G3 Plasmid Digestion

Participants: Dr. Benjamin Bartelle (mentor)

Date: **Saturday, July 15, 2023**

Protocol:

1. **Added 0.5 µL DpnI to each PCR tube**
2. **Incubated samples for 1 hour at room temperature**

Results: N/A

Conclusion: N/A

DpnI destroys methylated DNA – should not eat synthesized DNA from PCR

Transformation of E. coli with G3

Participants: Dr. Benjamin Bartelle (mentor)

Date: Saturday, July 15, 2023

Protocol:

1. Thawed **what kind? NEB** E. coli cells on ice for 10 minutes
2. Added 25 µL E. coli cells to a microfuge tube
3. Added 2 µL G3 to the tube
4. Flicked tube to mix and briefly centrifuged the samples down
5. Placed tubes on ice for 30 minutes
6. Heat shocked E. coli in a 42 °C water bath for 30 seconds
7. Placed tubes on ice for 5 minutes
8. Added 475 µL outgrowth medium to each tube
9. Incubated E. coli at 37 °C and shook at 400 rpm for 1 hour

Results: N/A

Conclusion: N/A

Was there a control? Idk if this protocol is correct. If it's BL21, it was done differently by Bella and Brianna

Plating G3 Transformed E. coli

Participants: Dr. Benjamin Bartelle (mentor)

Date: Saturday, July 15, 2023

Protocol:

1. Warmed an LB-agar kanamycin plate in a 37 °C incubator
2. Serially streaked G3 transformed E. coli onto the plate using a sterilized streaking tool
3. Incubated at 37 °C over the weekend

Results: ??

Conclusion: Growth on the plate