#### **EWAS-fusion**

# Epigenomewide association statistics (EWAS) for Functional Summary-based Imputation (FUSION) association and joint/conditional analyses

Transcriptomewide association statistic (TWAS) was originally proposed for gene expression data. For a given Trait of interest T for which GWAS summary statistics  $\mathbf{z}_T$  is available, the corresponding Wald statistic for TWAS is defined such that

$$z_{\text{TWAS}} = \frac{w'_{\text{ge}} z_T}{\sqrt{w'_{\text{ge}} V w_{\text{ge}}}}$$

where  $w_{\rm ge}$  is a weight associated with gene expression and V covariance matrix for  $z_T$ , respectively. By analogy, an epigenomewide association statistic (EWAS) is defined through methylation data so that

$$z_{\text{EWAS}} = \frac{w'_{\text{me}} z_T}{\sqrt{w'_{\text{me}} V w_{\text{me}}}}$$

where  $w_{\rm me}$  is the weight associated with methylation. Both approaches allow for imputation using GWAS summary statistics. The derivation of these weights and imputation were done using methods as described in Gusev et al. (2016) called TWAS as well as in Mancuso et al. (2016) called Functional Summary-based Imputation (FUSION). The TWAS statistics from both approaches agreed very well.

A total of 442,920 CpG sites based on Illumina humanmethylation450 chips on 1,.146 individuals in EPIC-Norfolk study were available. Among these, 1,117 individuals also had genotype data from Affymetrix BioBank Axiom chips. HapMap2 SNPs from genetic data of these individuals were extracted via PLINK2 according to cis-positions of each probe and subsequently used to build weight analogous to gene expression data as implemented in computer software TWAS. We filtered probes according to their heritabilities estimated from software GCTA at significant level of 0.01. We then performed EWAS for given GWAS summary statistics. The weight generation and methylation imputation was implemented in software called TWAS-pipeline, which allows for whole epigenome computation. After filtering, 78,133 probes reached significant level 0.01.

The FUSION framework has several advantages: First, it integrates heritability estimation and covariate adjustment for whole-chromosomes with additional models such as LASSO, elastic net, BLUP. Second, it offers cross-validation, joint/conditional analyses with the output also informing top hit SNPs and inferred methylation quantitative trait locus (meQTL). Besides, the new software uses modified GCTA software (gcta\_nr\_robust) leading to higher yield of probes with heritabilities reaching statistical significance, GEMMA giving BSLMM estimates and ability to align strands with reference panels. As both the increased number of models and cross-validation led to excessive computing time, we dropped BSLMM models and conducted five cross-validations. As a result our reference panel for EWAS imputation contains 79,569 probes reaching the heritability p value threshold of

0.01. The association as well as joint/conditional analysis using our weights and LD panel is implemented in software called EWAS-fusion. Like the original TWAS, our implementation will enable a range of GWAS summary statistics to be used coupled with downstream analysis.

## **Requirements**

To begin, the software FUSION including dependencies such as plink2R and reshape is required. The latest version also requires jlimR. Other facilities to be required are

- 1. Sun grid engine (sge) or GNU parallel for Linux clusters.
- 2. Weight files based on epigenetic data.

#### FILE Description

EWAS/ directory for EWAS weights

EWAS.bim SNP information file

glist-hg19 Probe list

LDREF/ Reference for LD RDat.pos Definition of regions

RDat.profile\* Probe profiles

### Input

The input file contains GWAS summary statistics similar to .sumstats as in LDSC with the following columns.

Column	Name	Description
1	SNP	RS id of SNPs
2	A1	Effect allele (first allele)
3	A2	Other allele (second allele)
4	Z	Z-scores, taking sign with repect to A1

# Use of the programs

ewas-fusion.sh input-file

These will send jobs to the Linux clusters. The sge error and output, if any, should be called EWAS.e and EWAS.o in your HOME directory.

## Output

The results will be in input-file.tmp/ directory.

<sup>\*</sup> It contains information about the probes but not directly involved in the association analysis. For annotation of the results, it is assumed that HumanMethylation450\_15017482\_v1-2.csv is available from the directory containing ewasannotate.R.

#### **Annotation**

This is furnished with contribution from Dr Alexia Cardona, alexia.cardona@mrc-epid.cam.ac.uk, as follows,

```
ewas-annotate.R input-file.tmp
```

This reads HumanMethylation450\_15017482\_v1-2.csv from directory containing ewas-annotate.R but this can be at different location

```
ewas-annotate.R input-file.tmp manifest location=/at/different/location
```

Q-Q and Manhattan plots using R/gap can be obtained from

```
ewas-plot.R input-file.tmp
```

## **Example**

The script test.sh uses height data from GIANT. It downloads and generates an input file called height to ewas-fusion.sh.

```
ewas-fusion.sh height
```

The results will be in height.tmp/ once it is done.

The annotation is done with

```
ewas-annotate.R height.tmp
```

The Q-Q and Manhattan plots are generated with

```
ewas-plot.R height.tmp
```

# Weight generation

This is a revised and much simplified implementation of codes available from TWAS-pipeline. Under our sge it is furnished with

```
qsub get_weight.qsub
```

or

qsub get\_weight.qsub 22

for chromosome 22.

Inputs to these are summarised as follows,

File Description

FUSION.pheno PLINK phenotype file containing data for all probes
FUSION.covar PLINK covariate file containing covariates such as PCs
CpG.txt CpG ID, missing data indicator, chromosome and position

In addition, PLINK binary pedigree file for each CpG also requires to be prepared, as in files. Although it was done, it is possible to use code as in 1KG.sh to get around gerneration of these individual files by using a combined one. Note the setup takes advantage of the compact storage of non-genetic data.

The results will be available from the weights directory to be profiled and used for association analysis above.

## **Acknowledgements**

We wish to thank colleagues and collaborators for their invaluable contributions to make this work possible.

#### References

Gusev A, et al. (2016). Integrative approaches for large-scale transcriptome-wide association studies. Nature Genetics, 48, 245-252

Mancuso N, et al. (2017). Integrating gene expression with summary association statistics to identify susceptibility genes for 30 complex traits. American Journal of Human Genetics, 2017, 100, 473-487, http://www.cell.com/ajhg/fulltext/S0002-9297(17)30032-0.

Turner SD (2014). qqman: an R package for visualizing GWAS results using Q-Q and manhattan plots. biorXiv DOI: 10.1101/005165

Wood AR, et al. (2014). Defining the role of common variation in the genomic and biological architecture of adult human height (2014). Nature Genetics 46, 1173-1186.

Zhao JH (2007). gap: Genetic Analysis Package. Journal of Statistical Software 23(8):1-18, http://www.jstatsoft.org/v23/i08 (version at CRAN).

# **Appendix**

Additional information for Illumina infinium humanmethylation450 beadchip as in Illumina website

Column Name Description Index Probe Index

TargetID Identifies the probe

name. Also used as a key column for data

import.

ProbeID\_A Illumina identifier for

probe sequence A

ProbeID B Illumina identifier for

probe sequence B

IlmnID Unique CpG locus

identifier from the

Illumina CG database

Name Unique CpG locus

identifier from the Illumina CG database

Address A\_ID Address of probe A

AlleleA\_ProbeSeq Sequence for probe A AddressB\_ID Address of probe B

AlleleB\_ProbeSeq Sequence for probe B

Infinium\_Design\_Type Defines Assay type -

Infinium I or Infinium

II

Next\_Base Base added at SBE

step - Infinium I assays only

Color\_Channel Color of the

incorporated baseá (Red or Green) -

Infinium I assays only

Forward\_Sequence Sequence (in 5'-3'

orientation) flanking

query site

Genome\_Build Genome build on

which forward sequence is based

CHR Chromosome -

genome build 37

MAPINFO Coordinates - genome

build 37

SourceSeq Unconverted design

sequence

Chromosome - Chromosome -

genome build 36

Coordinate\_36 Coordinates - genome

build 36

Strand Design strand

Probe\_SNPs Assays with SNPs

present within probe >10bp from query

site

Probe\_SNPs\_10 Assays with SNPs

present within probe ?10bp from query

site (HM27 carryover

or recently discovered)

Random\_Loci Loci which were

chosen randomly in the design process

Methyl27\_Loci Present or absent on

HumanMethylation27

array

UCSC\_RefGene\_Name Gene name (UCSC)
UCSC\_RefGene\_Accession Accession number

(UCSC)

UCSC\_RefGene\_Group Gene region feature

category (UCSC)

UCSC\_CpG\_Islands\_Name CpG island name

(UCSC)

Relation\_to\_UCSC\_CpG\_Island Relationship to

Canonical CpG Island: Shores - 0-2 kb from CpG island; Shelves -2-4 kb from CpG

island.

Phantom FANTOM-derived

promoter

DMR Differentially

methylated region (experimentally determined)

Enhancer Enhancer element

(informatically-determined)

HMM\_Island Hidden Markov

Model Island

Regulatory\_Feature\_Name Regulatory feature

(informatically determined)

Regulatory\_Feature\_Group Regulatory feature

category

DHS DNAse

hypersensitive site (experimentally determined)