

# Introduction to the shell and task automation

2015-04-22

# What is the shell?

## In brief. . .

- Command line interface, looks old-fashioned but very convenient
- Main interface when you want to login to **CSC servers** or **remote servers**
- Also present in **Linux** distributions for personal computers and **Mac**
- With **Windows**, the cmd prompt is a bit similar (text-based) but not as powerful

## Usage

- Often the only interface for remote connections
- Powerful built-in commands
- **Automate repetitive tasks**
- Shell scripts to **reproduce** data manipulation

# Where can we find the shell?

## To find a shell...

- On **GNU/Linux** and **MacOS** systems: open a **terminal**. This will provide you with a Unix-like shell on both systems
- On **Windows**: run `cmd.exe` or `cmd`. This shell is quite different from the Unix-like shell found in Linux and MacOS. To obtain a Unix shell on Windows, one can install the Cygwin tools.
- It is strongly recommended to learn how to use a **Unix shell** since it is very likely it is this type of shell you will be exposed to when you connect to a **remote server**.

# Where can we find the shell?

## One shell or several shells?

- A shell: a program providing an **interface** between the user and the computer. **Different shells exist.**
- The most popular and widely used shell is probably **bash**. It is the default shell in most GNU/Linux distributions.
- If you learn how to use **bash**, you will be able to use most **remote servers** you'll have to connect to, and also the **terminal** from MacOS or the **Cygwin** tools on Windows

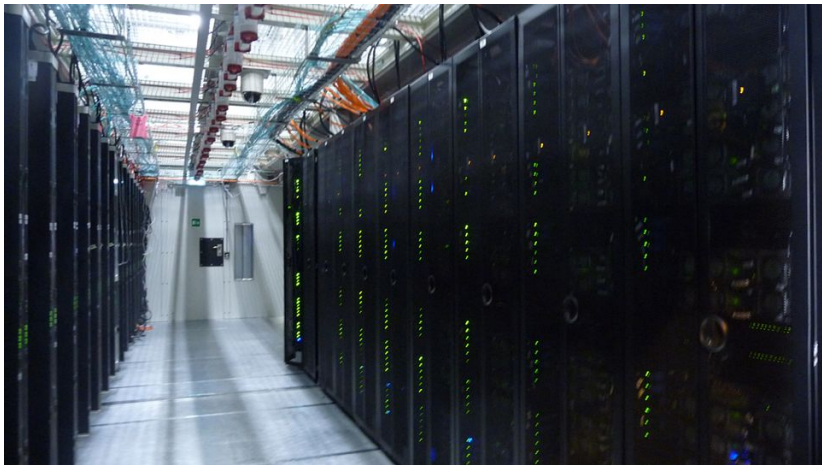
## One word on terminology

- During the course, we will often say interchangeably "the terminal", "the shell" or "bash".

# The CSC center in Kajaani



# Meet the Taito cluster ([taito.csc.fi](http://taito.csc.fi))



# A word about CSC servers

## Available servers

- Taito: 19152 cores (16 cores per nodes)
- Sisu: 39408 cores, for massively parallel jobs

## Job submission

- CPU-intense calculations have to be submitted through a queue system
- Server load
- We can also run some simple commands directly at login

## Module system

- Many softwares installed
- Sometimes different versions of a software
- User has to explicitly load **modules**

# Connection to a remote shell

## The plan

- Using the CSC server Taito in Kajaani (student account)
- Tools: **putty** (windows) or **ssh** (Mac and GNU/Linux)
- A word about **ssh** and the **security of connections**?

## Student account

- Logins: jyybio01 to jyybio20
- Password: on the whiteboard

## Connection

- From a terminal (Mac or GNU/Linux):  
`ssh jyybioxx@taito.csc.fi`  
where xx is your student number.
- From Putty: ask a teacher if needed



# First contact with the shell

## Just after connection

- What you see after connection in the **shell prompt**. It tells you the shell is ready to receive your input:  
`jyybioxx@taito-login3$`
- `jyybioxx` is your username, `taito-login` is the host server to which you are connected. The number after `taito-login` can vary because Taito has several login nodes.

# First contact with the shell

## Execute a command (`ls`)

The shell **reads** and **executes** commands you enter at the prompt, and **prints** the output. Type `ls` and press RETURN. You should see:

```
appl_taito
```

You just ran the `ls` command which produces an output: the list of files and folders present in the current directory.

- Try another command: `whoami`. What does this command do?

# First contact with the shell

## Execute a command (pwd)

- When you login to a server, you are automatically sent to your home folder. You can see where you are by typing `pwd`, which produces:  
`/homeappl/home/jyybioxx`
- So you are now in the folder `jyybioxx`, which is itself contained in `home`, which is contained in `homeappl`, which is at the root of the file system (`/`, there is no parent directory above).

# Adding options to a command

## Using `ls` options

- You can add options to a command with the dash sign `-`:

```
ls -l
```

(this is `-l`, not `-1`)

- This runs the `ls` command with the `-l` option, which produces a detailed output:

```
total 4
```

```
drwx----- 2 jyybio20 jyybio 4096 Apr 15 12:15 appl_taito
```

Now you can see the date of last modification of the folders and some other information.

# A word about rights

## The rights system

- In a Unix system, every file has an **owner** and belongs to a **group**
- Every file has rights for **reading**, **writing** and **execution**
- Those rights are set for three categories of users: **owner**, **group** and **others**

## ls -l output

```
drwx----- 2 jyybio20 jyybio 4096 Apr 15 12:15 appl_taito
```

- The three first letters are rights for the owner, the next three rights for the group, and the last three rights for others.
- If a letter is replaced by a dash, the right is not granted

```
-rwx-----
```

```
-r--r--r--
```

```
-rwxr--r--
```

```
drwxr-xr-x
```

# Clone the Git repository for the practicals

## Clone the Git repository

- Before going further, you should clone a Git repository containing the data which was prepared for you (Git is installed on Taito). The repository is hosted on GitHub.
- Check that you are in your home folder with `pwd`. You should see:  
`/homeappl/home/jyybioxx`  
If not, go back to your home folder by typing simply `cd` without any argument.
- Clone the Git repository with (all on one line):

```
git clone  
https://github.com/mdjbru-teaching-material/practicals.git
```

- Run `ls`. What happened?

# Data content and motivation

## The data files

- Each **file** corresponds to **one *Escherichia coli* strain** for which a complete or draft genome sequence is available. Each file contains the **peptide sequences** from all translations resulting from Ensembl known or novel gene predictions for that strain.
- Files are in the FASTA format. The original address is `ftp://ftp.ensemblgenomes.org/pub/current/bacteria/fasta/`.

## Motivation

We want to determine the **amino acid content** of **all proteins of each strain**, and compare the results between strains. We already have a Python script ready which can determine the amino-acid composition for protein sequences.

# Basic folder navigation

## cd command

- We can navigate from folder to folder using the `cd` command:

```
cd practicals
ls
cd ecoli-data
ls
```

- We could have gone directly to the second subfolder with:

```
cd practicals/ecoli-data
```

- You can see there are already some files in this folder. Let's ask for more details with `ls -l`
- How many files are there? How large are they?



## Combining options for `ls`

- We can ask for more human-readable sizes with:

```
ls -l -h
```

- Can you see the difference with `ls -l`? What does `ls -h` do?
- We could also combine both options to `ls`: `ls -lh`

# Basic folder navigation

## Moving to the parent directory

- We can go back through the parent folders using `cd ..`

```
pwd      # Where are you at this point?
```

```
cd ..
```

```
pwd      # And now?
```

```
ls
```

```
cd ..
```

```
pwd      # And here?
```

```
ls
```

# Basic folder navigation

## Going back to the home directory

- A faster way to go back to your home directory, from any starting directory, is just to type `cd` without any argument.
- Go back to the `ecoli-data` subfolder and back again to your home directory using `cd`.

## Shortcut for the home folder

- Another way to go to the home folder is to use the `~` character: this is automatically replaced by the path to your home folder by `bash`.

```
cd      # Back to your home folder
cd practicals
cd ~    # Bash understands "~" as "/homeappl/home/jyybioxx"
cd appl_taito
cd ~/practicals
```

# Creating folders

## The `mkdir` command

- Go back to the `practicals` folder and create a new folder in it:

```
cd ~/practicals
mkdir results
cd results
ls
```

## Exercise

- Create the following directory structure:

```
~/practicals/scripts/python/modules/seqAnalysis
```

- Go back to your home folder.

# Auto-completion

## The magic TAB key

- Let's go into seqAnalysis folder, but let's be lazy:

```
cd      # Start from your home folder  
cd pr   # Press TAB at this point
```

- Do you understand what happened?
- Use this feature to quickly go to seqAnalysis. What is the minimum number of keystrokes you have to use to go there from your home folder?

## Remember!

When you press **TAB**, the shell tries to complete what you just typed by itself. This **auto-completion feature** of the shell is very convenient and will save you a lot of typing!

## Test auto-completion

- Now create a folder:

```
~/practicals/scripts/python/modifiedSources
```

- Go back to your home folder, and go into `modifiedSources` using the TAB completion as much as you can. What do you notice?

## Double TAB

- Now create the folder

```
~/practicals/scripts/python/modularComponents
```

- Type:

```
cd ~/practicals/scripts/python/mod # Press =TAB= twice here  
# Type "ule" and press =TAB= again
```

- Do you understand how TAB completion works? This also works for command names.

# Copying, moving and removing files

## Creating an empty file

- Go to the seqAnalysis folder and type:

```
touch DNA-analysis.py  
ls
```

- What happened?

## Moving a file

- Now type:

```
mv DNA-analysis.py ../modularComponents
```

- What happened? Did you use the TAB key? (you should!)
- Explore the directory structure to find DNA-analysis.py again.



# Copying, moving and removing files

## Copying a file

- Go to the modularComponents subfolder and type:

```
cp DNA-analysis.py ../modules
```

- What happened?

## Removing a file

- From modularComponents folder, type:

```
rm DNA-analysis.py
```

- What happened?

# Creating a directory hierarchy

## Moving a folder

- From the scripts folder, move modularComponents into modules:

```
mv modularComponents modules  
tree
```

- What does tree do?

## Copying a folder

- Go to the practicals folder and make a copy of scripts:

```
cp -r scripts scripts-backup
```

- Note the `-r` option used for recursive copy inside the directories.

# Creating a directory hierarchy

## Removing a folder

- Remove the newly created folder with:

```
rm -r scripts-backup
```

- Again, note the `-r` option to work on folders.

# Creating a directory hierarchy

## Exercise

- Now that you have experience, create the exact following directory structure (only folders shown):

```
+-- appl_taito
|-- practicals
    +-- ecoli-data
        |   '-- [...]
    +-- results
        |   '-- 2015-04-22
    '-- scripts
        +-- python
            |   +-- popGenetics
            |   +-- proteinStructure
            |   '-- seqAnalysis
        '-- R
```

# Viewing a file

## cat command

- Go to the `ecoli-data` folder and type:

```
cat README
```

- Try also `cat` on one of the fasta files. What happened?

## head and tail commands

```
head Escherichia_coli_o5_k4_l_h4_str_atcc_23502.GCA_000333195.
```

```
tail Escherichia_coli_o5_k4_l_h4_str_atcc_23502.GCA_000333195.
```

```
head -n 30 Escherichia_coli_o5_k4_l_h4_str_atcc_23502.GCA_000333195.
```

```
tail -n 3 Escherichia_coli_o5_k4_l_h4_str_atcc_23502.GCA_000333195.
```

- Do you understand what those commands do?

# Viewing a file

## less command

- less is very useful to examine large file. You can navigate using the up and down arrows or B and SPACE keys, and you can exit with Q.

```
less Escherichia_coli_o5_k4_l_h4_str_atcc_23502.GCA_000333195.
```

# Useful tools: `wc`

## `wc` to count words

- Go to the `ecoli-data` folder and type:

```
wc Escherichia_coli_o55_h7_str_06_3555.GCA_000617385.1.26.pep
```

which produces:

```
26318    51865 1824223 Esch...
```

- We can have only the number of lines with `wc -l` (try it).

## Wildcards

- Try:

```
wc -l *.fa
```

- What happened?

# Redirection

## The > operator

- When a command produces some output, it can be redirected to a file instead of to the terminal:

```
wc -l *.fa > lineCounts  
cat lineCounts
```

- > is a **redirection** operator, and automatically creates a new file or erases an existing file.

## The >> operator

- To redirect output and append it to an existing file, we can use the >> operator:

```
wc -l README >> lineCounts  
cat lineCounts
```



# Useful tools: grep

## grep to search for matches

```
grep "flagellin" Escherichia_coli_o55_h7_str_06_3555.GCA_00061
grep --color=always "flagellin" Escherichia_coli_o55_h7_str_06
grep -n --color=always "flagellin" Escherichia_coli_o55_h7_str
grep -c --color=always "flagellin" Escherichia_coli_o55_h7_str
```

- Do you understand what each of the grep options do?

## Exercise

- Use grep to extract all the sequence names from one of the fasta file and store them in a file called proteinNames.

# Useful tools: grep

grep is versatile

```
grep -c flagellin *.fa
```

```
grep -c flagel *.fa
```

- Do you understand the output?

## Exercise

- How would you count the number of proteins in each fasta file?

## Useful tools: cut

### cut to get columns

```
grep -c flagel *.fa > flagelCounts
```

```
cat flagelCounts
```

```
cut -d "_" -f 1 flagelCounts
```

```
cut -d "_" -f 3 flagelCounts
```

```
cut -d "_" -f 3,5 flagelCounts
```

```
cut -d ":" -f 2 flagelCounts
```

- Do you understand what cut does and the roles of the -d and -f options?

# Useful tools: sort

## sort to sort things

- Use sort to sort the line counts from lineCounts:

```
sort lineCounts
```

- Is everything correct? What if you try `sort -n lineCounts`? Can you see a difference?
- Try also `sort -r lineCounts`. What is the difference?

## Exercise

- Using `grep` and `sort` and an intermediate files, sort the bacterial proteomes by decreasing number of proteins.
- Hint: `sort` supports two interesting options, `-t` to specify a field separator and `-k` to specify which field to use for sorting.

# Combining tools with pipes

## Pipes can connect an output and an input streams

- When we did sort `lineCounts`, we used `sort` on the output of `wc`, but we used an intermediate file. The shell offers a powerful way to connect directly the output of a command to the input of another: the **pipe operator**:

```
wc -l *.fa | sort -n
```

## Exercises

- The `w` output the list of connected users on the server. Try it and then try:

```
w | head
```

```
w | less
```

- Use a pipe to find all the users whose login contains "jyy".
- Extend the same pipe to count how many there are.

# Python script to determine amino acid composition

## Test the Python script

- The script `seqComposition.py` takes a fasta file and produces a table containing the amino-acid composition of each protein in the file.
- To run the script, type:

```
module load python-env/3.4.1
# This module loading step is specific to the server
python3 seqComposition.py myFastaFile
# Use the fasta file you wish instead of "myFastaFile"
```

The output is sent to the terminal.

- Propose at least two practical ways to have a look at this output.

# Python script to determine amino acid composition

## Exercise

- Using only the **Unix tools** you know, the **Python script** and **pipes**, determine the distribution of the number of histidines per protein in the proteome of the strain of your choice.
- More clearly stated: for a given strain, determines how many proteins have one histidine, how many have two, how many have three, ...

# One step towards wizardry: shell scripts

## Reusing your tool pipeline

- Let's use nano to store your pipeline in a file:

```
nano getHistDistrib.sh
```

(the usage of nano will be demonstrated live)

- The idea is to be able to produce the histidine distribution results just by typing:

```
bash getHistDistrib.sh myFastaFile
```

## Test your pipeline with a few files

- Test your pipeline for 5 strains. How would you feel about doing it for 2000 strains?



# One step towards wizardry: shell scripts

## Making a general purpose listing script

- Create a shell script (testListing.sh) with this content:

```
listFiles='ls *.fa'
echo $listFiles
for myFile in $listFiles; do
    echo $myFile
    echo $myFile.results
done
```

- Run it with bash. What does this do?

## Exercise: final script

- Combine the script with your pipeline and the listing script into a single script to get the histidine distribution for all the fasta files in this folder.