## E. coli G2-D DNA Miniprep

Participants: Brianna Branson, Isabella (Bella) Lirtzman

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## **Protocol:**

Note: these steps follow those outlined in the Monarch® Plasmid DNA Miniprep Kit Protocol (NEB #T1010) General Guidelines by New England Biolabs.

- 1. Transferred 5 mL of the liquid culture into a microfuge tube
- 2. Centrifuged at 13,000 rpm for 30 seconds and discarded supernatant
- 3. Resuspended pelleted cells in 200 µL Plasmid Resuspension Buffer
- 4. Added 200  $\mu$ L Plasmid Lysis Buffer to each tube and gently inverted the tubes until the solution turned dark pink
- 5. Incubated cells on the bench for 1 minute
- 6. Added  $400~\mu L$  Neutralization Buffer to each tube and inverted the tubes until the solution turned yellow and a precipitate formed
- 7. Incubated cells on the bench for 2 minutes
- 8. Centrifuged tube at 13,000 rpm for 5 minutes
- 9. Inserted a spin column into a collection tube and transferred the supernatant into the column
- 10. Centrifuged the tube at 13,000 rpm for 1 minute and discarded flow-through
- 11. Added 200  $\mu$ L Plasmid Wash Buffer 1 to each column and centrifuged all of the columns at 13,000 rpm for 1 minute
- 12. Added 400  $\mu$ L Plasmid Wash Buffer 2 to each column and centrifuged all of the columns at 13,000 rpm for 1 minute
- 13. Transferred each column to a clean 1.5 mL microfuge tube
- 14. Added 30 µL Elution Buffer to each tube
- 15. Waited 1 minute before centrifuging all of the tubes at 13,000 rpm for 1 minute

**Results:** N/A

**Conclusion:** N/A