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# set working directory
setwd("D:/STORAGE/College Work/year 4/Y4Q3/MATH 189/hw/hw3")
# read data
data <- read.table("hcmv.txt", header=TRUE)</pre>
head (data)
# Investigation 1
# global variables
numPalindromes <- 296
dnaLen <- 229354
binSeq < - seq(0, 232000, 4000)
# Graphing the observed distribution
hist(data[['location']], breaks = binSeq, ylim =c(0, 15),
     col = 'grey',
     main = "Distribution of Counts of Palindromes Bins=4000",
     xlab = 'Count intervals')
abline(a=296/58, b=0, lw = 3)
# Graphing several uniform distributions
# H1
sampleUnif = runif(numPalindromes, min = 0, max = length(binSeq))
h1 <- hist(sampleUnif, breaks= length(binSeq), plot = FALSE,</pre>
           main = 'Sample from Uniform Dist',
           xlab = 'Palindrome Occurrences per ')
# H2
sampleUnif = runif(numPalindromes, min = 0, max = length(binSeq))
h2 <- hist(sampleUnif, breaks= length(binSeq), plot = FALSE,
           main = 'Sample from Uniform Dist',
           xlab = 'Palindrome Occurrences per ')
# H3
sampleUnif = runif(numPalindromes, min = 0, max = length(binSeq))
h3 <- hist(sampleUnif, breaks= length(binSeq), plot = FALSE,
           main = 'Sample from Uniform Dist',
           xlab = 'Palindrome Occurrences per ')
# H4
sampleUnif = runif(numPalindromes, min = 0, max = length(binSeq))
h4 <- hist(sampleUnif, breaks= length(binSeq), plot = FALSE,
           main = 'Sample from Uniform Dist',
           xlab = 'Palindrome Occurrences per ')
# H5
sampleUnif = runif(numPalindromes, min = 0, max = length(binSeq))
h5 <- hist(sampleUnif, breaks= length(binSeq), plot = FALSE,
           main = 'Sample from Uniform Dist',
           xlab = 'Palindrome Occurrences per ')
# Setting up transparent colors for graph
c1 <- col2rgb("lightblue")</pre>
mycol1 \leftarrow rgb(c1[1], c1[2], c1[3], max = 255, alpha = 70, names = "blue50")
c1 <- col2rgb("pink")</pre>
mycol2 < - rgb(c1[1], c1[2], c1[3], max = 255, alpha = 70, names = "blue50")
c1 <- col2rab("lightvellow")</pre>
mycol3 \leftarrow rgb(c1[1], c1[2], c1[3], max = 255, alpha = 70, names = "blue50")
c1 <- col2rgb("lightgreen")</pre>
mycol4 \leftarrow rgb(c1[1], c1[2], c1[3], max = 255, alpha = 70, names = "blue50")
c1 <- col2rgb("violet")</pre>
mycol5 <- rgb(c1[1], c1[2], c1[3], max = 255, alpha = 70, names = "blue50")
plot(h1, col = mycol1, ylim = c(0, 18), xlab = 'Bins (Size 4000 Intervals)')
plot(h2, col = mycol2, add = TRUE)
plot(h3, col = mycol3, add = TRUE)
plot(h4, col = mycol4, add = TRUE)
plot(h5, col = mycol5, add = TRUE)
abline (a=296/58, b=0, lw = 3)
# Investigation 2
# Complementary palidrome is one type of pattern in DNA that
\ensuremath{\sharp} can be indiciative of an important site in DNA, such as the origin of
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# replication.
# A is complementary to T
# G is complementary to C
# A complementary palindrome is a sequence of letters that
# read in reverse as the complement of the forward sequence.
library(lattice)
stripplot(data$location, pch=1, cex=0.25)
hist(data$location, breaks=nrow(data)*400)
# Goals: Graphically examine the distribution of your sample spacings
# There are 3 types of spacings to examine:
    spacings between consecutive palindromes
   spacings between palindromes with one in between
      (i.e. sums of pairs of consecutive spacings)
    spacings between palindromes with two in between
      (i.e. sums of triplets of consecutive spacings)
# Graphically compare these 3 types of spacings to those
# that come from random uniform scatter (using empirical cdf or histograms)
# create a random uniform scatter
N <- 200000
n <- nrow(data)
gene <- seq(1, N)
# set.seed(100)
site.random <- sort(sample.int(N, size=n, replace=FALSE))</pre>
stripplot(site.random, pch=16, cex=0.25)
rand scatt = diff(site.random)
# stripplot(rand_scatt, pch=1, cex=0.25)
# hist(rand_scatt)
barplot(rand_scatt)
plot(ecdf(rand_scatt))
graph_mixed_plot <- function(data, title, subt) {</pre>
  data = diff(data)
 hist(data, xlim=c(0, max(data)), main=paste(title, '\n', subt))
  # mtext(sode=3, line=3, at=-0.07, adj=0, cex=1, title)
  # mtext(sode=3, line=3, at=-0.07, adj=0, cex=1, title)
  par(new = T)
  plot(ecdf(data),
       xlim=c(0, max(data)),
       col=rgb(0, 0, 0, alpha=0),
       axes=F.
       xlab=NA,
       ylab=NA,
      main=NA)
  lines(ecdf(data))
# spacings between consecutive palindromes
palin_consec = data$location
graph mixed plot (palin consec,
                 "Consecutive palindrome distance distribution",
                 "Population data")
# Random uniform scatter comparisons
site.random <- sort(sample.int(N, size=n, replace=FALSE))</pre>
graph_mixed_plot(site.random,
                 "Consecutive palindrome distance distribution",
                 "Uniform Sampled data 1")
site.random <- sort(sample.int(N, size=n, replace=FALSE))</pre>
graph_mixed_plot(site.random,
                 "Consecutive palindrome distance distribution",
                 "Uniform Sampled data 2")
pnorm((mean(site.random) - mean(palin_consec)) / (sd(palin_consec) / length(palin_consec)))
mean(site.random) - mean(palin consec)
# spacings between palindromes with one in between
palin one sep = data$location[seq(0, nrow(data), 2)]
graph mixed plot(palin one sep,
                 "Distribution of palindrome spacing with 1 between",
                 "Population data")
# Random uniform scatter comparisons
site.random <- sort(sample.int(N, size=n, replace=FALSE))[seq(0, nrow(data), 2)]</pre>
graph mixed plot(site.random,
                 "Distribution of palindrome spacing with 1 between",
                 "Uniform Sampled data 1")
site.random <- sort(sample.int(N, size=n, replace=FALSE))[seq(0, nrow(data), 2)]</pre>
graph_mixed_plot(site.random,
                 "Distribution of palindrome spacing with 1 between",
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"Uniform Sampled data 2")
# spacings between palindromes with two in between
palin two sep = data$location[seq(0, nrow(data), 3)]
graph_mixed_plot(palin_two_sep,
                 "Distribution of palindrome spacing with 2 between",
                 "Population data")
# Random uniform scatter comparisons
site.random <- sort(sample.int(N, size=n, replace=FALSE))[seq(0, nrow(data), 3)]</pre>
graph_mixed_plot(site.random,
                 "Distribution of palindrome spacing with 2 between",
                 "Uniform Sampled data 1")
site.random <- sort(sample.int(N, size=n, replace=FALSE))[seq(0, nrow(data), 3)]</pre>
graph_mixed_plot(site.random,
                 "Distribution of palindrome spacing with 2 between",
                 "Uniform Sampled data 2")
# Investigation 3
get bin counts <- function(data vec, bins, subt) {</pre>
 bin counts <- c()
  bin_names <- c()</pre>
 bin_size <- (max(data_vec) / bins)</pre>
  for (i in seq(bins)) {
    lower_bound = (bin_size * (i - 1))
    upper bound = (bin size * i)
   bin_counts = append(bin_counts,
                        length(data vec[(data vec <= upper bound) & (data vec > lower bound)]))
   bin names = append(bin names, as.character(round(upper bound)))
  }
 barplot(bin_counts, names.arg=bin_names, las=2, main=paste("Location counts\n", subt))
  axis(side = 1, labels = FALSE)
 return (bin size)
bins = 100
par(mfrow=c(2, 2), pin=c(5, 3))
bin_size = get_bin_counts(data$location, bins, "Population data")
plot.new()
sample count plots <- function(data vec, bin size, subt){</pre>
 bin counts <- c()
 bin_names <- c()
  for (i in seq(bins)) {
   lower bound = (bin size * (i - 1))
    upper_bound = (bin_size * i)
    bin counts = append(bin counts,
                        length(data_vec[(data_vec <= upper_bound) & (data_vec > lower_bound)]))
   bin_names = append(bin_names, as.character(round(upper_bound)))
 barplot(bin counts, names.arg=bin names, las=2, main=paste("Location counts\n", subt))
  axis(side = 1, labels = FALSE)
site.random <- sort(sample.int(N, size=n, replace=FALSE))</pre>
sample_count_plots(site.random, bin_size, "Sample data 1")
site.random <- sort(sample.int(N, size=n, replace=FALSE))</pre>
sample_count_plots(site.random, bin_size, "Sample data 2")
qqnorm(data$location, pch = 1, frame = FALSE, main="Population Q-Q Plot")
qqnorm(site.random, pch = 1, frame = FALSE, main="Sample Uniform Distribution Q-Q Plot")
chisq.test(data$location)
chisq.test(site.random)
### Investigation 4 BIGGEST CLUSTER ###
#Does the interval with the greatest number of palindromes
#indicate a potential origin of replication? Be careful in
#making your intervals, for any small, but significant
\#deviations from random scatter, such as a tight cluster of
#a few palindromes, could easily go undetected if the regions
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#examined are too large. Also, if the regions are too small,
#a cluster of palindromes may be split between adjacent

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#intervals and not appear as a high-count interval.
locations <- read.table('hcmv.txt', header=TRUE)$location</pre>
head(locations)
# Given (see slide 13-14).
N <- 229354 \# Base pairs, or length of CMV genome
n <- 296 # Palindromes
intervals <- c(2500,4000,5500,7000)
k_{intervals} \leftarrow ceiling(N / intervals)
k_lambda_hat <- c()</pre>
k_max_count <- c()</pre>
k_p_value <- c()
for(k in k_intervals) {
  k_{\text{count}} \leftarrow \text{as.vector(table(cut(locations, breaks = seq(0, N, length.out = k+1), include.lowest = TRUE)))}
  lambda_hat <- mean(k_count)</pre>
  k lambda hat <- c(k lambda hat, lambda hat)
  k_max_count <- c(k_max_count, max(k_count))</pre>
  library(hash)
  dict <- hash()
  for (i in 0:max(k count)) {
    key <- toString(i)</pre>
   dict[[key]] <- 0
  key <- toString(max(k_count)+1)</pre>
  dict[[key]] <- 0
  for (c in k count) {
   key <- toString(c)</pre>
    dict[[key]] \leftarrow dict[[key]] + 1
  k_counts_observed <- c()</pre>
  for (i in 0:max(k_count)) {
   key <- toString(i)</pre>
    k_counts_observed <- c(k_counts_observed, dict[[key]])</pre>
  key <- toString(max(k_count)+1)</pre>
  k_counts_observed <- c(k_counts_observed, dict[[key]])</pre>
  k_expected_poisson <- c()</pre>
  for (i in 0:max(k count)) {
    \label{eq:k_expected_poisson} $$ k_expected_poisson, dpois(i, lambda_hat))$
  \verb|k_expected_poisson| <- c(k_expected_poisson, 1 - sum(k_expected_poisson))|
  \verb|k_counts_expected| <- |k_expected_poisson| * |k|
  k_chi2<- sum((k_counts_observed - k_counts_expected)^2 / k_counts_expected)</pre>
  k_{chi2} = 0.95, df = max(k_{count}) - 2
  k_p_value <- c(k_p_value, pchisq(k_chi2, df = max(k_count) - 2, lower.tail = FALSE))</pre>
result <- data.frame(intervals, k_intervals, k_lambda_hat, k_max_count, k_p_value)
result
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