# WordCloud in Biology with tm and ggwordcloud

#### Camila Farias Amorim

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## 1) Load required packages, color palette and gg theme

```
library(tm) # for text mining
library(ggwordcloud) # for plotting the wordcloud
library(tidyverse)

library(jcolors)
pal9 <- jcolors("pal9")
theme_set(theme_minimal())</pre>
```

### 2) AC1 - Import the members (genes) and text mining

```
term1 <- "Signaling by Interleukins"
## Load the text:
AC1 <- readLines("import_export/annotation_cluster1")</pre>
## [1] "IL11, ANXA1, CXCL8, CSF2, MMP1, CCL3L3, IL24, OSM, LILRA3, LILRA5, IL1A, IL1B, CCL4, CCL3"
## [2] "IL21, IL11, ANXA1, CXCL8, CSF2, MMP1, CCL3L3, IL24, OSM, IL1A, IFNG, IL1B, CCL4, CCL3, S100A12"
## [3] "IL1A, CXCL8, CSF2, IL1B, CCL3L3, CCL4, CCL3"
## [4] "IL21, IL11, TNFRSF6B, ANXA1, CXCL8, CSF2, MMP1, CCL3L3, IL24, OSM, ISG15, IL1A, IFNG, IL1B, CCL
## [5] "IL1A, CXCL8, ANXA1, MMP1, IL1B, OSM"
## [6] "IL1A, IL1B, OSM"
## [7] "IL1A, CXCL8, IL1B, CCL3L3, IL24, OSM"
## [8] "IL1A, IL11, IL1B, OSM"
AC1 <- Corpus(VectorSource(AC1)) # Load the data as a corpus (a modified list)
inspect(AC1)
## <<SimpleCorpus>>
## Metadata: corpus specific: 1, document level (indexed): 0
## Content: documents: 8
## [1] IL11, ANXA1, CXCL8, CSF2, MMP1, CCL3L3, IL24, OSM, LILRA3, LILRA5, IL1A, IL1B, CCL4, CCL3
## [2] IL21, IL11, ANXA1, CXCL8, CSF2, MMP1, CCL3L3, IL24, OSM, IL1A, IFNG, IL1B, CCL4, CCL3, S100A12
## [3] IL1A, CXCL8, CSF2, IL1B, CCL3L3, CCL4, CCL3
## [4] IL21, IL11, TNFRSF6B, ANXA1, CXCL8, CSF2, MMP1, CCL3L3, IL24, OSM, ISG15, IL1A, IFNG, IL1B, CCL4
## [5] IL1A, CXCL8, ANXA1, MMP1, IL1B, OSM
## [6] IL1A, IL1B, OSM
## [7] IL1A, CXCL8, IL1B, CCL3L3, IL24, OSM
```

```
## [8] IL1A, IL11, IL1B, OSM
## Text transformation:
\#toSpace \leftarrow content \ transformer(function (x, pattern), qsub(pattern, "", x))
#AC1 <- tm map(AC1, toSpace, "/")
AC1 <- tm_map(AC1, removePunctuation) # Remove punctuation
inspect(AC1)
## <<SimpleCorpus>>
## Metadata: corpus specific: 1, document level (indexed): 0
## Content: documents: 8
##
## [1] IL11 ANXA1 CXCL8 CSF2 MMP1 CCL3L3 IL24 OSM LILRA3 LILRA5 IL1A IL1B CCL4 CCL3
## [2] IL21 IL11 ANXA1 CXCL8 CSF2 MMP1 CCL3L3 IL24 OSM IL1A IFNG IL1B CCL4 CCL3 S100A12
## [3] IL1A CXCL8 CSF2 IL1B CCL3L3 CCL4 CCL3
## [4] IL21 IL11 TNFRSF6B ANXA1 CXCL8 CSF2 MMP1 CCL3L3 IL24 OSM ISG15 IL1A IFNG IL1B CCL4 CCL3 S100A12
## [5] IL1A CXCL8 ANXA1 MMP1 IL1B OSM
## [6] IL1A IL1B OSM
## [7] IL1A CXCL8 IL1B CCL3L3 IL24 OSM
## [8] IL1A IL11 IL1B OSM
## Build a term-document matrix and model frequency data
AC1 <- TermDocumentMatrix(AC1) # Calculates sparsity and word frequencies
inspect(AC1)
## <<TermDocumentMatrix (terms: 19, documents: 8)>>
## Non-/sparse entries: 72/80
                     : 53%
## Sparsity
## Maximal term length: 8
## Weighting
                  : term frequency (tf)
## Sample
                      :
##
           Docs
## Terms
           1 2 3 4 5 6 7 8
     anxa1 1 1 0 1 1 0 0 0
##
##
     ccl3
           1 1 1 1 0 0 0 0
     ccl313 1 1 1 1 0 0 1 0
##
##
    ccl4 1 1 1 1 0 0 0 0
##
    csf2 1 1 1 1 0 0 0 0
     cxcl8 1 1 1 1 1 0 1 0
##
##
     il11 1 1 0 1 0 0 0 1
     illa 1111111
##
##
    il1b 1 1 1 1 1 1 1 1
##
    osm
           1 1 0 1 1 1 1 1
AC1 <- as.matrix(AC1)
rownames(AC1) <- str_to_upper(rownames(AC1)) # convert back to capital
AC1 <- sort(rowSums(AC1), decreasing=TRUE)
AC1 <- data.frame(word = names(AC1), freq=AC1) # new modeled data
AC1$freq <- as.numeric(AC1$freq)
AC1 <- arrange(AC1, desc(freq))
AC1$word <- as.factor(AC1$word)
AC1
##
                word freq
```

## IL1A

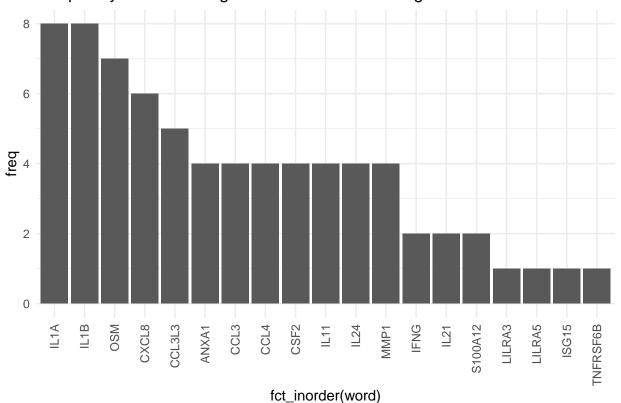
IL1A

```
## IL1B
                 IL1B
                          8
## OSM
                  OSM
                          7
## CXCL8
                CXCL8
## CCL3L3
               CCL3L3
                          5
## ANXA1
                ANXA1
                          4
## CCL3
                 CCL3
                          4
## CCL4
                 CCL4
                          4
## CSF2
                 CSF2
                          4
## IL11
                 IL11
                          4
                 IL24
## IL24
## MMP1
                 MMP1
                          2
## IFNG
                 IFNG
## IL21
                 IL21
                          2
## S100A12
              S100A12
                          2
## LILRA3
               LILRA3
                          1
## LILRA5
               LILRA5
                          1
## ISG15
                ISG15
                          1
## TNFRSF6B TNFRSF6B
```

## 3) AC1 - plotting

```
AC1 %>%
  ggplot(., aes(x=fct_inorder(word), y=freq)) +
  geom_bar(stat = "identity") +
  theme(axis.text.x = element_text(angle = 90, hjust = 1, vjust = 0.5)) +
  ggtitle("Frequency in which the gene was annotated to a geneset in AC1")
```

#### Frequency in which the gene was annotated to a geneset in AC1



```
AC1_plot <- AC1 %>%
  add_column(Gene = "yes") %>%
  ggplot(., aes(label = word, size = freq, color=Gene)) +
  scale_radius(range = c(1, 10), limits = c(0, NA)) +
  geom_text_wordcloud(area_corr = TRUE) +
  scale_size_area(max_size = 24) +
  scale_color_manual(values = pal9[1]) +
  ggtitle(term1) +theme(plot.title = element_text(hjust = 0.5, color = pal9[1]))
AC1_plot
```

Signaling by Interleukins



## 4) AC2 and AC3

```
# AC2
term2 <- "Innate immune responses and Neutrophil degranulation"
AC2 <- readLines("import_export/annotation_cluster2")
AC2 <- Corpus(VectorSource(AC2))
AC2 <- tm_map(AC2, removePunctuation)
AC2 <- TermDocumentMatrix(AC2) #from tm
AC2 <- as.matrix(AC2)
rownames(AC2) <- str_to_upper(rownames(AC2))</pre>
AC2 <- sort(rowSums(AC2), decreasing=TRUE)
AC2 <- data.frame(word = names(AC2), freq=AC2)
AC2$freq <- as.numeric(AC2$freq)
AC2 <- arrange(AC2, desc(freq))
AC2$word <- as.factor(AC2$word)
AC2_plot <- AC2 %>%
  add column(Gene = "yes") %>%
  ggplot(., aes(label = word, size = freq, color=Gene)) +
  scale_radius(range = c(1, 10), limits = c(0, NA)) +
  geom_text_wordcloud(area_corr = TRUE) +
  scale_size_area(max_size = 24) +
  scale_color_manual(values = pal9[2]) +
  ggtitle(term2) +theme(plot.title = element_text(hjust = 0.5, color = pal9[2]))
```

```
# AC3:
term3 <- "Chemotaxis"
AC3 <- readLines("import_export/annotation_cluster3")
AC3 <- Corpus(VectorSource(AC3))
AC3 <- tm_map(AC3, removePunctuation)
AC3 <- TermDocumentMatrix(AC3) #from tm
AC3 <- as.matrix(AC3)
rownames(AC3) <- str to upper(rownames(AC3))</pre>
AC3 <- sort(rowSums(AC3), decreasing=TRUE)
AC3 <- data.frame(word = names(AC3), freq=AC3)
AC3$freq <- as.numeric(AC3$freq)
AC3 <- arrange(AC3, desc(freq))
AC3$word <- as.factor(AC3$word)
AC3_plot <- AC3 %>%
  add_column(Gene = "yes") %>%
  ggplot(., aes(label = word, size = freq, color=Gene)) +
  scale_radius(range = c(1, 10), limits = c(0, NA)) +
  geom_text_wordcloud(area_corr = TRUE) +
  scale_size_area(max_size = 24) +
  scale_color_manual(values = pal9[3]) +
  ggtitle(term3) +theme(plot.title = element_text(hjust = 0.5, color = pal9[3]))
```

#### 7) Proposed final image

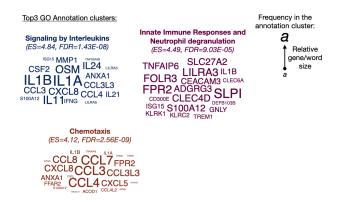


Figure 1: .

#### 8) Highlight specific genes of interest

```
AC1 %>%
  add_column(Gene = "yes") %>%
  mutate(IL1_cytokines = case_when(
    word %in% c("IL1A", "IL1B") ~ "biomarker",
    TRUE ~ "not_biomarker")) %>%
  ggplot(., aes(label = word, size = freq, color=IL1_cytokines)) +
  scale_radius(range = c(1, 10), limits = c(0, NA)) +
  geom_text_wordcloud(area_corr = TRUE) +
  scale_size_area(max_size = 24) +
```

```
scale_color_manual(values = c(pal9[4],pal9[6])) +
ggtitle(term1) + theme(plot.title = element_text(hjust = 0.5))
```

#### Signaling by Interleukins



#### Session Info

Session Info: R version 4.1.2 (2021-11-01) Platform: x86\_64-apple-darwin17.0 (64-bit) Running under: macOS Big Sur 10.16

Matrix products: default BLAS: /Library/Frameworks/R.framework/Versions/4.1/Resources/lib/libRblas.0.dylib LAPACK: /Library/Frameworks/R.framework/Versions/4.1/Resources/lib/libRlapack.dylib

locale: [1] en\_US.UTF-8/en\_US.UTF-8/en\_US.UTF-8/C/en\_US.UTF-8/en\_US.UTF-8

attached base packages: [1] stats graphics grDevices utils datasets methods base

other attached packages: [1] jcolors\_0.0.4 forcats\_0.5.1 stringr\_1.4.0

- [4] dplyr\_1.0.8 purrr\_0.3.4 readr\_2.1.2
- [7] tidyr\_1.2.0 tibble\_3.1.6 tidyverse\_1.3.1
- [10] ggwordcloud 0.5.0.9000 ggplot2 3.3.5 tm 0.7-8
- [13] NLP\_0.2-1

loaded via a name space (and not attached): [1] Rcpp\_1.0.8.3 lubridate\_1.8.0 png\_0.1-7 assert that\_0.2.1 [5] digest\_0.6.29 utf8\_1.2.2 slam\_0.1-50 R6\_2.5.1

- [9] cellranger 1.1.0 backports 1.4.1 reprex 2.0.1 evaluate 0.15
- [13] highr 0.9 httr 1.4.2 pillar 1.7.0 rlang 1.0.2
- [17] readxl\_1.3.1 rstudioapi\_0.13 rmarkdown\_2.13 labeling\_0.4.2
- [21] munsell 0.5.0 gridtext 0.1.4 broom 0.7.12 compiler 4.1.2
- [25] modelr\_0.1.8 xfun\_0.30 pkgconfig\_2.0.3 htmltools\_0.5.2 [29] tidyselect\_1.1.2 fansi\_1.0.2 crayon\_1.5.0 tzdb\_0.2.0
- [33] dbplyr\_2.1.1 withr\_2.5.0 grid\_4.1.2 jsonlite\_1.8.0
- [37] gtable 0.3.0 lifecycle 1.0.1 DBI 1.1.2 magrittr 2.0.2
- [41] scales\_1.1.1 cli\_3.2.0 stringi\_1.7.6 farver\_2.1.0
- [45] fs\_1.5.2 xml2\_1.3.3 ellipsis\_0.3.2 generics\_0.1.2
- [49] vctrs\_0.3.8 tools\_4.1.2 glue\_1.6.2 markdown\_1.1
- [53] hms\_1.1.1 parallel\_4.1.2 fastmap\_1.1.0 yaml\_2.3.5
- [57] colorspace\_2.0-3 rvest\_1.0.2 knitr\_1.37 haven\_2.4.3