# CMP 464/788 Lecture Notes

Experimental Testing of Algorithms
Introduction to Computational Biology
30 October 2003

## **Testing Methods Empirically**

- How accurate are the methods at reconstructing trees?
- In biological applications, the true, historical tree is almost never known, which makes assessing the quality of phylogenetic reconstruction methods problematic. (an exception: Hillis '92 created an evolutionary tree in the laboratory)
- Simulation is used instead to evaluate methods, given a model of evolution.

#### **Simulation Studies**

1. Construct a

"model" tree.

2. "Evolve"

sequences down

the tree.

3. Reconstruct

the tree using

method.

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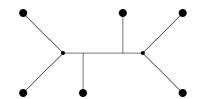
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## Simulating Data: Choosing Trees

Usually chosen from a random distribution on trees:
 Uniform, or Yule-Harding (birth-death trees)



- Can view this as two different random processes:
  - generate the tree shape, and then
  - assign weights or branch lengths to the shape.

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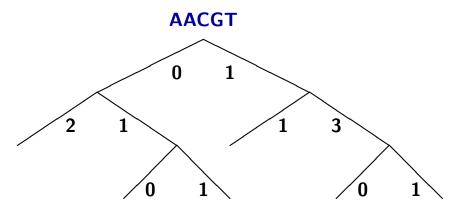
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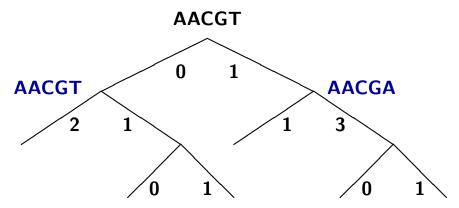
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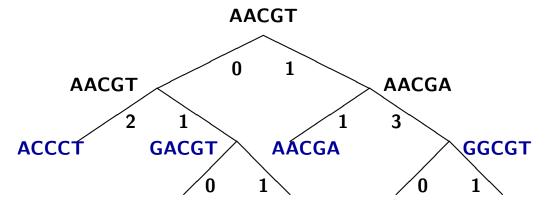
- The *Jukes-Cantor* (JC) model is the simplest Markov model of biomolecular sequence evolution.
- A DNA sequence (a string over  $\{A, C, T, G\}$ ) at the root evolves down a rooted binary tree T.



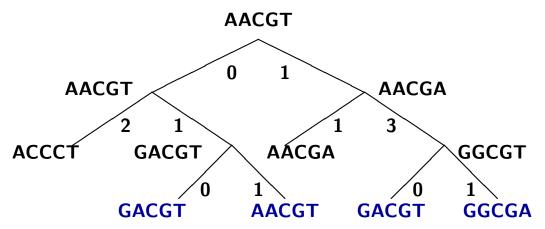
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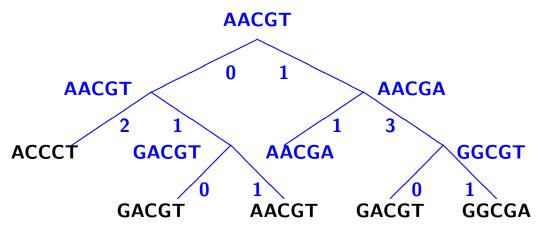
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- The assumptions of the model are:
  - 1. the sites (i.e., the positions within the sequences) evolve independently and identically
  - 2. if a site changes state it changes with equal probability to each of the remaining states, and
  - 3. the number of changes of each site on an edge e is a Poisson random variable with expectation  $\lambda(e)$  (this is also called the "length" of the edge e).

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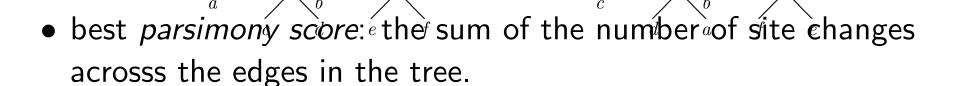
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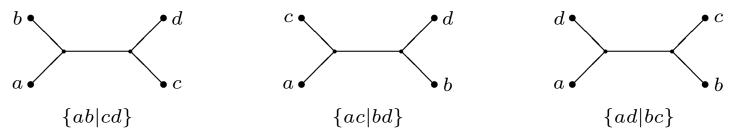
## **Evaluating Accuracy**

• To compare reconstructed tree to model tree, the *Robinson-Foulds Score* is often used:



# Case Study: Quartet Methods

• A quartet is an unrooted binary tree on four taxa:



• Let Q(T)= all quartets that agree with T. [Erdős et al. 1997]: T can be reconstructed from Q(T) in polynomial time.

## Case Study: Quartet Methods

- Quartet-based methods operate in two phases:
  - Construct quartets on all four taxa sets.
  - Combine these quartets into a tree.
- Running time:
  - For most optimizations, determining a quartet is fast.
  - There are  $\Theta(n^4)$  quartets, giving  $\Omega(n^4)$  running time.
  - In practice, the input quality is insufficient to ensure that all quartets are accurately inferred.
  - Quartet methods have to handle incorrect quartets.

#### Popular Quartet Methods

- Q\* or Buneman Method [Berry & Gascuel '97, Buneman '71]:
   Only add edges that agree with all input quartets.
   Doesn't tolerate errors— outputs conservative, but unresolved tree.
- Quartet Cleaning (QC) [Berry et al. 1999]: Add edges with a small number of errors proportional to  $q_e$ . Many variants: all handle a small number of errors.
- Quartet Puzzling [Strimmer & von Haeseler 1996]: "Order taxa randomly, greedily add edges, repeat 1000 times." Output majority tree.
  - Most popular with biologists.

# Standard Method: Neighbor Joining (NJ)

- [Saitou & Nei 1987]: very popular and fast:  $O(n^3)$ .
  - Based on the distance between nodes, join neighboring leaves, replace them by their parent, calculate distances to this node, and repeat.
  - This process eventually returns a binary (fully resolved) tree.
  - Joining the leaves with the minimal distance does not suffice, so subtract the averaged distances to compensate for long edges.
  - Experimental work shows that NJ trees are reasonably accurate,
     given a rate of evolution is neither too low nor too high.

## **Previous Experimental Studies**

- Berry et al. [1999] studied various QC methods:
  - Showed that QC methods outperform the  $Q^*(didn't compare to any other methods)$
  - By design, the QC methods recover all edges recovered by  $Q^*$ . Noteworthy that the QC methods *obtained* additional edges.
  - Varied evolutionary rates and sequence lengths, but studied only 10 taxa trees.
  - The theoretical bounds we derive and experiments on larger n suggest that performance on very large n may be poor.

# **Our Study**

- A detailed, large-scale experimental study of quartet methods and NJ under the Jukes-Cantor model of evolution:
  - Our results indicate that NJ always outperforms the quartet-based methods we examined, in terms of both accuracy and speed.
  - Give new theory about convergence rates of quartet-based methods which helps explain our observations.

## **Experimental Design**

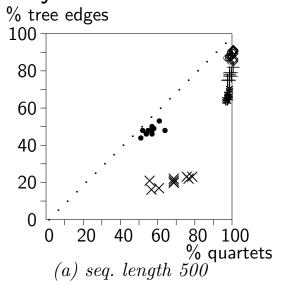
- Generated a large number of datasets, varying number of taxa, rates of evolution, and sequence lengths.
- For each dataset generated, we computed
  - the NJ and QP trees on the entire dataset, and
  - two sets of quartets,  $Q_{ML}$ , and  $Q_{NJ}$ .
- We applied cleaning methods to  $Q_{ML}$  and  $Q_{NJ}$  and compared quartets of  $Q_{ML}$ , of  $Q_{NJ}$ , and of the reconstructed trees against the model tree for accuracy.

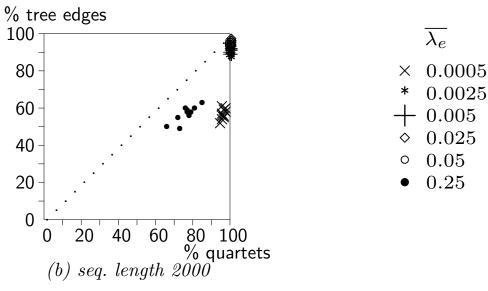
#### **Experimental Design: Parameter Space**

- In all, our study used 16,000 datasets and required many months of computation on the two clusters.
  - Taxa: 5, 10, 20, 40.
  - 8 expected evolutionary rates: from  $5\times 10^{-5}$  to  $5\times 10^{-1}$  per tree edge.
  - For each, we generated 100 tree shapes, grouped into 10 runs of 10 trials.
  - Sequence lengths: 500, 2,000, 8,000, and 32,000.

## Measuring Accuracy: Quartets and Edges

 Topological accuracy is a more demanding criterion than quartet accuracy.

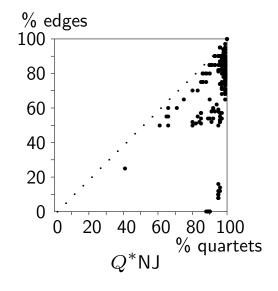


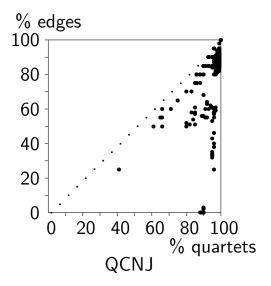


(Percent of true tree edges recovered by Quartet Puzzling for 40 taxa and two sequence lengths)

## **Sensitivity to Input Quality**

 Methods that estimate quartets and then combine them into a single tree can be greatly affected by the quality of the input quartets.





#### **Running Times**

- NJ was clearly the fastest method tested.
- QCML and QP were by far the slowest of the methods tested, slow enough that running them on more than a fifty taxa appears infeasible at present.
- With default settings, QP takes more than 200 days of computation to analyze ten runs of ten trials each for a single set of parameters on 80 taxa with a sequence length of 500. (30 minutes for NJ).