

Intermediate course: Meta-analysis of data from animal studies in R

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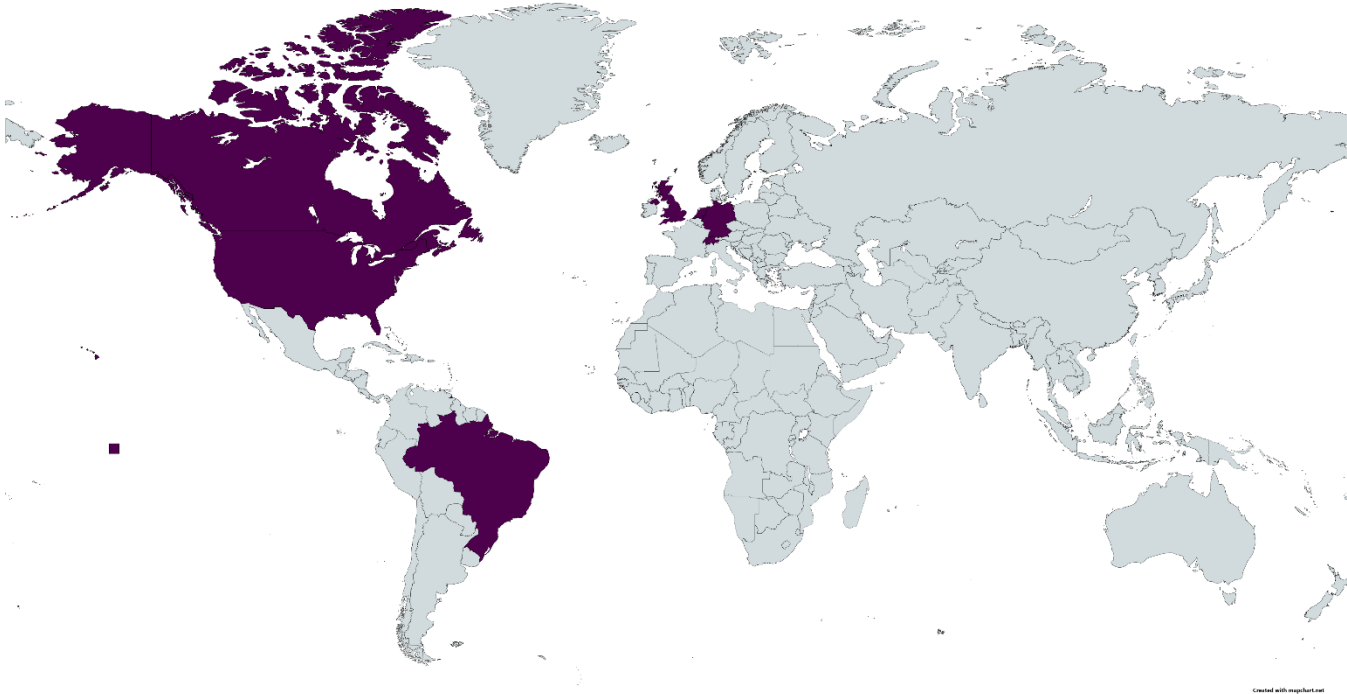
BIH Quest Center

Welcome!

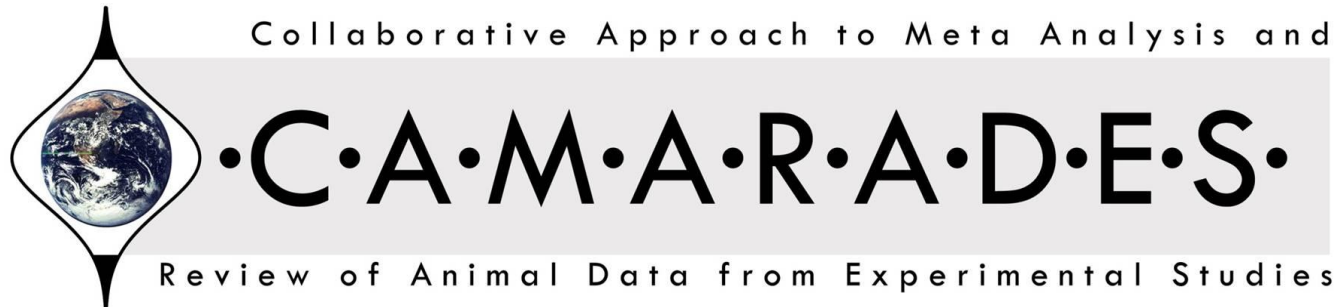


CAMARADES

Collaborative Approach to Meta-Analysis and Review of Animal Data from Experimental Studies



1. Preclinical meta-research
2. Supporting framework for researchers involved in systematic review and meta-analysis



CAMARADES Berlin - Resources

One-to-one methodological advice

Support for your SR project from research question to meta-analysis
Weekly **drop-in session**: Monday 12-13pm CET

Contact the Helpdesk: [**CAMARADES.berlin@charite.de**](mailto:CAMARADES.berlin@charite.de)

Preclinical Systematic Review Software

Free to use online, unlimited projects and reviewers.
[**http://SyRF.org.uk/**](http://SyRF.org.uk/)

Preclinical Systematic Review wiki website

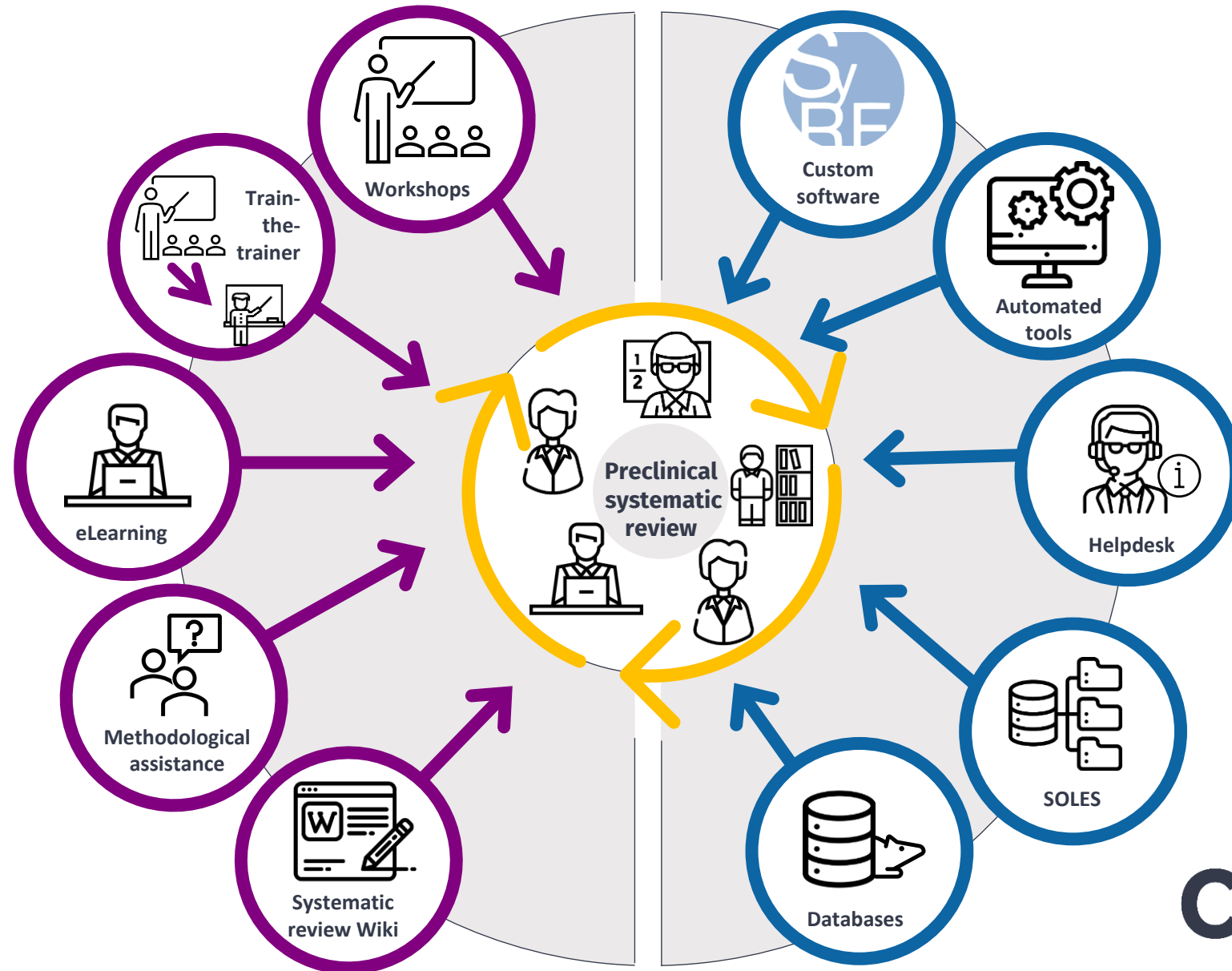
Guide to conducting all steps of a preclinical systematic review.
Information and instructions, links, resources, and further reading.
[**https://www.CAMARADES.de**](https://www.CAMARADES.de)

Communities for Open Research Synthesis

Community

Education &
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Infrastructure .



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www.cores-hub.io



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Regular updates on education formats, talks, and project developments

<https://www.linkedin.com/company/communities-for-open-research-synthesis/>



Newsletter

Sign-up to receive updates on the community news, events, workshops, and resources!

<http://eepurl.com/h4hsMv>



mailchimp



Course structure

1st Day

- *Introduction*
- *Which kind of data? Clarifying data types, outcomes and effect sizes*
- *Data extraction from primary animal studies*

BREAK

- *Calculating effect sizes manually and in `metafor`*
- *Run meta-analysis in R - understanding default settings*

BREAK

- *Plotting: Default forest plots & basic customizations*
- *Day 1 wrap-up & Q&A*

Course structure

2nd Day

- *Heterogeneity in meta-analysis of animal studies*
- *Meta regression:*
 - *Regression model & Explaining heterogeneity*

BREAK

- *Running models in R + Plotting*
- *Obstacles*

BREAK

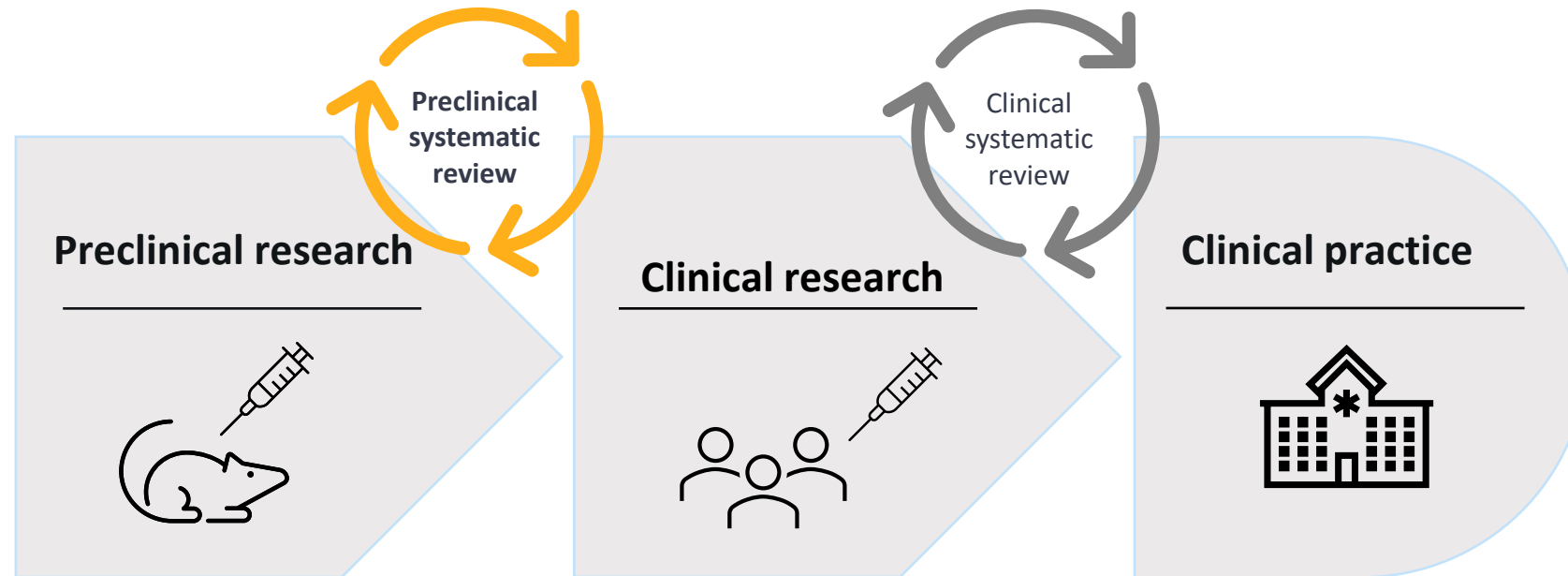
- *Exploring the impacts of Risks of Bias in R*
- *Funnel plotting*
- *Workshop Conclusion & Wrap-up*

Prior knowledge?

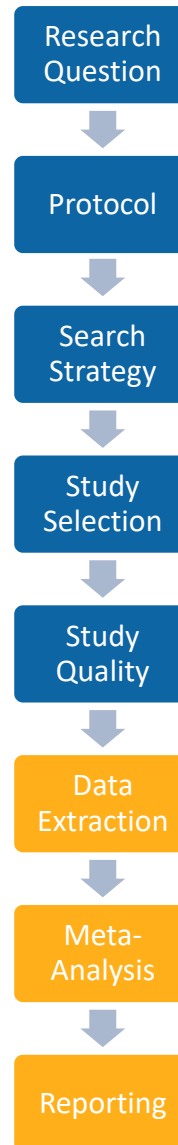
- R
 - Rstudio
 - Markdown
-
- Plenty of info on metafor's capabilities:
<https://www.metafor-project.org/doku.php/>

Systematic review recap

- Systematic review is not routinely used to guide decisions in preclinical research
- Evidence is not synthesized efficiently



Systematic review recap



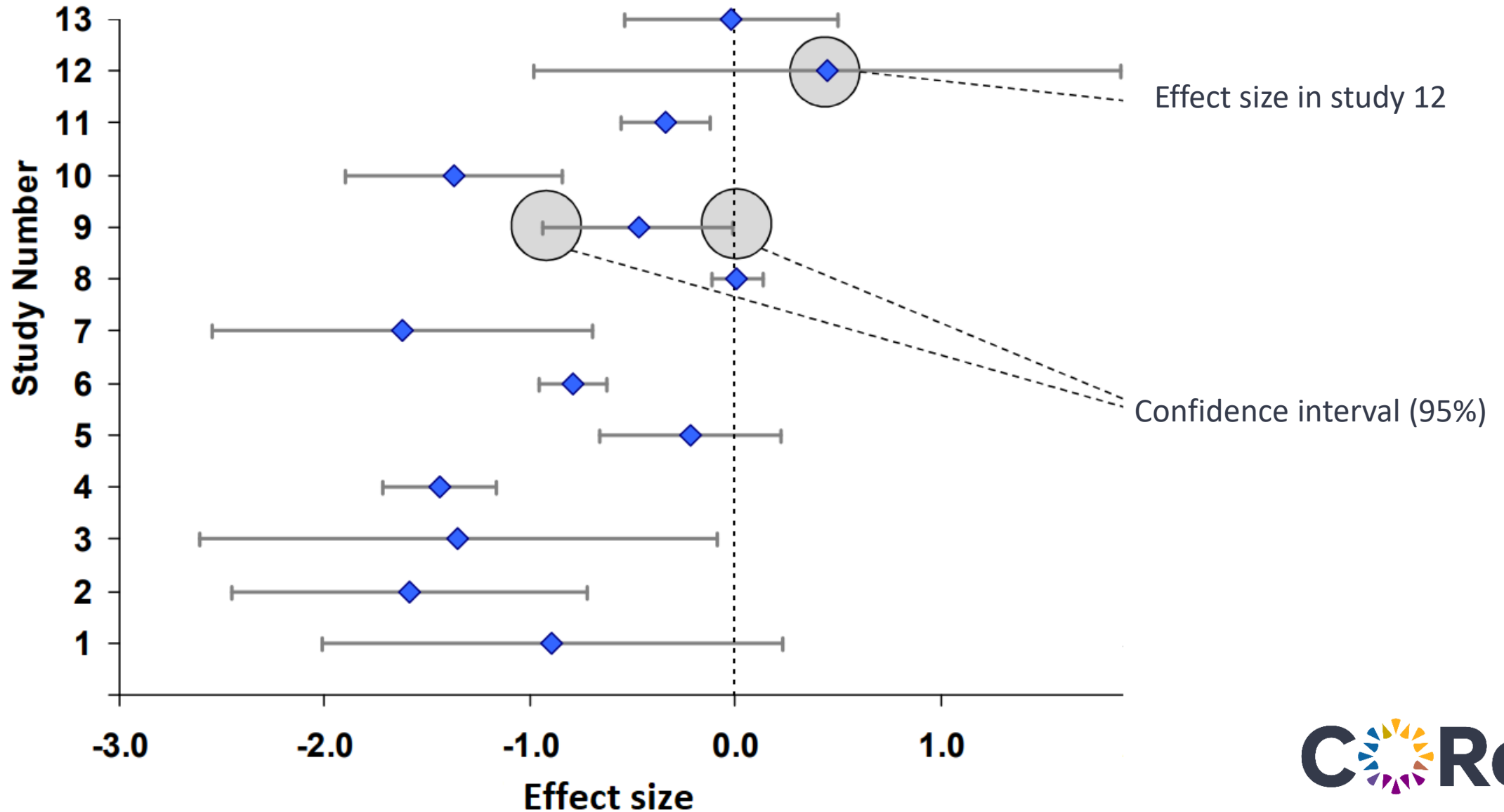
Systematic review recap

- Clarifying research question
- Search terms
- Quantitative analysis?
- Inclusion / exclusion criteria | PRISMA
- Protocol --> Prospero
- Stack of papers – what next?

1

Data types and effect measures

Meta-analysis



Meta-analysis

For quantitatively aggregating across many studies, we need:

- The same measure of effect for every study
- Uncertainty in the effect measure (standard error; 95%-CI; variance)

Data type

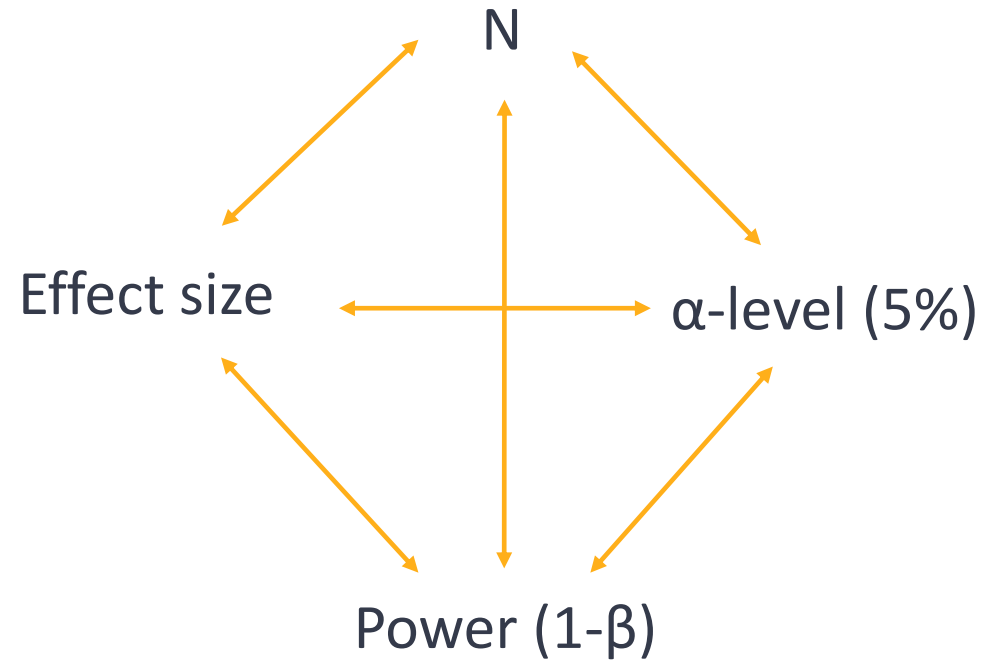
Scale:

- Continuous measures (time, size,)
- Binary / categorical outcomes (death, drop-out, therapy response, ...)
- Time-to-event (survival, onset of disease, ...)

Measure of:

- Difference (groups)
- Association

Effect size



The higher each of these, the likelier a „significant“ result

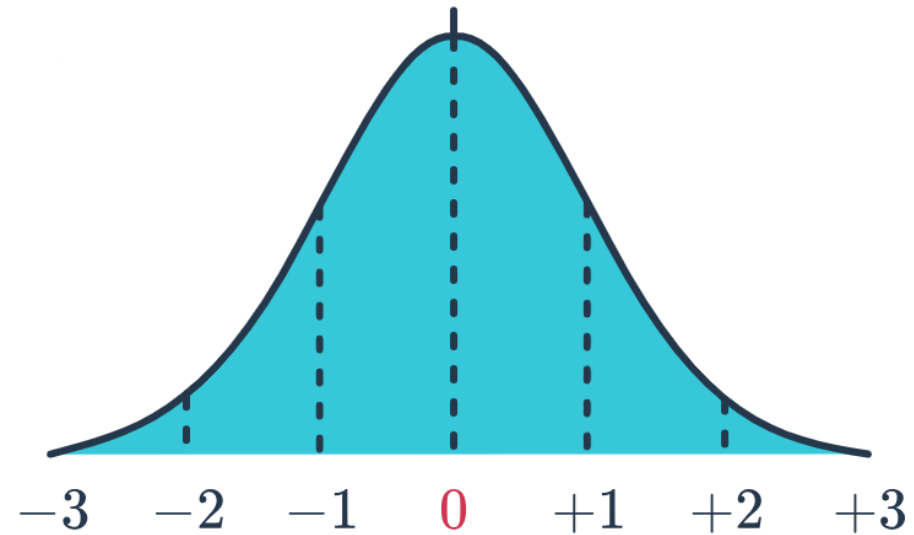
Raw mean difference

- The outcome is reported on a meaningful scale and all studies in the analysis use the same scale
- Meta-analysis is performed directly on the raw difference in means
- E.g. blood pressure – measured in (or converted to) mmHg
- difference = 0 -> no effect

Standardized mean difference (SMD)

- Aka Hedge's g (or Cohen's d, without correction for small samples)
- Logic: Normalizing mean differences with a standard deviation
- SMD = 0 -> no effect

$$\frac{\bar{x}_1 - \bar{x}_2}{s}$$



Standardized mean difference (SMD)

Advantages:

- Applicable to all continuous variables
- Same metric/size for any outcome (cave!)

Disadvantages:

- Abstract metric
- Clinical / biological relevance of effect not self-evident

value	Interpretation
$\leq .2$	“small” difference
$\approx .5$	“medium” difference
$\geq .8$	“large” difference

Standardized mean difference (SMD)

$$ES_i = \frac{\bar{x}_c - \bar{x}_{rx}}{S_{\text{pooled}}} \times \left(1 - \frac{3}{4N - 9}\right)$$

$$S_{\text{pooled}} = \sqrt{\frac{(n'_c - 1)SD_c^2 + (n_{rx} - 1)SD_{rx}^2}{N - 2}}$$

$$VAR = \frac{N}{n_{rx} \times n'_c} + \frac{ES_i^2}{2N}$$

Normalized mean difference

- Can be used when outcomes are on a ratio scale, **where the score on a 'control' or 'sham' animal is known.**
- Advantages: Original unit of measurement is retained (eg tumor or infarct volume mm³)

Normalized mean difference

$$ES_i = 100 \times \frac{(\bar{x}_c - \bar{x}_{\text{sham}}) - (\bar{x}_{\text{rx}} - \bar{x}_{\text{sham}})}{\bar{x}_c - \bar{x}_{\text{sham}}}$$

$$SD_{c^*} = 100 \times \frac{SD_c}{\bar{x}_c - \bar{x}_{\text{sham}}}$$

$$SD_{\text{rx}^*} = 100 \times \frac{SD_{\text{rx}}}{\bar{x}_c - \bar{x}_{\text{sham}}}$$

$$VAR = \frac{SD_{c^*}^2}{n'_c} + \frac{SD_{\text{rx}^*}^2}{n_{\text{rx}^*}}$$

Odds ratio

- Difference / association between to binary variables
- How do the odds for O differ between groups G?

Advantages:

- Invariant when reorganizing cross tab
- (directly related to logistic regression)

Disadvantages:

- Abstract metric
- Odds \neq risk

	O1	O2	
G1	a	b	n_1
G2	c	d	n_2

Odds ratio

$$OR_i = \frac{a_i \times d_i}{b_i \times c_i}$$

- $OR = 1$ -> no effect. $OR = 0.5$ corresponds to $OR = 2$
- For aggregating, $\ln(OR)$ is used (0 -> no effect, $\ln(OR=0.5) = -0.69$, $\ln(OR=2) = 0.69$)

$$VAR = (1/a_i) + (1/b_i) + (1/c_i) + (1/d_i)$$

Further effect measures

mean difference	$\bar{x}_1 - \bar{x}_2$	$\mu_1 - \mu_2$	$s_p^2(1/n_1 + 1/n_2)$
response ratio	$\ln[\bar{x}_1 / \bar{x}_2]$	$\ln[\mu_1 / \mu_2]$	$\frac{s_1^2}{n_1 \bar{x}_1^2} + \frac{s_2^2}{n_2 \bar{x}_2^2}$
risk difference	$p_1 - p_2$	$\pi_1 - \pi_2$	$p_1(1-p_1)/n_1 + p_2(1-p_2)/n_2$
risk ratio	$\ln[p_1 / p_2]$	$\ln[\pi_1 / \pi_2]$	$\frac{1}{p_1 n_1} - \frac{1}{n_1} + \frac{1}{p_2 n_2} - \frac{1}{n_2}$
correlation coefficient	r	ρ	$\frac{(1-r^2)^2}{n-1}$
transformed correlation	$\frac{1}{2} \ln \left[\frac{1+r}{1-r} \right]$	$\frac{1}{2} \ln \left[\frac{1+\rho}{1-\rho} \right]$	$1/(n-3)$
proportion	p	π	$p(1-p)/n$
log odds	$\ln[p/(1-p)]$	$\ln[\pi/(1-\pi)]$	$1/[np(1-p)]$
mean	\bar{x}	μ	s^2 / n

2

**Introduction to our working example:
Centrofobic rats (in pain)**

Centrofobic rats in pain

PLOS ONE

RESEARCH ARTICLE

A systematic review and meta-analysis of thigmotactic behaviour in the open field test in rodent models associated with persistent pain

<https://journals.plos.org/plosone/article?id=10.1371/journal.pone.0290382>

Zhang XY, Diaz-delCastillo M, Kong L, Daniels N, MacIntosh-Smith W, Abdallah A, Domanski D, Sofrenovic D, Yeung TP, Valiente D, Vollert J. A systematic review and meta-analysis of thigmotactic behaviour in the open field test in rodent models associated with persistent pain. Plos one. 2023 Sep 8;18(9):e0290382.

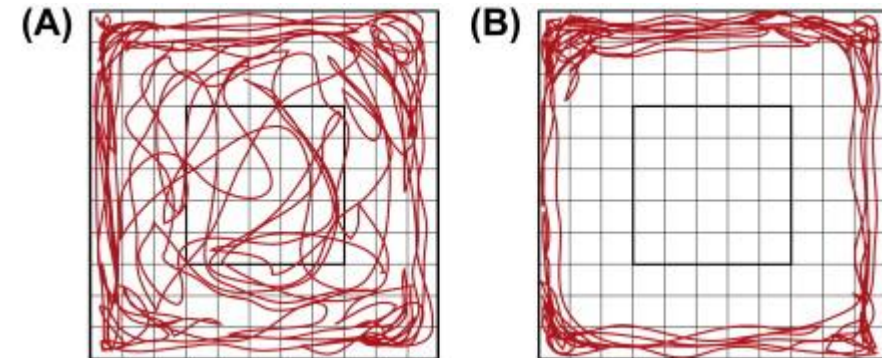
Centrofobic rats in pain

Failure of translating preclinical findings on new analgesic drugs to the clinic¹

What is a good model for pain? What is a good outcome measure?

- Limb withdrawal
- **Open field test** (-> thigmotaxis)

-> Stress? Anxiety? Pain?



<https://www.sciencedirect.com/topics/biochemistry-genetics-and-molecular-biology/open-field-test>

¹ Borsook D, Hargreaves R, Bountra C, Porreca F. Lost but making progress—Where will new analgesic drugs come from? Science Translational Medicine. 2014; 6(249):249sr3–sr3. <https://doi.org/10.1126/scitranslmed.3008320>

Is thigmotaxis a valid measure of pain?

“This systematic review aimed to 1) assess whether thigmotaxis can be affected by injury and disease models associated with **persistent pain** and analgesic drug treatments in rodents;”

- (Spinal) Nerve injury
- Head injury
- Chemotherapy
- Migraine models
-



Comparing
to sham

Our example

Does the model work? Is pain reflected in the open field test?

We will focus on:

- Rats
- Exp. vs sham control
- Thigmotaxis as measured by time in center / forays to center
- -> Fig 2 in paper by Zhang et al.

Data

Raw data: <https://osf.io/rmt97/>

Prepared data set:

Data Zhang Fig 2.xlsx - Excel																									
Datei Start Einfügen Seitenlayout Formeln Daten Überprüfen Ansicht Hilfe Was möchten Sie tun?																									
Einfügen Ausschneiden Kopieren Format übertragen Zwischenablage																									
Calibri 11 A A Textumbruch Standard Bedingte Formatierung Als Tabelle																									
F K U % 000 0,00 % Standard Gut Neutral Schlecht Ausgabe Berechnung Eingabe Erklärender ...																									
Einfügen Löschen Format Zellen																									
AutoSumme Ausfüllen Löschen Sortieren Filtern Bearbeiten																									
Q14 1,469																									
	A	B	C	D	E	F	G	H	I	J	K	L	M	N	O	P	Q	R	S	T	U	V	W	X	
1	StudyId	Author	year	CohortId	Experiment	Units	Average	Time	Average	SD	Model	Label	Outcome	Label	Cohort	Label	Mean	Se	Mc	Sc	Ne	Nc	N.total	Model	Label
2	053a1a1f-	Saffarpour et al., 20	2020	624deb4c-	91e15487-s	Mean	6	0	0,375	2,674843	CCI	Time in centre	Model		0,375	2,674843	10,409	2,493581	6	6	12	Chronic cc	Neuropat	Rat	
3	0767bd07-	Missig et al., 2017	2017	448d5488-	e3dc6732-Number	Mean	5	0	3,71	1,565248	CCI	Entries to the centre	CCI + Vehi		3,71	1,565248	7,5	1,565248	5	1,666667	6,666667	Chronic cc	Neuropat	Rat	
4	0cd5ef20-	Amorim et al., 2014	2014	33cdc3c4-	9830fe43-%	Mean	6	0	6,397	2,334364	Arthritis	Time in centre	Model		6,397	2,334364	10,782	4,242516	6	6	12	Kaolin and	Arthropat	Rat	
5	0cef4a1c-	Bree et al., 2020	2020	c5c74ecc-	b5131d79-s	Mean	8	3	156,867	64,86149	CHI 450 g	Time in centre	Model (45		156,867	64,86149	522,161	273,5768	8	8	16	Closed he	Neuropat	Rat	
6	0cef4a1c-	Bree et al., 2020	2020	504ec31e-	244de311-s	Mean	8	3	318,188	165,907	CHI 150 g	Time in centre	Model (15		318,188	165,907	490,177	317,3788	8	10	18	Closed he	Neuropat	Rat	
7	0cef4a1c-	Bree et al., 2020	2020	66867a95-	2d0aefc2-s	Mean	8	3	343,631	189,7592	CHI 150 g repeat	Time in centre	Model (15		343,631	189,7592	521,363	207,1512	8	8	16	Closed he	Neuropat	Rat	
8	11dcb928-	Gerasimova et al., 2	2021	9193fb2c-	9e0012e1-s	Mean	15	0	2,6	1,549193	hHCY (hyperhon	Time in centre	Model						15	15	30	hHCY (hyp	Hyperhon	Rat	
9	18f7151c-	Huang et al., 2013	2013	cf44eb2f-	8d738b65-s	Mean	12	0	3,218	2,857884	d4T	Time in centre	Model		3,218	2,857884	11,38	5,909757	12	12	24	Antiretrov	Neuropat	Rat	
10	21ab4cd6-	Yan et al., 2017	2017	2109961e-	be65beb3-Percentag	Mean	8	0	2,677	1,291	Saline;nLPS	Time in centre	nLPS-aSal		2,677	1,291	3,697	2,159	8	2,333333	10,33333	Saline (m	Inflamma	Rat	
11	21ab4cd6-	Yan et al., 2017	2017	473d09fb-	f3406a62-Percentag	Mean	8	0	2,832	1,586	Saline;nLPS	Time in centre	nLPS-aSal		2,832	1,586	1,469	1,215	8	2,333333	10,33333	Saline (m	Inflamma	Rat	
12	21ab4cd6-	Yan et al., 2017	2017	8d46313f-	f3406a62-Percentag	Mean	8	0	1,582	1,223	aLPS;nLPS	Time in centre	nLPS-aLPS		1,582	1,223	1,469	1,215	8	2,333333	10,33333	LPS (lipop	Inflamma	Rat	
13	21ab4cd6-	Yan et al., 2017	2017	fe9ca88b-	be65beb3-Percentag	Mean	7	0	3,939	2,179	Saline;aLPS	Time in centre	nSaline-a		3,939	2,179	3,697	2,159	7	2,333333	9,333333	Saline (m	Inflamma	Rat	

Download the dataset

- Github: <https://github.com/camaradesberlin/Intermediate-MA-workshop>
- Download the “R exercise” folder
- Unzip the folder
- Open the Rproject by clicking the "Intermediate MA.Rproj" file

3

Extracting data from primary studies

What data to extract?

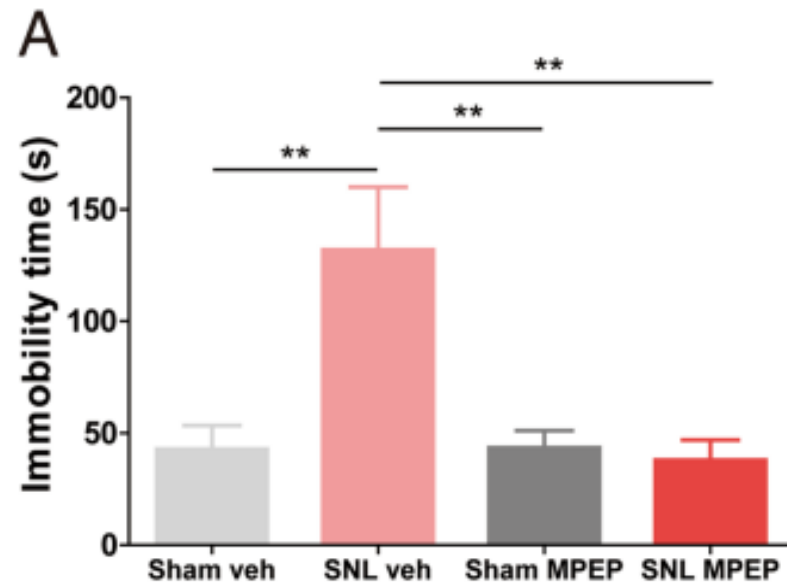
$$ES_i = \frac{\bar{x}_c - \bar{x}_{\text{rx}}}{S_{\text{pooled}}} \times \left(1 - \frac{3}{4N - 9}\right)$$

$$S_{\text{pooled}} = \sqrt{\frac{(n'_c - 1)SD_c^2 + (n_{\text{rx}} - 1)SD_{\text{rx}}^2}{N - 2}}$$

$$VAR = \frac{N}{n_{\text{rx}} \times n'_c} + \frac{ES_i^2}{2N}$$

Sources of data

- Figures
- Tables
- Text



Variable	Intervention group mean (s.e.)	Control group mean (s.e.)
Activities of daily living (ADCS ADL total score)	T0: 60.55 (0.91)	T0: 60.53 (0.91)
	T1: 62.35 (0.91)	T1: 57.47 (0.91)
	T2: 61.26 (1.00)	T2: 53.50 (0.94)
Neuropsychiatric symptom profiles (NPI total score)	T0: 11.25 (1.26)	T0: 11.77 (1.26)
	T1: 10.05 (1.26)	T1: 15.71 (1.26)
	T2: 10.40 (1.38)	T2: 16.09 (1.29)
Executive function and language ability (semantic word fluency, number of words)	T0: 13.60 (0.65)	T0: 13.92 (0.65)
	T1: 15.27 (0.65)	T1: 12.46 (0.65)
	T2: 14.15 (0.69)	T2: 12.05 (0.67)

Sources of data

- When SEM is given instead of SD:

$$SD_c = SEM_c \times \sqrt{n_c}$$

- When median & range (or IQR) are given instead of SD:
- Transformation to mean and SD via R package metamedian (Hozo et al.)

Variable	Intervention group mean (s.e.)	Control group mean (s.e.)
Activities of daily living (ADCS ADL total score)	T0: 60.55 (0.91)	T0: 60.53 (0.91)
	T1: 62.35 (0.91)	T1: 57.47 (0.91)
	T2: 61.26 (1.00)	T2: 53.50 (0.94)
Neuropsychiatric symptom profiles (NPI total score)	T0: 11.25 (1.26)	T0: 11.77 (1.26)
	T1: 10.05 (1.26)	T1: 15.71 (1.26)
	T2: 10.40 (1.38)	T2: 16.09 (1.29)
Executive function and language ability (semantic word fluency, number of words)	T0: 13.60 (0.65)	T0: 13.92 (0.65)
	T1: 15.27 (0.65)	T1: 12.46 (0.65)
	T2: 14.15 (0.69)	T2: 12.05 (0.67)

Hozo SP, Djulbegovic B, Hozo I. Estimating the mean and variance from the median, range, and the size of a sample. BMC medical research methodology. 2005 Dec;5:1-0.

3a Exercise

Extract data from Gerasimova et al.

- First identify the relevant experiment / comparison
- Find means, SDs
- Hints:
 - N = 15 in both groups
 - The numbers are in the text

Gerasimova E, Burkhanova G, Chernova K, Zakharov A, Enikeev D, Khaertdinov N, Giniatullin R, Sitdikova G. Hyperhomocysteinemia increases susceptibility to cortical spreading depression associated with photophobia, mechanical allodynia, and anxiety in rats. Behavioural brain research. 2021 Jul 9;409:113324.

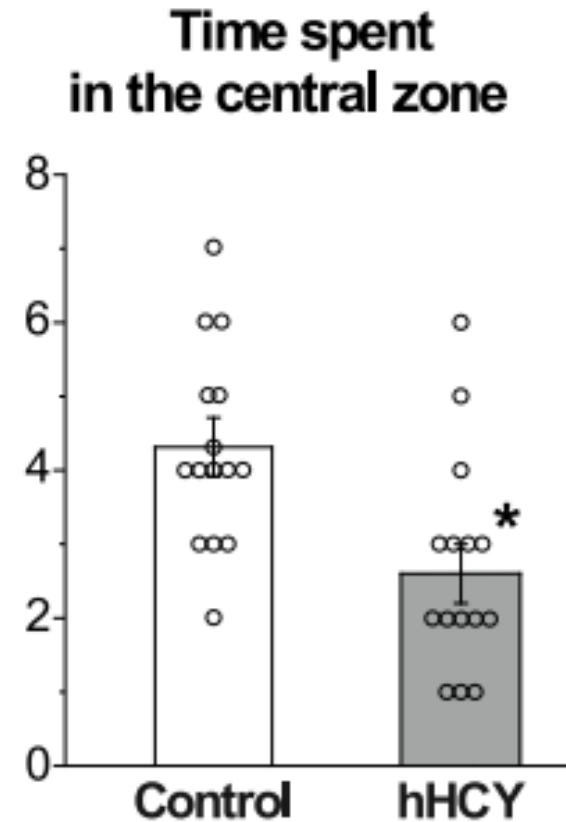


Supporting software

- SyRF



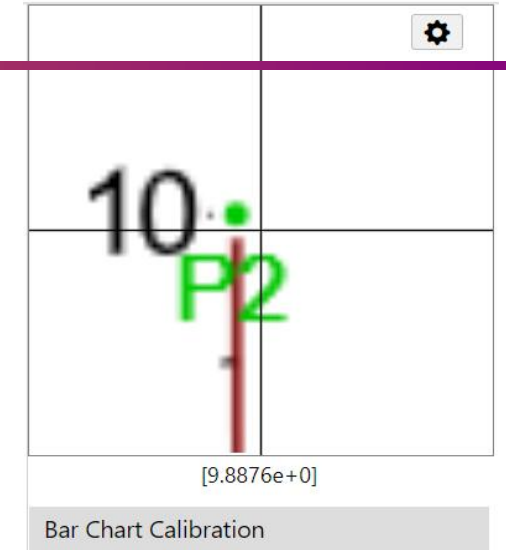
- Measuring stuff:
 - Universal Desktop Ruler
 - Adobe Desktop Ruler
 - Webplot Digitizer
- Further reading: <https://osf.io/np3az>



Gerasimova et al.

WebPlotDigitizer

1. Load the PDF + navigate to the pdf page
2. Calibrate Axes
3. Assign points on the graph
4. View Data and export data



Acquired Data

Dataset:

Variables: Label, Value

```
Bar0, 5.617977528089887
Bar1, 6.01123595505618
```

Sort by: Order:

Format

Number Formatting: Digits:

Column Separator:

*Plotly is a secure data analysis and graphing site with data sharing and access controls. Visit <http://plot.ly> for details.

4

Calculating effect sizes

Standardized mean difference (SMD)

$$ES_i = \frac{\bar{x}_c - \bar{x}_{rx}}{S_{pooled}} \times \left(1 - \frac{3}{4N - 9}\right)$$

$$S_{pooled} = \sqrt{\frac{(n'_c - 1)SD_c^2 + (n_{rx} - 1)SD_{rx}^2}{N - 2}}$$

$$VAR = \frac{N}{n_{rx} \times n'_c} + \frac{ES_i^2}{2(N - 3.49)}$$

N control

	A	B	C	D	E	F	G	H	I	J
1	StudyIdSt	Author	Me	Se	Mc	Sc	Ne	Nc	N.total	ModelN
2	053a1a1f-	Saffarpour et al., 20	0,375	2,674843	10,409	2,493581	6	6	12	Chronic
3	0767bd07-	Missig et al., 2017	3,71	1,565248	7,5	1,565248	5	1,666667	6,666667	Chronic
4	0cd5ef20-	Amorim et al., 2014	6,397	2,334364	10,782	4,242516	6	6	12	Kaolin a
5	0cef4a1c-f	Bree et al., 2020	156,867	64,86149	522,161	273,5768	8	8	16	Closed f
6	0cef4a1c-f	Bree et al., 2020	318,188	165,907	490,177	317,3788	8	10	18	Closed f
7	0cef4a1c-f	Bree et al., 2020	343,631	189,7592	521,363	207,1512	8	8	16	Closed f
8	11dcb928-	Gerasimova et al., 2					15	15	30	hHCY (h
9	18f7151c-7	Huang et al., 2013	3,218	2,857884	11,38	5,909757	12	12	24	Antiretr
10	21ab4cd6-	Yan et al., 2017	2,677	1,291	3,697	2,159	8	2,333333	10,33333	Saline (r

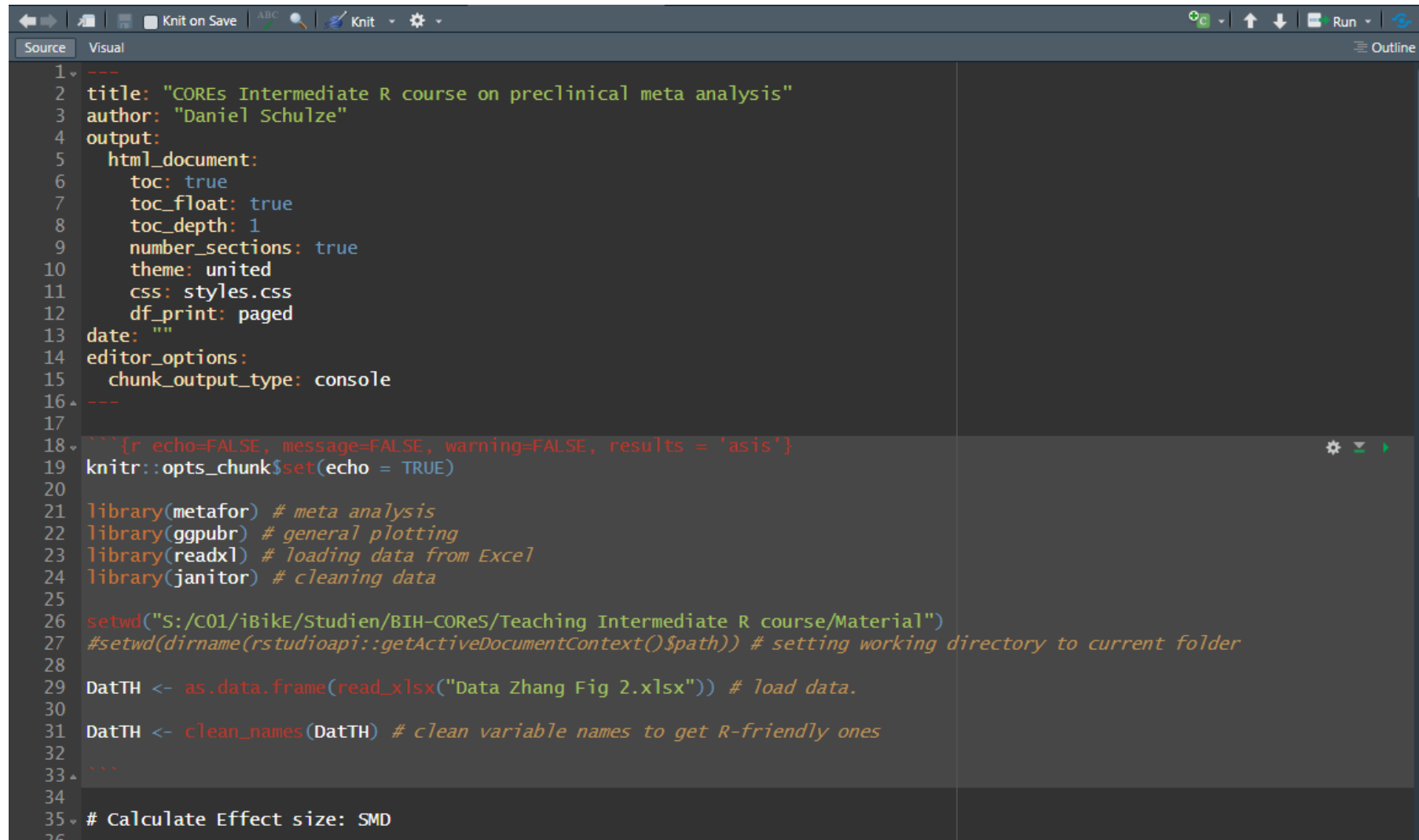
N control

A single experiment can contain a number of comparisons. If the control cohort is serving more than one treatment group, we correct the number of animals reported in the control cohort by the number of treatment groups.

$$n'_c = \frac{n_c}{\text{num. treatment groups}}$$

$$N = n_{\text{rx}} + n'_c$$

Intro to R & R Markdown



```
1 ---
2 title: "COREs Intermediate R course on preclinical meta analysis"
3 author: "Daniel Schulze"
4 output:
5   html_document:
6     toc: true
7     toc_float: true
8     toc_depth: 1
9     number_sections: true
10    theme: united
11    css: styles.css
12    df_print: paged
13 date: ""
14 editor_options:
15   chunk_output_type: console
16 ---
17
18 ```{r echo=FALSE, message=FALSE, warning=FALSE, results = 'asis'}
19 knitr::opts_chunk$set(echo = TRUE)
20
21 library(metafor) # meta analysis
22 library(ggpubr) # general plotting
23 library(readxl) # loading data from Excel
24 library(janitor) # cleaning data
25
26 setwd("S:/C01/iBikE/Studien/BIH-COREs/Teaching Intermediate R course/Material")
27 #setwd(dirname(rstudioapi::getActiveDocumentContext()$path)) # setting working directory to current folder
28
29 DatTH <- as.data.frame(read_xlsx("Data Zhang Fig 2.xlsx")) # load data.
30
31 DatTH <- clean_names(DatTH) # clean variable names to get R-friendly ones
32
33 ```
34
35 # Calculate Effect size: SMD
36
```

4a Exercise

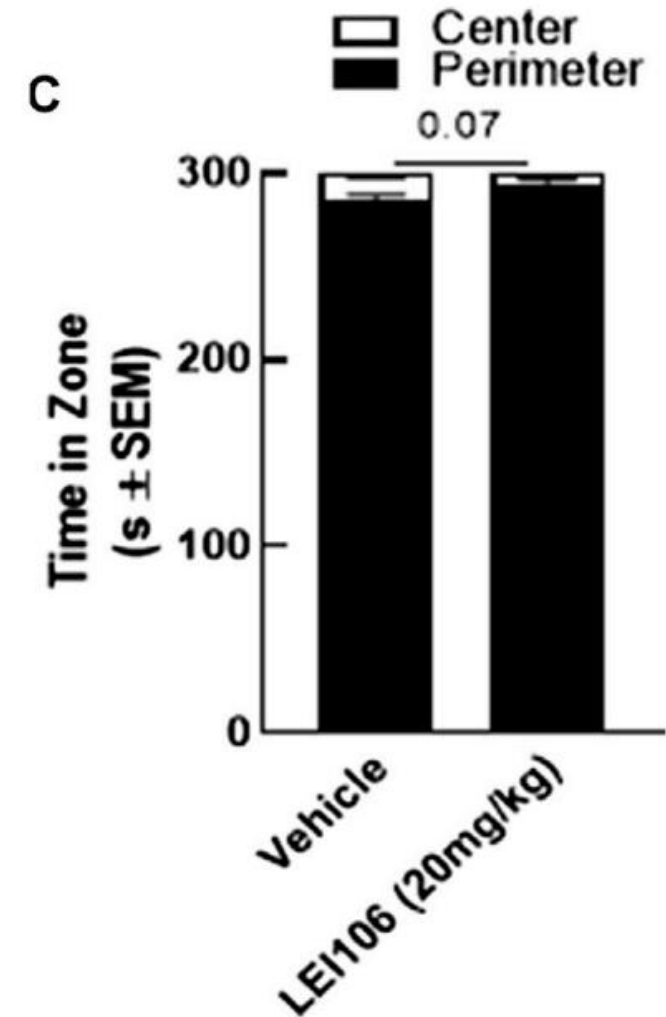
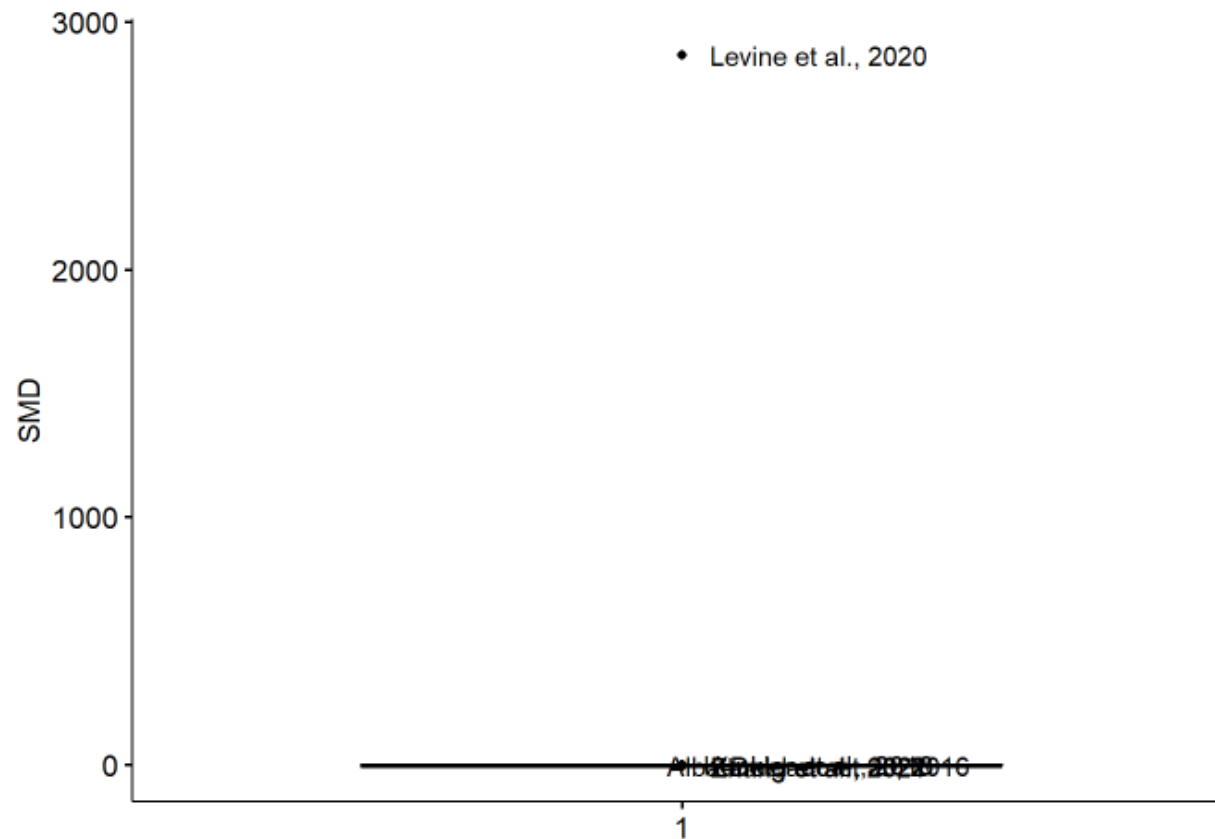
R Markdown Script

Calculate effect sizes manually

- Pooled SD
- SMD
- Variance



Outlier in the data



Levine A, Liktor-Busa E, Karlage KL, Giancotti L, Salvemini D, Vanderah TW, Largent-Milnes TM. DAGL α inhibition as a non-invasive and translational model of episodic headache. *Frontiers in pharmacology*. 2021 Jan 12;11:615028.

4b Exercise

R Markdown Script

Calculate effect sizes in metafor

- `escalc()`



Specifics for NMD

- Not part of `escalc()`
- Has to be done manually
- Pay attention to the direction of effect!

Specifics for Odds Ratio

- How to deal with 0 events?

$$OR_i = \frac{a_i \times d_i}{b_i \times c_i}$$

Two approaches:

- Adding 0.5 to all zero cells in the cross table
- Arcsin transformation: "AS" in `escalc()` (cave: abstract measure of effect!)

$$\arcsin\sqrt{(a/a + b)} - \arcsin\sqrt{(c/c + d)}$$

5

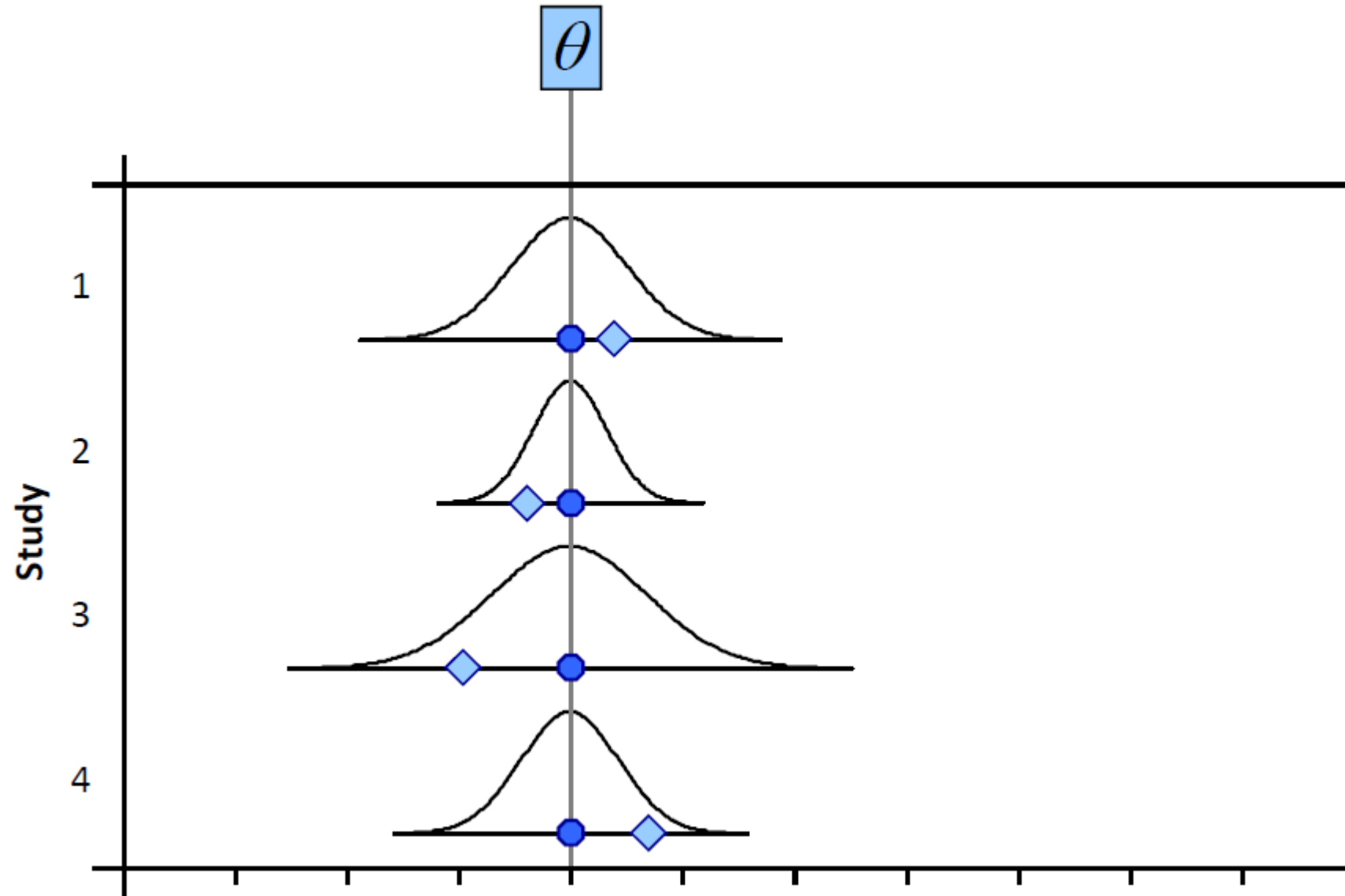
Meta-analytical models

MA = Weighted average of effects

- Logic: Weight study results by study specific uncertainty (size, SD, effect)
- Studies get a larger weight, when:
 - Larger N
 - Smaller SD
 - Larger group difference

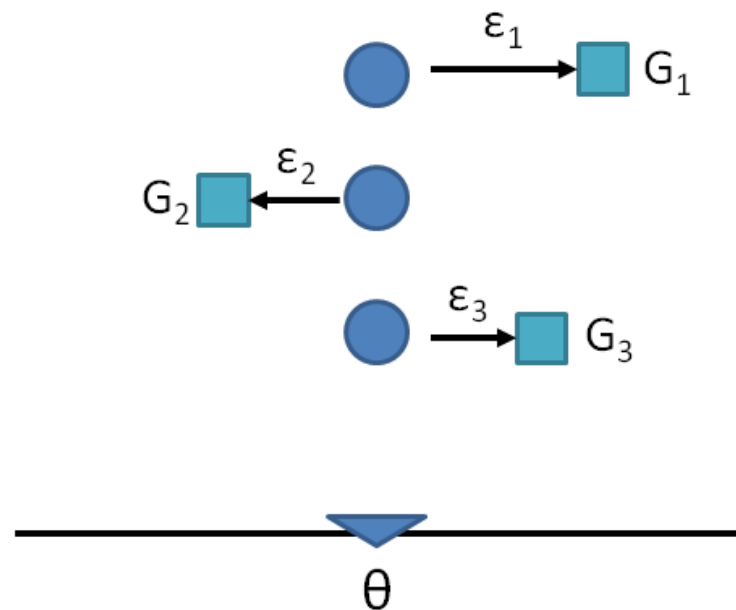
$$\hat{\theta} = \frac{\sum w_i y_i}{\sum w_i} \quad w_i = \frac{1}{VAR}$$

Equal/fixed effects model



Equal/fixed effects model

The Fixed-Effect Model

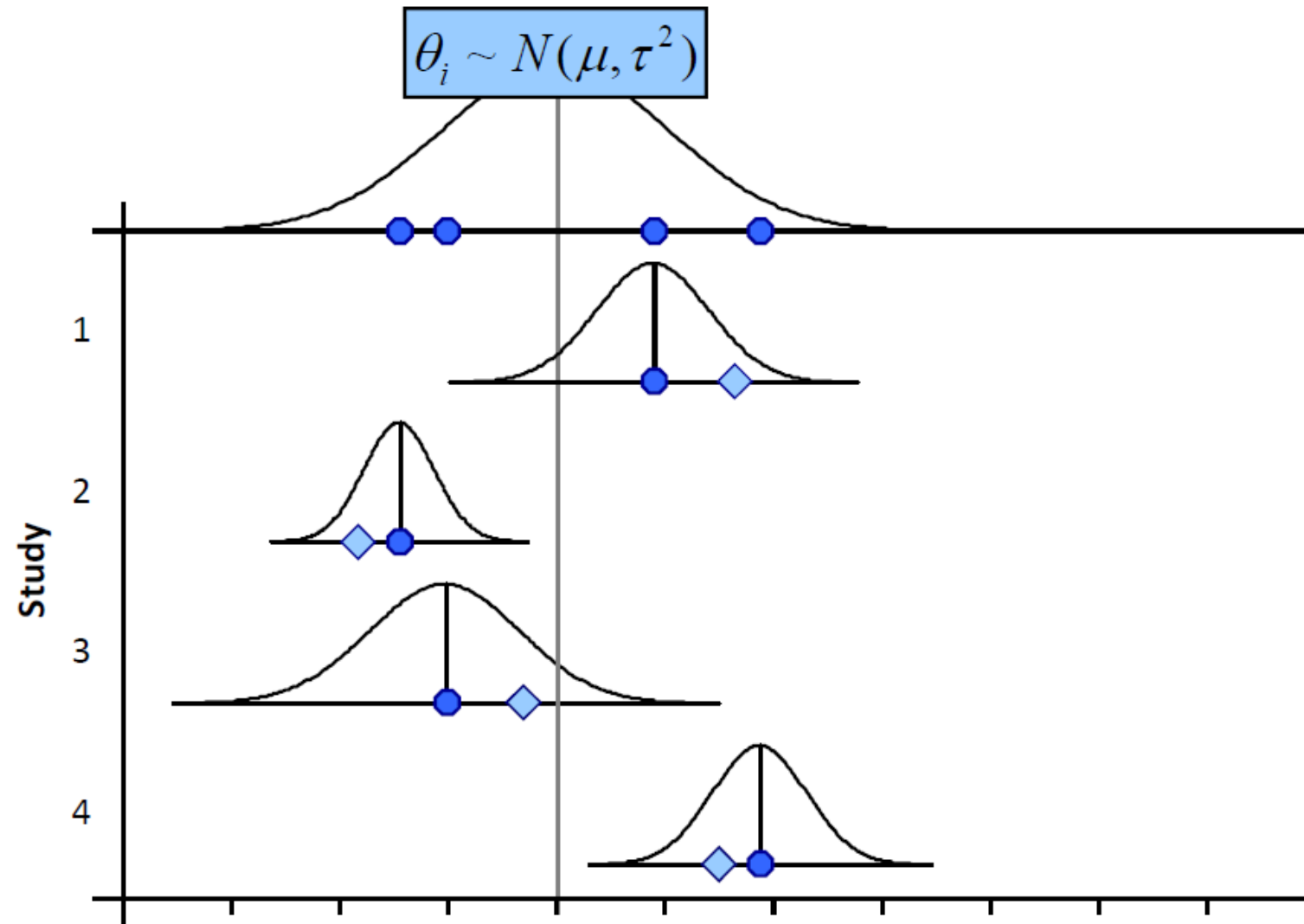


We assume that all of studies (blue circles) share a common effect-size (blue triangle; θ). Thus, the only reason our studies differ in their results is sampling error (ϵ_i).

Thus, the observed effect (G_i) is assumed to be a function of:

$$G_i = \theta + \epsilon_i$$

Random effects model



Random effects model

The Random-Effects Model

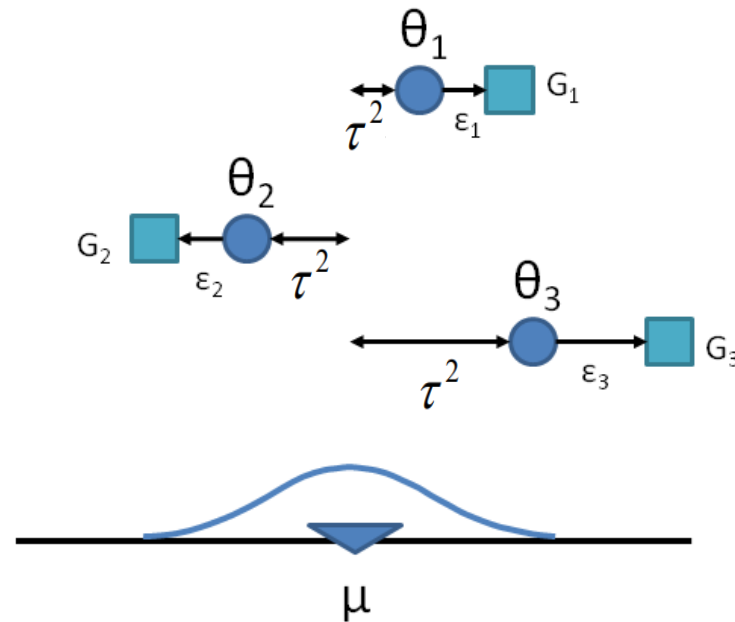
Now we have multiple sources of error:

τ^2 is the true variation in effect sized. Or, the distance between the population effect for each study (θ_i) and summary effect (μ).

ε_i , the sampling error for each observed effect (G_i).

Thus:

$$G_i = \mu + \tau^2 + \varepsilon_i$$



Underlying effects (θ) are normally distributed around some average effect (μ).

Random effects model

Model

$$y_i = \overbrace{\mu + u_i}^{\theta_i} + \varepsilon_i \quad u_i \sim N(0, \tau^2)$$

Parameter
Estimate

$$\hat{\mu} = \frac{\sum w_i y_i}{\sum w_i} \quad w_i = \frac{1}{v_i + \hat{\tau}^2}$$

Var and SE of
the Estimate

$$Var[\hat{\mu}] = \frac{1}{\sum w_i} \quad SE[\hat{\theta}] = \sqrt{\frac{1}{\sum w_i}}$$

Inference

$$z = \frac{\hat{\mu}}{SE[\hat{\mu}]} \quad \hat{\mu} \pm 1.96 SE[\hat{\mu}]$$

Random effects model

- Is the standard nowadays, default in metafor
- Estimator: Restricted maximum likelihood (REML), default in metafor
- Confidence intervals: Via method by Knapp-Hartung, set in metafor via `test = "knha"`

5a Exercise

R Markdown Script
Run Meta-Analysis

- `rma()`
- Keep in mind to specify `test = „knha“`



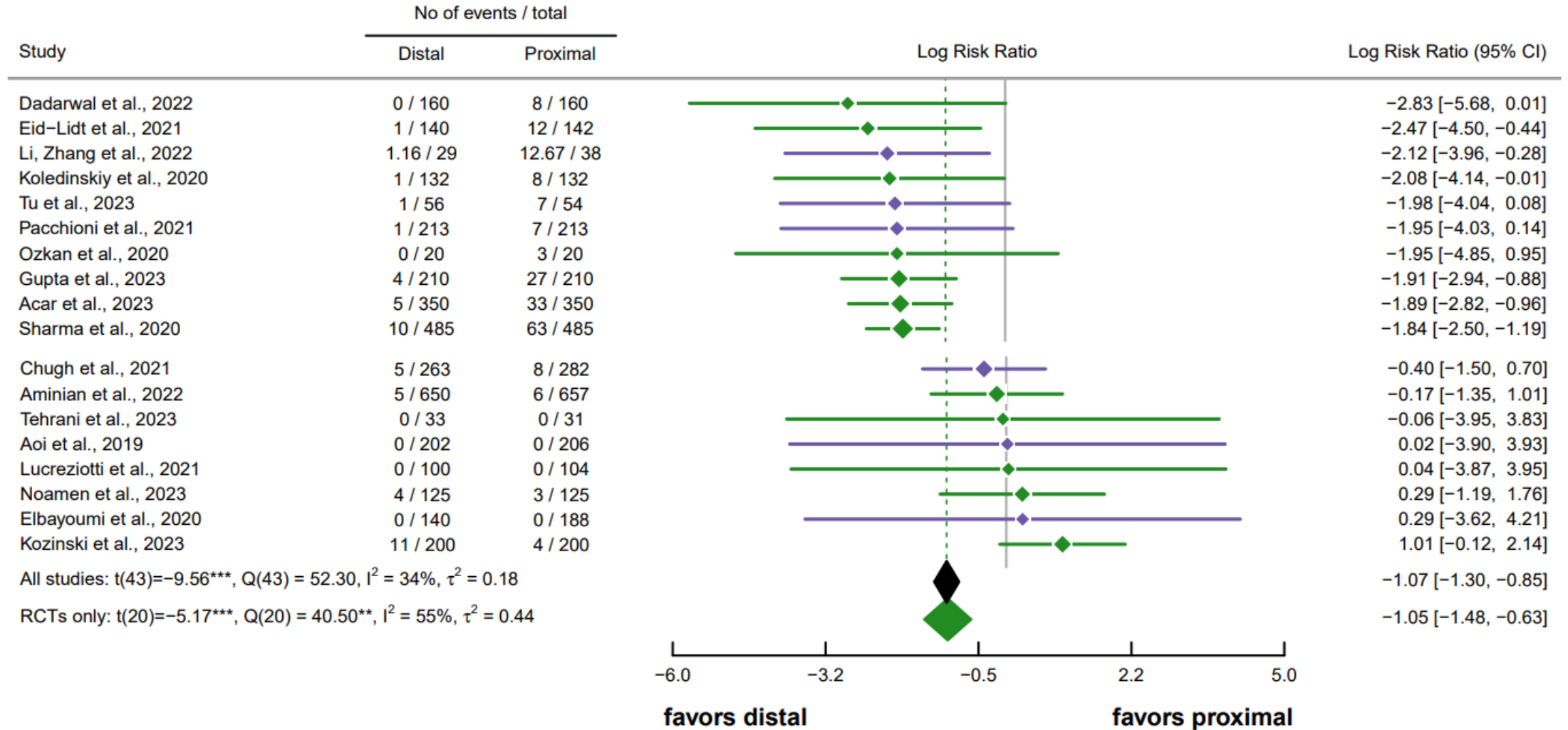
Random effects model

```
##
## Random-Effects Model (k = 90; tau^2 estimator: REML)
##
##      logLik    deviance      AIC      BIC      AICc
## -137.0998    274.1997    278.1997    283.1769    278.3392
##
## tau^2 (estimated amount of total heterogeneity): 0.7711 (SE = 0.1664)
## tau (square root of estimated tau^2 value):      0.8781
## I^2 (total heterogeneity / total variability):    73.38%
## H^2 (total variability / sampling variability):    3.76
##
## Test for Heterogeneity:
## Q(df = 89) = 311.7118, p-val < .0001
##
## Model Results:
##
## estimate      se      tval  df    pval    ci.lb    ci.ub
## -1.1448  0.1176  -9.7344  89  <.0001  -1.3785  -0.9111  ***
##
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

6

Forest Plot

Forest plot



6a Exercise

R Markdown Script

Create a forest plot

- `forest()`
- The `order` argument gives control over the sequence of studies
- The `slab` argument allows the inclusion of study labels



Modifying

k+3=16

k+2=15

k+1=14

k=13

12

11

10

9

8

7

6

5

4

3

2

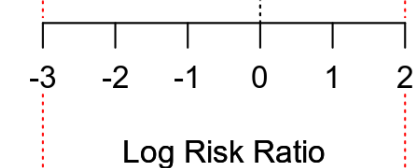
1

0

-1

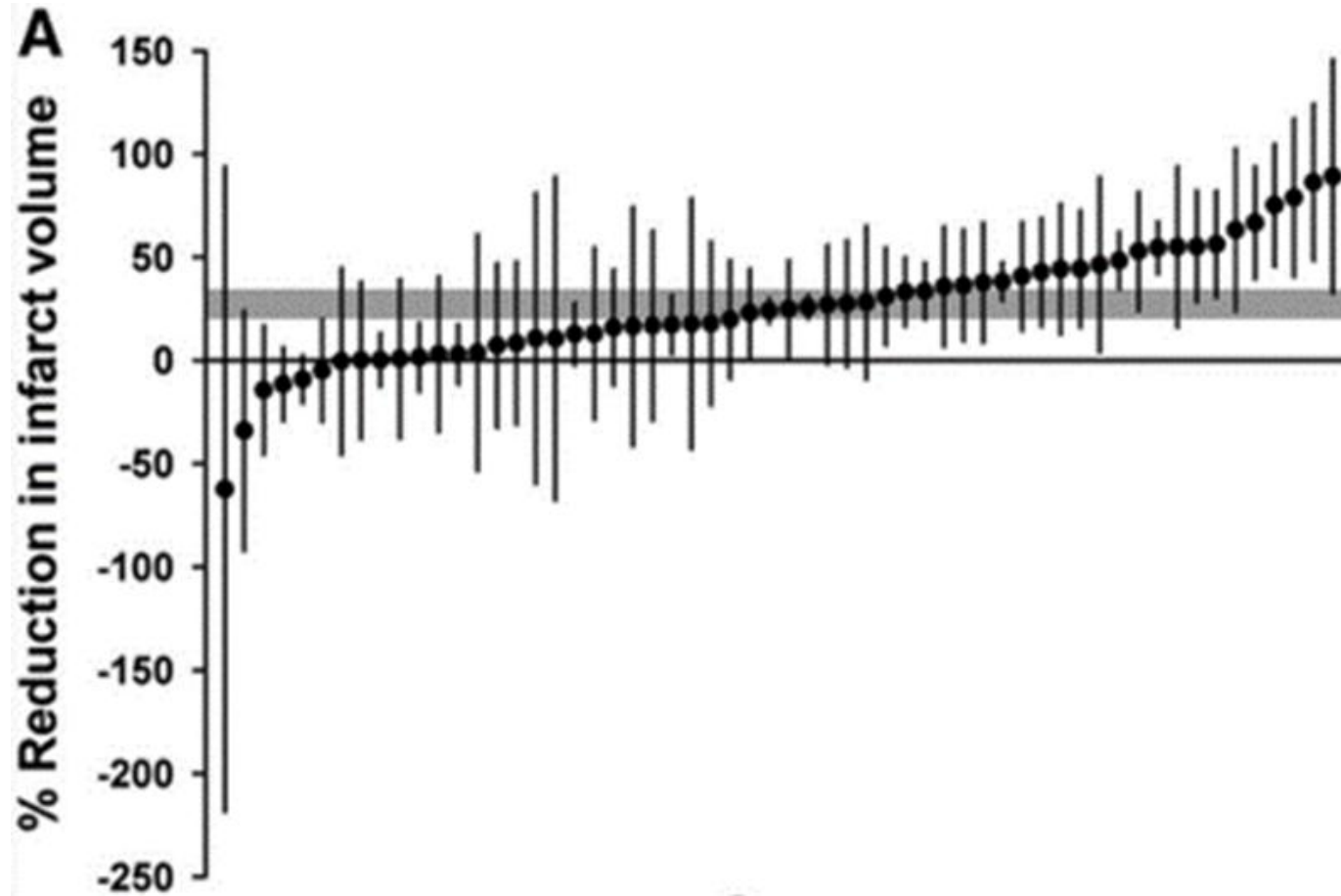
Author(s) and Year	Vaccinated		Control		Log[RR] [95% CI]
	TB+	TB-	TB+	TB-	
Aronson, 1948	4	119	11	128	-0.89 [-2.01, 0.23]
Ferguson & Simes, 1949	6	300	29	274	-1.59 [-2.45, -0.72]
Rosenthal et al, 1960	3	228	11	209	-1.35 [-2.61, -0.08]
Hart & Sutherland, 1977	62	13536	248	12619	-1.44 [-1.72, -1.16]
Frimodt-Moller et al, 1973	33	5036	47	5761	-0.22 [-0.66, 0.23]
Stein & Aronson, 1953	180	1361	372	1079	-0.79 [-0.95, -0.62]
Vandiviere et al, 1973	8	2537	10	619	-1.62 [-2.55, -0.70]
TPT Madras, 1980	505	87886	499	87892	0.01 [-0.11, 0.14]
Coetzee & Berjak, 1968	29	7470	45	7232	-0.47 [-0.94, -0.00]
Rosenthal et al, 1961	17	1699	65	1600	-1.37 [-1.90, -0.84]
Comstock et al, 1974	186	50448	141	27197	-0.34 [-0.56, -0.12]
Comstock & Webster, 1969	5	2493	3	2338	0.45 [-0.98, 1.88]
Comstock et al, 1976	27	16886	29	17825	-0.02 [-0.54, 0.51]
RE Model					-0.71 [-1.07, -0.36]

Labels to the right of xlim[1].



Annotations to the left of xlim[2].

Timber/caterpillar plot



Timber/caterpillar plot

```
forest(yi, vi, xlim=c(-2.5,3.5), ### adjust horizontal plot region limits
       order="obs", ### order by size of yi
       slab=NA, annotate=FALSE, ### remove study labels and annotations
       efac=0, ### remove vertical bars at end of CIs
       pch=19, ### changing point symbol to filled circle
       col="gray40", ### change color of points/CIs
       psize=2, ### increase point size
       cex.lab=1, cex.axis=1, ### increase size of x-axis title/labels
       lty=c("solid","blank")) ### remove horizontal line at top of plot
```

Welcome back!



Course structure

2nd Day

- *Heterogeneity in meta-analysis of animal studies*
- *Meta regression:*
 - *Regression model & Explaining heterogeneity*

BREAK

- *Running models in R + Plotting*
- *Obstacles*

BREAK

- *Exploring the impacts of Risks of Bias in R*
- *Funnel plotting*
- *Workshop Conclusion & Wrap-up*

7

Heterogeneity

Heterogeneity

- “Statistical heterogeneity manifests itself in the... [study] effects being more different from each other than one would expect due to random error (chance) alone”

Cochrane Handbook

= unexpected amount of variation in effect sizes between studies

- We can calculate the amount of variation we would expect due to sampling error (ϵ)
- Heterogeneity is measured by:
 - Q-test statistic
 - τ^2
 - I^2

Heterogeneity: Q-test

- If the effect sizes are really homogeneous, then Q follows a chi-square distribution
- -> p-value of a significance test

$$H_0 : \theta_1 = \theta_2 = \dots = \theta_k$$

$$w_i = 1 / v_i$$

$$Q = \sum w_i (y_i - \hat{\theta})^2$$

Heterogeneity: τ^2

- τ^2 estimates the total amount of variability (heterogeneity) among the effect sizes
- heterogeneity may be due to random or systematic differences between the θ_i 's
- τ^2 does not differentiate between sources
- Hard to interpret (is a raw variance): When is it large?

Heterogeneity: I^2

- I^2 estimates (in %) how much of the total variability in the effect size estimates is due to heterogeneity among the true effects
- Rough guidelines (Cochrane):
 - 0% to 40%: might not be important
 - 30% to 60%: moderate heterogeneity
 - 50% to 90%: substantial heterogeneity
 - 75% to 100%: considerable heterogeneity

$$I^2 = 100\% \times \frac{\hat{\tau}_{RE}^2}{\hat{\tau}_{RE}^2 + s^2}$$
$$= 100\% \times \frac{Q - (k - 1)}{Q}$$
$$s^2 = \frac{(k - 1) \sum w_i}{(\sum w_i)^2 - \sum w_i^2}$$

7a Exercise

R Markdown Script
Check heterogeneity

- Run `rma()` again
- Adding heterogeneity measures to the forest plot



Example: Heterogeneity

```
##
## Random-Effects Model (k = 90; tau^2 estimator: REML)
##
##      logLik    deviance      AIC      BIC      AICc
## -137.0998    274.1997    278.1997    283.1769    278.3392
##
## tau^2 (estimated amount of total heterogeneity): 0.7711 (SE = 0.1664)
## tau (square root of estimated tau^2 value):      0.8781
## I^2 (total heterogeneity / total variability):    73.38%
## H^2 (total variability / sampling variability):    3.76
##
## Test for Heterogeneity:
## Q(df = 89) = 311.7118, p-val < .0001
##
## Model Results:
##
## estimate      se      tval  df    pval    ci.lb    ci.ub
##  -1.1448  0.1176  -9.7344  89   <.0001  -1.3785  -0.9111  ***
##
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Heterogeneity – a bad thing?

- In clinical MAs, heterogeneity is traditionally a nuisance
- By understanding the sources of heterogeneity, we can better understand mechanisms at work
- Especially important in the light of translational failure in preclinical research
- Heterogeneity might thus be a desirable property of our MA
- Heterogeneity in experimental setups increases replicability (Usui et al, 2021)

Usui T, Macleod MR, McCann SK, Senior AM, Nakagawa S. Meta-analysis of variation suggests that embracing variability improves both replicability and generalizability in preclinical research. PLoS Biology. 2021 May 19;19(5):e3001009.

8

Exploring Heterogeneity - Meta-Regression

Heterogeneity in preclinical studies

- Heterogeneity is typically greater in preclinical than in clinical MAs
- Study-level variables that may influence the direction/size of the outcome
 - substantive variables (characteristics of the treatment, context, subjects)
 - methodological variables (e.g., randomized versus non-randomized study)
 - extrinsic variables (e.g., publication year, published/unpublished)
- In preclinical data:
 - Study samples (e.g. species, sex)
 - Interventions or outcomes (dose, outcome types)
 - Methodology: outcome measures used, quality etc.

Meta-Regression

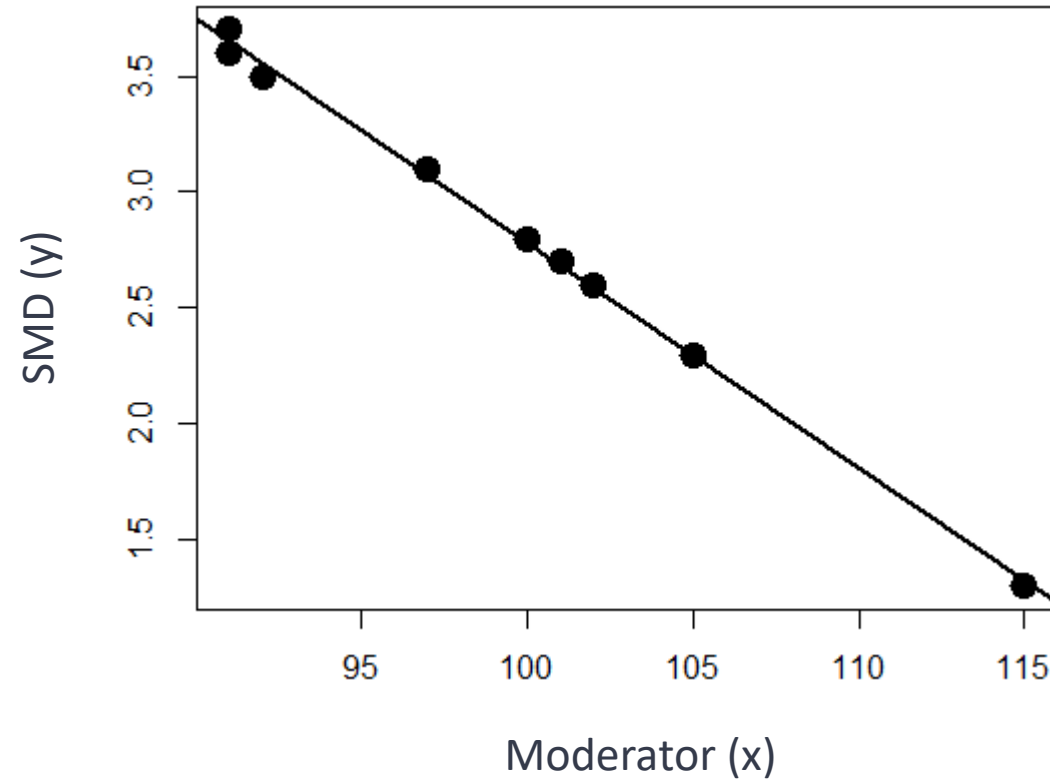
- Using regression analysis to evaluate the relevance of these study-specific features for the found effect
- The variables are also called “**moderators**” of the effect

Intro to Regression

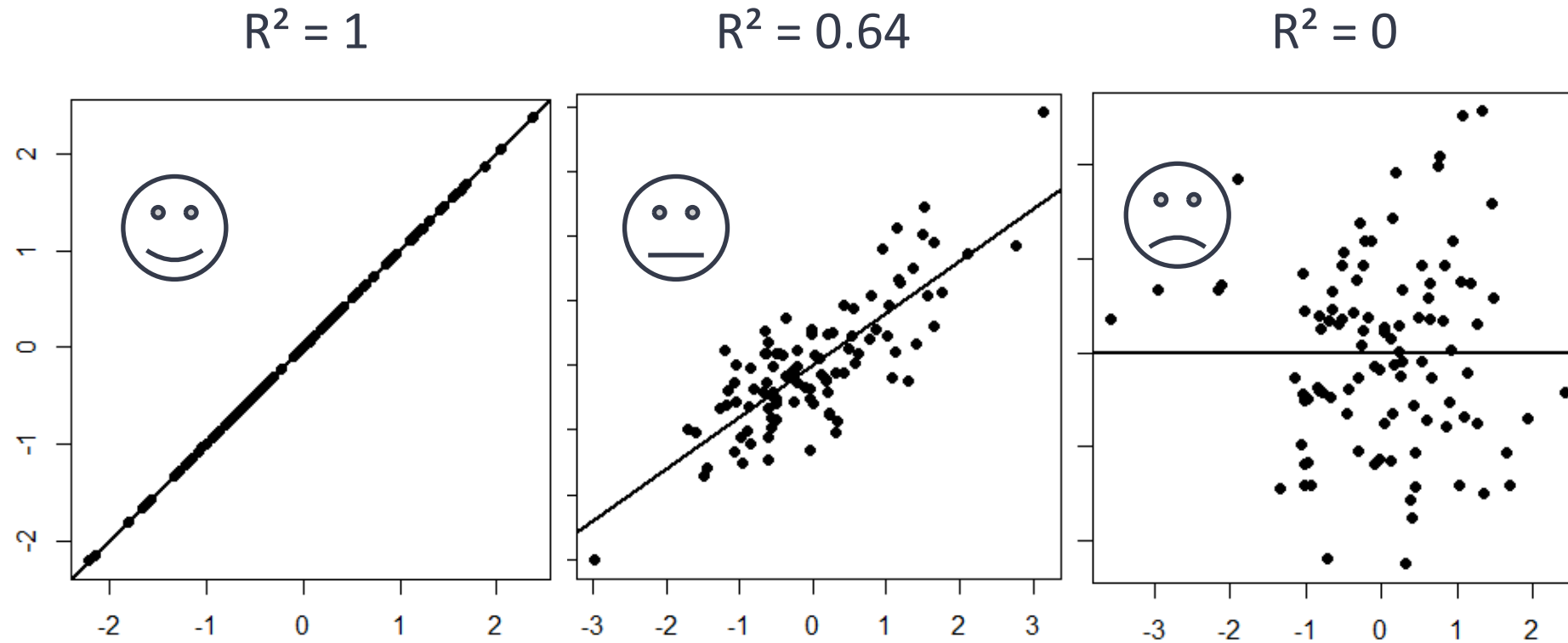
Describing a scatter plot:

$$\hat{y} = a + b \times x$$

- a= intercept
- b = slope



Intro to Regression



Meta-Regression

A linear regression is described by:

- An equation for the line: $\hat{y} = a + b \times x$
- R^2 as measure of quality: How much variance in y can be predicted by x ?
- I^2 is also a variance proportion!
- -> How much variance (I^2) can be explained by the moderator?

8a Exercise

R Markdown Script
Meta-regression

- `rma()`, once again
- Adding a moderator:
year of publication

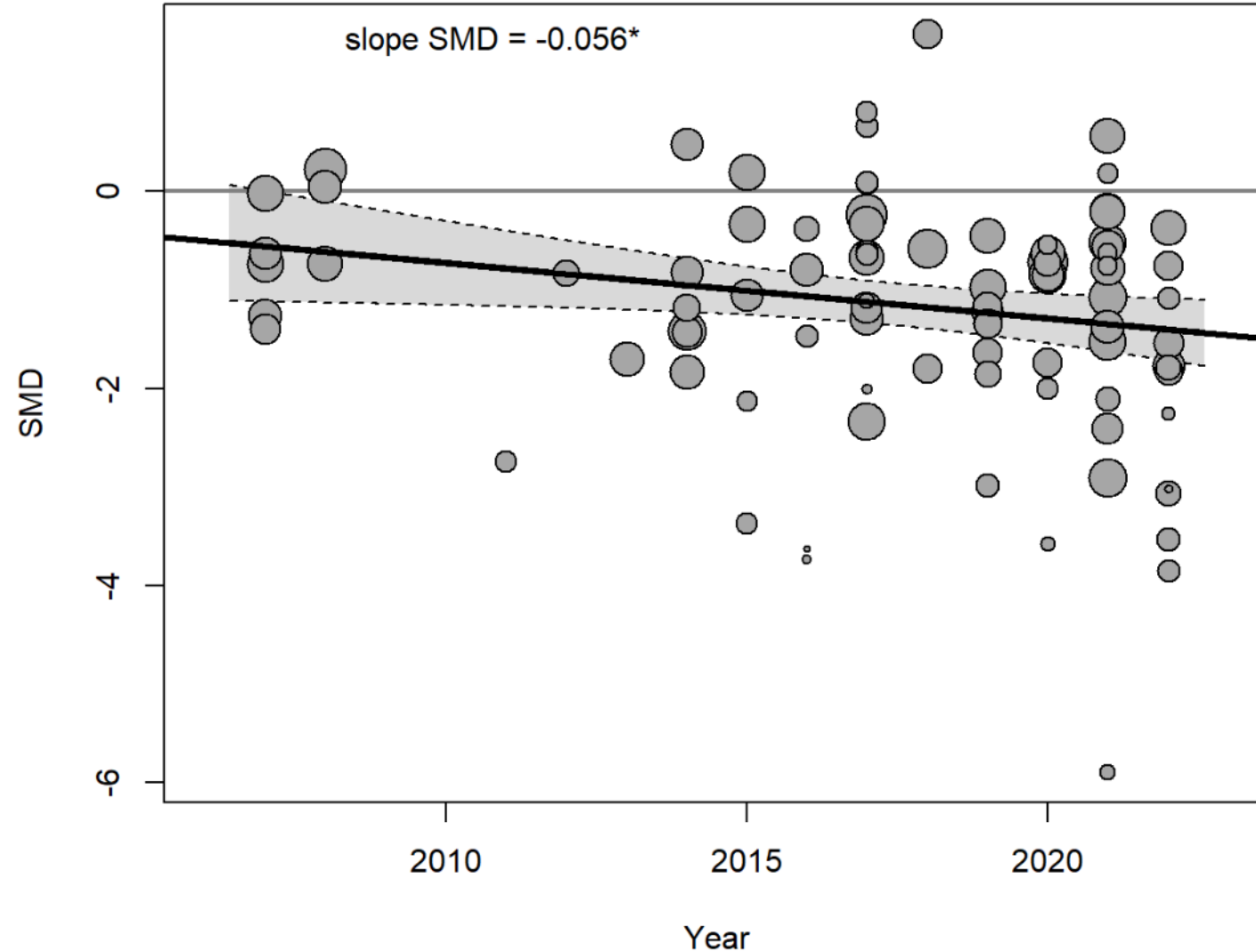


Meta-Regression

```
##
## Mixed-Effects Model (k = 90; tau^2 estimator: REML)
##
##      logLik    deviance      AIC      BIC      AICc
## -133.7690    267.5381    273.5381    280.9701    273.8238
##
## tau^2 (estimated amount of residual heterogeneity):    0.7199 (SE = 0.1594)
## tau (square root of estimated tau^2 value):          0.8484
## I^2 (residual heterogeneity / unaccounted variability): 71.85%
## H^2 (unaccounted variability / sampling variability):  3.55
## R^2 (amount of heterogeneity accounted for):          6.64%
##
## Test for Residual Heterogeneity:
## QE(df = 88) = 289.5407, p-val < .0001
##
## Test of Moderators (coefficient 2):
## QM(df = 1) = 4.9019, p-val = 0.0268
##
## Model Results:
##
##      estimate      se      zval      pval      ci.lb      ci.ub
## intrcpt  112.6325  51.3878   2.1918   0.0284  11.9143  213.3507  *
## year      -0.0564   0.0255  -2.2140   0.0268  -0.1063  -0.0065  *
##
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Plotting Meta-Regression

- Bubble plot:



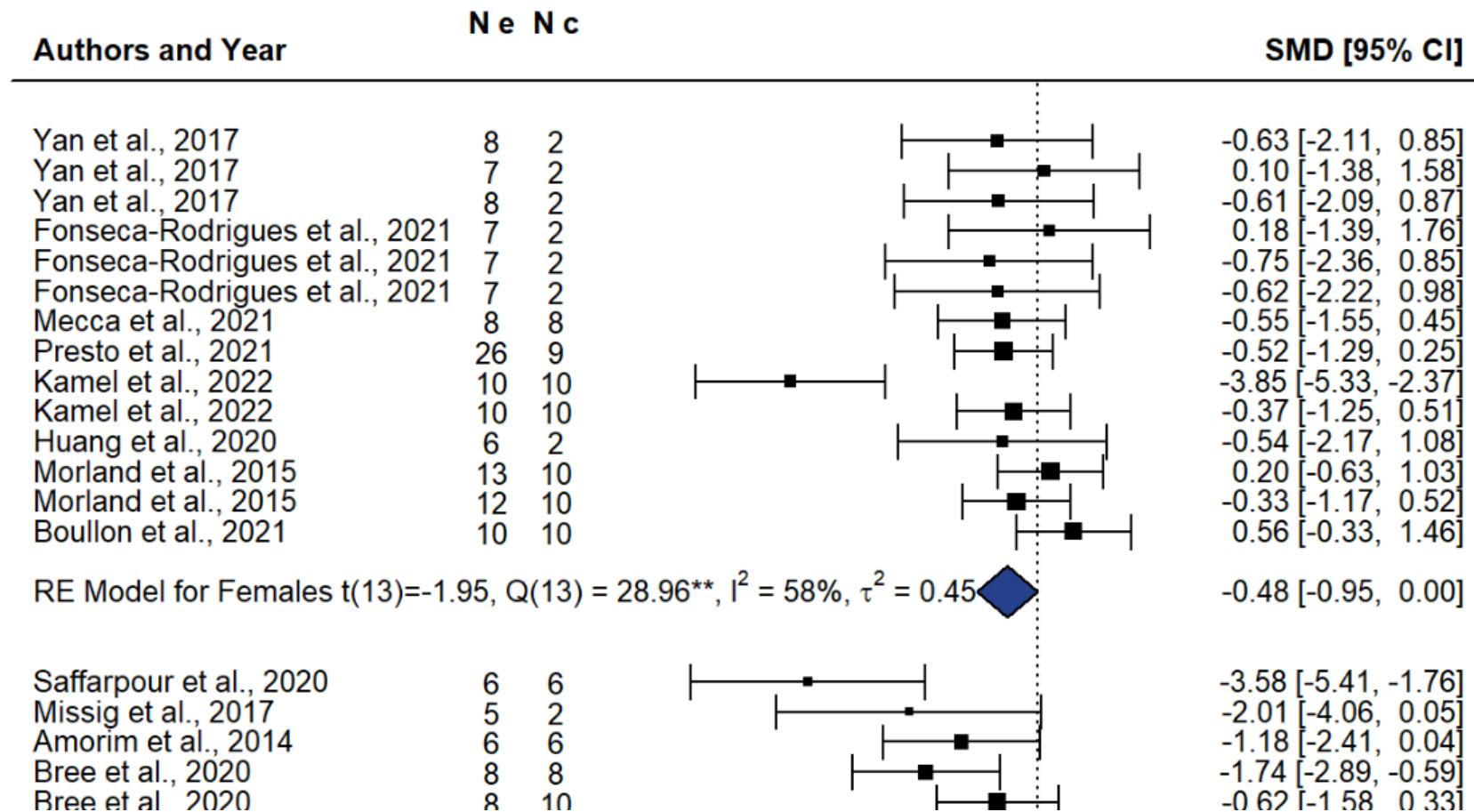
Meta-Regression with binary predictors

- Nothing changes, however, predictors have to be coded as factor (or 0 and 1) in R

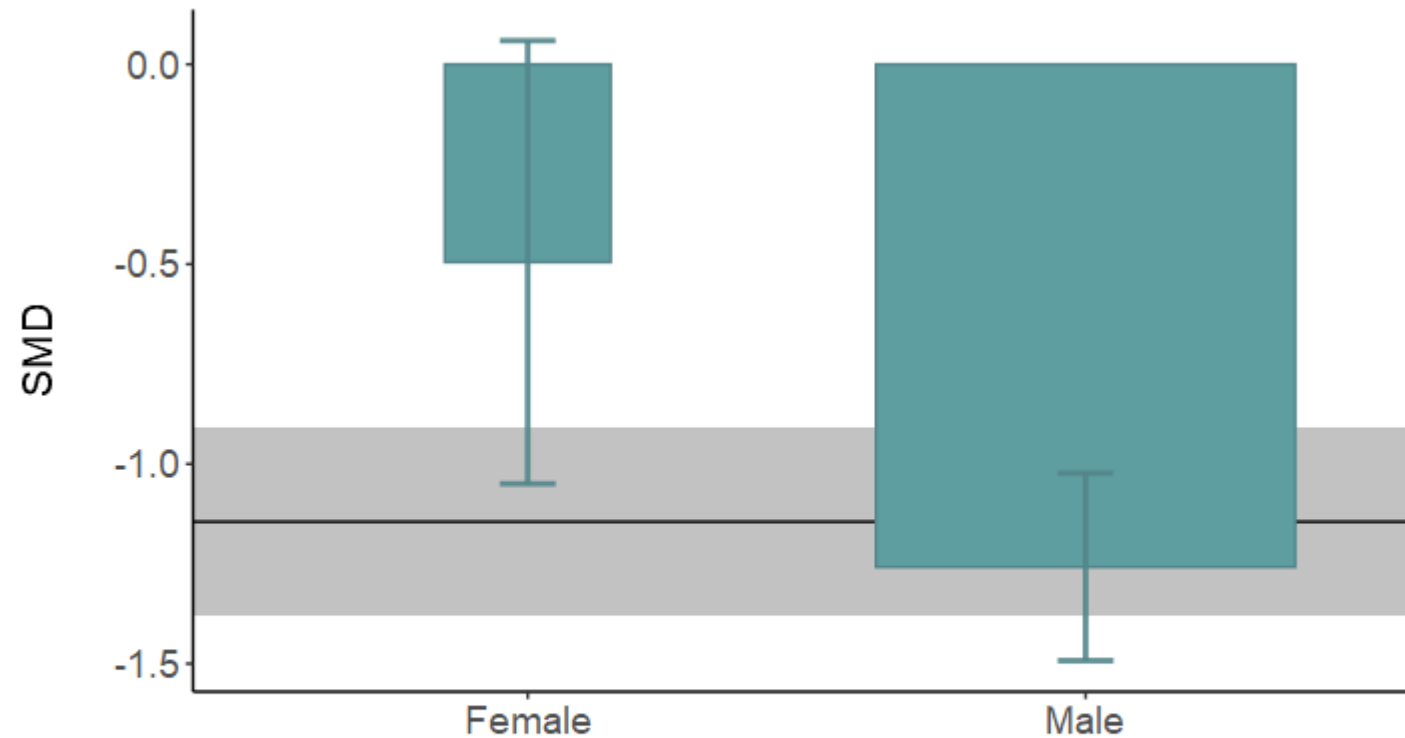
```
##
## Mixed-Effects Model (k = 89; tau^2 estimator: REML)
##
##      logLik    deviance      AIC      BIC      AICc
## -132.2240    264.4480    270.4480    277.8458    270.7372
##
## tau^2 (estimated amount of residual heterogeneity):    0.7227 (SE = 0.1611)
## tau (square root of estimated tau^2 value):          0.8501
## I^2 (residual heterogeneity / unaccounted variability): 71.86%
## H^2 (unaccounted variability / sampling variability):  3.55
## R^2 (amount of heterogeneity accounted for):          8.43%
##
## Test for Residual Heterogeneity:
## QE(df = 87) = 293.4408, p-val < .0001
##
## Test of Moderators (coefficient 2):
## QM(df = 1) = 6.1679, p-val = 0.0130
##
## Model Results:
##
##      estimate      se      zval      pval      ci.lb      ci.ub
## intrcpt  -0.4952  0.2829  -1.7503  0.0801  -1.0497  0.0593 .
## sexMale  -0.7629  0.3072  -2.4835  0.0130  -1.3649 -0.1608 *
##
## ---
## Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

Sub-group Analysis (less used in preclinical)

- Forest plot (and meta-analysis) separated by group:



Plotting categorical moderators



The horizontal grey bar represents the 95% CIs of the global estimate.
The width of the bars is proportional to the number of animals in each subgroup.

8b Exercise

R Markdown Script
Meta-regression

- Run meta-regression for the strain variable



Obstacles in Meta-Regression

- Small number of studies: Run only if there are > 10 studies (?)
- Grouping categories: reduce number of categories

8

Detecting Small Study Effects (publication bias)

Publication Bias

- Publication bias occurs when the results of published and unpublished studies differ systematically
- Neutral and negative studies:
 - Take longer to publish
 - Remain unpublished
 - Less likely to be identified in systematic review
 - Leads to the overstatement of efficacy in meta-analysis
- Selective outcome reporting
- Selective analysis reporting

Publication Bias

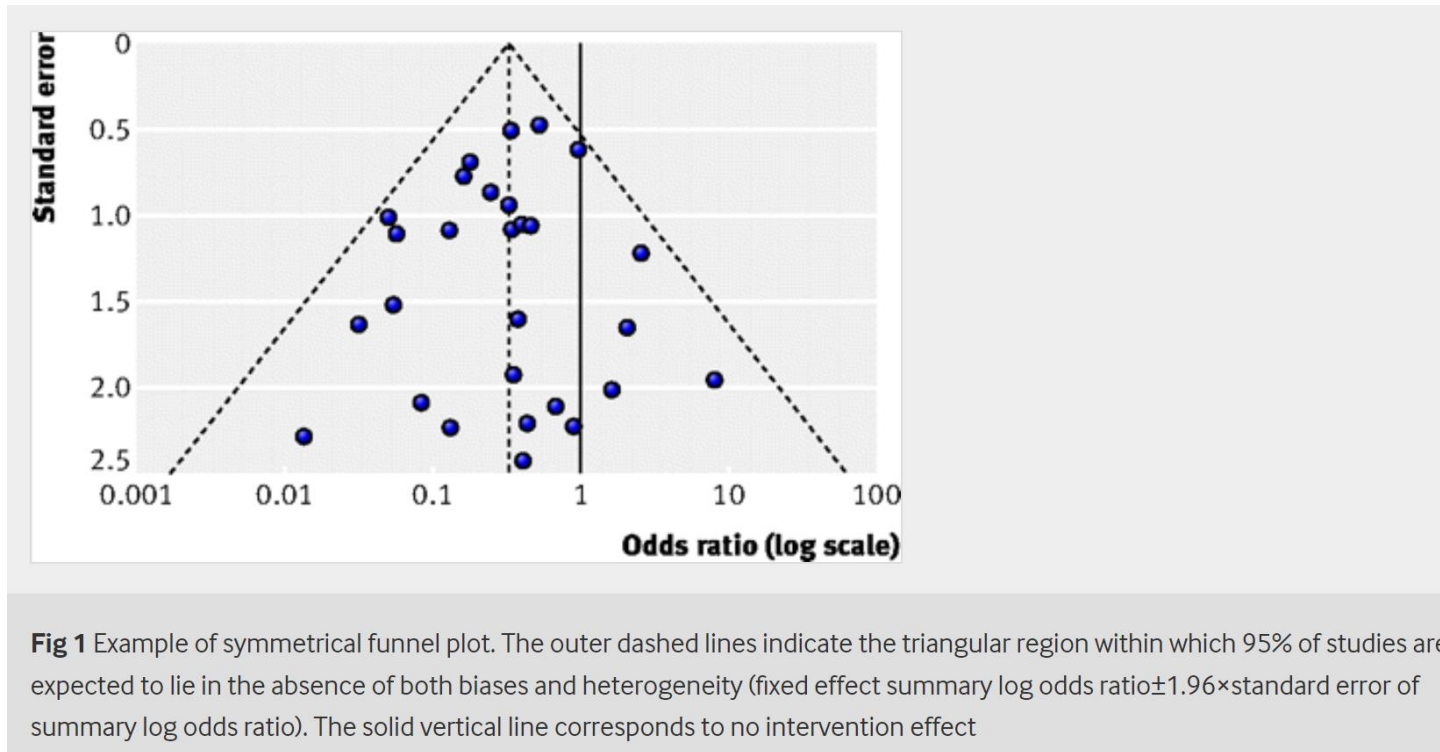
Small study effects assumptions:

- Larger studies with more commitment of resources and time → more likely to be published
- Smaller studies have greater risk of generating non-significant findings
- Smaller studies therefore are more likely to “remain in the file drawer”



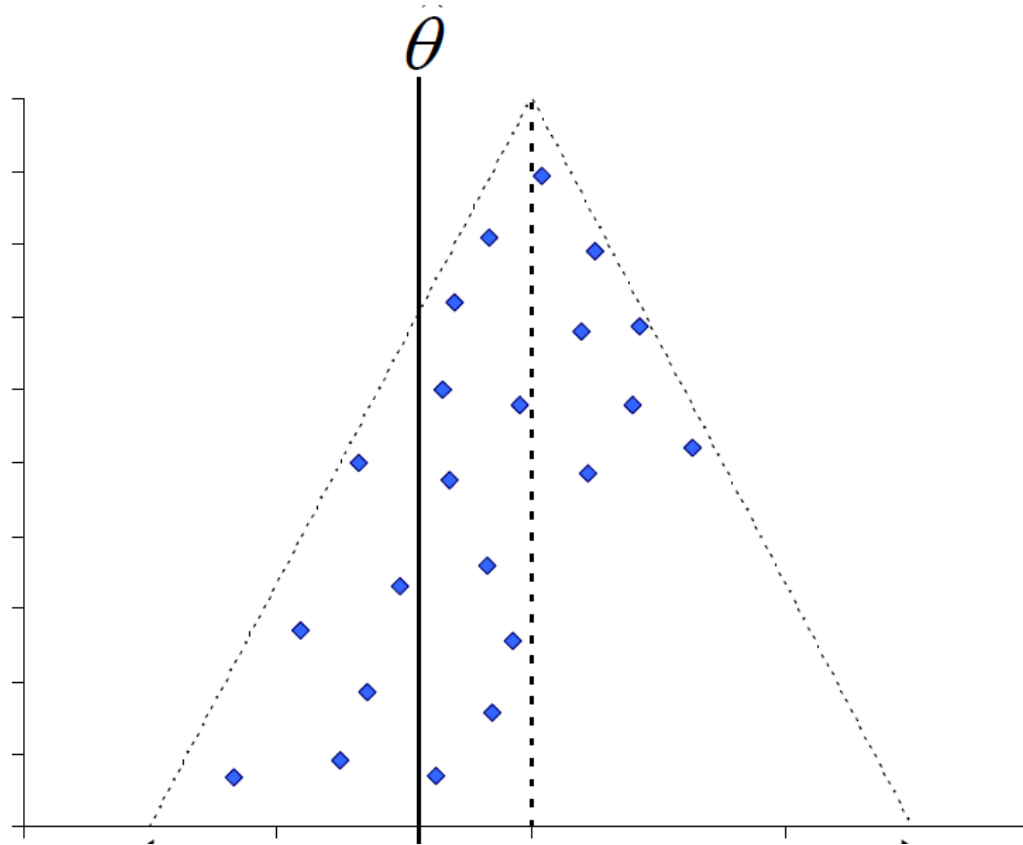
Funnel plot

- In the absence of bias and between study heterogeneity, the scatter will be due to sampling variation alone and the plot will resemble a symmetrical inverted funnel

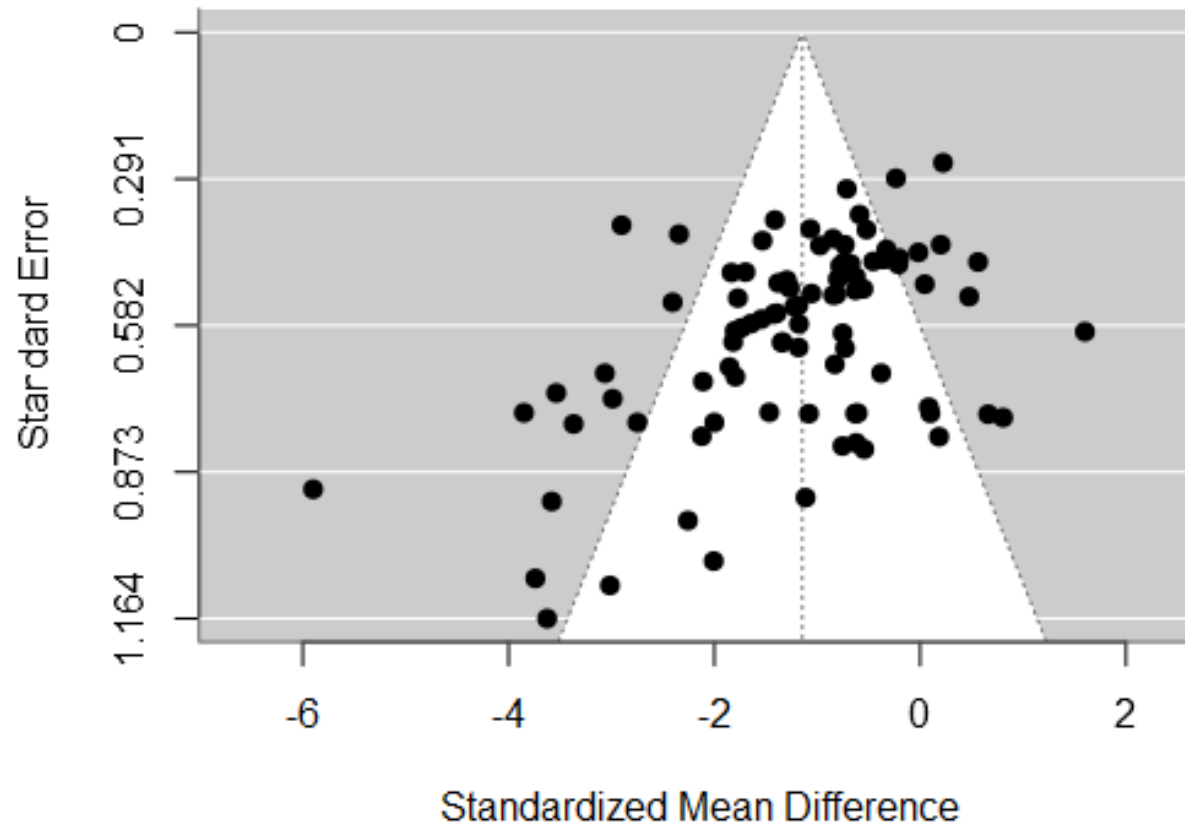


Funnel plot

- Problematic example:



Funnel plot for our data



Funnel plot in preclinical data

- In small samples and when using SMD, funnel plots tend to overrepresent differences (Zwetsloot et al.)
- Artificial biases
- Alternative: use other measure of precision but SE
- E.g. sample size (-> inverse of N)

Zwetsloot PP, Van Der Naald M, Sena ES, Howells DW, IntHout J, De Groot JA, Chamuleau SA, MacLeod MR, Wever KE. Standardized mean differences cause funnel plot distortion in publication bias assessments. *elife*. 2017 Sep 8;6:e24260.

9a Exercise

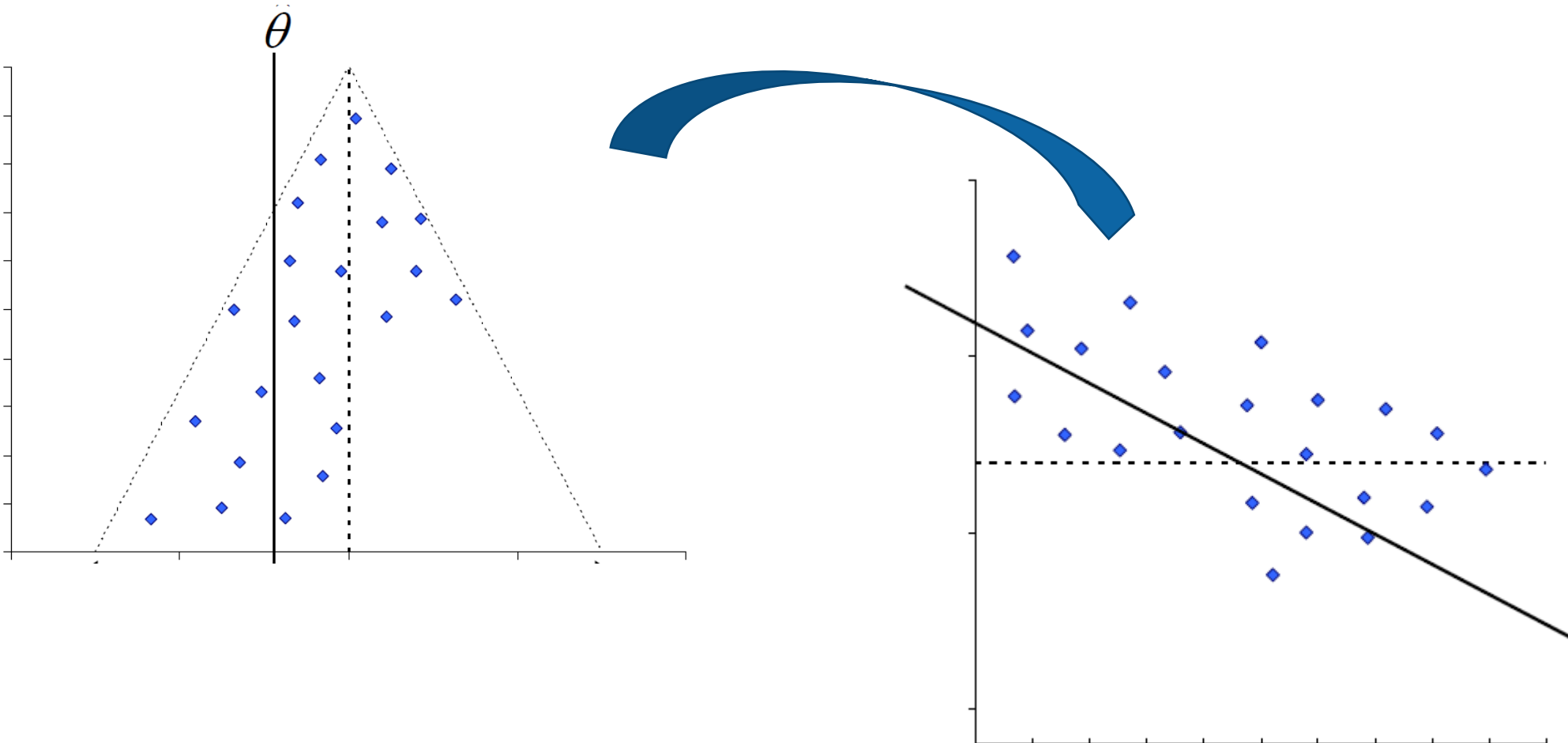
R Markdown Script
Funnel plotting

- Use the inverse:
`yaxis = "sqrtninv"`



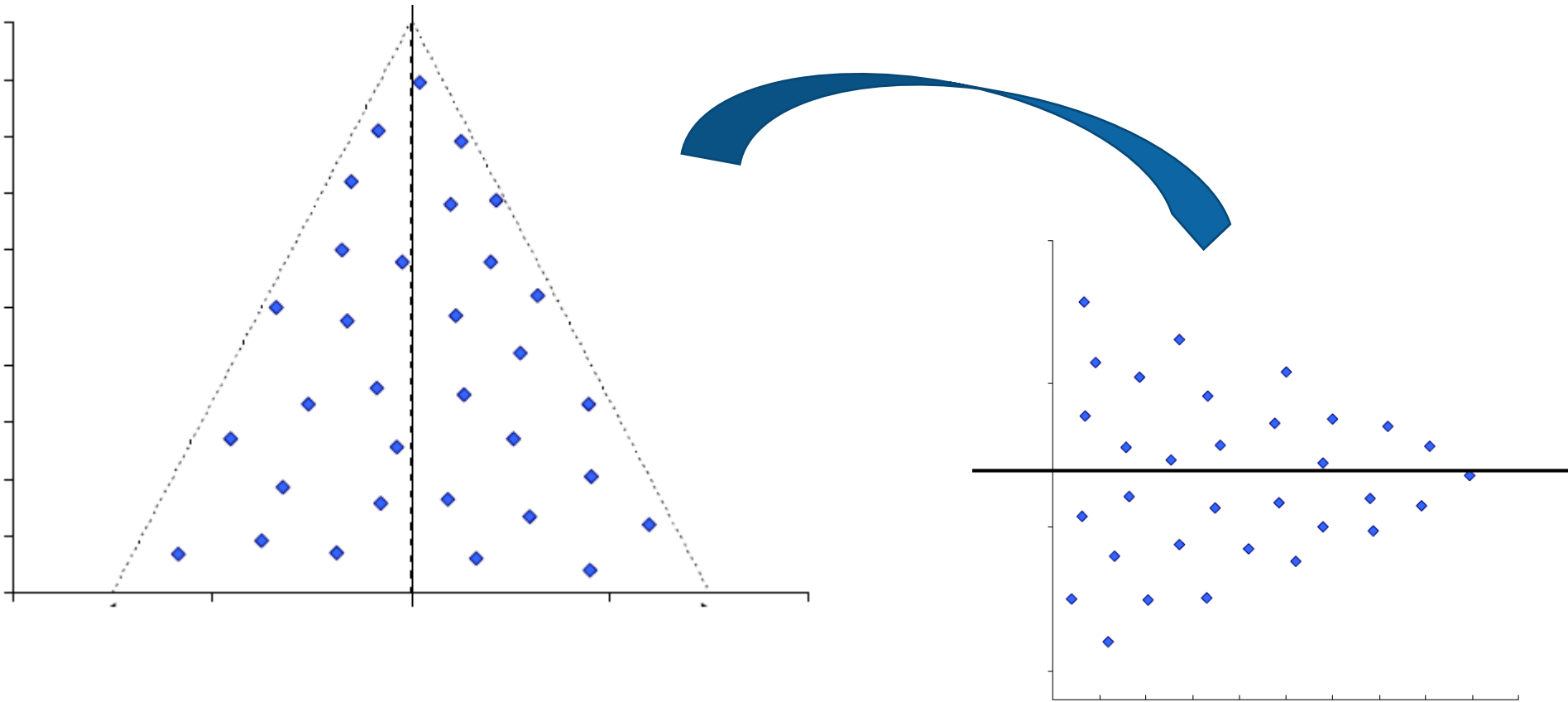
Egger's Regression

A regression line thru the flipped funnel plot (y = effect size, x = precision)



Egger's Regression

Without bias, the regression line is flat (= no slope, slope = 0)



Exercise

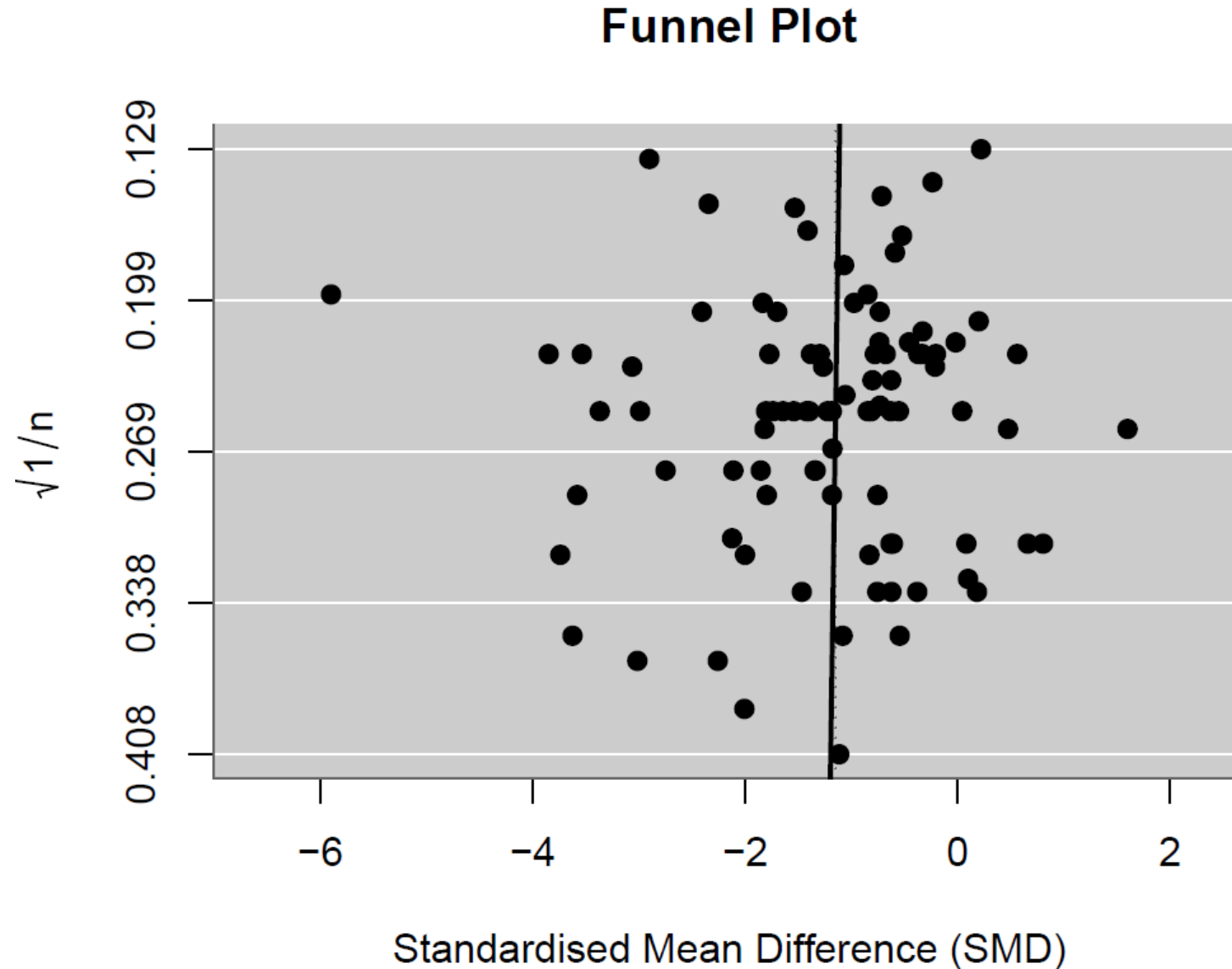
R Markdown Script

Calculate an Egger's regression

- `regtest()`
- Add the line to the funnel (code is prepared)



Egger's Regression



Trim & Fill

- How many studies would be needed to balance an unbalanced funnel plot?
- -> simulate additional study results to fill the gaps