

Meeting May 19, 2020

From Lyden:

This is an awesome start. A lot of the data, e.g. site, animal ID, will already be in the REDCap database. Machine and image acquisition parameters 'should' all be in the file header of the dicom files, assuming all sites comply with the upload instructions.

I want to argue against any manual readouts, such as 'hemorrhage' or 'midline shift' because we are valiantly trying to automate all of the image analysis. The less human intervention the better, I think. Having said that, hemorrhage is important, and we have not previously discussed it. For the RHAPSODY study, we used a novel protocol from Mark Haacke called SPIN that automatically quantitated blood. It's a little late in the game now, but maybe you all can discuss and see if there is an automated process you could implement, if not in time for STAGE I perhaps by Stage 2.

Variable name:

Data type (numeric vs text vs date, etc):

Variable Length (number of digits or characters):

Allowable range:

Any notes:

For example, let's take volume of stroke as an example.

Variable name: Lesion_Volume

Type: numeric

Length: 7 digits - nnnn.nnn

Range: 0ml to 40ml

Notes: volume is calculated as the measurable void on T2

From Karen:

For the scan acquisition info we do extract metadata from the image headers (see attached from example scans uploaded to SPAN) and store in the database. For analysis results, if they are in CSV format they could potentially be pushed into the repository and linked assuming that the results dataset contains the image ID.

<https://s3-us-west-2.amazonaws.com/secure.notion-static.com/3ac252fb-8003-40bc-b9aa-e7b4069b3ee9/span.csv>

From Joe:

Thanks Ryan. Everything looks pretty good.

A few questions and possible additions to consider now for acquisition and analysis;

1) In our last zoom, you mentioned the potential problem of ADC normalization within a lesion in at least one of the mice. My assumption from the beginning of this has been that ADC

will not be reliable at 48 hours, and so I was thinking of this simply as a method to eliminate CSF, but not as something to be used for lesion segmentation. This is partly/mostly a biology question, I think:

what do we do when we see clearly elevated T2 in the core of the lesion but ADC does discriminate that area? I was thinking we would ignore ADC in that instance. What do others think?

2) I have some concern about too-small surface coils potentially not covering the space adequately (see the last example in your zip). If we get to the spatial normalization soon enough,

we may want to add a metric of minimum SNR without the brain mask as a way of seeing drop-off problems.

3) If we are going to use spatial normalization, which makes sense to me (and which I first suggested but then we dropped because it seemed potentially difficult),

we probably want to specify a stroke side and that animals must be registered in the scanners correctly (e.g., don't accidentally swap "feet first" vs "head first").

It would simplify spatial normalization and segmentation to always have the lesion on the same side in images. In principle, one could swap coordinate parities, but this is an additional step.

4) Do you have spatial normalization methods that you think will work well in rodents given large strokes and the signal roll-off from surface coils?

I have some experience with this, so I'd be happy to chat about it. There probably are a variety of methods that would work but the typical strategy of brain extraction usually isn't very good in rodents.

From Ryan:

Thanks Joe – all seem like good considerations! Here are a few thoughts that come to mind:

1) I noticed this issue in the data too, where T2 and ADC disagree in some cases. I'd have to do some reading to weigh in myself, and happy to update things as folks see fit. I think we can also adjust the ADC threshold to do what you describe.

2) Yes, it does seem like there is inhomogeneity that affects the lesion, and I bet this would be problematic for brain extraction and atlas registration. I'm using N4 bias field correction, but it can only do so much for the farthest signal drop.

3) I did some tests with ANTs, and the results seemed encouraging, but it did fail in about

t a quarter of the cases. I used a standard T2 template, but I'd guess the differences in field of view and contrast may be an issue. We could possibly make a study-specific (or perhaps site-specific) template once the protocol is nailed down and there is more data

4) Would be happy to brainstorm! I've used ANTs in other rodent datasets, but they don't seem to work as well here. Seems like we can segment the stroke without template registration, but I think we would need it to measure brain/csf volume and to subdivide by hemisp here.

From Joe:

I typically have not relied upon brain extraction for spatial normalization in rodent species, mostly due to non-standard image contrasts and unpredictable RF bias. Instead I simply register into a template-based brain mask to eliminate extra-cerebral signal. The upside is that it eliminates the brain extraction step, and the downside is that the "capture range" on automated alignment is reduced (it typically needs to get "close", like 1/4 FOV, for automated alignment to capture it). For RF bias, I divide each image (template/source) by a low-passed version of self, so one can enforce an arbitrary degree of flattening.

One possible strategy would be to segment the lesion in native space and then initialize normalization using the lesion centroid. Then we could align using a series of progressively increasing flattening to solve the bias and lesion issues. I think in the end the template should be the normalized average of lesioned mice, so there will be some iteration. Then final processing occurs in normalized space.

I haven't experimented with ANTs. It may well be the case that ANTs just works when you use a template based upon lesioned mice. I feel like we shouldn't have to use site-specific templates, but I guess that would be ok if it enables using some existing method.

Meeting Notes:

- Add bias field quality control metric

https://s3-us-west-2.amazonaws.com/secure.notion-static.com/4bda01cc-e95c-4440-8b08-0b650b3c035f/MRI_database_entries.xlsx