
    target=ax,
    layout="fruchterman_reingold",
    vertex_shape = ["circle" if type == "Dendritic fragment" else "square" if
    ↪type == "Cell body in EM volume" else "triangle" for type in triangle.
    ↪vs["type1"]],
    vertex_size=[50 if type == "Characterized pyramidal neuron" else 50 for
    ↪type in triangle.vs["type1"]],
    vertex_label=triangle.vs["id"],
    vertex_color=["green" if type == "Postsynaptic excitatory target" else
    ↪"red" if type == "Postsynaptic inhibitory target" else "pink" for type in
    ↪triangle.vs["type2"]],
    edge_width = 1
)

plt.gcf().set_facecolor('none')
plt.savefig("../Plots/triangle.png", transparent=True)
plt.show()
```



## 2.2.4 Modularity

### Question

*Are neurons with lot of connections of the same type?*

Indeed we have one particular type of node, which is the *Characterized pyramidal neuron* that have the highest out-degree and the lowest in-degree.

We expect to observe disassortative mixing, because in our case the stimulus starts from a *Characterized pyramidal neuron* and ends up to a *dendritic fragment* or to a *cell body in EM volume*.

The function we use to calculate the modularity coefficient is **not** normalized.

Documentation of the method can be found [here](#).

```
[10]: # We have to binarize the qualitative attributes first.
pyramidal = [1 if x == "Characterized pyramidal neuron" else 0 for x in neurons.
            vs["type1"]]
dendritic = [1 if x == "Dendritic fragment" else 0 for x in neurons.vs["type1"]]
cell = [1 if x == "Cell body in EM volume" else 0 for x in neurons.vs["type1"]]

# Now we can calculate modularity with respect to each qualitative attribute
pyramidal_modularity = neurons.modularity(pyramidal)
dendritic_modularity = neurons.modularity(dendritic)
cell_modularity = neurons.modularity(cell)

print(f"Modularity for Characterized pyramidal neuron is: {pyramidal_modularity:
    .5f}")
```

```
print(f"Modularity for Dendritic fragment is: {dendritic_modularity:.5f}")
print(f"Modularity for Cell body in EM volume is: {cell_modularity:.5f}")
```

Modularity for Characterized pyramidal neuron is: -0.00830  
 Modularity for Dendritic fragment is: -0.00271  
 Modularity for Cell body in EM volume is: 0.00000

The modularity coefficient that we use is strictly less than 1.

We just interpret the first one, *Modularity for Characterized pyramidal neuron* since the other two are not useful for our analysis. This coefficient is negative, so it indicates disassortative mixing as we expected. We can conclude that *Characterized pyramidal neuron* **don't** form ties between them.

### 2.2.5 Recap of the network statistics

We plot a table that summarizes all the network statistics calculated until now.

```
[11]: import pandas as pd

network_stats_dic = {
    "network_statistics": ["Density", "Reciprocity", "Transitivity",
↪ "Modularity"],
    "values": [density, reciprocity, transitivity, pyramidal_modularity]
}

network_stats = pd.DataFrame(network_stats_dic)
network_stats.set_index("network_statistics", inplace=True)
network_stats.T.head()
```

```
[11]: network_statistics  Density  Reciprocity  Transitivity  Modularity
values                0.005657             0.0         0.004687    -0.008298
```

## 2.3 Nodal Statistics

We can now use some centrality measure to spot important nodes given a specific criteria.

We will use the following centrality measures:

- Degree
- Closeness
- Betweenness
- Eigenvector

All these centrality measures will be also incorporated to the corresponding centralization index. The interpretation varies depending on the measure used.

Centralization index is not provided in python-igraph, however is easy to implement and we provide these methods in the following module.

For simplify the visualization and the comparison of vertex centrality measures, we create a DataFrame to store information of the nodes and the relative measurements.



```
[12]: data = {
        "id": neurons.vs["id"],
        "type1": neurons.vs["type1"],
        "type2": neurons.vs["type2"]
    }
    centrality = pd.DataFrame(data)
    centrality.tail(2)
```

```
[12]:      id      type1      type2
193  n193  Dendritic fragment  Postsynaptic excitatory target
194  n194  Dendritic fragment  Postsynaptic inhibitory target
```

The index of the rows corresponds to the attribute id of the nodes.

### 2.3.1 Degree Centrality

We define a node important if it is connected to many other nodes.

We have a directed graph, so we can distinguish between in and out degree, or we can consider the support of the graph.

In our case it makes more sense to consider a node important if it has high **in** degree, because every stimulus starts from a known pyramidal with high out degree, so it doesn't make sense to consider important those kind of nodes.

The nodes with high in degree are the ones more targeted to propagate or inhibit the stimulus. We consider this type of nodes important.

Base method documentation can be found [here](#).

```
[13]: centrality["in_degree"] = neurons.degree(centrality.index,mode="in")
    centrality["in_degree_std"] = centrality["in_degree"]/centrality["in_degree"].
    ↪max()
    print(f"The maximum in-degree is: {centrality["in_degree"].max()}")
    centrality.sort_values(by=["in_degree"], ascending=False).
    ↪head(3)[["type1","type2","in_degree_std"]]
```

The maximum in-degree is: 4

```
[13]:      type1      type2  in_degree_std
149  Cell body in EM volume  Postsynaptic inhibitory target      1.00
82   Cell body in EM volume  Postsynaptic inhibitory target      1.00
102  Cell body in EM volume  Postsynaptic inhibitory target      0.75
```

We can see that **in\_degree** is relatively low and the maximal value is 4.

It's important to note that the most important nodes, based on in degree centrality, are all Postsynaptic inhibitory target.

```
[14]: from centralization.centralization_index import degree_centralization

    degree = degree_centralization(neurons,"in")
```

```
print(f"Network degree centralization (standardized): {degree:.03}")
```

Network degree centralization (standardized): 0.015

Based on the network degree centralization, we can conclude that there are no extremely important nodes. That makes sense because if there were few extremely important neurons, “loosing” them implies that all the visual cortex will become unusable and that is a too fragile system.

### 2.3.2 Closeness Centrality

We define a node important if it is close to many other nodes.

For calculating this measure we have to calculate every shortest path. This computation is expensive and there are many tricks to approximate this measure, (i.e.) using sketches. In our case, since the graph is small, we rely on the exact measure provided by python-igraph.

Since we have a directed graph we have to choose if the calculation must be applied considering in-edges, out-edges or the undirected case. Undirected case will give us that “Characterized pyramidal neuron” is indeed important, but we already know that. For this reason we choose to consider in-edges important.

Base method documentation can be found [here](#).

```
[15]: centrality["in_closeness_std"] = neurons.closeness(mode="in",normalized=True)
centrality.sort_values(by=["in_closeness_std"], ascending=False).
↳head(3)[["type1","type2","in_closeness_std"]]
```

```
[15]:
```

	type1	type2	in_closeness_std
184	Cell body in EM volume	Postsynaptic inhibitory target	1.0
185	Cell body in EM volume	Postsynaptic inhibitory target	1.0
19	Dendritic fragment	Postsynaptic excitatory target	1.0

Here things are interesting, because the majority of nodes ( $\approx \frac{3}{4}$ ) have standardized closeness equal to 1.

If all nodes in a graph have the same closeness centrality along a specific path, it could indicate a well-balanced network structure where all nodes are equally important and interconnected. This scenario suggests strong connectivity and a balanced distribution of the nodes’ roles in transmitting information along the specified path.

### 2.3.3 Betweenness Centrality

This centrality measure can be interpreted as a *reflection of power* in a social network perspective, since a high value indicates that the node is *between* a lot of shortest paths.

All our paths (the majority) have length 1. We expect to observe most of the measure to 0.

```
[16]: centrality["betweenness"] = neurons.betweenness(directed=True)
centrality["betweenness_std"] = centrality["betweenness"] /
↳centrality["betweenness"].max()
centrality.sort_values(by=["betweenness_std"], ascending=False).
↳head(3)[["type1","type2","betweenness_std"]]
```

```
[16]:
```

	type1	type2	betweenness_std
7	Characterized pyramidal neuron	NA	1.000000
3	Characterized pyramidal neuron	NA	0.888889
1	Dendritic fragment	NA	0.555556

These 3 neurons are the only ones with centrality not equals to zero and the **ONLY** “*signal generators*” that have in edges.

### 2.3.4 Eigenvector Centrality

For the eigenvector centrality a node is important if it is connected to many important nodes. We calculate this quantity using the method `eigvals` of numpy, which gives us the eigenvalues of the adjacency matrix.

```
[17]: import numpy as np
from numpy import linalg as LA

centrality["eigenvector"] = LA.eigvals(np.array(neurons.get_adjacency()).data)
# gives all NaN (0/0)
centrality["eigenvector_std"] = centrality["eigenvector"] /
    ↪centrality["eigenvector"].max()
centrality.sort_values(by=["eigenvector"], ascending=False).
    ↪head(3)[["type1", "type2", "eigenvector"]]
```

```
[17]:
```

	type1	type2 \
194	Dendritic fragment	Postsynaptic inhibitory target
0	Characterized pyramidal neuron	NA
1	Dendritic fragment	NA

	eigenvector
194	0.0
0	0.0
1	0.0

As in the other measures we find that there are no “*dominant*” nodes, since the network is well and uniformly structured so that every nodes participates in the spread of information.

### 2.3.5 Nodal statistics summary

The only centrality measure that provides information is the **degree centrality**.

Our conclusion, based on this descriptive analysis is that the network is sparse, but it is well balanced with no evidence of important nodes.