Primer Preparation?

Participants: Brianna Branson, Rori Hoover, Patrick Jiang

Date: Thursday, July 13, 2023

Protocol:

1. Resuspended G2.1 plasmid in 266 µL **??**
2. Resuspended G2.2 plasmid in 252 µL **??**
3. Resuspended G3.1 plasmid in 213 µL **??**

Results: N/A

Conclusion: N/A

What are you doing with the primers? Please explain exactly why you needed to resuspend them and where the plasmids were stored in the meantime

Remove His Tags from G2 using

G4 is separate

10 mM Hexanoic Acid Stock Preparation

Participants: Pranav Bhavaraju

Date: Thursday, July 13, 2023

Protocol:

1. Added 10 mL DI water to a 15 mL test tube
2. Added 12.53 µL hexanoic acid to the tube in a fume hood

Results: N/A

Conclusion: N/A

Storage?

Native hexanoic acid is toxic to E. coli, so this will kill everything – need to saponify it

Plating Metabolic Construct and Wild-Type Transformed E. coli

Participants: Pranav Bhavaraju

Date: Thursday, July 13, 2023

Protocol:

1. Warmed 2 M9 plates in a 37 °C incubator **(when did you prepare the M9 plates? I only saw preparation of the solution mentioned in the notebook?)**
2. Pipetted 18.79 µL pure hexanoic acid onto each plate
3. Spread solution across plate using 3 glass beads for each one
4. Serially streaked G4.4 transformed E. coli onto one plate using a sterilized streaking tool
5. Serially streaked wild-type E. coli onto the second plate using a sterilized streaking tool
6. Incubated at 37 °C overnight

Results: ??

Conclusion: ??

G4.C for metabolic construct

M9 plates are

ON plates, there are yeast, sugar, and other heavy metals

M9 has no LB, no yeast, no carbon source – just the essential salts and plating with hexanoic acid to