Hepatoprotective effects of systemic ER activation

ER regulated genes and NAFLD classification models

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12 August, 2022

```
# source and library import
source('code/00_helper_functions.R')
library(tidyverse)
library(RColorBrewer)
library(ComplexHeatmap)
library(caret)
library(multiROC)
library(patchwork)
# color palettes
colPals <- list()</pre>
colPals$conditions <- setNames(c('#E98BB6', '#B02262', '#7F9AD7', '#2A2F72', '#7DC7D1', '#339ACD', '#35
                                c('CDf', 'HFDf', 'CDm', 'HFDm', 'DPN', 'DIP', 'E2', 'PPT'))
colPals$RdBu <- rev(RColorBrewer::brewer.pal(n=11, name = 'RdBu'))</pre>
colPals$UpDown <- setNames(colPals$RdBu[c(10,2)],
                            c('up', 'down'))
colPals$clusters <- setNames(c('#A9D265', '#82506D', '#FA9F1C', '#676A6E'),</pre>
                              c('1', '2', '3', '4'))
colPals$celltypes <- setNames(c('#B4272F', '#E5462D', '#FFD1D1', '#F4E54C', '#FBAA3E', '#AA654E', '#B58
                                 '#B3177E', '#CAC1DD', '#67227D','#36B449', '#82C349', '#A9D265', '#1994
                               c('Cholangiocytes', 'Endothelial cells', 'HsPCs', 'Stromal cells', 'Hepat
                                 'Kupffer cells', 'Monocytes & Monocyte-derived cells', 'T cells', 'NK c
                                 'ILC1s', 'B cells', 'cDC1s', 'cDC2s', 'Mig. cDCs', 'pDCs', 'Basophils',
colPals$inferno <- c('#FCFFA4', '#FCA50A', '#DD513A', '#932667', '#420A68', '#000004')
colPals$stage <- setNames(c('#F6F6F6','#FAD7A0','#7B241C'),</pre>
                           c('CTRL','NAFL','NASH'))
colPals$NAS <- setNames(c('#F6F6F6','#BFC9CA','#000000'),</pre>
                         c('low','mid','high'))
colPals$fibrosis <- setNames(c('#F6F6F6','#DF7B71','#000000'),</pre>
                              c('low','mid','high'))
```

Load data

```
# mouse-human orthologs
mouse_human_orthologs <- read.table(
  file = 'data/ensembl_mmus_hsap_sep2019_orthologs.tsv',
  sep = '\t',
  header = TRUE,
  quote = '')</pre>
```

```
DEGs <- readRDS('results/bulkRNAseq_mmus_DEGs.rds')</pre>
# RNAseq data
RNAseq <- readRDS('results/bulkRNAseq mmus data filt norm.rds')
# human cohort data from:
# Govaere et al. 2020 (https://doi.org/10.1126/scitranslmed.aba4448)
# Kozumi et al. 2021 (https://doi.org/10.1002/hep.31995)
# GTEx v8 (https://www.qtexportal.org/home/datasets)
cohort_data <- readRDS('data/bulkRNAseq_human_cohort_data.rds')</pre>
cohort_data$Govaere$cpm_filt <- cohort_data$Govaere$cpm %>%
  tibble::rownames_to_column(var = 'gene') %>%
  dplyr::filter(gene %in% mouse_human_orthologs$GeneID_human) %>%
  dplyr::mutate(gene = dplyr::recode(gene, !!!setNames(mouse_human_orthologs$GeneSymbol_human,
                                                         mouse_human_orthologs$GeneID_human))) %>%
  dplyr::filter(!duplicated(gene) & gene != "") %>%
  tibble::column_to_rownames(var = 'gene')
cohort_data$Kozumi$cpm_filt <- cohort_data$Kozumi$cpm %>%
  dplyr::filter(rownames(.) %in% mouse_human_orthologs$GeneSymbol_human
                & !duplicated(rownames(.)))
cohort_data$GTEx_liver_CTRL$cpm_filt <- cohort_data$GTEx_liver_CTRL$cpm %>%
  dplyr::filter(rownames(.) %in% mouse_human_orthologs$GeneSymbol_human
                & !duplicated(rownames(.)))
# gene sets
gene_sets <- list()</pre>
gene_sets[['ER_genes']] <- scan('results/ER_regulated_genes.txt', what = character(), quiet = T)</pre>
gene_sets[['NAS_markers']] <- scan('results/ER_regulated_genes_NAS_markers.txt', what = character(), qu</pre>
gene_sets[['fibrosis_markers']] <- scan('results/ER_regulated_genes_fibrosis_markers.txt', what = chara</pre>
gene_sets[['sulf2']] <- c('SULF2')</pre>
gene_sets[['thbs2']] <- c('THBS2')</pre>
gene_sets[['combined_sulf2_thbs2']] <- c('SULF2', 'THBS2')</pre>
```

consensus differentially expressed genes

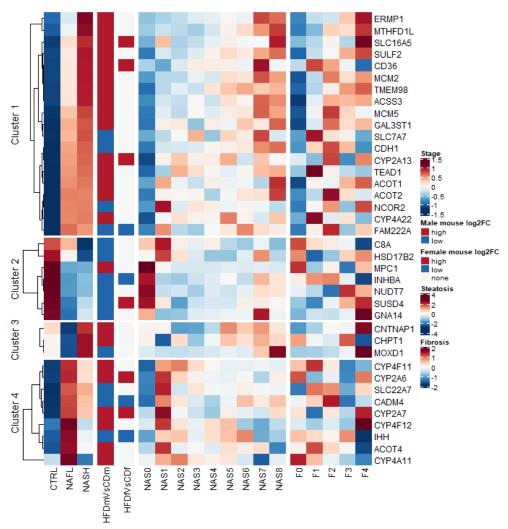
Integrated heatmap of ER regulated genes across human NAFLD spectrum

```
human_stage <- cohort_data$Govaere$cpm_filt %>%
    dplyr::filter(rownames(.) %in% gene_sets$ER_genes) %>%
    groupTransform(cohort_data$Govaere$meta$Stage, function(x) apply(x,1,median)) %>%
    scaleData(method = 'zscore') %>%
    dplyr::arrange(rownames(.))

human_steatosis <- cohort_data$Govaere$cpm_filt %>%
    dplyr::filter(rownames(.) %in% gene_sets$ER_genes) %>%
    groupTransform(cohort_data$Govaere$meta$NAS, function(x) apply(x,1,median)) %>%
    scaleData(method = 'zscore') %>%
    dplyr::arrange(rownames(.))
```

```
human_fibrosis <- cohort_data$Govaere$cpm_filt %>%
  dplyr::filter(rownames(.) %in% gene_sets$ER_genes) %>%
  groupTransform(cohort_data$Govaere$meta$Fibrosis, function(x) apply(x,1,median)) %%
  scaleData(method = 'zscore') %>%
  dplyr::arrange(rownames(.))
mouse_log2FC_HFDmVsCDm <- DEGs$filt$CDmVsHFDm %>%
  dplyr::rename(GeneSymbol mouse = external gene name) %>%
  dplyr::inner_join(mouse_human_orthologs, by = 'GeneSymbol_mouse') %>%
  dplyr::filter(GeneSymbol_human %in% gene_sets$ER_genes) %>%
  dplyr::filter(!duplicated(GeneSymbol_human)) %>%
  dplyr::select(GeneSymbol_human, log2FoldChange) %>%
  dplyr::mutate(log2FoldChange = log2FoldChange*-1) %>%
  dplyr::rename(HFDmVsCDm = log2FoldChange) %>%
  dplyr::mutate(HFDmVsCDm = factor(ifelse(HFDmVsCDm>0, 'high', 'low'), levels = c('high', 'low'))) %%
  tibble::column_to_rownames(var = 'GeneSymbol_human') %>%
  dplyr::arrange(rownames(.))
mouse_log2FC_HFDfVsCDf <- DEGs$filt$CDfVsHFDf %>%
  dplyr::rename(GeneSymbol_mouse = external_gene_name) %>%
  dplyr::inner_join(mouse_human_orthologs, by = 'GeneSymbol_mouse') %>%
  dplyr::filter(GeneSymbol_human %in% gene_sets$ER_genes) %>%
  dplyr::filter(!duplicated(GeneSymbol_human)) %>%
  dplyr::select(GeneSymbol_human, log2FoldChange) %>%
  dplyr::mutate(log2FoldChange = log2FoldChange*-1) %>%
  dplyr::rename(HFDfVsCDf = log2FoldChange) %>%
  dplyr::mutate(HFDfVsCDf = ifelse(HFDfVsCDf>0, 'high', 'low')) %>%
  dplyr::add_row(GeneSymbol_human = dplyr::setdiff(rownames(mouse_log2FC_HFDmVsCDm),
                                                   .$GeneSymbol_human),
                 HFDfVsCDf = 'none') %>%
  dplyr::mutate(HFDfVsCDf = factor(HFDfVsCDf, levels = c('high','low', 'none'))) %>%
  tibble::column_to_rownames(var = 'GeneSymbol_human') %>%
  dplyr::arrange(rownames(.))
set.seed(4)
clusters_ER_genes <- kmeans(human_stage, centers = 4, iter.max = 100)</pre>
Heatmap(human_stage, width = unit(24, "mm"), name = "Stage",
        split = clusters_ER_genes$cluster,
        col = circlize::colorRamp2(breaks=seq(-max(abs(human_stage)), max(abs(human_stage)), length.out
                                   colors=colPals$RdBu),
       row_title = c("Cluster 1", "Cluster 2", "Cluster 3", "Cluster 4"),
        cluster row slices = FALSE,
        column_order=c("CTRL", "NAFL", "NASH")) +
  Heatmap(mouse_log2FC_HFDmVsCDm, width = unit(8, "mm"), name = "Male mouse log2FC",
          col = colPals RdBu[c(10,2)]) +
  Heatmap(mouse_log2FC_HFDfVsCDf, width = unit(8, "mm"), name = "Female mouse log2FC",
          col = c(colPals RdBu[c(10,2)], '#F6F6F6')) +
  Heatmap(human_steatosis, width = unit(72, "mm"), name = "Steatosis",
          column_order=paste0('NAS', seq(0,8)),
          col = circlize::colorRamp2(breaks=seq(-max(abs(human_steatosis)), max(abs(human_steatosis)),
                                     colors=colPals$RdBu)) +
  Heatmap(human_fibrosis, width = unit(40, "mm"), name = "Fibrosis",
```

```
column_order=paste0('F', seq(0,4)),
col = circlize::colorRamp2(breaks=seq(-max(abs(human_fibrosis)), max(abs(human_fibrosis)), leg
colors=colPals$RdBu))
```



NAFLD classification models

NAFLD models

```
models[['NAS']] <- lapply(gene_sets, function(x) {</pre>
  buildModel(dat = t(cohort_data$Govaere$cpm_filt),
             group = factor(cohort_data$Govaere$meta$NAS_class, levels = c('low','mid','high')),
             method = 'glmnet',
             features = x,
             preproc = c('scale', 'center'),
             train_test_split = 0.6,
             tune_iter = 10,
             seed = 22)
})
models[['Fibrosis']] <- lapply(gene_sets, function(x) {</pre>
  buildModel(dat = t(cohort_data$Govaere$cpm_filt),
             group = factor(cohort_data$Govaere$meta$Fibrosis_class, levels = c('low', 'mid', 'high')),
             method = 'glmnet',
             features = x,
             preproc = c('scale', 'center'),
             train_test_split = 0.6,
             tune_iter = 10,
             seed = 22)
})
saveRDS(models, file = 'results/nafld_classification_models.rds')
```

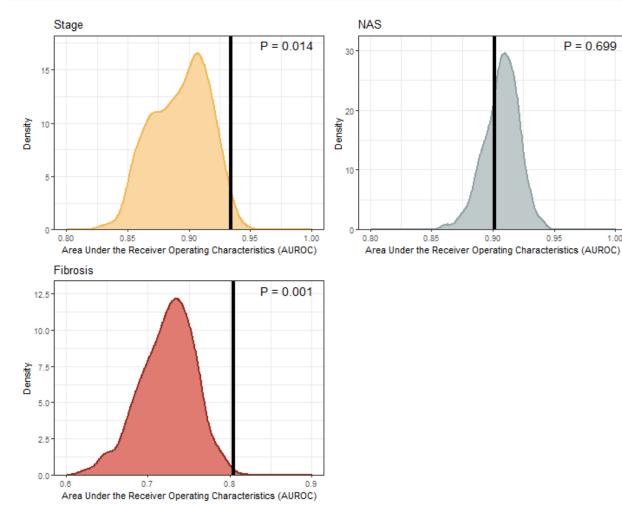
Random gene set models

```
# random gene sets for null distribution of prediction models
# all_genes <- rownames(cohort_data$Govaere$cpm_filt)</pre>
\# random_genes <- lapply(seq(1,1000), function(x) sample(x = all_genes, size = length(gene_sets$ER_gene
# saveRDS(random_genes, file = 'results/random_38gene_subsets.rds')
# load random gene sets for reproducibility
random_genes <- readRDS(file = 'results/random_38gene_subsets.rds')</pre>
# train random models (very long computing time!)
# random models <- list()</pre>
# random_models[['Stage']] <- lapply(random_genes, function(x) {</pre>
   buildModel(dat = t(cohort_data$Govaere$cpm_filt),
#
               group = factor(cohort\_data\$Govaere\$meta\$Stage, levels = c('CTRL', 'NAFL', 'NASH')),
#
               method = 'qlmnet',
#
               features = x,
#
               preproc = c('scale', 'center'),
#
               train\_test\_split = 0.6,
#
               tune_iter = 10,
#
               seed = 22)
# })
# random_models[['NAS']] <- lapply(random_genes, function(x) {</pre>
    buildModel(dat = t(cohort data$Govaere$cpm filt),
#
               group = factor(cohort\_data\$Govaere\$meta\$NAS\_class, levels = c('low', 'mid', 'high')),
#
               method = 'glmnet',
#
               features = x,
#
               preproc = c('scale', 'center'),
#
               train_test_split = 0.6,
               tune\_iter = 10,
```

```
#
                seed = 22)
# })
# random_models[['Fibrosis']] <- lapply(random_genes, function(x) {</pre>
    buildModel(dat = t(cohort_data$Govaere$cpm_filt),
#
               group = factor(cohort_data$Govaere$meta$Fibrosis_class, levels = c('low', 'mid', 'high')),
#
               method = 'glmnet',
#
               features = x,
#
               preproc = c('scale', 'center'),
#
               train_test_split = 0.6,
#
               tune_iter = 10,
#
               seed = 22)
# })
# saveRDS(random_models, file = 'Results/random_models_38genes.rds')
random_models <- readRDS(file = 'results/random_models_38genes.rds')</pre>
```

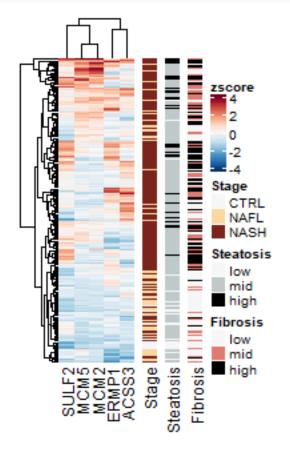
Predictive potential of ER regulated genes vs random gene sets

```
p <- list()
df <- data.frame(auc = lapply(random_models$Stage, function(x) x$resultsROC$AUC$model$micro) %>% unlist
val <- models$Stage$ER genes$resultsROC$AUC$model$micro</pre>
pval <- tailFraction(val, df$auc, tail = 'right')</pre>
p[[1]] \leftarrow ggplot(df, aes(x = auc)) +
  geom_density(lwd = 1, colour = '#F6B651',
               fill = '#FAD7AO', alpha = 1) +
  geom_vline(xintercept = val, lwd = 1.5) +
  scale_x_continuous(limits = c(0.8,1)) +
  scale_y = continuous(expand = expansion(mult = c(0, .1))) +
  annotate("text", x=Inf, y = Inf, label = paste('P =',pval), vjust=1.5, hjust=1.2, size=5) +
  xlab('Area Under the Receiver Operating Characteristics (AUROC)') +
  ylab('Density') +
  ggtitle('Stage') +
  theme bw()
df <- data.frame(auc = lapply(random_models$NAS, function(x) x$resultsROC$AUC$model$micro) %>% unlist()
val <- models$NAS$ER_genes$resultsROC$AUC$model$micro</pre>
pval <- tailFraction(val, df$auc, tail = 'right')</pre>
p[[2]] \leftarrow ggplot(df, aes(x = auc)) +
  geom_density(lwd = 1, colour = '#92A3A5',
               fill = '#BFC9CA', alpha = 1) +
  geom_vline(xintercept = val, lwd = 1.5) +
  scale_x_continuous(limits = c(0.8,1)) +
  scale_y_continuous(expand = expansion(mult = c(0, .1))) +
  annotate("text", x=Inf, y = Inf, label = paste('P =',pval), vjust=1.5, hjust=1.2, size=5) +
  xlab('Area Under the Receiver Operating Characteristics (AUROC)') +
  ylab('Density') +
  ggtitle('NAS') +
  theme_bw()
df <- data.frame(auc = lapply(random_models$Fibrosis, function(x) x$resultsROC$AUC$model$micro) %>% unl
val <- models$Fibrosis$ER_genes$resultsROC$AUC$model$micro</pre>
```

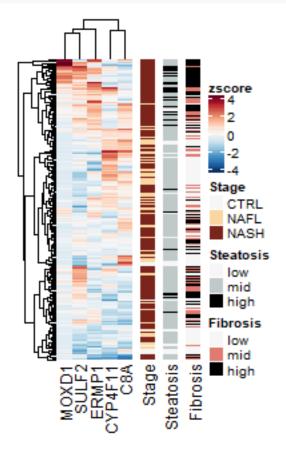


NAS markers (patient stratification)

```
df <- cohort_data$Govaere$cpm_filt %>%
  dplyr::filter(rownames(.) %in% gene_sets$NAS_markers) %>%
  scaleData(method = 'zscore') %>%
  t()
```

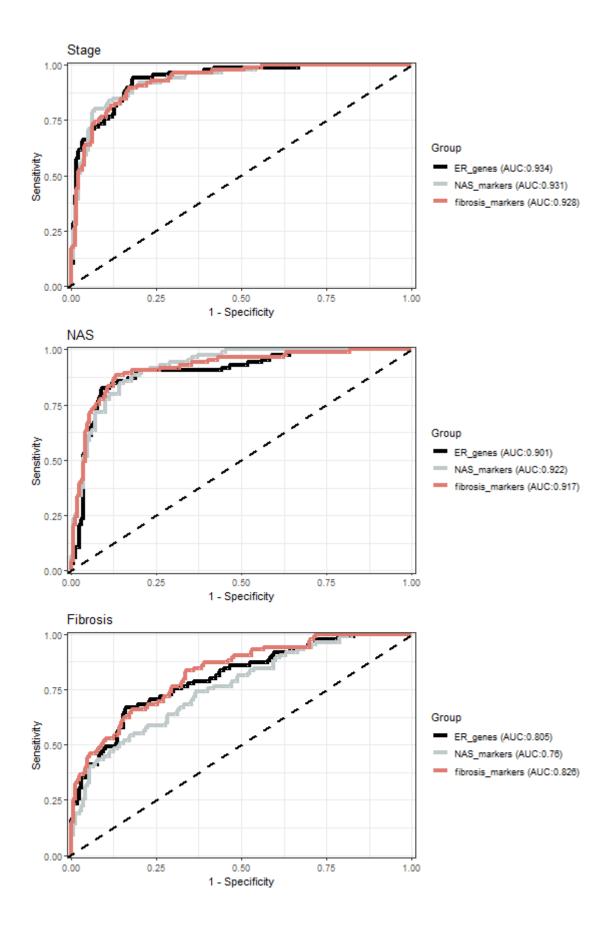


Fibrosis markers (patient stratification)



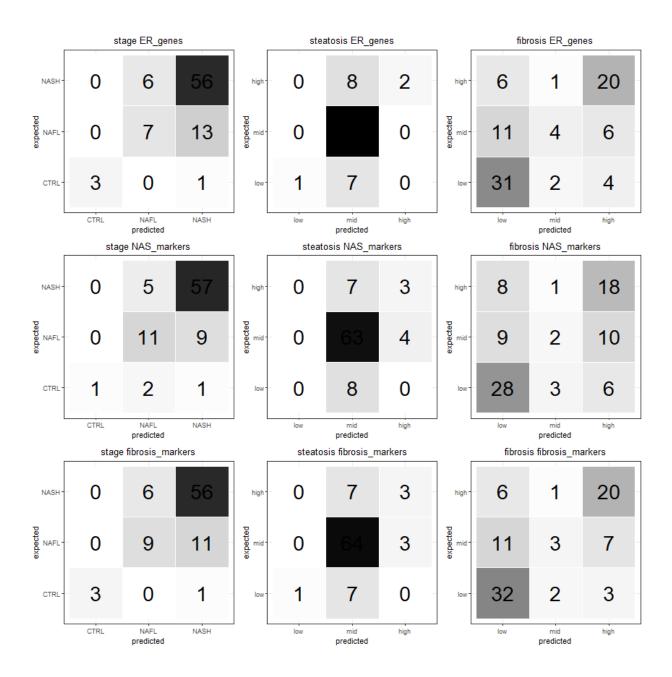
ROC curves

```
theme_bw()
df <- lapply(models$NAS[1:3], function(x) x$plotROC %>% dplyr::filter(Group == 'Micro'))
df <- lapply(names(models$NAS[1:3]), function(x) df[[x]] %>% dplyr::mutate(Group = x))
df <- dplyr::bind_rows(df) %>%
  dplyr::mutate(Group = factor(Group, levels = c('ER_genes', 'NAS_markers', 'fibrosis_markers')),
                label = pasteO(Group, ' (AUC:', round(AUC,3), ')'))
p[[2]] \leftarrow ggplot(df, aes(x=1-Specificity, y=Sensitivity)) +
  geom path(aes(color=Group), size=1.5) +
  geom_abline(intercept=0, slope=1, color='#000000', lwd=1, linetype = 'dashed') +
  scale x continuous(expand = expansion(mult = c(.01, .01))) +
  scale_y_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_color_manual(values = c('#000000','#BFC9CA','#DF7B71'), labels=unique(df$label)) +
  ggtitle('NAS') +
  theme_bw()
df <- lapply(models$Fibrosis[1:3], function(x) x$plotROC %>% dplyr::filter(Group == 'Micro'))
df <- lapply(names(models$Fibrosis[1:3]), function(x) df[[x]] %>% dplyr::mutate(Group = x))
df <- dplyr::bind_rows(df) %>%
  dplyr::mutate(Group = factor(Group, levels = c('ER_genes','NAS_markers','fibrosis_markers')),
                label = paste0(Group, ' (AUC:', round(AUC,3), ')'))
p[[3]] <- ggplot(df, aes(x=1-Specificity, y=Sensitivity)) +</pre>
  geom path(aes(color=Group), size=1.5) +
  geom abline(intercept=0, slope=1, color='#000000', lwd=1, linetype = 'dashed') +
  scale x continuous(expand = expansion(mult = c(.01, .01))) +
  scale_y_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_color_manual(values = c('#000000','#BFC9CA','#DF7B71'), labels=unique(df$label)) +
  ggtitle('Fibrosis') +
  theme_bw()
patchwork::wrap_plots(p, nrow=3, ncol=1, byrow=T)
```



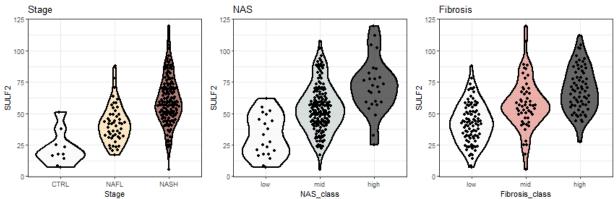
Confusion matrices

```
df <- lapply(names(models$Stage[1:3]), function(x) models$Stage[[x]]$confusionMat %>% dplyr::mutate(con
df <- c(df, lapply(names(models$NAS[1:3]), function(x) models$NAS[[x]]$confusionMat %>% dplyr::mutate(c
df <- c(df, lapply(names(models$Fibrosis[1:3]), function(x) models$Fibrosis[[x]]$confusionMat %>% dplyr
max_count <- lapply(df, function(x) x %>% dplyr::pull(count)) %>% unlist() %>% max()
df <- lapply(df, function(x) x %>% dplyr::mutate(color=paste0('grey', 100-round(count/max_count*100))))
p <- lapply(df, function(x) {</pre>
  ggplot(x, aes(predicted,expected)) +
    geom_tile(aes(fill = color), color = "white") +
    geom_text(aes(label = sprintf("%1.0f", count)), vjust = 0.5, size=10) +
    scale_fill_identity() +
    theme_void() +
    theme_bw() +
    ggtitle(paste(x$cond[1], x$set[1])) +
    theme(legend.position = "none",
          plot.title = element_text(hjust = 0.5))
})
patchwork::wrap_plots(p, nrow=3, ncol=3, byrow=F)
```



SULF2 expression across NAFLD stage, NAS and fibrosis categories

```
geom_violin(lwd=1, alpha=0.6, color='black') +
  ggbeeswarm::geom_quasirandom(width = 0.2, color='black') +
  scale_fill_manual(values = colPals$stage) +
  scale_color_manual(values = colPals$stage) +
  ggtitle('Stage') +
  theme_bw() +
  theme(legend.position='none')
p[[2]] <- ggplot(df, aes(x=NAS_class, y=SULF2, fill=NAS_class, color=NAS_class)) +
  geom_violin(lwd=1, alpha=0.6, color='black') +
  ggbeeswarm::geom_quasirandom(width = 0.2, color='black') +
  scale_fill_manual(values = colPals$NAS) +
  scale_color_manual(values = colPals$NAS) +
  ggtitle('NAS') +
  theme_bw() +
  theme(legend.position='none')
p[[3]] <- ggplot(df, aes(x=Fibrosis_class, y=SULF2, fill=Fibrosis_class, color=Fibrosis_class)) +
  geom_violin(lwd=1, alpha=0.6, color='black') +
  ggbeeswarm::geom_quasirandom(width = 0.2, color='black') +
  scale_fill_manual(values = colPals$fibrosis) +
  scale_color_manual(values = colPals$fibrosis) +
  ggtitle('Fibrosis') +
  theme_bw() +
  theme(legend.position='none')
patchwork::wrap_plots(p, nrow=1, ncol=3, byrow=T)
```

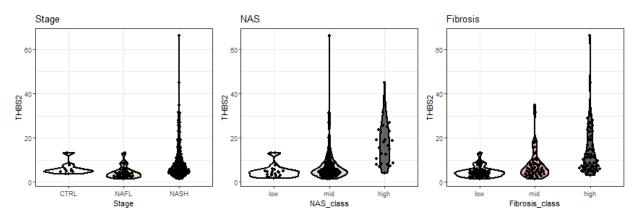


THBS2 expression across NAFLD stage, NAS and fibrosis categories

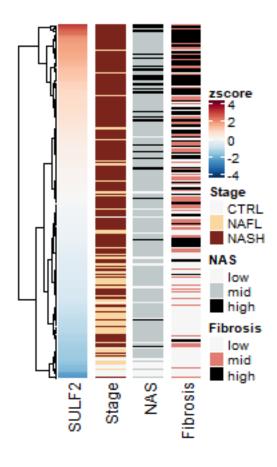
```
p <- list()

p[[1]] <- ggplot(df, aes(x=Stage, y=THBS2, fill=Stage, color=Stage)) +
    geom_violin(lwd=1, alpha=0.6, color='black') +
    ggbeeswarm::geom_quasirandom(width = 0.2, color='black') +
    scale_fill_manual(values = colPals$stage) +
    scale_color_manual(values = colPals$stage) +
    ggtitle('Stage') +
    theme_bw() +</pre>
```

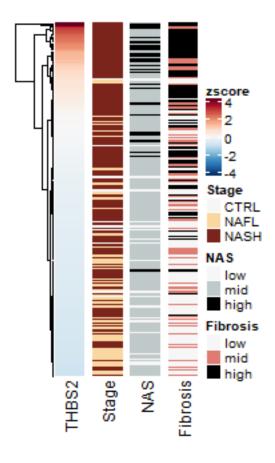
```
theme(legend.position='none')
p[[2]] <- ggplot(df, aes(x=NAS_class, y=THBS2, fill=NAS_class, color=NAS_class)) +
  geom_violin(lwd=1, alpha=0.6, color='black') +
  ggbeeswarm::geom_quasirandom(width = 0.2, color='black') +
  scale_fill_manual(values = colPals$NAS) +
  scale_color_manual(values = colPals$NAS) +
  ggtitle('NAS') +
  theme bw() +
  theme(legend.position='none')
p[[3]] \leftarrow ggplot(df, aes(x=Fibrosis\_class, y=THBS2, fill=Fibrosis\_class, color=Fibrosis\_class)) + fill=Fibrosis\_class, color=Fibrosis\_class)
  geom_violin(lwd=1, alpha=0.6, color='black') +
  ggbeeswarm::geom_quasirandom(width = 0.2, color='black') +
  scale_fill_manual(values = colPals$fibrosis) +
  scale_color_manual(values = colPals$fibrosis) +
  ggtitle('Fibrosis') +
  theme_bw() +
  theme(legend.position='none')
patchwork::wrap_plots(p, nrow=1, ncol=3, byrow=T)
```



SULF2 expression (patient stratification)



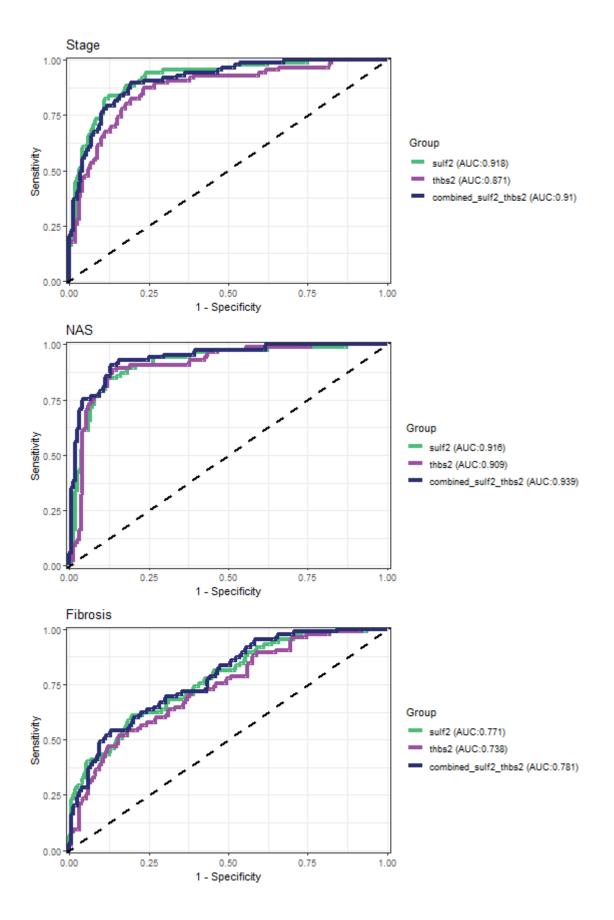
THBS2 expression (patient stratification)



ROC curves of SULF2 & THBS2 classification models

```
p <- list()
df <- lapply(models$Stage[4:6], function(x) x$plotROC %>% dplyr::filter(Group == 'Micro'))
df <- lapply(names(models$Stage[4:6]), function(x) df[[x]] %>% dplyr::mutate(Group = x))
df <- dplyr::bind_rows(df) %>%
  dplyr::mutate(Group = factor(Group, levels = c('sulf2','thbs2','combined_sulf2_thbs2')),
                label = paste0(Group, ' (AUC:', round(AUC,3), ')'))
p[[1]] \leftarrow ggplot(df, aes(x=1-Specificity, y=Sensitivity)) +
  geom_path(aes(color=Group), size=1.5) +
  geom_abline(intercept=0, slope=1, color='#000000', lwd=1, linetype = 'dashed') +
  scale_x_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_y_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_color_manual(values = c('#4DBC79','#9A509F','#2A2F72'), labels=unique(df$label)) +
  ggtitle('Stage') +
  theme_bw()
df <- lapply(models$NAS[4:6], function(x) x$plotROC %>% dplyr::filter(Group == 'Micro'))
df <- lapply(names(models$NAS[4:6]), function(x) df[[x]] %>% dplyr::mutate(Group = x))
df <- dplyr::bind_rows(df) %>%
  dplyr::mutate(Group = factor(Group, levels = c('sulf2', 'thbs2', 'combined_sulf2_thbs2')),
                label = paste0(Group, ' (AUC:', round(AUC,3), ')'))
```

```
p[[2]] \leftarrow ggplot(df, aes(x=1-Specificity, y=Sensitivity)) +
  geom_path(aes(color=Group), size=1.5) +
  geom_abline(intercept=0, slope=1, color='#000000', lwd=1, linetype = 'dashed') +
  scale_x_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_y_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_color_manual(values = c('#4DBC79','#9A509F','#2A2F72'), labels=unique(df$label)) +
  ggtitle('NAS') +
  theme bw()
df <- lapply(models$Fibrosis[4:6], function(x) x$plotROC %>% dplyr::filter(Group == 'Micro'))
df <- lapply(names(models$Fibrosis[4:6]), function(x) df[[x]] %% dplyr::mutate(Group = x))</pre>
df <- dplyr::bind_rows(df) %>%
  dplyr::mutate(Group = factor(Group, levels = c('sulf2', 'thbs2', 'combined_sulf2_thbs2')),
                label = pasteO(Group, ' (AUC:', round(AUC,3), ')'))
p[[3]] \leftarrow ggplot(df, aes(x=1-Specificity, y=Sensitivity)) +
  geom_path(aes(color=Group), size=1.5) +
  geom_abline(intercept=0, slope=1, color='#000000', lwd=1, linetype = 'dashed') +
  scale_x_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_y_continuous(expand = expansion(mult = c(.01, .01))) +
  scale_color_manual(values = c('#4DBC79','#9A509F','#2A2F72'), labels=unique(df$label)) +
  ggtitle('Fibrosis') +
  theme bw()
patchwork::wrap_plots(p, nrow=3, ncol=1, byrow=T)
```

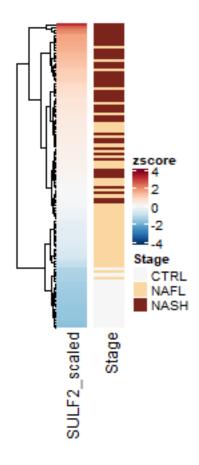


External validation

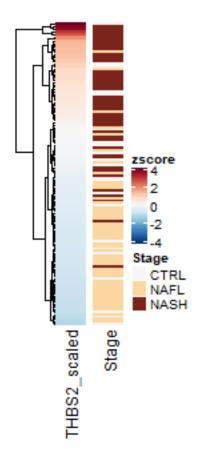
Integrate GTEx v8 (CTRL) & Kozumi et al. 2021 (NAFL/NASH) datasets

```
# merge cohort data from GTEx v8 (CTRL) & Kozumi et al. 2021 (NAFL/NASH)
eval_dat <- cohort_data$GTEx_liver_CTRL$cpm_filt %>%
  tibble::rownames_to_column(var = 'GeneSymbol') %>%
  dplyr::inner_join(tibble::rownames_to_column(cohort_data$Kozumi$cpm_filt, var = 'GeneSymbol'), by='GeneSymbol'), by='GeneSymbol'), by='GeneSymbol')
  tibble::column_to_rownames(var = 'GeneSymbol')
eval_dat_meta <- data.frame(Sample=c(colnames(cohort_data$Kozumi$cpm_filt),</pre>
                                  colnames(cohort_data$GTEx_liver_CTRL$cpm_filt)),
                         Stage=c(cohort_data$Kozumi$meta$Stage,
                                 rep('CTRL', dim(cohort_data$GTEx_liver_CTRL$meta)[1])),
                         Source=c(rep('Kozumi', dim(cohort_data$Kozumi$meta)[1]),
                                  rep('GTEx', dim(cohort_data$GTEx_liver_CTRL$meta)[1]))) %>%
  dplyr::arrange(factor(Sample, levels = colnames(eval_dat)))
# normalize expression across datasets using a stable houskeeping gene
# SRSF4 has been reported as stably expressed throughout NAFLD spectrum
# (https://doi.org/10.1111/j.1530-0277.2011.01627.x)
df <- eval dat %>%
  dplyr::filter(rownames(.) %in% c('SULF2', 'THBS2', 'SRSF4')) %>%
  t() %>%
  as.data.frame() %>%
  dplyr::mutate(SULF2_scaled=SULF2/(SRSF4/mean(SRSF4)),
                THBS2 scaled=THBS2/(SRSF4/mean(SRSF4)),
                SRSF4_scaled=SRSF4/(SRSF4/mean(SRSF4))) %>%
  t() %>%
  scaleData(method = 'zscore') %>%
  t() %>%
  cbind(eval_dat_meta) %>%
  dplyr::mutate(Stage=factor(Stage, levels=c('CTRL', 'NAFL', 'NASH')),
                Source=factor(Source, levels=c('Kozumi', 'GTEx')))
```

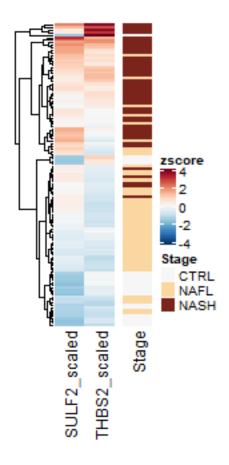
SULF2 expression (patient stratification)



THBS2 expression (patient stratification)

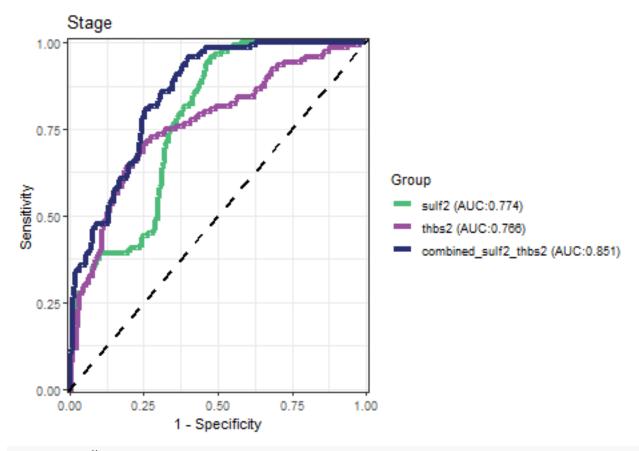


Combined SULF2 & THBS2 expression (patient stratification)



ROC curves for SULF2 & THBS2 model predictions

```
scaling_factors <- eval_dat %>%
  t() %>%
  as.data.frame() %>%
  dplyr::mutate(SRSF4=SRSF4/mean(SRSF4)) %>%
  dplyr::pull(SRSF4)
eval_dat_scaled <- t(t(eval_dat)/scaling_factors)</pre>
tests <- list()</pre>
tests[['Stage']][['sulf2']] <- testModel(models$Stage$sulf2$model,</pre>
                                           t(eval_dat_scaled),
                                           eval_dat_meta$Stage,
                                           preproc = c('scale', 'center'))
tests[['Stage']][['thbs2']] <- testModel(models$Stage$thbs2$model,</pre>
                                           t(eval_dat_scaled),
                                           eval_dat_meta$Stage,
                                           preproc = c('scale', 'center'))
tests[['Stage']][['combined_sulf2_thbs2']] <- testModel(models$Stage$combined_sulf2_thbs2$model,
                                                          t(eval_dat_scaled),
                                                           eval_dat_meta$Stage,
                                                          preproc = c('scale', 'center'))
df <- lapply(tests$Stage, function(x) x$plotROC %>% dplyr::filter(Group == 'Micro'))
```



sessionInfo()

```
## R version 4.0.5 (2021-03-31)
## Platform: x86_64-w64-mingw32/x64 (64-bit)
## Running under: Windows 10 x64 (build 19044)
##
## Matrix products: default
##
locale:
## [1] LC_COLLATE=English_United States.1252
## [2] LC_CTYPE=English_United States.1252
## [3] LC_MONETARY=English_United States.1252
```

```
## [4] LC NUMERIC=C
## [5] LC_TIME=English_United States.1252
## attached base packages:
## [1] grid
                 stats
                           graphics grDevices utils
                                                          datasets methods
## [8] base
## other attached packages:
   [1] patchwork 1.1.1
                             multiROC_1.1.1
                                                   caret_6.0-92
   [4] lattice_0.20-41
                             ComplexHeatmap_2.4.3 RColorBrewer_1.1-2
  [7] forcats_0.5.1
                              stringr_1.4.0
                                                   dplyr_1.0.3
## [10] purrr_0.3.4
                             readr_1.4.0
                                                   tidyr_1.2.0
## [13] tibble_3.1.4
                              ggplot2_3.3.3
                                                   tidyverse_1.3.0
##
## loaded via a namespace (and not attached):
## [1] ggbeeswarm_0.6.0
                              colorspace_2.0-0
                                                   rjson_0.2.20
## [4] ellipsis_0.3.2
                              class_7.3-18
                                                   circlize_0.4.12
## [7] GlobalOptions 0.1.2
                             fs 1.5.0
                                                   clue 0.3-58
## [10] rstudioapi_0.13
                                                   farver_2.0.3
                             listenv_0.8.0
## [13] prodlim_2019.11.13
                              fansi_0.4.2
                                                   lubridate_1.7.9.2
## [16] xml2_1.3.2
                              codetools_0.2-18
                                                   splines_4.0.5
## [19] knitr 1.31
                              jsonlite_1.7.2
                                                   pROC_1.18.0
## [22] broom_0.7.4
                              cluster_2.1.1
                                                   dbplyr_2.0.0
## [25] png 0.1-7
                              compiler_4.0.5
                                                   httr 1.4.2
## [28] backports_1.2.1
                              assertthat_0.2.1
                                                   Matrix_1.3-2
## [31] fastmap_1.1.0
                              cli_2.3.0
                                                   htmltools_0.5.2
## [34] tools_4.0.5
                              gtable_0.3.0
                                                   glue_1.4.2
## [37] reshape2_1.4.4
                              Rcpp_1.0.7
                                                   cellranger_1.1.0
## [40] vctrs_0.3.8
                             nlme_3.1-151
                                                   iterators_1.0.13
## [43] timeDate_3043.102
                             gower_1.0.0
                                                   xfun_0.31
## [46] globals_0.14.0
                             rvest_0.3.6
                                                   lifecycle_0.2.0
## [49] future_1.21.0
                             MASS_7.3-53
                                                   zoo_1.8-8
## [52] scales_1.1.1
                              ipred_0.9-12
                                                   hms_1.0.0
## [55] parallel_4.0.5
                                                   rpart_4.1-15
                              yaml_2.2.1
## [58] stringi_1.5.3
                             highr 0.8
                                                   foreach 1.5.1
## [61] e1071_1.7-4
                             hardhat_0.2.0
                                                   boot_1.3-26
## [64] lava 1.6.10
                              shape 1.4.5
                                                   rlang 0.4.10
## [67] pkgconfig_2.0.3
                              evaluate_0.14
                                                   recipes_0.2.0
## [70] labeling_0.4.2
                             tidyselect_1.1.0
                                                   parallelly_1.23.0
## [73] plyr_1.8.6
                             magrittr_2.0.1
                                                   R6_2.5.0
## [76] generics 0.1.2
                             DBI 1.1.1
                                                   pillar_1.6.2
## [79] haven 2.3.1
                             withr 2.4.1
                                                   survival 3.2-7
## [82] nnet 7.3-15
                              future.apply_1.7.0
                                                   modelr 0.1.8
## [85] crayon_1.4.0
                                                   rmarkdown_2.14
                              utf8_1.1.4
## [88] GetoptLong_1.0.5
                             readxl_1.3.1
                                                   data.table_1.13.6
## [91] ModelMetrics_1.2.2.2 reprex_1.0.0
                                                   digest_0.6.27
## [94] stats4 4.0.5
                             munsell_0.5.0
                                                   glmnet_4.1-4
## [97] beeswarm_0.4.0
                             vipor_0.4.5
```