

Cancers make their own luck: theories of cancer origins

Amir Jassim¹, Eric P. Rahrmann¹, Ben D. Simons^{2,3} & Richard J. Gilbertson^{1,4}✉

Abstract

Cancer has been a leading cause of death for decades. This dismal statistic has increased efforts to prevent the disease or to detect it early, when treatment is less invasive, relatively inexpensive and more likely to cure. But precisely how tissues are transformed continues to provoke controversy and debate, hindering cancer prevention and early intervention strategies. Various theories of cancer origins have emerged, including the suggestion that it is ‘bad luck’: the inevitable consequence of random mutations in proliferating stem cells. In this Review, we discuss the principal theories of cancer origins and the relative importance of the factors that underpin them. The body of available evidence suggests that developing and ageing tissues ‘walk a tightrope’, retaining adequate levels of cell plasticity to generate and maintain tissues while avoiding overstepping into transformation. Rather than viewing cancer as ‘bad luck’, understanding the complex choreography of cell intrinsic and extrinsic factors that characterize transformation holds promise to discover effective new ways to prevent, detect and stop cancer before it becomes incurable.

Sections

Introduction

Theories of cancer origins

Cell intrinsic factors

Cell extrinsic factors

The convergence of cancer risk factors

Conclusions

¹CRUK Cambridge Institute, University of Cambridge, Cambridge, UK. ²Wellcome Trust-Medical Research Council Cambridge Stem Cell Institute, Jeffrey Cheah Biomedical Centre, University of Cambridge, Cambridge, UK. ³Department of Applied Mathematics and Theoretical Physics, Centre for Mathematical Sciences, University of Cambridge, Cambridge, UK. ⁴Department of Oncology, University of Cambridge, Cambridge, UK. ✉e-mail: Richard.Gilbertson@cruk.cam.ac.uk

Introduction

The successful treatment of any disease requires an understanding of the biology at work in diseased tissues. During the nineteenth and early twentieth centuries, infectious diseases were the leading cause of death^{1,2} (Fig. 1). Early recognition that these diseases might be prevented through appropriate quarantine and sanitation³, followed by the discovery of bacteria and viruses as their causative agents, inspired basic research⁴, prevention⁵ and targeted chemotherapy programmes^{6,7}. This evolution in thinking brought about a fundamental change in our understanding, prevention and treatment of infection, and thereby a dramatic decline in morbidity and mortality (Fig. 1). Public health measures and the development of vaccines have been central to this success, preventing some infections from ever taking hold^{8,9}.

As infection control improved through the first half of the twentieth century, the evolving practice of modern medicine was applied to cancer, which had emerged as a major killer (Fig. 1). The discovery of chemicals that could kill rapidly dividing cells launched the field of cancer chemotherapy^{10,11}, whereas the understanding of oncogenes and crosstalk between cancer and the immune system has yielded effective molecule-targeted therapies¹² and immunotherapies^{13,14}, respectively.

Despite these advances, cancer remains a leading cause of death by disease. Every year, more than 19 million people are diagnosed with cancer and 10 million die of the disease, accounting for one in six deaths globally¹⁵. Two thirds of these deaths occur in lower and middle-income countries where cancer is often diagnosed late and access to treatment is limited¹⁵. Even in relatively developed countries, more than one third of patients diagnosed with cancer present as emergency cases with relatively late-stage disease and a significantly increased risk of dying within 12 months¹⁶. The annual global cancer burden is expected to rise to more than 28 million cases by 2040, with a larger increase in transitioning countries (from 64% to 95%) than in transitioned countries (from 32% to 56%)¹⁵. These dismal statistics have prompted worldwide calls to reduce cancer mortality by a third within the next decade^{17–19}. Increasing focus is being placed on detecting and treating cancer, or its precursors, as early as possible in the belief that this approach will dramatically increase patient survival while decreasing the invasiveness, cost and side effects of treatment^{20,21}.

Achieving this ambition will require a sea change in the way we study and manage cancer. Similar to the control of infectious disease, the greatest reductions in cancer mortality have been achieved through epidemiological research and primary prevention measures including tobacco control²², regulating occupational carcinogen exposure²³ and vaccines^{24,25}. Although much of this success has been achieved without detailed knowledge of the biology of cancer origins, further progress will require an evolution of our approach to the challenge of cancer. At least half of all cancers are still thought to be preventable²⁶, yet the majority of cancer research funding is invested in late-stage disease. Concerted, multidisciplinary research efforts engaging basic, epidemiological and clinical researchers armed with a better understanding of cancer origins will be required if we are to invent effective strategies that prevent cancer or detect and treat it early.

In this Review, we discuss the principal theories of cancer origins and the relative contribution of intrinsic and extrinsic factors to malignancy. It is envisaged that a better understanding of these processes will significantly accelerate our ability to diagnose cancer early, and treat it more precisely, when the disease is easier to control with less expensive and relatively non-toxic treatments.

Theories of cancer origins

Differences in exposure to risk factors and life expectancy result in global heterogeneity in the leading cancer types; but cancers are not distributed randomly across the body. This is most apparent in the comparison of paediatric²⁷ and adult¹⁹ malignancies (Fig. 2). Paediatric malignancies are relatively rare (1 in 440 children)^{27,28}, initiate during embryogenesis predominantly within ectodermal (for example, brain tumours) and mesodermal (for example, haematological malignancies) lineages, and have a relatively low mutational burden. Conversely, one in two adults develop cancer with a relatively high mutational burden, and almost entirely within epithelial tissues after the sixth decade of life¹⁹. These patterns suggest strongly that cancer is not a random process but one dictated by reproducible determinants in developing and ageing tissues.

Although the incidence and age of onset of many cancer types are well documented, our understanding of how cancers arise continues to provoke debate (Fig. 3 and Box 1). The observation that cancer incidence increases with age has been explained by the somatic mutation theory of cancer. First proposed almost 100 years ago, this theory posits that cancers arise in proliferating cell lineages that acquire six or seven ‘factors’ (now believed to be DNA mutations) during life^{29–32}. But certain observations do not fit this theory, including spontaneous or hormone-driven regression of paediatric and adult cancers^{33,34}, normalization of malignant teratomas injected into blastocysts³⁵ and evidence that many carcinogens do not damage DNA^{36,37}. Therefore, Soto and Sonnenschein³⁸ proposed an alternative tissue organization field theory of cancer (Fig. 3). The tissue organization field theory proposes that whole tissues are the target of carcinogens, disturbing the biophysical and biomechanical communication between the parenchyma and the mesenchyme or stroma. As a consequence, the proliferation and motility restraints imposed by normal tissue architecture are lost, inducing progressive metaplasia, dysplasia and carcinoma. Recently, two additional theories have been proposed – the bad luck³⁹ and ground state⁴⁰ theories – that draw on concepts underpinning both the somatic mutation and tissue organization field theories (Fig. 3).

The bad luck theory

Tomasetti and colleagues^{39,41} proposed a model in which random mistakes made during DNA replication in stem cells (R-mutations) result in the inevitable propagation of mutant clones leading to cancer (Fig. 3). This model distinguishes R-mutations from those that are heritable (H-mutations) or caused by environmental carcinogens (E-mutations). By comparing the incidence of 17 human cancer types reported in 423 cancer registries across 69 countries with estimated rates of stem cell division in the corresponding host tissues, the authors calculated that as many as two thirds of cancer-causing mutations are R-mutations.

Although similar to the somatic mutation theory, the bad luck theory – so called because R-mutations and stem cell divisions are an inevitable characteristic of tissues – is important as it provides a conceptual framework to understand the relative contributions of H, E and R-factors to cancer risk.

But elements of the bad luck theory are problematic. It is well established that cancer risk varies temporarily and geographically in a manner that cannot be attributed merely to the chance mutation of dividing stem cells⁴². Furthermore, the assumption that cancer risk is dictated entirely by the number of stem cell divisions throughout life does not adequately account for other cell intrinsic (for example, epigenetic states) or extrinsic (for example, immune microenvironment) factors that may modulate cancer susceptibility independent of

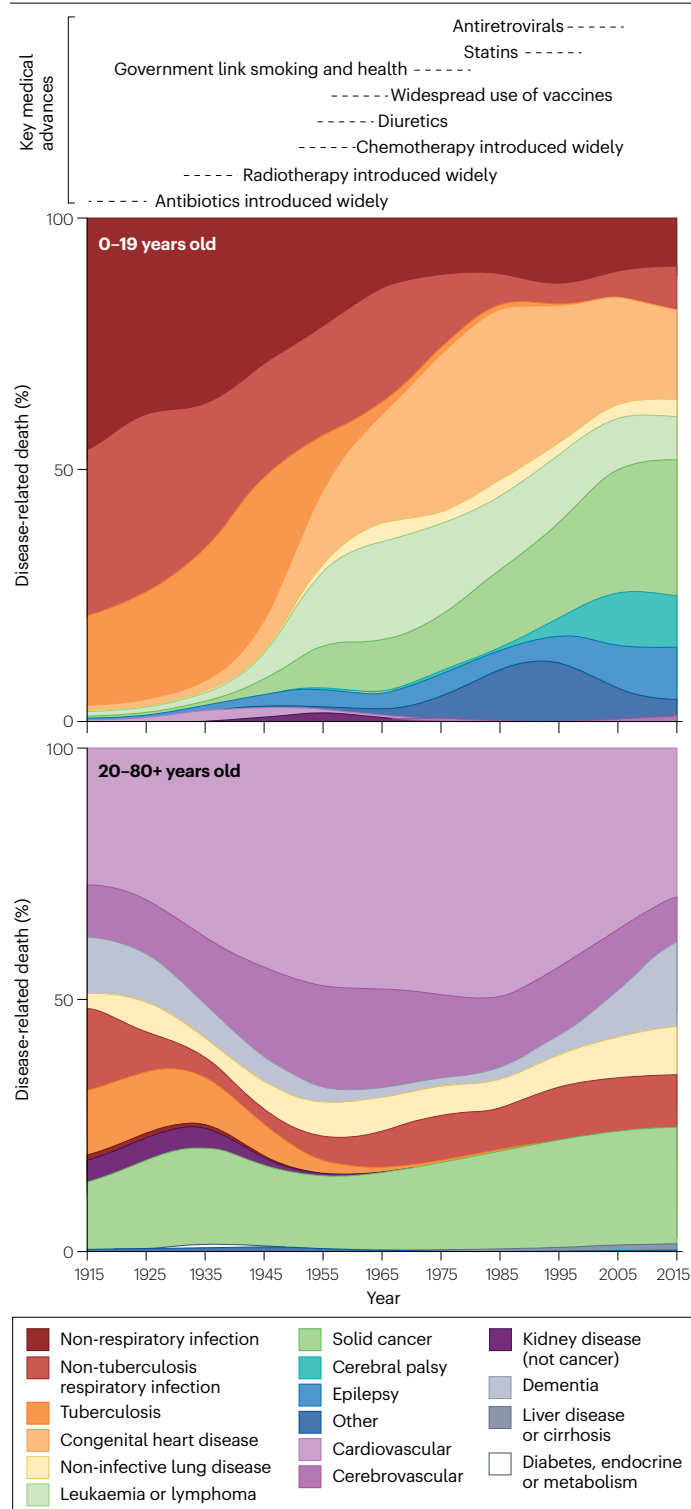


Fig. 1 | Major causes of disease-related death. The major causes of disease-related death recorded by the UK Office for National Statistics² between 1915 and 2015 in individuals 0–19 years of age (upper) or >20 years of age (lower). Hashed lines at top show the point at which the indicated major medical breakthroughs began to impact clinical practice.

cell division. Indeed, the mutation-centric nature of the theory assumes that extrinsic factors impact cancer risk merely through E-mutations rather than other processes such as changes in cell state (that is, metaplasia, discussed later in this Review) following tissue damage. This is particularly important in light of evidence that many carcinogens are not mutagenic³⁷, and mutational processes that are independent of cell division contribute substantially to somatic mutagenesis⁴³. These shortcomings – shared by the somatic mutation theory – in part inspired the tissue organization field theory of cancer.

There are also technical concerns with the data used to support the bad luck theory. By necessity, many of the stem cell division metrics used by Tomasetti and Vogelstein³⁹ were not measured directly but were derived from comparisons of the total number of cells in each tissue with estimates of the number of resident stem cells. This approach does not account for age and non-malignant disease-related variations in the stem cell state that might impact cancer risk.

The ground state theory

We have proposed an alternative theory for cancer origins that focuses on the functional state of a cell (its ‘ground state’) rather than its classification as a stem, lineage-committed progenitor, or other cell type⁴⁰ (Fig. 3). This concept is important as it accommodates the notion of cell plasticity in which developmental, ageing and injury factors can alter the susceptibility of cells to transformation independent of cell division. This theory accords with the observation that many carcinogens are not mutagenic^{37,44}, and mutational processes that are independent of cell division contribute substantially to somatic mutagenesis⁴³. Thus, the ground state theory builds on principles underpinning both the bad luck and tissue organization field theories, while emphasizing the convergence of extrinsic and intrinsic factors to generate the cell states that drive cancer. It is hoped that considering cancer origins in this manner will allow us to build on the success of epidemiological studies, and develop effective cancer prevention, early diagnosis and intervention strategies^{20,21,45,46}.

In contrast to the work of Tomasetti and colleagues, we formulated the ground state theory from observations made directly in genetically engineered mouse models⁴⁰. Using in vivo lineage tracing we first recorded the stem cell capacity of specific populations of cells marked with prominin 1 (PROM1; also known as CD133, a well-recognized marker of certain normal and malignant stem cells^{40,47}) in 14 major organs: lineage tracing is a gold standard in vivo test of stem cell function in which cells and their progeny are genetically labelled and tracked throughout life with a permanent fluorescent marker⁴⁸. Cells were lineage traced in both neonatal and adult mice to understand how stem cell function might vary with age. In a parallel set of experiments, we measured the susceptibility of these same PROM1⁺ cells to transformation in neonatal and adult mice by conditionally activating oncogenes (*Cttnb1* (which encodes β -catenin), *Kras* or *Notch1*) and/or inactivating tumour suppressor genes (*Trp53*, *Pten* or cyclin-dependent kinase inhibitor 2A (*Cdkn2a*)).

In agreement with the bad luck theory, the level of stem and/or progenitor cell function of any given PROM1⁺ cell correlated directly with its susceptibility to form cancer. This held true in the presence of multiple mutations regardless of the developmental stage at gene induction, strongly supporting the notion that stem cells dictate organ cancer risk. However, this risk varied markedly with age. On average, PROM1⁺ neonatal cells were 7-fold more resistant to transformation than their adult counterparts. Cancer resistance in neonates was independent of stem cell proliferation, organ site and

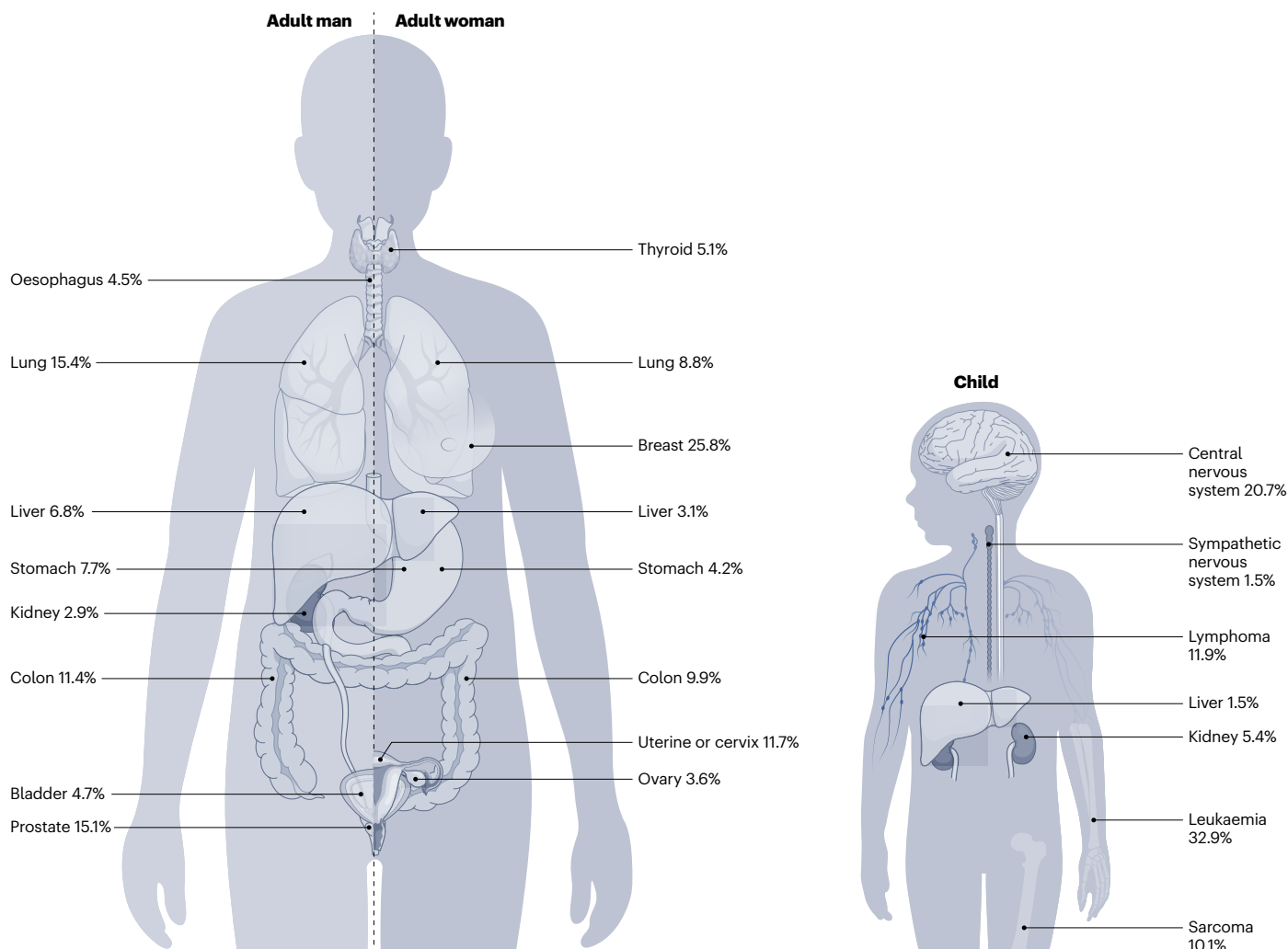


Fig. 2 | Organ sites of common human cancers. The major organ sites and types of adult (left) and childhood (right) cancer. Adult schematic reports male (left) and female (right) cancers. The percentage of the total cancer burden contributed by each cancer type is shown. Evidence of a cancer origin in stem and/or

progenitor cells for some of the depicted cancer types can be found in refs. [40,47,57,59–68,75–86,88,90](#). Adult data obtained from the [World Cancer Research Fund](#) website, and childhood data from ref. [27](#).

lifelong persistence of mutations. It is tempting to speculate that multi-organ species have evolved cancer resistance mechanisms to protect tissues from transformation during the extreme mitotic and differentiation stresses of early development^{49–51}. Thus, cancer risk is not merely an inevitable consequence of mutant stem cell proliferation but is also dictated by age-dependent, proliferation-independent variables in stem cells.

Comparative studies of embryonic, neonatal and adult haematopoietic stem cells (HSCs) support this notion, demonstrating the relative resistance of immature stem cells to transformation. For example, the FMS-like tyrosine kinase 3 (*FLT3*) internal tandem duplication mutation that is common in adult acute myeloid leukaemia (AML) but rare in childhood AML only induces transforming, self-renewal and myeloid commitment programmes once haematopoietic progenitors have transitioned from a fetal to an adult transcriptional state⁵². Similarly, differences in enhancer of zeste homologue 2

(EZH2)-dependent histone H3 lysine 27 (H3K27) trimethylation between leukaemias derived from the fetal liver and adult bone marrow restrict NOTCH1-driven autocrine insulin-like growth factor 1 (IGF1) signalling to fetal leukaemia stem cells, reducing their transplantability⁵³. Beyond the blood, human fetal neural, liver and intestinal stem cells accumulate mutations at much higher rates than those of their adult counterparts and yet are far less likely to undergo transformation^{54,55}.

Importantly, the correlation between stem cell function and cancer risk does not vary solely with development. Rather, as discussed later in this Review, we and others have shown that quiescent adult stem cells carrying oncogenic mutations rarely transform, but readily generate cancer when activated to repair in the face of tissue injury^{40,56}. Thus, in addition to developmental factors, cell extrinsic, environmental insults may impact cancer risk by changing the ground state of cells to a reparative, proliferative state.

But similar to the bad luck theory, there are caveats with observations underpinning the ground state theory. Species differences might limit the extrapolation of tumorigenesis from mice to humans, and mouse models that yield large numbers of cancers by simultaneously mutating millions of cells might not adequately recapitulate the transformation of human tissues that occurs through stochastic mutation of limited cell clones. Furthermore, the preselection of oncogenic mutations in mouse models might bias patterns of tumorigenesis. Nevertheless, the concepts and questions raised by the various theories of cancer origins provide a useful backdrop against which to debate the determinants of cancer risk and opportunities for cancer prevention.

Cell intrinsic factors

Cell identity

A fundamental element in understanding cancer origins is the identity of the cell(s) in each tissue that can undergo malignant transformation. As many leukaemias and solid tumours are hierarchically organized and sustained by a subpopulation of self-renewing cells, then stem cells – or cells that have acquired stem cell-like function, for example, in response to tissue damage – have been proposed as the origin of cancer in most tissues^{40,47,57–65} (Fig. 2). Mouse models in which oncogenic mutations have been targeted to cells in different states of differentiation provide some of the most compelling evidence that cancers arise from stem cells^{40,47,62,63,66–68}. Although some cancers can arise from committed progenitors or more differentiated cells, for example, certain leukaemias⁶⁹, it is generally agreed that cancers are propagated by populations of cells in a ‘state of stemness’^{70,71}.

But epidemiological and functional studies demonstrate that cancer is not merely the consequence of randomly mutating stem cells. Tissue-specific patterns of mutations in sporadic cancers and organ-restricted patterns of tumorigenesis in inherited cancer syndromes demonstrate that cells are not equally susceptible to transformation and that different tissues are transformed by different oncogenic mutations^{72,73}.

Arguably, the clearest examples of how cell context determines the risk of developing certain cancers are provided by childhood malignancies (Fig. 4). Childhood cancers are typically not seen in adults because they arise from progenitor cells found only in the embryo, and some cannot be modelled in mice because they arise from human-specific progenitors. For example, unique progenitor populations within the human embryonic rhombic lip likely predispose humans, but not other species, to develop certain forms of the brain tumour medulloblastoma⁷⁴. Studies in genetically modified mice have shown that anatomically, molecularly and clinically distinct subtypes of medulloblastoma and ependymoma – two relatively common forms of childhood brain tumour – arise from temporally and topographically restricted populations of neural stem and progenitor cells^{66,67,75–77}. In these studies, the introduction of cancer subtype-specific mutations into all proliferating neural stem cells in the central nervous system transformed only distinct lineages. These lineages generated tumours that recapitulated the corresponding human cancer subtype. Notably, the epigenome and transcriptome of these susceptible neural stem cells are very similar to those of their daughter tumours, suggesting their epigenetic state is conducive and permissive to transformation by the

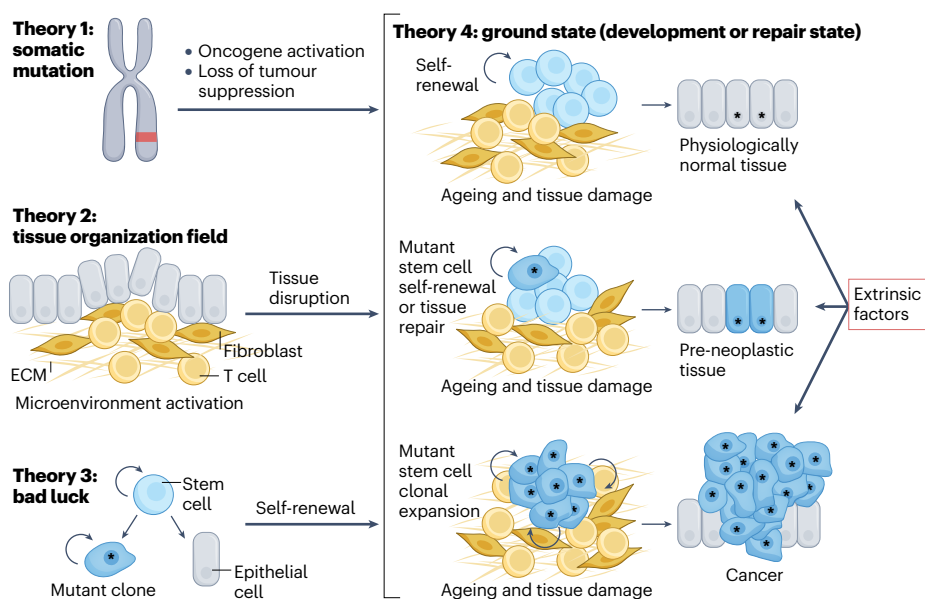


Fig. 3 | Principal theories of cancer origins. The somatic mutation theory (Theory 1) proposes that accumulating mutations in oncogenes and tumour suppressor genes transform cells, leading to unbridled proliferation and cancer. The tissue organization field theory (Theory 2) suggests that malignancies are driven at the tissue level by chronic abnormal interactions between the stroma and parenchyma. The stroma may include immune cells, extracellular matrix (ECM) and fibroblasts, among other elements. The bad luck theory (Theory 3) extends the somatic mutation theory, proposing that random mutations (marked with an asterisk) in self-renewing stem cells generate malignant,

self-renewing, daughters that propagate cancer. The ground state theory (Theory 4) unites elements of the somatic mutation, tissue organization field and bad luck theories, emphasizing the important contribution of each to determining cancer risk. Exposure of ageing tissue stem cells to extrinsic cancer risk factors (for example, radiation, ultraviolet (UV) light, alcohol and tobacco) may result in tolerance of mutations by physiologically normal mutant clones with a ground state not conducive to transformation (top), metaplasia and pre-neoplastic lesions that may not progress (middle), and full malignant transformation (bottom).

Box 1

Shared elements among the different theories of cancer origins

The various theories of cancer origins emphasize distinct elements required for tissue transformation. The body of available evidence suggests that each of these elements contributes to cancer, although their relative importance may vary with cell context and cancer type.

Cell susceptibility states

Not every cell in the body makes cancer. The existence of cell susceptibility states is evidenced by the different cancers that arise in children and adults and the non-random distribution of cancers across the body. Although these patterns are dictated, in part, by differences in exposure to risk factors, they also arise because of the 'ground state' of a particular cell, in a particular place, at a particular time that renders it susceptible to cancer. This concept encompasses the notion of cell plasticity in which developmental, ageing or injury factors remodel the epigenome and transcriptome of cells establishing a transformable state.

DNA abnormalities

Changes in DNA sequence that activate the function of oncogenes (for example, point mutations, amplifications and translocations)

or inactivate tumour suppressor genes (for example, nonsense mutations or deletions) have long been recognized as important for cell transformation. DNA abnormalities are important elements in all major theories of cancer origins. Historically, oncogenes and tumour suppressor genes were thought to predominantly impact fundamental aspects of the cancer phenotype such as cell proliferation and invasion. DNA abnormalities are now known to have much broader effects: remodelling the epigenome to generate a transforming, plastic cell state and cooperating with cell extrinsic factors, including tissue damage, to produce unique epigenomic states conducive to transformation.

Cell extrinsic factors

This element is central to the tissue field organization and ground state theories but emphasized less in the somatic mutation and bad luck theories. These include a plethora of factors ranging from physical mutagens to infective agents and tissue damage. A common theme includes the ability of these factors to induce metaplasia, producing a marked change in cell plasticity and increased risk of malignant transformation.

corresponding mutations^{66,67,75–77}. Unaffected lineages throughout the rest of the nervous system appeared to tolerate these mutations, giving rise to apparently normal tissues⁶⁷ (Fig. 4). In medulloblastoma, this lineage restriction can be released by deleting DEAD box protein 3, X-chromosomal (*Ddx3x*), which encodes an ATP-dependent RNA helicase that regulates rhombomere patterning in the developing hindbrain, suggesting that broadening out the permissive epigenetic state to a larger number of stem cells increases cancer risk⁷⁸. Subtypes of childhood high-grade glioma also appear to develop from stage and topographic-specific neural stem and progenitor cells^{79–83}.

Cell lineage-restricted susceptibility to cancer is not limited to childhood brain tumours but likely dictates the formation of most childhood leukaemias and solid tumours^{84–90}, as well as certain adult cancer types. For example, the *BRAF*^{V600E} mutation occurs in melanomas in which the tumour cells express a neural crest-like transcriptome, suggesting that this developmental state is competent for transformation^{91,92}. Although neural crest and melanoblast stages are readily transformed by *BRAF*^{V600E} in zebrafish and human pluripotent stem cell models, melanocytes are relatively resistant⁹³. The competency of neural crest cells and melanoblasts to transformation is dictated by stage-specific expression of the SRY-box 10 (SOX10) transcription factor and ATPase family AAA domain-containing protein 2 (ATAD2) chromatin factor that together promote the progenitor phenotype. Indeed, forced expression of ATAD2 in melanocytes renders them competent to transformation⁹³.

Together with evidence that neonatal stem cells are intrinsically resistant to cancer⁴⁰, these data underscore the identity of the cell of origin as a key determinant of cancer risk. This does not preclude the possibility that cancers result from random mutations of stem cells as suggested by the bad luck theory; however, as different stem cell

populations are transformed by different mutations, even within the same tissue, and stem cells appear to show age-related differences in their susceptibility to transformation independent of proliferation, then additional forces must be at work to determine the susceptibility of specific cells to specific mutations.

The epigenome

What are these additional forces and characteristics that dictate cell susceptibility to transforming mutations? Among cell intrinsic factors, the epigenome is a major determinant of cancer risk that is constantly remodelled in developing and ageing tissues^{94–96}. Indeed, promoter hypermethylation of developmental regulators characterizes transforming cells *in vitro*⁹⁷, and transient expression of reprogramming factors drives global changes in DNA methylation and tumorigenesis in transgenic mice⁹⁸. Remarkably, pluripotent stem cells derived from these tumours generate non-neoplastic cells when transplanted in mice⁹⁸, demonstrating that they have escaped irreversible genetic transformation and that epigenetic regulation alone might drive cancer in certain contexts.

At least two broad types of epigenetic change impact cell state and cancer susceptibility. As alluded to above, the first involves the normal remodelling of chromatin and histone marks that occurs during development and ageing. Within the embryo, specific configurations of the epigenome in temporally and topographically restricted progenitor cells prepare them to generate the diverse daughters that populate each organ in each anatomical context. But these specific epigenetic states portend a cellular pliancy that also renders them uniquely susceptible to specific mutations^{99,100}. Although we may have evolved mechanisms to suppress cancer during the intense mitotic and differentiation stress of early development, the requirement for

cellular plasticity in development may explain why cancer is rare, but not completely absent, during childhood.

Age-related remodelling of the epigenome may also contribute to the increased risk of cancer observed during ageing¹⁰¹. Changes in DNA methylation correlate strongly with chronological age in normal tissues, and there is some evidence that individuals who are 'epigenetically older' than their chronological age have an increased risk of cancer¹⁰². Epigenetic changes in ageing HSCs reinforce self-renewal and impede differentiation, establishing a genome landscape susceptible to transformation^{103,104} (Fig. 5). Similarly, studies of cancer-free breast biopsies have revealed a strong correlation between chronological age and methylation changes¹⁰⁵. Among 787 sites differentially methylated with age, many were in gene enhancer and transcription factor binding sites. Breast cancers displayed further deregulation of DNA methylation at these sites that were differentially methylated with age, rather than at alternative 'cancer-specific' sites.

A second group of changes to affect the epigenome and cancer risk includes mutations in histones and epigenetic regulators, as well as transcriptional silencing of tumour suppressors. Recurrent mutations in histones alter epigenomic patterning within gliomas, sarcomas and lymphomas, thereby disrupting fundamental DNA-templated processes including gene transcription and DNA damage repair¹⁰⁶. Mutations in epigenetic modifiers may themselves create an epigenetic state

permissive to transformation. Within the blood of ageing individuals, mutations in DNA methyltransferase 3A (*DNMT3A*), tet methylcytosine dioxygenase 2 (*TET2*) or *ASXL1*, which encodes a Polycomb group protein, lead to progressive expansions of haematopoietic clones (known as clonal haematopoiesis of indeterminate potential (CHIP)) and an increased risk of leukaemia^{107–110}. Mutations of *DNMT3A* in HSCs or progenitor cells are an early premalignant event¹¹¹ that causes CpG hypomethylation at gene-regulatory elements, upregulating genes important in mediating stemness¹⁰⁶. The subsequent accumulation of additional oncogenic mutations, facilitated by this shift in the epigenetic landscape, results in full transformation¹¹¹. Intriguingly, CHIP may also increase the risk of cancers in solid tissues, although this needs to be validated and the mechanism understood¹¹². With respect to the epigenetic silencing of tumour suppressor genes, this can include extensive regions of repressive chromatin that mimic large chromosomal deletions¹¹³. Thus, epigenetic remodelling that provides developing and ageing tissues with the plasticity needed to generate and maintain tissues may come with a price: the risk of priming these tissues for tumorigenesis.

DNA mutations: not the be-all and end-all?

Cancer has long been regarded as a disease of the genome¹¹⁴. The current body of massively parallel sequencing data derived from thousands

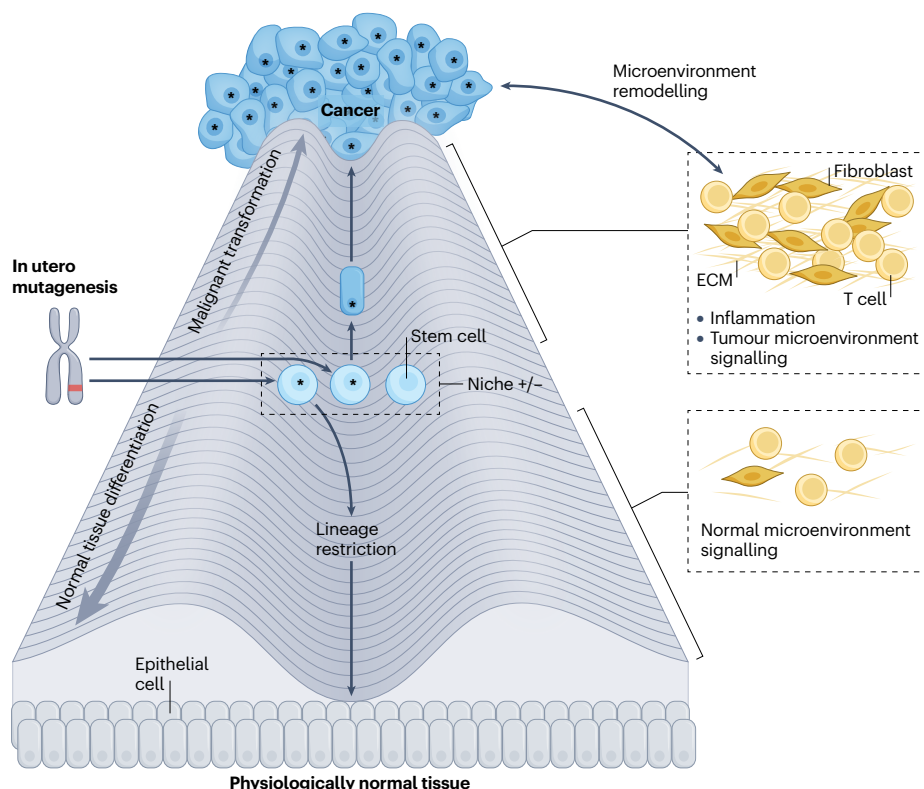


Fig. 4 | Origins of childhood cancer. Different types of tissue stem cells reside in stem cell niches that may have positive (+/-) roles in transformation. A specific somatic mutation (asterisk), typically acquired by in utero mutagenesis, is tolerated by the first stem cell type (left) that undergoes lineage-restricted differentiation to form physiologically normal tissue. As this is the natural lineage trajectory, the process is represented as a downhill path (modified from Waddington's epigenetic landscape analogy²¹¹). This process is

supported by normal microenvironmental signalling. In contrast, the ground state of the second (middle) stem cell is susceptible to cancer driven by this specific mutation, initiating transformation. As neonatal stem cells are relatively resistant to cancer, this is represented as an uphill path. Signalling between the transforming cells and microenvironment remodels and contributes to this process and the cancer phenotype. ECM, extracellular matrix.

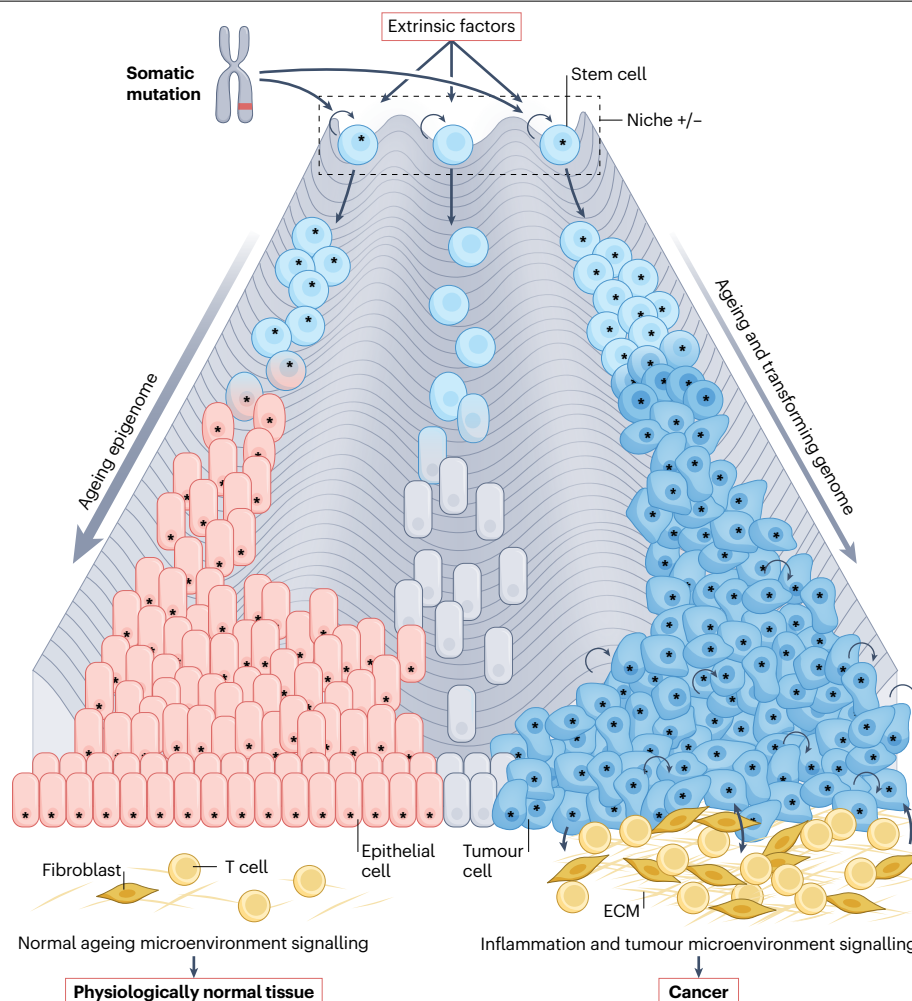


Fig. 5 | Origins of adult cancer. Different tissue stem cells, or cells that have acquired stemness characteristics through metaplasia, reside in stem cell niches that may have positive and negative (+/–) roles in transformation. A somatic mutation (asterisk), induced by extrinsic risk factors, is tolerated by the stem cell type on the left that subsequently undergoes lineage-restricted differentiation to form physiologically normal tissue. This natural lineage trajectory is represented as a downward slope (modified from Waddington's epigenetic landscape analogy)²¹¹ and might be further favoured by the ageing genome that increases tolerance of mutations to enhance the proliferative and/or repair

potential of ageing tissues. Non-mutant clones (stem cell lineage in the middle) are outcompeted by the mutant lineage (left), leading to aged tissues comprising mutant clones of physiologically normal tissue. Both lineages are supported by normal ageing microenvironmental signalling. The stem cell lineage on the right represents mutation of a susceptible stem cell, the accumulation of additional mutations and/or activation of a proliferative state that ultimately drives cancer formation. Signalling between the transforming cells and microenvironment remodels and contributes to this process and the cancer phenotype. ECM, extracellular matrix.

of human cancers has shown that tumours acquire an average of four or five 'driver' mutations and that this mutational burden increases with age. These data accord strikingly with predictions made decades ago from epidemiological studies that first inspired the somatic mutation theory of cancer and support the bad luck theory^{29,32}.

But the wealth of sequence data now available is unmasking a far more complex relationship between mutations and cancer risk. Historically, synonymous mutations – those that do not alter the protein sequence – have been thought of as mere passengers in cancer, whereas non-synonymous mutations have been regarded as 'drivers' of the disease. Indeed, large-scale CRISPR engineering studies in yeast suggest that synonymous and non-synonymous mutations similarly impact

cell fitness, although this remains to be validated more broadly^{115,116}. Furthermore, selection of synonymous mutations in oncogenes can impact RNA splicing and transcription in cancer^{117,118}. Environmental influences are thought to determine which of these mutations are propagated by yeast. If similar environmental pressures operate to select oncogenic mutations in cancer, then this complicates the view that cancer is an inevitable consequence of mutating stem cells. Indeed, human stem cell-derived small intestine, colon and liver organoids acquire mutations at very similar rates despite marked differences in cancer incidence among their host tissues¹¹⁹. Furthermore, whereas juvenile tissues are resistant to cancer relative to those in adults, fetal neural, liver and intestinal stem cells accumulate mutations at much

greater rates than their adult counterparts^{54,55} and have greater proliferative capacity⁴⁰. Thus, factors other than mutations and stem cell proliferation likely determine whether or not a cell is susceptible to transformation.

Observations that large numbers of oncogenic mutations can be tolerated by physiologically normal tissues add weight to this argument. Although the skin of the eye lid rarely forms cancer, this apparently normal tissue harbours numerous driver-mutant clones¹²⁰, as do the ageing lung, oesophagus and colon^{64,121–123} that are frequent sites of cancer. Although counterintuitive, the accumulation of such mutant clones is quite possibly a beneficial, even ‘normal’ feature of ageing epithelium (Fig. 5). The emergence of such clones in the oesophagus has been shown to have a surprising anti-tumorigenic role through the purging of early tumours by cell competition, thereby preserving tissue integrity⁶⁴. Similarly, mutations in genes such as *PKD1*, which encodes

polycystin 1, histone-lysine *N*-methyltransferase 2D (*KMT2D*) and AT-rich interactive domain-containing 1A (*ARID1A*) expand cell clones within the damaged human liver, and heterozygous deletion of these genes in mice is hepatoprotective against liver injury¹²⁴.

The choreography of cancer risk is therefore far more complex than previously appreciated: one in which the shifting landscape of developing, ageing and damaged stem cells determine their susceptibility to transforming mutations – a process compatible with the tissue organization field and ground state theories of cancer. This does not mean that the observations underpinning the bad luck theory are wrong, as the proliferative capacity of stem cells likely correlates closely with other facets of their identity. But understanding the precise characteristics of stem cells that render them susceptible to transformation, rather than attributing this merely to propagation of mutations through proliferation, is key if we are to develop effective cancer prevention strategies.

Box 2

Predicting cancer risk and more

Epidemiological research has identified chemicals and infective agents that cause cancer, leading to highly successful cancer prevention programmes. When combined with fundamental understanding of the biology of cancer origins, this knowledge can serve as the basis of systematic tools to stratify cancer risk. For example, a combination of genetic, lifestyle and imaging risk factors predicts 50% of women in the UK population with the highest risk of breast cancer, encompassing ~80% of all breast cancer cases in a 5-year period²¹². But a significant proportion of individuals at risk of cancer remain invisible to current prediction tools. The inclusion of additional biological risk factors might further improve predictive power. For example, detecting cancer-specific epigenetic profiles in accessible tissues, such as the uterine cervix or lymphocytes, or identifying such signatures in circulating cell free DNA is currently being investigated^{96,213}. Alternatively, tracking expansions of premalignant clones might provide insights into the origin and early detection of malignancy. This work is most advanced in blood. Recent studies of >200,000 UK Biobank participants has enabled the mapping of inherited predisposition to clonal haematopoiesis – the clonal expansion of a blood stem cell and its progeny driven by somatic driver mutations^{109,110}. This work has not only enabled the detection of clones years before the emergence of cancer but also the genes likely involved in this process, including those encoding regulators of DNA damage repair (poly(ADP-ribose) polymerase 1 (*PARP1*), ataxia telangiectasia mutated (*ATM*), checkpoint kinase 2 (*CHEK2*)) and haematopoietic stem cell (HSC) migration and homing (*CD164*), as well as known somatic drivers of myeloid oncogenesis (SET-binding protein 1 (*SETBP1*)). Remarkably, genetic predisposition to clonal haematopoiesis is not only associated with an increased risk of leukaemia but also of solid cancers and even non-malignant disorders such as atrial fibrillation¹¹⁰. Thus, rather than merely focusing on cancer risk, future epidemiological and biology-based strategies might consider disease risk more holistically, guiding patients through health pathways that seek to prevent and intervene in constellations of disease for which they are at particular risk.

Cell extrinsic factors

Extrinsic cancer risk factors – agents originating outside cells that can increase their risk of malignant transformation – have been recognized for more than 260 years¹²⁵. Understanding how these agents increase cancer risk is important as this knowledge is central to cancer prevention (Box 2). Publication of the bad luck theory prompted vigorous debate because it raised concerns that it would deprioritize research of extrinsic factors and cancer prevention^{126–131}. Subsequent studies published to redress the balance of debate suggested that intrinsic risk factors, including stem cell proliferation, contribute less than a third of lifetime risk to cancer development¹³². Protagonists on both sides of this argument have since published a consensus statement, agreeing that both cell intrinsic and cell extrinsic factors are important considerations in any comprehensive cancer prevention and treatment programme¹³³. But transformational progress in cancer prevention will require not only an understanding of what intrinsic and extrinsic factors increase cancer risk but precisely how these interact in each tissue, developmental and ageing context to generate cancer.

Tumour microenvironment

The immediate surroundings of cells profoundly impact their behaviour. This is well illustrated by the niches that protect and regulate stem cells¹³⁴. As stem cells, or cells that have acquired self-renewal properties, are the likely cell of origin of many cancers, these specialized niches are likely important regulators of cancer risk¹³⁵ (Figs. 4 and 5). Similar to normal neural stem cells, malignant stem cells in brain tumours occupy perivascular niches that regulate their function and are important for their survival^{136,137}. Removal of these niches can inhibit tumour growth directly¹³⁶, whereas their retention protects brain cancer stem cells during treatment, allowing them to propagate disease relapse following radiotherapy¹³⁸. Given the widespread distribution of these niches throughout tumours, it is likely that cancers can create these self-sustaining microenvironments. Evidence of this can be seen in mouse models of squamous cell carcinoma in which an interleukin-33 (IL-33)–transforming growth factor- β (TGF β) feedback loop between stem-like tumour initiating cells (TICs) and macrophages creates a niche crucial for tumour progression¹³⁹.

Although corruptible, niches might also suppress stem cell transformation. The balance of collagens, proteoglycans and glycoproteins in the extracellular matrices (ECMs) that line stem cell niches constrains their transformation^{140–143}. Indeed, communication between tumour stroma and malignant cells can remodel the ECM, dictating whether

Glossary

Apolipoprotein B mRNA-editing enzyme catalytic polypeptide

(APOBEC). An enzyme that edits mRNA species by deaminating cytosine to uracil.

Barrett oesophagus

A precursor condition for oesophageal cancer in which there is an abnormal (metaplastic) change in the mucosal cells lining the lower portion of the oesophagus, from stratified squamous epithelium to simple columnar epithelium.

Blastocysts

Clusters of dividing cells made by a fertilized egg, comprising the early stage of an embryo.

Developmental regulators

Genes that play an important role in the control of normal tissue development.

Dysplasia

The presence of abnormal cells within a tissue that may represent the precursor of malignant change.

Ependymoma

The third most common brain tumour of children arising from radial glia throughout the neural axis.

Internal tandem duplication

Duplication of sections of DNA adjacent to the original sequence.

Medulloblastoma

The most common malignant brain tumour to affect children, arising in the hindbrain from progenitor cells of the upper or lower rhombic lips.

Melanoblast

A neural crest-derived precursor cell of melanocytes, the cells that make pigment in the skin.

Metaplasia

The emergence of new cell types or disproportionate numbers of normal cell types.

Reprogramming factors

Transcription factors including OCT3 and OCT4, SOX2, MYC and KLF4 that can convert a differentiated somatic cell state into a pluripotent embryonic-like state.

Rhombomere

A transiently divided segment of the developing neural tube within the hindbrain.

Telomerase reverse transcriptase

(TERT). Part of a distinct subgroup of RNA-dependent polymerases that lengthen telomeres (the ends of DNA strands).

cancers progress^{144,145} (Figs. 4 and 5). In mice with breast cancer, malignant mammary epithelial cells and breast fibroblasts interact through the formation of a PTEN–ETS2 signalling axis that suppresses breast cancer through the extensive remodelling of the ECM. Loss of *PTEN* from these stromal fibroblasts accelerates tumour initiation and progression in a manner dependent on *ETS2* expression within tumour cells^{144,145}. The embryonic ECM has also been suggested to suppress cancer¹⁴⁶. Thus, the relative resistance of embryonic and neonatal stem cells to cancer likely involves a complex interaction between cell extrinsic and intrinsic properties⁴⁰.

Immune cells that survey and remove sick and infected cells from tissues are also important modulators of cancer risk¹⁴⁷. This notion is supported by the increased incidence of cancer in patients who are immunosuppressed¹⁴⁸; the infiltration of aggressive cancers with specific immune cell subsets¹⁴⁹; the more efficient development of tumours in mice with deficient CD8⁺ cytotoxic, CD4⁺ T helper 1 (T_H1)

and/or natural killer cells^{150,151}; and the success of therapies that enable cancer cell killing by the immune system^{13,14}. Immune cells are thought to survey tissues constantly, recognizing and removing ‘non-self’ mutant cancer cells¹⁴⁷. Although cancers can escape this surveillance by evading immune recognition^{152,153} and/or developing an immune-tolerant microenvironment^{154,155}, a question of relevance to cancer origins is whether stem cells are peculiarly susceptible, or resistant, to this surveillance. Elegant systems that measure antigen-dependent interactions between T cells and tissue stem cells are beginning to provide answers to this question¹⁵⁶. One such study has shown that whereas intestinal, ovarian and mammary adult stem cells are eliminated by activated T cells, quiescent stem cells in other tissues resist T cell killing¹⁵⁷. This appears to be an intrinsic property of quiescent stem cells that downregulate antigen presenting machinery – a property that is reversed when stem cells re-enter the cell cycle.

Communication between transforming cells and their microenvironment is therefore likely to modulate the capacity of epigenetically primed and mutated stem cells to generate cancer. This includes complex relationships with niche environments that cancers may corrupt and/or create, as well as interactions with the host immune system.

Infections and the microbiome

Microorganisms that invade tissues have long been recognized as important extrinsic determinants of cancer risk^{158–161}. The bacterium *Helicobacter pylori* is the most common infection-related cause of cancer¹⁶¹; the next four most frequent are viruses¹⁶⁰, including human papilloma virus (HPV), hepatitis B virus (HBV), hepatitis C virus and Epstein–Barr Virus. Similar to other cancer risk factors, these infections can create an epigenome permissive to transformation, create a genome instability that leads to oncogenic mutations or remodel the microenvironment to a state conducive to cancer formation.

HBV promotes hepatocellular carcinogenesis by inducing host genome instability and epigenetic remodelling following viral integration; activating cancer-related signalling pathways; and remodelling the immune microenvironment by inducing chronic inflammation¹⁶². Epstein–Barr Virus, the first isolated human tumour virus¹⁶³, remodels the host cell genome, methylating and downregulating tumour suppressor genes¹⁶⁴. HPV encodes various proteins, notably E6 and E7, that degrade, or interfere with the function of, tumour suppressor proteins¹⁶⁵.

Next-generation sequencing of human cancers has provided further understanding of the mechanisms underpinning viral-mediated transformation. Telomerase reverse transcriptase (TERT) – an established driver of carcinogenesis – is frequently upregulated by integration of HBV at the gene’s promoter site^{160,166}, whereas HPV-integrated cancers are characterized by apolipoprotein B mRNA-editing enzyme catalytic polypeptide (APOBEC)-associated mutations¹⁶⁰. As APOBEC changes viral genome sequences as a cellular defence against viruses¹⁶⁷, its activation following viral integration might introduce cancer-causing mutations within the host tissue genome.

In addition to bacteria and viruses that cause infectious disease, there is increasing evidence that commensal microorganisms – collectively termed the microbiota – influence cancer risk¹⁶⁸. This has been demonstrated most convincingly with the intestinal microflora. For example, *Fusobacterium nucleatum* promotes colorectal cancer by direct binding of cancer cells through the bacterial adhesin FadA, which in turn leads to upregulation of β -catenin signalling and promotion of a pro-inflammatory microenvironment^{169,170}.

When considering cancer origins, it is important to determine whether all cells in a tissue, or just rare subpopulations such as stem cells, are susceptible to commensal and/or infection-mediated transformation. Evidence from HPV-associated cancer suggests that viral-mediated transformation might be cell selective. Although HPV can infect the entire genital mucosa, malignant transformation occurs most commonly at the junction of the columnar endocervix and the squamous ectocervix¹⁷¹. This region comprises two types of specialized epithelial cells with stem-like properties, which regenerate the endocervix and ectocervix. Furthermore, HPV transformation is far less common at the transformation zones of the vulva, vagina and anus that comprise differentiated and multilayered epithelia¹⁷². Thus, the self-renewal capacity and residence within an immune-privileged niche may contribute to the susceptibility of these cells to transformation¹⁷³.

Mutagens

Massive parallel sequencing has not only identified which genes are mutated in cancer and how often but also enabled the segregation of these into >40 specific signatures likely caused by distinct mutagenic processes^{174–176}. These signatures include signatures of single-base substitutions associated with exposure to chemotherapies, ultraviolet (UV) light, occupational carcinogens or endogenous enzymatic mutagenesis, for example, via the DNA cytidine deaminase APOBEC3 family. These signatures may be used to predict potential causative carcinogens in specific cancer types.

Evidence suggests that two of these signatures, referred to as ‘clock-like’, accumulate steadily throughout life from the fertilized egg to the cancer cell¹⁷⁶. The inevitability of such mutations accords with the bad luck theory of cancer origins. Indeed, some of these mutations predominate in cancers derived from highly proliferative epithelia, for example, the stomach and colorectum¹⁷⁶. But ‘clock-like’ mutations are not inevitably propagated by lifelong stem cell proliferation. Their incidence varies markedly among cancer types in a manner that does not always correlate with lifelong proliferative capacity. Indeed, two different embryonal tumours of the developing nervous system are among those with the highest (neuroblastoma) and lowest (medulloblastoma) incidence of these mutations¹⁷⁶. Thus, at least in some contexts, alternative, proliferation-independent mechanisms are likely to underpin the generation of these mutations.

Notably, the impact of mutagens on cancer risk can also be modified by the ground state of the cell. Alcohol is a known carcinogen that increases the risk of several human cancer types^{177–179}. The alcohol-derived metabolite acetaldehyde causes DNA double-stranded breaks and chromosome rearrangements in HSCs in mice¹⁸⁰. Acetaldehyde-damaged HSCs are repaired by the Fanconi anaemia cross-link and non-homologous end-joining DNA repair pathways and removed by the p53 response pathway. Deletion of *Trp53* rescues HSC defects and increases the pool of mutant HSCs. Thus, intrinsic properties of stem cells including genome stability, DNA repair and cell death pathways might modify the ultimate impact of extrinsic factors on cancer risk.

The convergence of cancer risk factors

Given the close similarities between cancer and the physiological states that have evolved to maintain and repair ageing tissues, it is not surprising that one in two of us eventually develop some form of malignancy¹⁸¹. Whether a particular cell, in a particular place, at a particular time departs from its normal lineage to produce malignant tissue is likely to be determined by the convergence of context-specific cell intrinsic and

extrinsic risk factors. This process is enabled by permissive epigenetic, plastic cell states that have evolved to support normal development and ageing. This susceptibility is likely characterized by existing or acquired self-renewal – the process by which stem cells divide to make more stem cells, ensuring that their population is maintained or expanded for long-term clonal growth^{70,71,182}. Remarkably diverse but predictable patterns of DNA mutations, acquired through enzymatic, infective, chemical or physical mutagens, hardwire and corrupt self-renewal capacity. Remodelling of communication between cells evolving towards a malignant state and their immediate microenvironment progresses the tumour.

Although it is helpful to consider cancer determinants as either cell intrinsic or cell extrinsic, exploiting this knowledge to diagnose and intervene early in the disease process will require understanding of how these factors interact to determine cancer risk.

The interface of intrinsic and extrinsic risk factors

Berenblum and Shubik¹⁸³ first suggested 75 years ago that cancers are formed through a carcinogen-driven initiation phase that is followed by an irritant-driven promotion phase when latent tumour cells are provoked to proliferate. Research conducted over the following decades has highlighted the importance of tissue damage and inflammation in cancer risk¹⁸⁴. This encompasses overt injury associated with chemical or infective agents as well as the subtle wear and tear associated with ageing^{185–189}.

Metaplasia is a key feature of the tissue damage response¹⁹⁰. Although metaplasia can take different forms in different tissues, it is characterized by a marked change in cell plasticity. Lineage-restricted progenitors in hair follicles that do not normally produce skin epithelial cells can repair epithelial lineages following extensive skin damage¹⁹¹; and differentiated secretory cells of the lung de-differentiate to replace damaged airway basal stem cells¹⁹². Metaplasia can even involve the emergence of new cell types not seen in the normal tissue, as observed in Barrett oesophagus¹⁹⁰. Similar to cell plasticity associated with development and ageing, metaplasia is important for repairing damaged tissues through the expansion of cell populations with stem cell-like properties; but it carries the risk of an increased likelihood of transformation¹⁹⁰.

Studies of genetically engineered mice support this hypothesis. Cooperation between oncogenic mutations in *Kras* and tissue injury remodel the pancreatic epigenome, producing thousands of chromatin accessibility changes not caused by mutant *Kras* or injury alone¹⁹³. Interestingly, this process also involves IL-33 that is a key element in the generation of TIC niches in squamous cell carcinoma discussed above¹³⁹. Thus, oncogenic mutations and tissue injury can together remodel chromatin to promote neoplasia-specific transcriptional programmes. Similarly, adult liver stem cells carrying oncogenic mutations are quiescent and resistant to cancer, but are activated following tissue damage, increasing cancer risk 40-fold (ref. 40); and air pollutants promote the proliferation of latent clones of *Kras* or epithelial growth factor receptor (*Egfr*)-mutant lung epithelia to generate adenocarcinomas⁵⁶.

Changes in the microenvironment may also contribute to this perfect storm of cancer risk factors. For example, changes in the physical stiffness of tissues that may result from cycles of damage and repair activate Yes-associated protein (YAP) and transcriptional co-activator with PDZ-binding motif (TAZ) target genes to increase cell susceptibility to transformation¹⁹⁴. In the lung, tissue damage appears to increase cancer risk only in animals with a competent immune system⁵⁶.

This may well be mediated by macrophage-derived secretion of IL-1 β (refs. 56,195). Thus, whereas the immune system plays a key role in cancer surveillance by removing mutant cells with malignant potential¹⁴⁷, it might also promote cancer when responding to tissue damage and inflammation.

In many tissues, damage activation of adult stem cells is associated with a partial reversion to an ancestral, immature state^{40,56,196–199}. This seems logical as embryonic-like stem cells might regenerate tissues optimally. But the reparative capacity and cancer-suppressing properties of activated adult stem cells is limited relative to their juvenile counterparts^{40,200,201}. Thus, comparisons of embryonic and damage-activated adult stem cell transcriptomes might unmask novel cancer-suppressing processes.

This concept is not unique to cancer. Although neonatal mammals can readily regenerate heart muscle, this capacity is lost in adults²⁰². Similar to the cancer suppression programmes that may operate in neonates and are lost in adults, injured adult heart reactivates a fetal-like programme that produces injury-induced hypertrophy but not myocardial regeneration²⁰². Full reactivation of cardiac regeneration in the injured adult heart could transform the treatment of myocardial infarction. Thus, interdisciplinary collaborations that seek to reactivate embryonic regenerative programmes safely could transform medicine, enabling restoration of damaged adult tissues to a fully functioning, disease-free state.

Metastasis

Most cancer-related deaths are not caused by the primary tumour but are the consequence of metastatic spread^{203,204}. Thus, if one considers cancer risk in terms of its threat to human health, then the risk lies not merely in the transformation of tissues but in its propensity to metastasize. Understanding and preventing this process would markedly reduce the risk posed by cancer.

Similar to the cell susceptibility states required for tumour initiation, circulating tumour cells capable of seeding metastases are thought to be stem-like^{203,205,206}. This plasticity has focused largely on studies of epithelial-to-mesenchymal transition (EMT) and mesenchymal-to-epithelial transition (MET) that occur when tumour cells leave the primary cancer, enter the bloodstream, travel to distant sites and re-establish a tumour mass²⁰⁷. Although EMT is also a feature of gastrulation in the embryo²⁰⁸, in the context of cancer this process is thought to be wholly abnormal and a pathological consequence of transformation.

However, increasing evidence suggests that the metastatic cascade can be divorced from upstream tumorigenesis^{209,210}. Initial studies demonstrated that untransformed mouse mammary epithelial cells injected into mice seed morphologically normal microcolonies in the lung²⁰⁹. Activation of inducible oncogenes in these cells then results in the formation of metastases. Subsequently, we have identified the sodium leak channel non-selective protein (NALCN) as a key regulator of cancer metastasis and non-malignant cell dissemination²¹⁰. Deletion of *Nalcn* from malignant or normal epithelia in mice equally mobilized epithelial stem cells into the blood: these cells trafficked to distant organs to make metastatic cancer or apparently normal tissues, respectively. We propose that this process occurs throughout life to supply reparative stem cells to distant damaged organs. Therefore, metastasis might be regarded as an otherwise normal process that is hijacked, rather than ‘created’, by cancer. If this mechanism is validated in human cancer, then these findings present an even more complex picture of cancer origins in

which tissue damage, stem cell activation and mutations conspire to promote tumorigenesis and metastasis.

Conclusions

Preventing cancer, or treating it before it becomes incurable, will require a full understanding of the intrinsic and extrinsic factors that increase the risk and spread of malignancy. This understanding could transform cancer from the uniformly feared disease it is today into one that is manageable and not life-limiting.

A unifying element in the pathophysiology of cancer may prove to be cell plasticity and stem cell self-renewal machinery that enable cancers to propagate malignant clones unchecked, providing fertile ground for discovering new early cancer diagnostic and intervention strategies. Emerging evidence that ageing epithelia comprise ever expanding numbers of mutant stem cells, perhaps to retain tissue fitness, indicates that our view of oncogenic mutations needs to be far more sophisticated than that currently held. It must recognize and understand how ageing organs ‘walk a tightrope’, constantly balancing the need to maintain declining tissues by adjusting their genomes towards a pro-repair state, while avoiding the risk of overstepping into transformation. Evidence that juvenile stem cells are intrinsically resistant to cancer relative to their adult counterparts might provide a navigable route through this complexity, as direct comparison of these states could unmask novel cancer suppression mechanisms that have evolved to protect immature developing tissues. If these can be safely resurrected in ageing stem cells, then the impact on cancer prevention could be profound and highly effective.

Published online: 24 July 2023

References

- Centers for Disease Control. *Leading Causes of Death 1900–1998* https://www.cdc.gov/nchs/data/dvs/lead1900_98.pdf (2000).
- Office for National Statistics. *Causes of Death Over 100 Years* <https://www.ons.gov.uk/peoplepopulationandcommunity/birthsdeathsandmarriages/deaths/articles/causesofdeathover100years/2017-09-18> (2017).
- Institute of Medicine (US) Committee for the Study of the Future of Public Health. *The Future of Public Health* (National Academies Press (US), 1988).
- Koch, R. Die aetologie der milzbrand-krankheit, begründet auf die entwicklungsgeschichte des bacillus anthracis [German]. *Beitr. Z. Biol. Pflanz.* **1**, 277–308 (1876).
- Smith, P. W., Watkins, K. & Hewlett, A. Infection control through the ages. *Am. J. Infect. Control.* **40**, 35–42 (2012).
- Ehrlich, P. & Berthelm, A. Über das salzsaure 3.3'-Diamino-4.4'-dioxo-arsenobenzol und seine nächsten Verwandten [German]. *Ber. der Dtsch. chemischen Ges.* **45**, 756–766 (1912).
- Fleming, A. On the antibacterial action of cultures of a penicillium, with special reference to their use in the isolation of *B. influenzae*. *Br. J. Exp. Pathol.* **10**, 226–236 (1929).
- Plotkin, S. History of vaccination. *Proc. Natl Acad. Sci. USA* **111**, 12283–12287 (2014).
- Voysey, M. et al. Safety and efficacy of the ChAdOx1-nCoV-19 vaccine (AZD1222) against SARS-CoV-2: an interim analysis of four randomised controlled trials in Brazil, South Africa, and the UK. *Lancet* **397**, 99–111 (2021).
- King, M. L. & Sullivan, M. M. The similarity of the effect of podophyllin and colchicine and their use in the treatment of condylomata acuminata. *Science* **104**, 244–245 (1946).
- Pappenheimer, A. M. & Vance, M. The effects of intravenous injections of dichloroethylsulfide in rabbits, with special reference to its leucotoxic action. *J. Exp. Med.* **31**, 71–94 (1920).
- Sawyers, C. Targeted cancer therapy. *Nature* **432**, 294–297 (2004).
- Cable, J. et al. Frontiers in cancer immunotherapy — a symposium report. *Ann. N. Y. Acad. Sci.* **1489**, 30–47 (2021).
- Waldman, A. D., Fritz, J. M. & Lenardo, M. J. A guide to cancer immunotherapy: from T cell basic science to clinical practice. *Nat. Rev. Immunol.* **20**, 651–668 (2020).
- Sung, H. et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* **71**, 209–249 (2021). **This work presents global statistics of cancer incidence and mortality, emphasizing the human health emergency posed by cancer and the need for earlier intervention.**
- McPhail, S. et al. Risk factors and prognostic implications of diagnosis of cancer within 30 days after an emergency hospital admission (emergency presentation): an International Cancer Benchmarking Partnership (ICBP) population-based study. *Lancet Oncol.* **23**, 587–600 (2022).

17. Danaei, G., vander Hoorn, S., Lopez, A. D., Murray, C. J. L. & Ezzati, M. Causes of cancer in the world: comparative risk assessment of nine behavioural and environmental risk factors. *Lancet* **366**, 1784–1793 (2005).
18. National Health Services. *NHS Long Term Plan for Cancer* <https://www.longtermplan.nhs.uk/areas-of-work/cancer/> (2019).
19. WHO. WHO report on cancer: setting priorities, investing wisely and providing care for all. *World Health Organization* <https://www.who.int/publications/i/item/9789240001299> (2022).
20. Crosby, D. et al. Early detection of cancer. *Science* **375**, aay9040 (2022).
21. Fitzgerald, R. C. et al. Cytosponge-trefoil factor 3 versus usual care to identify Barrett's oesophagus in a primary care setting: a multicentre, pragmatic, randomised controlled trial. *Lancet* **396**, 333–344 (2020).
22. Gredner, T., Mons, U., Niedermaier, T., Brenner, H. & Soerjomataram, I. Impact of tobacco control policies implementation on future lung cancer incidence in Europe: an international, population-based modeling study. *Lancet Reg. Health Eur.* **4**, 100074 (2021).
23. Health and Safety Executive. *Mesothelioma Statistics for Great Britain, 2023* <https://www.hse.gov.uk/statistics/causdis/mesothelioma/mesothelioma.pdf> (2023).
24. Chang, M. H. et al. Decreased incidence of hepatocellular carcinoma in hepatitis B vaccinees: a 20-year follow-up study. *J. Natl Cancer Inst.* **101**, 1348–1355 (2009).
25. Roden, R. B. S. & Stern, P. L. Opportunities and challenges for human papillomavirus vaccination in cancer. *Nat. Rev. Cancer* **18**, 240–254 (2018).
26. GBD 2019 Cancer Risk Factors Collaborators. The global burden of cancer attributable to risk factors, 2010–19: a systematic analysis for the Global Burden of Disease Study 2019. *Lancet* **400**, 563–591 (2022).
27. Force, L. M. et al. The global burden of childhood and adolescent cancer in 2017: an analysis of the Global Burden of Disease Study 2017. *Lancet Oncol.* **20**, 1211–1225 (2019).
28. Johnston, W. T. et al. Childhood cancer: estimating regional and global incidence. *Cancer Epidemiol.* **71**, 101662 (2021).
29. Nordling, C. O. A new theory on the cancer-inducing mechanism. *Br. J. Cancer* **7**, 68–72 (1953).
30. Boveri, T. concerning the origin of malignant tumors. *J. Cell Sci.* **7**, 68–72 (1929).
31. Knudson, A. G. Mutation and cancer: statistical study of retinoblastoma. *Proc. Natl Acad. Sci. USA* **68**, 820–823 (1971).
32. Armitage, P. & Doll, R. The age distribution of cancer and a multi-stage theory of carcinogenesis. *Br. J. Cancer* **8**, 1–12 (1954).
- Together with Nordling (1953), Boveri (1929) and Knudson (1971), this study details the mutation theory of cancer in which malignant transformation is proposed to arise as a consequence of several changes in DNA that confer proliferative and invasive cell properties.**
33. Wiley Online Library. Complete pathologic maturation and regression of stage IVS neuroblastoma without treatment. *Cancer* **62**, 818–925 (1967).
34. Huggins, C. Endocrine-induced regression of cancers. *Science* **156**, 1050–1054 (1967).
35. Mintz, B. & Illmensee, K. Normal genetically mosaic mice produced from malignant teratocarcinoma cells. *Proc. Natl Acad. Sci. USA* **72**, 3585–3589 (1975).
36. Committee on Chemical Environmental Mutagens, Board on Toxicology and Environmental Health Hazards, Commission on Life Sciences & National Research Council. *Identifying and Estimating the Genetic Impact of Chemical Mutagens* <https://nap.nationalacademies.org/read/19435/chapter/1> (National Academy Press, 1983).
37. Riva, L. et al. The mutational signature profile of known and suspected human carcinogens in mice. *Nat. Genet.* **52**, 1189–1197 (2020).
38. Soto, A. M. & Sonnenschein, C. The tissue organization field theory of cancer: a testable replacement for the somatic mutation theory. *BioEssays* **33**, 332–340 (2011).
- This work details the tissue organization field theory of cancer in which cell extrinsic properties of cells are identified as key regulators of transformation.**
39. Tomasetti, C. & Vogelstein, B. Cancer etiology. Variation in cancer risk among tissues can be explained by the number of stem cell divisions. *Science* **347**, 78–81 (2015).
40. Zhu, L. et al. Multi-organ mapping of cancer risk. *Cell* **166**, 1132–1146.e7 (2016).
- This work uses Cre-conditional mouse models for lifelong lineage tracing and cancer induction studies and demonstrates that cancer in multiple organs results from the mutation of stem cells in a cancer-susceptible state that varies with age, damage and site, underpinning the ground state theory of cancer.**
41. Tomasetti, C., Li, L. & Vogelstein, B. Stem cell divisions, somatic mutations, cancer etiology, and cancer prevention. *Science* **355**, 1330–1334 (2017).
- Together with Tomasetti and Vogelstein (2015), this study describes the 'bad luck' theory of cancer in which transformation is proposed to result from the chance mutation of proliferating stem cells.**
42. Davey Smith, G., Relton, C. L. & Brennan, P. Chance, choice and cause in cancer aetiology: individual and population perspectives. *Int. J. Epidemiol.* **45**, 605–613 (2016).
43. Abascal, F. et al. Somatic mutation landscapes at single-molecule resolution. *Nature* **593**, 405–410 (2021).
44. Lopez-Bigas, N. & Gonzalez-Perez, A. Are carcinogens direct mutagens? *Nat. Genet.* **52**, 1137–1138 (2020).
45. Boshart, M. et al. A new type of papillomavirus DNA, its presence in genital cancer biopsies and in cell lines derived from cervical cancer. *EMBO J.* **3**, 1151–1157 (1984).
46. Koutsky, L. A. et al. A controlled trial of a human papillomavirus type 16 vaccine. *N. Engl. J. Med.* **347**, 1645–1651 (2002).
47. Zhu, L. et al. Prolamin 1 marks intestinal stem cells that are susceptible to neoplastic transformation. *Nature* **457**, 603–607 (2009).
48. Kretzschmar, K. & Watt, F. M. Lineage tracing. *Cell* **148**, 33–45 (2012).
49. Cairns, J. Mutation selection and the natural history of cancer. *Nature* **255**, 197–200 (1975).
50. Casás-Selves, M. & Degregori, J. How cancer shapes evolution, and how evolution shapes cancer. *Evolution* **4**, 624–634 (2011).
51. Campisi, J. Aging, tumor suppression and cancer: high wire-act! *Mech. Ageing Dev.* **126**, 51–58 (2005).
52. Porter, S. N. et al. Fetal and neonatal hematopoietic progenitors are functionally and transcriptionally resistant to Flt3-ITD mutations. *eLife* **5**, e18882 (2016).
53. Giambra, V. et al. Epigenetic restoration of fetal-like IGF1 signaling inhibits leukemia stem cell activity. *Cell Stem Cell* **23**, 714–726.e7 (2018).
54. Bae, T. et al. Different mutational rates and mechanisms in human cells at pregastrulation and neurogenesis. *Science* **359**, 550–555 (2018).
55. Kuijk, E. et al. Early divergence of mutational processes in human fetal tissues. *Sci. Adv.* **5**, eaaw1271 (2019).
56. Hill, W. et al. Lung adenocarcinoma promotion by air pollutants. *Nature* **616**, 159–167 (2023).
- This combination of human epidemiological and tissue studies as well as mouse models demonstrates that principles of the ground state theory of cancer operate in determining lung adenocarcinoma risk.**
57. Visvader, J. E. Cells of origin in cancer. *Nature* **469**, 314–322 (2011).
58. Hanahan, D. & Weinberg, R. A. Hallmarks of cancer: the next generation. *Cell* **144**, 646–674 (2011).
59. Zhang, M., Lee, A. V. & Rosen, J. M. The cellular origin and evolution of breast cancer. *Cold Spring Harb. Perspect. Med.* **7**, a027128 (2017).
60. Wang, X. et al. A luminal epithelial stem cell that is a cell of origin for prostate cancer. *Nature* **461**, 495–500 (2009).
61. Kim, C. F. B. et al. Identification of bronchioalveolar stem cells in normal lung and lung cancer. *Cell* **121**, 823–835 (2005).
62. Barker, N. et al. Crypt stem cells as the cells-of-origin of intestinal cancer. *Nature* **457**, 608–611 (2009).
63. Barker, N. et al. Lgr5⁺ stem cells drive self-renewal in the stomach and build long-lived gastric units in vitro. *Cell Stem Cell* **6**, 25–36 (2010).
64. Colom, B. et al. Mutant clones in normal epithelium outcompete and eliminate emerging tumours. *Nature* **598**, 510–514 (2021).
65. Hope, K. J., Jin, L. & Dick, J. E. Acute myeloid leukemia originates from a hierarchy of leukemic stem cell classes that differ in self-renewal capacity. *Nat. Immunol.* **5**, 738–743 (2004).
66. Johnson, R. A. et al. Cross-species genomics matches driver mutations and cell compartments to model ependymoma. *Nature* **466**, 632–636 (2010).
67. Gibson, P. et al. Subtypes of medulloblastoma have distinct developmental origins. *Nature* **468**, 1095–1099 (2010).
- Together with Johnson et al. (2010), this study demonstrates that neural lineages in the embryonic brain are susceptible to different oncogenic mutations, resulting in the development of the various subtypes of childhood brain tumours observed in the clinic.**
68. Huntly, B. J. P. et al. MOZ-TIF2, but not BCR-ABL, confers properties of leukemic stem cells to committed murine hematopoietic progenitors. *Cancer Cell* **6**, 587–596 (2004).
69. Cozzio, A. et al. Similar MLL-associated leukemias arising from self-renewing stem cells and short-lived myeloid progenitors. *Genes Dev.* **17**, 3029 (2003).
70. Kreso, A. & Dick, J. E. Evolution of the cancer stem cell model. *Cell Stem Cell* **14**, 275–291 (2014).
71. Magee, J. A., Piskounova, E. & Morrison, S. J. Cancer stem cells: impact, heterogeneity, and uncertainty. *Cancer Cell* **21**, 283–296 (2012).
72. Schneider, G., Schmidt-Suppran, M., Rad, R. & Saur, D. Tissue-specific tumorigenesis: context matters. *Nat. Rev. Cancer* **17**, 239–253 (2017).
73. Haigis, K. M., Cichowski, K. & Elledge, S. J. Tissue-specificity in cancer: the rule, not the exception. *Science* **363**, 1150–1151 (2019).
74. Hendrikse, L. D. et al. Failure of human rhombic lip differentiation underlies medulloblastoma formation. *Nature* **609**, 1021–1028 (2022).
75. Vladoiu, M. C. et al. Childhood cerebellar tumours mirror conserved fetal transcriptional programs. *Nature* **572**, 67–73 (2019).
76. Taylor, M. D. et al. Radial glia cells are candidate stem cells of ependymoma. *Cancer Cell* **8**, 323–335 (2005).
77. Hovestadt, V. et al. Resolving medulloblastoma cellular architecture by single-cell genomics. *Nature* **572**, 74–79 (2019).
78. Patmore, D. M. et al. DDX3X suppresses the susceptibility of hindbrain lineages to medulloblastoma. *Dev. Cell* **54**, 455–470.e5 (2020).
79. Filbin, M. G. et al. Developmental and oncogenic programs in H3K27M gliomas dissected by single-cell RNA-seq. *Science* **360**, 331–335 (2018).
80. Nagaraja, S. et al. Transcriptional dependencies in diffuse intrinsic pontine glioma. *Cancer Cell* **31**, 635–652.e6 (2017).
81. Pathania, M. et al. H3.3^{K27M} cooperates with Trp53 loss and PDGFRA gain in mouse embryonic neural progenitor cells to induce invasive high-grade gliomas. *Cancer Cell* **32**, 684–700.e9 (2017).
82. Funato, K., Major, T., Lewis, P. W., Allis, C. D. & Tabar, V. Use of human embryonic stem cells to model pediatric gliomas with H3.3K27M histone mutation. *Science* **346**, 1529–1533 (2014).
83. Silveira, A. B. et al. H3.3 K27M depletion increases differentiation and extends latency of diffuse intrinsic pontine glioma growth in vivo. *Acta Neuropathol.* **137**, 637–655 (2019).

84. Riggi, N. et al. EWS-FLI-1 expression triggers a Ewing's sarcoma initiation program in primary human mesenchymal stem cells. *Cancer Res.* **68**, 2176–2185 (2008).
85. Sole, A. et al. Unraveling Ewing sarcoma tumorigenesis originating from patient-derived mesenchymal stem cells. *Cancer Res.* **81**, 4994–5006 (2021).
86. Krivtsov, A. V. et al. Cell of origin determines clinically relevant subtypes of MLL-rearranged AML. *Leukemia* **27**, 852–860 (2013).
87. Custers, L. et al. Somatic mutations and single-cell transcriptomes reveal the root of malignant rhabdoid tumours. *Nat. Commun.* **12**, 1407 (2021).
88. Coorens, T. H. H. et al. Embryonal precursors of wilms tumor. *Science* **366**, 1247–1251 (2019).
89. Abarrategi, A. et al. Osteosarcoma: cells-of-origin, cancer stem cells, and targeted therapies. *Stem Cell Int.* **2016**, 3631764 (2016).
90. Jansky, S. et al. Single-cell transcriptomic analyses provide insights into the developmental origins of neuroblastoma. *Nat. Genet.* **53**, 683–693 (2021).
91. White, R. M. et al. DHODH modulates transcriptional elongation in the neural crest and melanoma. *Nature* **471**, 518–522 (2011).
92. Kaufman, C. K. et al. A zebrafish melanoma model reveals emergence of neural crest identity during melanoma initiation. *Science* **351**, aad2197 (2016).
93. Baggiolini, A. et al. Developmental chromatin programs determine oncogenic competence in melanoma. *Science* **373**, eabc1048 (2022).
94. Horvath, S. & Raj, K. DNA methylation-based biomarkers and the epigenetic clock theory of ageing. *Nat. Rev. Genet.* **19**, 371–384 (2018).
95. Booth, L. N. & Brunet, A. The aging epigenome. *Mol. Cell* **62**, 728–744 (2016).
96. Widschwendter, M. et al. Epigenome-based cancer risk prediction: rationale, opportunities and challenges. *Nat. Rev. Clin. Oncol.* **15**, 292–309 (2018).
97. Xie, W. et al. DNA methylation patterns separate senescence from transformation potential and indicate cancer risk. *Cancer Cell* **33**, 309–321.e5 (2018).
98. Ohnishi, K. et al. Premature termination of reprogramming in vivo leads to cancer development through altered epigenetic regulation. *Cell* **156**, 663–677 (2014).
- This study using mouse models shows that epigenetic reprogramming can drive tumorigenesis but this transformation is reversible, demonstrating that irreversible genetic change is not required for tumour formation.**
99. Chen, X., Pappo, A. & Dyer, M. A. Pediatric solid tumor genomics and developmental pliancy. *Oncogene* **34**, 5207–5215 (2015).
100. Gilbertson, R. J. Mapping cancer origins. *Cell* **145**, 25–29 (2011).
101. White, M. C. et al. Age and cancer risk: a potentially modifiable relationship. *Am. J. Prev. Med.* **46**, S7–S15 (2014).
102. Morales Berstein, F. et al. Assessing the causal role of epigenetic clocks in the development of multiple cancers: a Mendelian randomization study. *eLife* **11**, e75374 (2022).
103. Rodrigues, C. P., Shvedunova, M. & Akhtar, A. Epigenetic regulators as the gatekeepers of hematopoiesis. *Trends Genet.* **37**, 125–142 (2021).
104. Sun, D. et al. Epigenomic profiling of young and aged HSCs reveals concerted changes during aging that reinforce self-renewal. *Cell Stem Cell* **14**, 673–688 (2014).
105. Johnson, K. C., Houseman, E. A., King, J. E. & Christensen, B. C. Normal breast tissue DNA methylation differences at regulatory elements are associated with the cancer risk factor age. *Breast Cancer Res.* **19**, 81 (2017).
106. Nacev, B. A. et al. The expanding landscape of 'oncohistone' mutations in human cancers. *Nature* **567**, 473–478 (2019).
107. Xie, M. et al. Age-related mutations associated with clonal hematopoietic expansion and malignancies. *Nat. Med.* **20**, 1472–1478 (2014).
108. Jaiswal, S. et al. Age-related clonal hematopoiesis associated with adverse outcomes. *N. Engl. J. Med.* **371**, 2488–2498 (2014).
109. Genovese, G. et al. Clonal hematopoiesis and blood-cancer risk inferred from blood dna sequence. *N. Engl. J. Med.* **371**, 2477–2487 (2014).
110. Kar, S. P. et al. Genome-wide analyses of 200,453 individuals yield new insights into the causes and consequences of clonal hematopoiesis. *Nat. Genet.* **54**, 1155–1166 (2022).
111. Yang, L., Rau, R. & Goodell, M. A. DNMT3A in haematological malignancies. *Nat. Rev. Cancer* **15**, 152–165 (2015).
112. Reed, S. C., Croessmann, S. & Park, B. H. CHIP happens: clonal hematopoiesis of indeterminate potential and its relationship to solid tumors. *Clin. Cancer Res.* **29**, 1403–1411 (2023).
113. Frigola, J. et al. Epigenetic remodeling in colorectal cancer results in coordinate gene suppression across an entire chromosome band. *Nat. Genet.* **38**, 540–549 (2006).
114. Campbell, P. J. et al. Pan-cancer analysis of whole genomes. *Nature* **578**, 82–93 (2020).
115. Shen, X., Song, S., Li, C. & Zhang, J. Synonymous mutations in representative yeast genes are mostly strongly non-neutral. *Nature* **606**, 725–731 (2022).
116. Kruglyak, L. et al. Insufficient evidence for non-neutrality of synonymous mutations. *Nature* **616**, E8–E9 (2023).
117. Supek, F., Miñana, B., Valcárcel, J., Gabaldón, T. & Lehner, B. Synonymous mutations frequently act as driver mutations in human cancers. *Cell* **156**, 1324–1335 (2014).
118. Benisty, H., Weber, M., Hernandez-Alías, X., Schaefer, M. H. & Serrano, L. Mutation bias within oncogene families is related to proliferation-specific codon usage. *Proc. Natl Acad. Sci. USA* **117**, 30848–30856 (2020).
119. Blokzijl, F. et al. Tissue-specific mutation accumulation in human adult stem cells during life. *Nature* **538**, 260–264 (2016).
120. Martincorena, I. et al. High burden and pervasive positive selection of somatic mutations in normal human skin. *Science* **348**, 880–886 (2015).
121. Martincorena, I. et al. Somatic mutant clones colonize the human esophagus with age. *Science* **362**, 911–917 (2018).
122. Yokoyama, A. et al. Age-related remodelling of oesophageal epithelia by mutated cancer drivers. *Nature* **565**, 312–317 (2019).
123. Yizhak, K. et al. RNA sequence analysis reveals macroscopic somatic clonal expansion across normal tissues. *Science* **364**, eaaw0726 (2019).
124. Zhu, M. et al. Somatic mutations increase hepatic clonal fitness and regeneration in chronic liver disease. *Cell* **177**, 608–621.e12 (2019).
- Together with Martincorena et al. (2015), Martincorena et al. (2018), Yokoyama et al. (2019) and Yizhak et al. (2019), this study describes the accumulation of large clones harbouring potentially oncogenic mutations in otherwise apparently normal ageing human tissues.**
125. Hill, J. *Cautions Against the Immoderate Use of Snuff: Founded on the Known Qualities of the Tobacco Plant: and the Effects It Must Produce When This Way Taken into the Body: and Enforced by Instances of Persons Who Have Perished Miserably of Diseases, Occasioned, or Rendered Incurable by Its Use* <http://resource.nlm.nih.gov/2166041R> (1761).
126. Song, M. & Giovannucci, E. L. Cancer risk: accuracy of literature. *J. Natl Cancer Inst.* **66**, 1784 (1981).
127. O'callaghan, M. Cancer risk: accuracy of literature. *Science* **347**, 729 (2015).
128. Wild, C. et al. Cancer risk: role of chance overstated. *Science* **347**, 728 (2015).
129. Potter, J. D. & Prentice, R. L. Cancer risk: tumors excluded. *Science* **347**, 727 (2015).
130. Gotay, C., Dummer, T. & Spinelli, J. Cancer risk: prevention is crucial. *Science* **347**, 729 (2015).
131. Ashford, N. A. et al. Cancer risk: role of environment. *Science* **347**, 729–731 (2015).
132. Wu, S., Powers, S., Zhu, W. & Hannun, Y. A. Substantial contribution of extrinsic risk factors to cancer development. *Nature* **529**, 43–47 (2016).
133. Song, M., Vogelstein, B., Giovannucci, E. L., Willett, W. C. & Tomasetti, C. Cancer prevention: molecular and epidemiologic consensus. *Science* **361**, 1317–1318 (2018).
134. Jones, D. L. & Wagers, A. J. No place like home: anatomy and function of the stem cell niche. *Nat. Rev. Mol. Cell Biol.* **9**, 11–21 (2008).
135. Plaks, V., Kong, N. & Werb, Z. The cancer stem cell niche: how essential is the niche in regulating stemness of tumor cells? *Cell Stem Cell* **16**, 225–238 (2015).
136. Calabrese, C. et al. A perivascular niche for brain tumor stem cells. *Cancer Cell* **11**, 69–82 (2007).
137. Gilbertson, R. J. & Rich, J. N. Making a tumour's bed: glioblastoma stem cells and the vascular niche. *Nat. Rev. Cancer* **7**, 733–736 (2007).
138. Hambardzumyan, D. et al. PI3K pathway regulates survival of cancer stem cells residing in the perivascular niche following radiation in medulloblastoma in vivo. *Genes Dev.* **22**, 436–446 (2008).
139. Taniguchi, S. et al. Tumor-initiating cells establish an IL-33-TGF- β ; niche signaling loop to promote cancer progression. *Science* **369**, eaay1813 (2020).
140. Winkler, J., Abisoye-Ogunniyan, A., Metcalf, K. J. & Werb, Z. Concepts of extracellular matrix remodelling in tumour progression and metastasis. *Nat. Commun.* **11**, 5120 (2020).
141. Bissell, M. J. & Hines, W. C. Why don't we get more cancer? A proposed role of the microenvironment in restraining cancer progression. *Nat. Med.* **17**, 320–329 (2011).
142. Watt, F. M. & Huck, W. T. S. Role of the extracellular matrix in regulating stem cell fate. *Nat. Rev. Mol. Cell Biol.* **14**, 467–473 (2013).
143. Frantz, C., Stewart, K. M. & Weaver, V. M. The extracellular matrix at a glance. *J. Cell Sci.* **123**, 4195–4200 (2010).
144. Jones, C. E. et al. Stromal PTEN regulates extracellular matrix organization in the mammary gland. *Neoplasia* **21**, 132–145 (2019).
145. Trimboli, A. J. et al. Pten in stromal fibroblasts suppresses mammary epithelial tumours. *Nature* **461**, 1084–1091 (2009).
146. Stoker, A. W., Hatier, C. & Bissell, M. J. The embryonic environment strongly attenuates v-src oncogenesis in mesenchymal and epithelial tissues, but not in endothelia. *J. Cell Biol.* **111**, 217–228 (1990).
147. Gonzalez, H., Hagerling, C. & Werb, Z. Roles of the immune system in cancer: from tumor initiation to metastatic progression. *Genes. Dev.* **32**, 1267–1284 (2018).
148. Saluzzo, S. et al. Delayed antiretroviral therapy in HIV-infected individuals leads to irreversible depletion of skin- and mucosa-resident memory T cells. *Immunity* **54**, 2842–2858.e5 (2021).
149. Gentles, A. J. et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat. Med.* **21**, 938–945 (2015).
150. Teng, M. W. L., Swann, J. B., Koebel, C. M., Schreiber, R. D. & Smyth, M. J. Immune-mediated dormancy: an equilibrium with cancer. *J. Leukoc. Biol.* **84**, 988–993 (2008).
151. Kim, H.-J. & Cantor, H. CD4 T-cell subsets and tumor immunity: the helpful and the not-so-helpful. *Cancer Immunol. Res.* **2**, 91–98 (2014).
152. McGranahan, N. et al. Allele-specific HLA loss and immune escape in lung cancer evolution. *Cell* **171**, 1259–1271.e11 (2017).
153. Chowell, D. et al. Patient HLA class I genotype influences cancer response to checkpoint blockade immunotherapy. *Science* **359**, 582–587 (2018).
154. Mantovani, A., Marchesi, F., Malesci, A., Laghi, L. & Allavena, P. Tumour-associated macrophages as treatment targets in oncology. *Nat. Rev. Clin. Oncol.* **14**, 399–416 (2017).
155. Böttcher, J. P. et al. NK cells stimulate recruitment of cDC1 into the tumor microenvironment promoting cancer immune control. *Cell* **172**, 1022–1037.e14 (2018).
156. Agudo, J. et al. GFP-specific CD8 T cells enable targeted cell depletion and visualization of T-cell interactions. *Nat. Biotechnol.* **33**, 1287–1292 (2015).
157. Agudo, J. et al. Quiescent tissue stem cells evade immune surveillance. *Immunity* **48**, 271–285.e5 (2018).

158. Parkin, D. M. The global health burden of infection-associated cancers in the year 2002. *Int. J. Cancer* **118**, 3030–3044 (2006).
159. Plummer, M. et al. Global burden of cancers attributable to infections in 2012: a synthetic analysis. *Lancet Glob. Health* **4**, e609–e616 (2016).
160. Zapata, M. et al. The landscape of viral associations in human cancers. *Nat. Genet.* **52**, 320–330 (2020).
161. Thrift, A. P., Wenker, T. N. & El-Serag, H. B. Global burden of gastric cancer: epidemiological trends, risk factors, screening and prevention. *Nat. Rev. Clin. Oncol.* **20**, 338–349 (2023).
162. Jiang, Y., Han, Q., Zhao, H. & Zhang, J. The mechanisms of HBV-induced hepatocellular carcinoma. *J. Hepatocell. Carcinoma* **8**, 435 (2021).
163. Abhik, S. & S. R. E. Mechanisms of B-cell oncogenesis induced by Epstein–Barr virus. *J. Virol.* **93**, e00238-19 (2019).
164. Saha, A., Jha, H. C., Upadhyay, S. K. & Robertson, E. S. Epigenetic silencing of tumor suppressor genes during in vitro Epstein–Barr virus infection. *Proc. Natl Acad. Sci. USA* **112**, E5199–E5207 (2015).
165. Martinez-Zapien, D. et al. Structure of the E6/E6AP/p53 complex required for HPV-mediated degradation of p53. *Nature* **529**, 541–545 (2016).
166. Sung, W.-K. et al. Genome-wide survey of recurrent HBV integration in hepatocellular carcinoma. *Nat. Genet.* **44**, 765–769 (2012).
167. Roberts, S. A. et al. An APOBEC cytidine deaminase mutagenesis pattern is widespread in human cancers. *Nat. Genet.* **45**, 970–976 (2013).
168. Elinav, E., Garrett, W. S., Trinchieri, G. & Wargo, J. The cancer microbiome. *Nat. Rev. Cancer* **19**, 371–376 (2019).
169. Rubinstein, M. R. et al. *Fusobacterium nucleatum* promotes colorectal carcinogenesis by modulating E-cadherin/β-catenin signaling via its FadA adhesin. *Cell Host Microbe* **14**, 195–206 (2013).
170. Kostic, A. D. et al. *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. *Cell Host Microbe* **14**, 207–215 (2013).
171. Schiffman, M. et al. Carcinogenic human papillomavirus infection. *Nat. Rev. Dis. Prim.* **2**, 16086 (2016).
172. Yang, E. J. et al. Microanatomy of the cervical and anorectal squamocolumnar junctions: a proposed model for anatomical differences in HPV-related cancer risk. *Mod. Pathol.* **28**, 994–1000 (2015).
173. Budhwani, M., Lukowski, S. W., Porceddu, S. V., Frazer, I. H. & Chandra, J. Dysregulation of stemness pathways in hpv mediated cervical malignant transformation identifies potential oncotherapy targets. *Front. Cell Infect. Microbiol.* **10**, 307 (2020).
174. Alexandrov, L. B. et al. Signatures of mutational processes in human cancer. *Nature* **500**, 415–421 (2013).
175. Burns, M. B., Temiz, N. A. & Harris, R. S. Evidence for APOBEC3B mutagenesis in multiple human cancers. *Nat. Genet.* **45**, 977–983 (2013).
176. Alexandrov, L. B. et al. Clock-like mutational processes in human somatic cells. *Nat. Genet.* **47**, 1402–1407 (2015).
177. Roswall, N. & Weiderpass, E. Alcohol as a risk factor for cancer: existing evidence in a global perspective. *J. Prev. Med. Public Health* **48**, 1–9 (2015).
178. Bagardi, V. et al. Alcohol consumption and site-specific cancer risk: a comprehensive dose–response meta-analysis. *Br. J. Cancer* **112**, 580–593 (2015).
179. Rumgay, H. et al. Global burden of cancer in 2020 attributable to alcohol consumption: a population-based study. *Lancet Oncol.* **22**, 1071–1080 (2021).
180. Garaycoechea, J. I. et al. Alcohol and endogenous aldehydes damage chromosomes and mutate stem cells. *Nature* **553**, 171–177 (2018).
181. Ahmad, A. S., Ormiston-Smith, N. & Sasieni, P. D. Trends in the lifetime risk of developing cancer in Great Britain: comparison of risk for those born from 1930 to 1960. *Br. J. Cancer* **112**, 943–947 (2015).
182. van Velthoven, C. T. J. & Rando, T. A. Stem cell quiescence: dynamism, restraint, and cellular idling. *Cell Stem Cell* **24**, 213–225 (2019).
183. Berenblum, I. & Shubik, P. A new, quantitative, approach to the study of the stages of chemical carcinogenesis in the mouse's skin. *Br. J. Cancer* **1**, 383–391 (1947).
184. Coussens, L. M. & Werb, Z. Inflammation and cancer. *Nature* **420**, 860–867 (2002).
185. Walter, D. et al. Exit from dormancy provokes DNA-damage-induced attrition in haematopoietic stem cells. *Nature* **520**, 549–552 (2015).
186. Insinga, A. et al. DNA damage in stem cells activates p21, inhibits p53, and induces symmetric self-renewing divisions. *Proc. Natl Acad. Sci. USA* **110**, 3931–3936 (2013).
187. Behrens, A., van Deursen, J. M., Rudolph, K. L. & Schumacher, B. Impact of genomic damage and ageing on stem cell function. *Nat. Cell Biol.* **16**, 201–207 (2014).
188. Tian, H. et al. A reserve stem cell population in small intestine renders Lgr5-positive cells dispensable. *Nature* **478**, 255–259 (2011).
189. Amcheslavsky, A., Jiang, J. & Ip, Y. T. Tissue damage-induced intestinal stem cell division in *Drosophila*. *Cell Stem Cell* **4**, 49–61 (2009).
190. Lin, B. et al. Modulating cell fate as a therapeutic strategy. *Cell Stem Cell* **23**, 329–341 (2018).
191. Ito, M. et al. Stem cells in the hair follicle bulge contribute to wound repair but not to homeostasis of the epidermis. *Nat. Med.* **11**, 1351–1354 (2005).
192. Tata, P. R. et al. Dedifferentiation of committed epithelial cells into stem cells in vivo. *Nature* **503**, 218–223 (2013).
193. Alonso-Curbelo, D. et al. A gene–environment-induced epigenetic program initiates tumorigenesis. *Nature* **590**, 642–648 (2021).
194. Panciera, T. et al. Reprogramming normal cells into tumour precursors requires ECM stiffness and oncogene-mediated changes of cell mechanical properties. *Nat. Mater.* **19**, 797–806 (2020).
195. Hiraiwa, K. & van Eeden, S. F. Contribution of lung macrophages to the inflammatory responses induced by exposure to air pollutants. *Mediators Inflamm.* **2013**, 619523 (2013).
196. Poplawski, G. H. D. et al. Injured adult neurons regress to an embryonic transcriptional growth state. *Nature* **581**, 77–82 (2020).
197. Fazilat, H. et al. Tracing colonic embryonic transcriptional profiles and their reactivation upon intestinal damage. *Cell Rep.* **36**, 109484 (2021).
198. Vercauteren Drubbel, A. et al. Reactivation of the Hedgehog pathway in esophageal progenitors turns on an embryonic-like program to initiate columnar metaplasia. *Cell Stem Cell* **28**, 1411–1427.e7 (2021).
199. Miao, Q. et al. SOX11 and SOX4 drive the reactivation of an embryonic gene program during murine wound repair. *Nat. Commun.* **10**, 4042 (2019).
200. Gurtner, G. C., Werner, S., Barrandon, Y. & Longaker, M. T. Wound repair and regeneration. *Nature* **453**, 314–321 (2008).
201. Rolfe, K. J. & Grobbelaar, A. O. A review of fetal scarless healing. *ISRN Dermatol.* **2012**, 698034 (2012).
202. Sadek, H. & Olson, E. N. Toward the goal of human heart regeneration. *Cell Stem Cell* **26**, 7–16 (2020).
203. Ganesh, K. & Massagué, J. Targeting metastatic cancer. *Nat. Med.* **27**, 34–44 (2021).
204. Diliekås, H., Rogers, M. S. & Straume, O. Are 90% of deaths from cancer caused by metastases? *Cancer Med.* **8**, 5574–5576 (2019).
205. Massagué, J. & Ganesh, K. Metastasis-initiating cells and ecosystems. *Cancer Discov.* **11**, 971–994 (2021).
206. Massagué, J. & Obenauf, A. C. Metastatic colonization by circulating tumour cells. *Nature* **529**, 298–306 (2016).
207. Yang, J. et al. Guidelines and definitions for research on epithelial–mesenchymal transition. *Nat. Rev. Mol. Cell Biol.* **21**, 341–352 (2020).
208. Nájera, G. S. & Weijer, C. J. The evolution of gastrulation morphologies. *Development* **150**, dev200885 (2023).
209. Podsypanina, K. et al. Seeding and propagation of untransformed mouse mammary cells in the lung. *Science* **321**, 1841–1844 (2008).
210. Rahrmann, E. P. et al. The NALCN channel regulates metastasis and nonmalignant cell dissemination. *Nat. Genet.* **54**, 1827–1838 (2022).

Metastasis is the single biggest risk factor for cancer death: this study identifies NALCN as a regulator of cell shedding from solid tissues independent of cancer, divorcing this process from tumorigenesis and unmasking a potential new target for antimetastatic therapies.

211. Waddington, C. H. *The Strategy of the Genes: A Discussion of Some Aspects of Theoretical Biology* (Routledge, 2014).
212. Lee, A. et al. BOADICEA: a comprehensive breast cancer risk prediction model incorporating genetic and nongenetic risk factors. *Genet. Med.* **21**, 1708–1718 (2019).
213. Jamshidi, A. et al. Evaluation of cell-free DNA approaches for multi-cancer early detection. *Cancer Cell* **40**, 1537–1549.e12 (2022).

Acknowledgements

The authors are grateful to B. Simons for helpful discussions during writing of the manuscript. R.J.G. is supported by Major Centre Core and Children's Brain Tumour Centre of Excellence grants from Cancer Research UK, The Brain Tumour Charity, and P01CA96832 and U54CA243125 from the US National Cancer Institute.

Author contributions

All authors researched data for the article and contributed substantially to discussion of the content. R.J.G. and A.J. wrote the article. All authors reviewed and/or edited the manuscript before submission.

Competing interests

The authors declare no competing interests.

Additional information

Peer review information *Nature Reviews Cancer* thanks the anonymous reviewers for their contribution to the peer review of this work.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

Related links

World Cancer Research Fund: <https://www.wcrf.org/cancer-trends/worldwide-cancer-data/>

© Springer Nature Limited 2023