# Sequence Classification Model Report

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Artificial Intelligence

Prepared for

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### 1. Introduction

12] A promoter is a DNA region where the transcription process of a gene starts. It controls the binding of RNA polymerase which transcribes DNA to mRNA that is eventually translated into a functioning protein. In other words, the promoter region is responsible for controlling when and where in the organism a specific gene is expressed.

Studying and analyzing Promoters has been extremely important in understanding many different diseases especially the ones of genetic nature and the reason for that is:

that a 4] several of these diseases are caused by mutations or variations in the promoter region \_and not in the coding region as previously thought\_ for example:

- 4 Asthma 67
- ♣ Diabetes [8]
- ♣ Beta thalassemia [9]
- Huntington's disease [10]
- ♣ Cancer [8]

[5] which will open a pathway to correct that specific variance leading to the treatment of the resulting disease by interference (editing) of the structure or the number of promoter-bound proteins leaving the other genes that are associative with the target gene intact

[5][4]That's why the classification of sequences to promoters and non-promoters in increasingly gaining the interest and attention of many bioinformatics researchers and due to their importance, there was several studies dedicated to classifying promoters and non- promoters regions, but the opportunity for further improvement is present.

### To Achieve this We considered using:

– Deep Learning (Neural Networks):-

Issues with that approach were:

- >Inconvenient in Hardware as it's (Hardware-Dependent)
- >Not being able to show the problem to the network in an easy and accurate way
- >The time duration isn't known
- Machine learning:
- support vector machine: Issues with that approach were:
  - >Time complexity was remarkably high and gave the approximately the same results as the perceptron
- 2. k-Nearest Neighbor classifier model:-

Issues with that approach were:

- >Lower accuracy (59.4%)
- >Sensitive to noise and missing data (Nan)
- 3. Random Forest classifier model

Issues with that approach were:

- >lower accuracy (68.33%)
- >inconvenient storage and memories usage
- 4. Tree classifier model

Issues with that approach were:

>lower accuracy (62%)

### eventually we opted for:

Perceptron classifier model

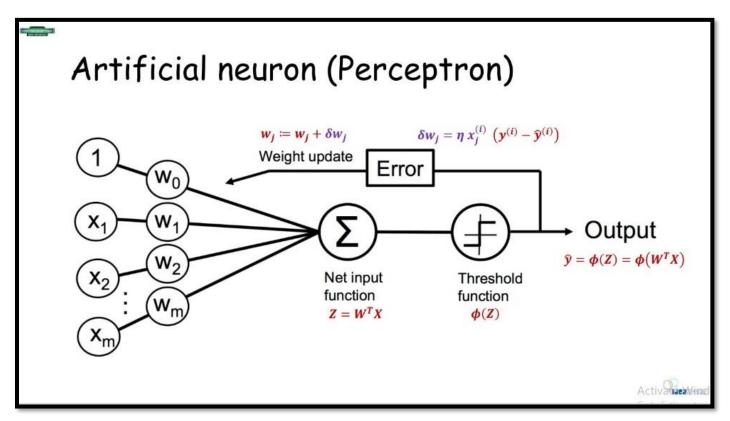
Which gave the best results in the fastest time

### 2. Methodology:

### 1. Perceptron Algorithm:

[11][12]

it is linear machine learning(a supervised learning) algorithm for making binary classification (promoter and non-promoter)



it lets artificial neurons learn and process certain data in the training set one at a time.

The Perceptron algorithm learns the weights for the input signals so it can draw a linear boundary

which let us to identify between the two linearly separable classes Non promoter (0) and promoter(1).

### **Perceptron Learning Rule:**

- -Perceptron Learning Rule states that the algorithm automatically learns the optimal weight coefficients.
- -Then The input features are multiplied with these weights to determine wither a neuron fires or not.

-The Perceptron receives several inputs, and if the sum of the input signals surpasses a certain threshold, it either outputs a signal or does not output anything .

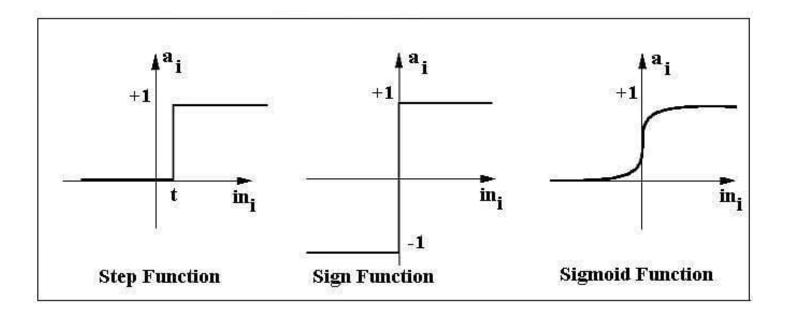
that's how the perceptron (after supervised learning and classification) can be used to predict the label of a sample

### **Perceptron main Function:**

Perceptron is a function that maps its input "x," after being multiplied with the learning weight coefficient; then generates an output value "f(x)"

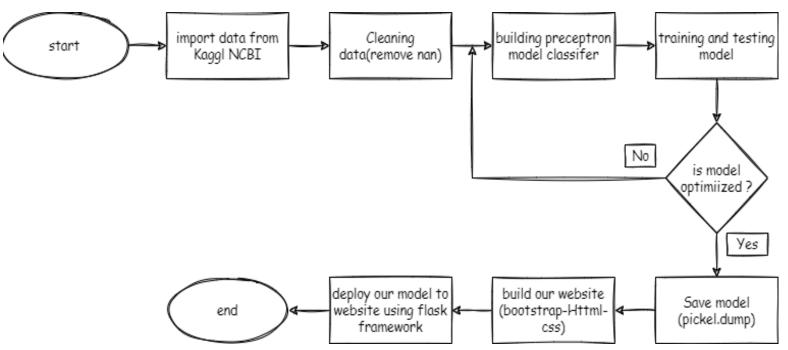
### **Activation Functions of Perceptron:**

The activation function has a step rule (change the output into +1 or -1) to check whether or not the output of the weighting function is greater than zero



Step function is triggered above a specific value of the output; else it outputs 0 . Sign Function outputs +1 or -1 depending on if or not the neuron output is greater than 0 . Sigmoid function outputs a value between 0 and 1.

### 2.Project Flowchart:



#### 3. Pseudocode:

### [13]

end

pg. 6

```
BEGIN
Import pandas as pd and numpy libraries as np from sklearn import datasets
from sklearn.model_selection import train_test_split
from sklearn.preprocessing import StandardScaler
from sklearn.metrics import accuracy_score
from sklearn.linear_model import Perceptron
from sklearn.svm import SVC
from sklearn.model_selection import train_test_split
from sklearn.feature_extraction.text import CountVectorizer
Dfl=pd.read(promoter dataset text)
Df1.dropna(subset=['unnamed:0'],how=all,inplace=true)
dfl.reset_index(inplace = True)
dfl.drop(['EP 1 (+) mt:Col_1; range -400 to -100.', 'index'], axis = 1, inplace=True)
dfl.rename(columns={'Unnamed: 0': "sequence"}, inplace = True)
df1['label'] = 0
same thinge in promoter dataset
df = pd.concat([df1, df2], axis = 0)
for seq in df['sequence']:
begin
  if 'N' in seq:
begin
    display(df.loc[df['sequence'] == seq])
end
```

```
df.drop([1822], inplace = True)
df3=df
display df3
begin
def Kmers_funct(seq, size=4):
  return [seq[x:x+size].lower() for x in range(len(seq) - size + 1)]
end
df['words'] = df.apply(lambda x: Kmers_funct(x['sequence']), axis=1)
df = df.drop('sequence', axis=1)
Y = df.iloc[:, 0].values
X = list(df['words'])
for item in range(len(X)):
begin
  X[item] = ''.join(X[item])
end
from sklearn.feature_extraction.text import CountVectorizer
cv = CountVectorizer(ngram_range=(4,4))
X = cv.fit\_transform(X)
X_train, X_test, y_train, y_test = train_test_split(X,y,test_size=0.20,random_state=1)
ppn = Perceptron(eta0=.01, max_iter=75)
ppn.fit(X_train, y_train)
df3['words'] = df3.apply(lambda x: Kmers_funct(x['sequence']), axis=1)
df3 = df3.drop('sequence', axis=1)
X2 = list(df3['words'])
for item in range(len(X2)):
begin
  X2[item] = ''.join(X2[item])
end
cv = CountVectorizer(ngram_range=(4,4))
X2 = cv.fit_transform(X2)
Display ppn.predict(X2[0])
import pickle
pickle.dump(ppn,open('model.pkl','wb'))
model=pickle.load(open('model.pkl','rb'))
END
```

## 3. Experimental Simulation

1. Python programming language:

as it is open-source language that includes a huge library collection that makes it and best language in dealing with machine learning convenience wise.

2.Perceptron learning algorithm:

as it archived the highest accuracy with the lowest run time as it is a linear classifier with time complexity O(t)

- 3. Biopython to work with sequences
- 4.Pandas and NumPy libraries for cleaning the dataset
- 5.Sklearn for:
  - training, testing, and splitting the dataset
  - calculating accuracy score
  - **4** import models
- 6.Kaggle and NCBI for acquiring our dataset
- 7.Google Collab editor for accessing faster GPUs, TPUs and more RAM.
- 8.Bootstrap (html, CSS, JavaScript) to create a web page
- 9.Flask framework to link our model to the web page

Test cases: We used we use 20% from dataset in test and 80% in train.

#### **Code details:**

- import pandas and numpy library
- 2. Reading non-promoter data and cleaning it from missing data

```
[ ] import pandas as pd
import numpy as np
import os

df1 = pd.read_csv('/content/drive/MyDrive/ML/NonPromoterSequence.txt', sep = '>', )
    df1.dropna(subset=['Unnamed: 0'], how='all', inplace=True)
    df1.reset_index(inplace = True)
    df1.drop(['EP 1 (+) mt:CoI_1; range -400 to -100.', 'index'], axis = 1, inplace=True)
    df1.rename(columns={'Unnamed: 0': "sequence"}, inplace = True)
    df1['label'] = 0
```

3. Reading promoter data and cleaning it from missing data

```
df2 = pd.read_csv('/content/drive/MyDrive/ML/PromoterSequence.txt', sep = '>', )
df2.dropna(subset=['Unnamed: 0'], how='all', inplace=True)
df2.reset_index(inplace = True)
df2.drop(['EP 1 (+) mt:CoI_1; range -100 to 200.', 'index'], axis = 1, inplace=True)
df2.rename(columns={'Unnamed: 0': "sequence"}, inplace = True)
df2['label'] = 1
```

4. Link two dataset together in one dataframe.

```
[ ] df = pd.concat([df1, df2], axis = 0 )
    df.shape
(22600, 2)
```

- 5. Function to Divide Data to 4-mer
- 6. Calling the Kmers\_funct and dividing the data

```
[ ] def Kmers_funct(seq, size=4):
        return [seq[x:x+size].lower() for x in range(len(seq) - size + 1)]
[ ]
    df['words'] = df.apply(lambda x: Kmers_funct(x['sequence']), axis=1)
    df = df.drop('sequence', axis=1)
   df
    df['words']
             [taat, aatt, atta, ttac, taca, acat, catt, att...
             [attt, tttt, tttt, ttta, ttac, taca, acaa, caa...
             [agag, gaga, agat, gata, atag, tagg, aggt, ggt...
             [tatg, atgt, tgta, gtat, tata, atat, tata, ata...
             [agaa, gaaa, aaat, aata, ataa, taat, aata, ata...
    11295 [cgac, gaca, acaa, caaa, aaag, aagt, agtt, gtt...
    11296
             [cata, atat, tatc, atct, tcta, ctac, taca, aca...
    11297
            [atac, tacc, accg, ccgc, cgcg, gcgg, cgga, gga...
    11298 [atta, ttat, tatt, attc, ttcc, tccg, ccga, cga...
    11299 [aatt, attc, ttca, tcat, catt, attt, ttta, tta...
    Name: words, Length: 22598, dtype: object
```

7. Converting Charcters to int64 matrcies using CountVectorizer

```
[ ] Y = df.iloc[:, 0].values

[ ] X = list(df['words'])
    for item in range(len(X)):
        X[item] = ' '.join(X[item])

[ ] from sklearn.feature_extraction.text import CountVectorizer
    cv = CountVectorizer(ngram_range=(4,4)) #The n-gram size of 4 is previously determined by testing
    X = cv.fit_transform(X)
```

### 8. Importing sklearn

### 9. Splitting the Data into Test and Train

```
import numpy as np
    import pandas as pd
    from sklearn import datasets
    from sklearn.model_selection import train_test_split
    from sklearn.preprocessing import StandardScaler
    from sklearn.metrics import accuracy_score
    from sklearn.linear_model import Perceptron
    from sklearn.linear_model import LogisticRegression
    from sklearn.svm import SVC
    from sklearn.tree import DecisionTreeClassifier
    from sklearn.ensemble import RandomForestClassifier
    from sklearn.neighbors import KNeighborsClassifier
[ ] # Splitting the human dataset into the training set and test set
    from sklearn.model_selection import train_test_split
    X_train, X_test, y_train, y_test = train_test_split(X,
                                                         test_size = 0.20,
                                                         random_state=1)
```

### 10. Testing the model prediction on a single sequence

11. Importing Pickle and saving the model

```
df3.at[0,'sequence']="ATTTTTACAAGAACAAGACATTTAACTTTAACTTTAGCTTTACCTTTATGATTATGTTTTATATTATATGGATG
[ ]
    df3['words'] = df3.apply(lambda x: Kmers_funct(x['sequence']), axis=1)
     df3 = df3.drop('sequence', axis=1)
[ ] X2 = list(df3['words'])
    for item in range(len(X2)):
        X2[item] = ' '.join(X2[item])
    from sklearn.feature_extraction.text import CountVectorizer
     cv = CountVectorizer(ngram_range=(4,4)) #The n-gram size of 4 is previously determined by testing
    X2 = cv.fit_transform(X2)
                                                                       + Code
                                                                                   + Text
[ ] ppn.predict(X2[0])
    array([0])
[ ] import pickle
    pickle.dump(ppn,open('model.pkl','wb'))
    model=pickle.load(open('model.pkl','rb'))
```

### 4.Results and Technical Discussion:

1. dataset after cleaning from nan



22598 rows × 2 columns

- 2. Training the Perceptron
- 3. Printing the accuracy score in both testing and training da

```
ppn = Perceptron(eta0=.01, max_iter=75)
ppn.fit(X_train, y_train)

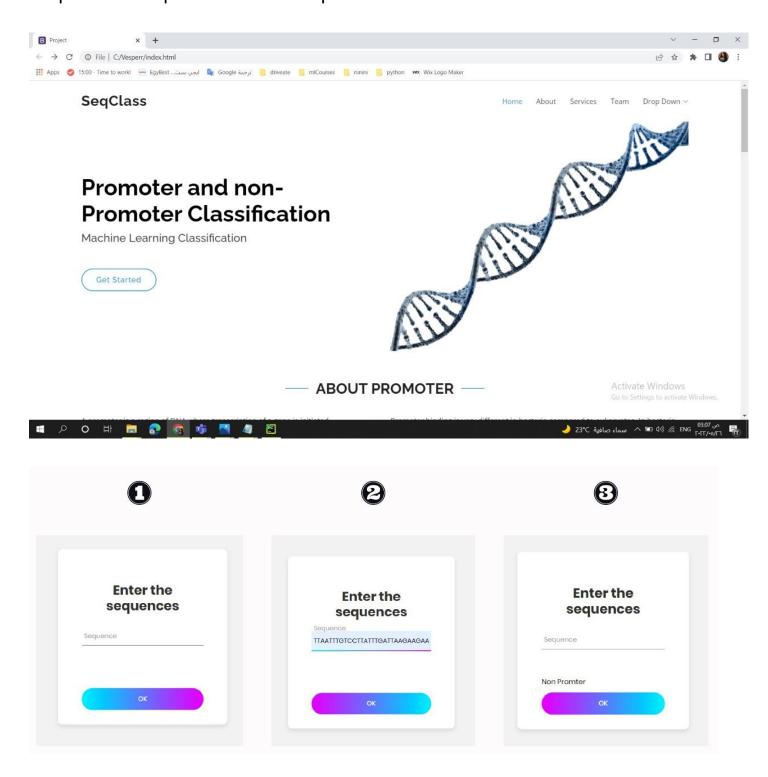
print("Test = " ,accuracy_score(ppn.predict(X_test), y_test))
print("Train = " ,accuracy_score(ppn.predict(X_train), y_train))

Test = 0.7331858407079646
Train = 0.9945790463546853
```

The main result: is how accurate the perceptron model \_after training\_ in classifying DNA sequences into a promoter and non-Promoter region -The model gave the highest accuracy on train data and 73.33% on test data

### Final product:

A website linked with the perceptron model to predict whether any given DNA sequence is a promoter or non promoter



### 5.Conclusions:

- to further study and analyze promoter and non-promoter regions in order to treat certain genetic diseases there needed to be an optimized method that is able to classify them with high accuracy
- that's way we build a perceptron model to do just that and it resulted in 73.33 % accuracy score and a time complexity of o(t) as it's a liner classifier
  - # then we proceeded to build a full functioning responsive website
  - that can predict whether the inputted sequence is a promoter or nonpromoter

### 6.References:

- Sharan, Roded (4 January 2007). "Analysis of Biological Networks:
   Transcriptional Networks Promoter Sequence Analysis" (PDF). Tel Aviv
   University. Retrieved 30 December 2012.
- 2. Asgari E., Mofrad M. R. K. (2015). Continuous distributed representation of biological sequences for deep proteomics and genomics. PLoS ONE 10: e0141287. 10.1371/journal.pone.0141287
- 3. NCBI, article: PMC6848157
- 4. www.genome.gov/genetics-glossary/Promoter
- 5. Copland JA, Sheffield-Moore M, Koldzic-Zivanovic N, Gentry S, Lamprou G, Tzortzatou-Stathopoulou F, Zoumpourlis V, Urban RJ, Vlahopoulos SA (June 2009). "Sex steroid receptors in skeletal differentiation and epithelial neoplasia is tissue-specific intervention possible"
- 6. Hobbs K, Negri J, Klinnert M, Rosenwasser LJ, Borish L (December 1998). "Interleukin-10 and transforming growth factor-beta promoter polymorphisms in allergies and asthma". American Journal of Respiratory and Critical Care Medicine.
- 7. Burchard EG, Silverman EK, Rosenwasser LJ, Borish L, Yandava C, Pillari A, Weiss ST, Hasday J, Lilly CM, Ford JG, Drazen JM (September 1999).
  "Association between a sequence variant in the IL-4 gene promoter and

- FEV (1) in asthma". American Journal of Respiratory and Critical Care Medicine
- 8. www.frontiersin.org/articles/10.3389/fbioe.2019.00305/full
- 9. Kulozik AE, Bellan-Koch A, Bail S, Kohne E, Kleihauer E (May 1991).
  "Thalassemia intermedia: moderate reduction of beta globin gene transcriptional activity by a novel mutation of the proximal CACCC promoter element"
- 10. Petrij F, Giles RH, Dauwerse HG, Saris JJ, Hennekam RC, Masuno M, Tommerup N, van Ommen GJ, Goodman RH, Peters DJ (July 1995).

  "Rubinstein-Taybi syndrome caused by mutations in the transcriptional co-activator CBP". Nature. 376 (6538): 348–51
- 11. www.simplilearn.com/tutorials/deep-learning-tutorial/perceptron
- 12. www.javatpoint.com/perceptron-in-machine-learning
- 13. www.code4example.com/pseudocode/pseudocode-examples/

## 7.Appendix A:

#### Source code:

https://colab.research.google.com/drive/IfGTjUtyMEY3jWmSJCMxp4wjXcHWT0xg?usp=sharing

### Website (local hosting):

https://drive.google.com/drive/folders/lhwFEjyQf8Rt3FFJeaserfLvlRcg8Vnp9?usp=sharing