

# Bioinformatics Algorithms

## COS-BIOL-530/630

### Lecture03

| <b>Days &amp; Times</b> | <b>Room</b>                    | <b>Meeting Dates</b>    |
|-------------------------|--------------------------------|-------------------------|
| Tu 2:00PM - 3:50PM      | Thomas Gosnell Hall (GOS)-2178 | 01/13/2025 - 04/28/2025 |
| Th 2:00PM - 3:50PM      | Thomas Gosnell Hall (GOS)-2178 | 01/13/2025 - 04/28/2025 |

Instructor:

Fernando Rodriguez

email: [frvsbi@rit.edu](mailto:frvsbi@rit.edu)

Office: Orange Hall 1311

# Bioinformatics Algorithms

## COS-BIOL-530/630

### Lecture03

#### Announcements

##### **Week3:**

Lecture03

Lab03

- Discussion 3
- Activity 3

Activity2 (Filamin-A Dotplot) due on Thursday

Quiz 3 opens on Friday. Due on Tuesday 2pm

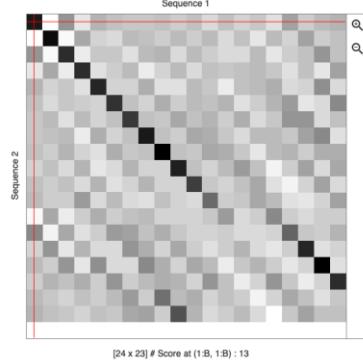
- Lab00 -> PHYLIP (next week)
- **Exam2: Tuesday April 15<sup>th</sup> (Schedule updated)**
- Oedipus?

>Sequence1

BIQINFORMATICSALGQRITHMS

>Sequence2

BIQINFORMATICEURITHMICS



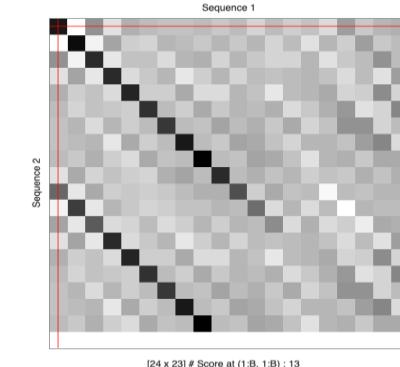
Local alignment  
(match)

>Sequence1

BIQINFORMATICSALGQRITHMS

>Sequence2

BIQINFORMATICINFORMATIC



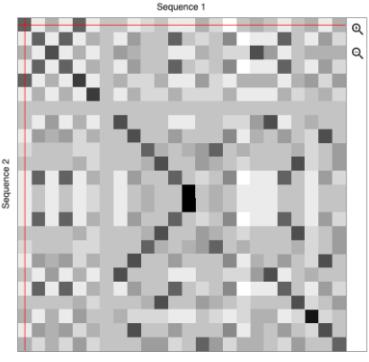
Repeat

>Sequence1

BIQINFORMATICSALGQRITHMS

>Sequence2

BIQINFORMATICCITAMRITHMS



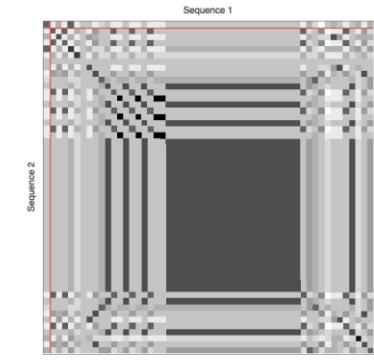
Inverted repeat

>Sequence1

BIQINFORMATICTCTCC|TTTTTTTTTTTTTTTTT|ISALGQRITHMS

>Sequence2

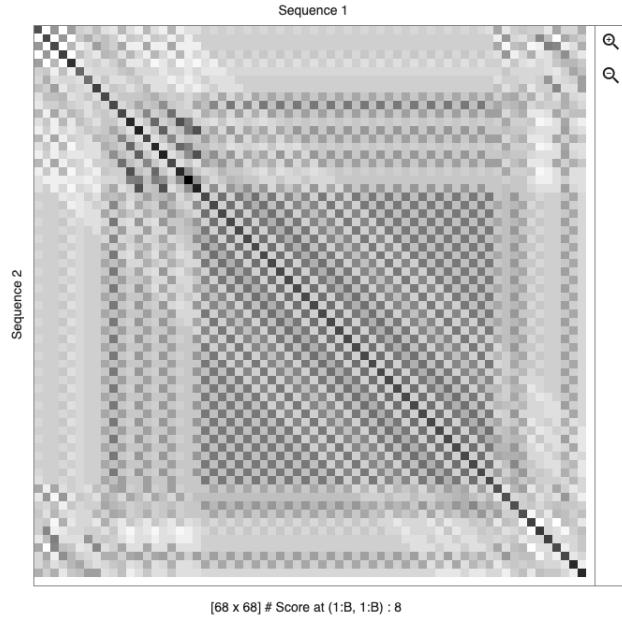
BIQINFORMATICTCTCT|TTTTTTTTTTTTTTTTT|ITAMRITHMS



Repeat (TIC)

Low complexity (T's)

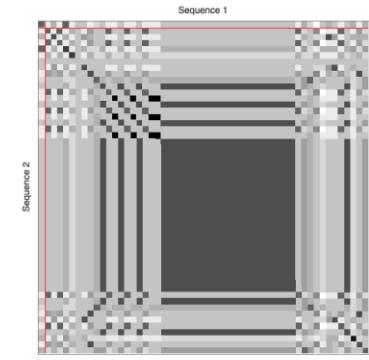
window size = 1



```
>Sequence1  
BIQINFORMATICTICTICCTATATATATATATATATATATATATATATATAISALGQRITHMS  
>Sequence2  
BIQINFORMATICTICTICCTATATATATATATATATATATATATATATAISALGQRITHMS
```

window size = 2

```
>Sequence1  
BIQINFORMATICTICTICCTTTTTTTTTTTTTTTTTTISALGQRITHMS  
>Sequence2  
BIQINFORMATICTICTICCTTTTTTTTTTTTTTTTTTITAMRITHMS
```



## Repeat (TIC)

## Low complexity (T's)

window size = 1

# Alignments algorithms

## - Lecture02 & Lecture03 -

- Pairwise Alignment
  - Global
  - Local
  - Scoring system
- Multiple Sequence Alignment (MSA)
- Heuristic – Database search

# Dynamic programming algorithm (Quiz2 –Q1)

Global alignment: Needleman-Wunsch algorithm

Example from Discussion 2.2

|                   |          |
|-------------------|----------|
| Subject sequence: | GACGCAGT |
| Query sequence:   | ACCCAT   |



Let's assume the Score Matrix:

- ❖ Match = 5
- ❖ Mismatch = -2
- ❖ Gap penalty = -6

  
GACGCAGT  
\*\*|\*\* \*  
\_ACCCA\_T

Let's assume the Score Matrix:

- ❖ Match = 4
- ❖ Mismatch = -6
- ❖ Gap penalty = -9

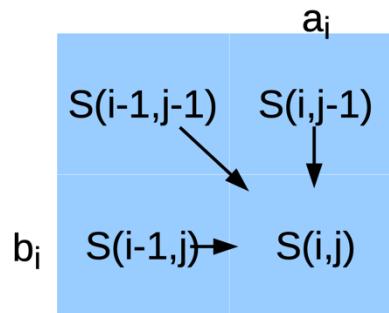
  
GACGCAGT  
\*\*|\*\* \*  
\_ACCCA\_T

Alignment Score?

# Dynamic programming algorithm

- Local alignment: **Smith-Waterman algorithm**  
It is just a variation of Global (Needleman-Wunsch algorithm)

- Performed the same way (steps) as global alignment
- To find the best, overall, alignment of subsets of sequences
- Negative scores = 0
- Find the best score within the matrix
- Traceback the best score to the first zero



# Dynamic programming algorithm

- Local alignment: **Smith-Waterman algorithm**  
It is just a variation of Global (Needleman-Wunsch algorithm)

Example from Discussion 2.2

Sequence1: GACGCAGT  
Sequence2: ACCCAT

Scoring system:  
Match: 5  
Mismatch: -2  
Gap: -6



| <i>S</i>             |   | <b>G<sub>1</sub></b> | <b>A<sub>2</sub></b> | <b>C<sub>3</sub></b> | <b>G<sub>4</sub></b> | <b>C<sub>5</sub></b> | <b>A<sub>6</sub></b> | <b>G<sub>7</sub></b> | <b>T<sub>8</sub></b> |
|----------------------|---|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|----------------------|
|                      | 0 | 0                    | 0                    | 0                    | 0                    | 0                    | 0                    | 0                    | 0                    |
| <b>A<sub>1</sub></b> | 0 | 0                    | 5                    | 0                    | 0                    | 0                    | 5                    | 0                    | 0                    |
| <b>C<sub>2</sub></b> | 0 | 0                    | 0                    | 10                   | 4                    | 5                    | 0                    | 3                    | 0                    |
| <b>C<sub>3</sub></b> | 0 | 0                    | 0                    | 5                    | 8                    | 9                    | 3                    | 0                    | 1                    |
| <b>C<sub>4</sub></b> | 0 | 0                    | 0                    | 5                    | 3                    | 13                   | 7                    | 1                    | 0                    |
| <b>A<sub>5</sub></b> | 0 | 0                    | 5                    | 0                    | 3                    | 7                    | 18                   | 12                   | 6                    |
| <b>T<sub>6</sub></b> | 0 | 0                    | 0                    | 3                    | 0                    | 1                    | 12                   | 16                   | 17                   |
| Score: 18            |   |                      |                      |                      |                      |                      |                      |                      |                      |



ACCCA  
\* \* | \* \*  
ACGCA

# Alignments algorithms

## – Lecture02 & Lecture03 –

- Pairwise Alignment
  - Global
  - Local
  - Scoring system
- Multiple Sequence Alignment (MSA)
- Heuristic – Database search

# Multiple sequence alignments (MSA)

- Alignments that contain more than two sequences.
- Multiple alignments improve:
  - Accuracy of alignments between the sequence pairs.
  - Reveals patterns of conserved residues that are not obvious when looking at only two sequences.
- But:
  - Required more computing resources.
  - If alignment is incorrect in an active site residue, any function inference may be wrong.
  - Sometimes, we need “pre-aligned” sequences to generate multiple alignments.

# Multiple sequence alignments

|            |  |
|------------|--|
| HBB_HUMAN  | -----VHLTPEEKSAVTALWGKVNV--VDEVGGEALGRLLVVYPWTQRFFESFGDLST       |
| HBB_HORSE  | -----VQLSGEEKAAVLALWDKVNV--EEEVGGEALGRLLVVYPWTQRFFDSFGDLSN       |
| HBA_HUMAN  | -----VLSPADKTNVKAAGKVGAGAHAGEYGAELERMFLSFPTTKTYFPHF-DLS-         |
| HBA_HORSE  | -----VLSAADKTNVKAAWSKVGGHAGEYGAELERMFLGFPTTKTYFPHF-DLS-          |
| GLB5_PETMA | PIVDTGSVAPLSAAEKTKIRSAWAPVYSTYETSGVDILVKFFTSTPAAQEFFPKFKGLTT     |
| MYG_PHYCA  | -----VLSEGEWQLVLHVWAKVEADVAGHGQDILIRLFKSHPTLEKFDRFKHLKT          |
| LGB2_LUPLU | -----GALTESQAALVKSSWEENANIPKHTHRFFILVLEIAPAACKDLFSFLKGTE         |
|            | * : : : * . : : * : * : .  |
| HBB_HUMAN  | PDAVMGNPKVKAHGKIVLGAFSDGLAHLDN-----LKGTFAATLSELHCDKLHVDPENFRL    |
| HBB_HORSE  | PGAVMGNPKVKAHGKIVLHSFGEGVHLDN-----LKGTFAALSELHCDKLHVDPENFRL      |
| HBA_HUMAN  | ---HGSAQVKGHGKIVADALTNAVAHVD-----MPNALSAISDLHAKLRLVDPVNFKL       |
| HBA_HORSE  | ---HGSAQVKAHGKIVGDALTLAVGHLD-----LPGALSNLSDLHAKLRLVDPVNFKL       |
| GLB5_PETMA | ADQLKKSSADVRWAHERITNAVNDAVASMDT-----EKMSMKLRDLSGKHAKSFQVDPQYFKV  |
| MYG_PHYCA  | EAEMKASEDLKKHGVTVLTALGAILKKKGH-----HEAEELKPLAQSHATKHKIPIKYLEF    |
| LGB2_LUPLU | VP-----QNNPELQAHAGKIVFKLVYEAAIQLQVTGVVVTDATLKNLGSVHVSKG-VADAHFPV |
|            | . : : * . : . : * . * . : : .                                    |
| HBB_HUMAN  | LGNVLVCVLAHHFGKEFTPQVAAQKVVAGVANALAHKYH-----                     |
| HBB_HORSE  | LGNVLVVVLARHFGKDFTPELQASYQKVVAGVANALAHKYH-----                   |
| HBA_HUMAN  | LSHCLLVTLA AHLPAEFTP AVHASLDKFLASVSTVLT SKYR-----                |
| HBA_HORSE  | LSHCLLSTLAVHLPNDFTPAVHASLDKFLSSVSTVLT SKYR-----                  |
| GLB5_PETMA | LAAVIADTVAA G-----DAGFEKLMSM CILLRSAY-----                       |
| MYG_PHYCA  | I SEAI IHVILHSRHPGDFGADAQGAMNKALELFRKDIAAKYKELGYQG               |
| LGB2_LUPLU | VKEAILKTIKEVVGAKWSEELNSAWTIAYDELAIVIKKEMNDAA-----                |
|            | : : : . . . . : .  |

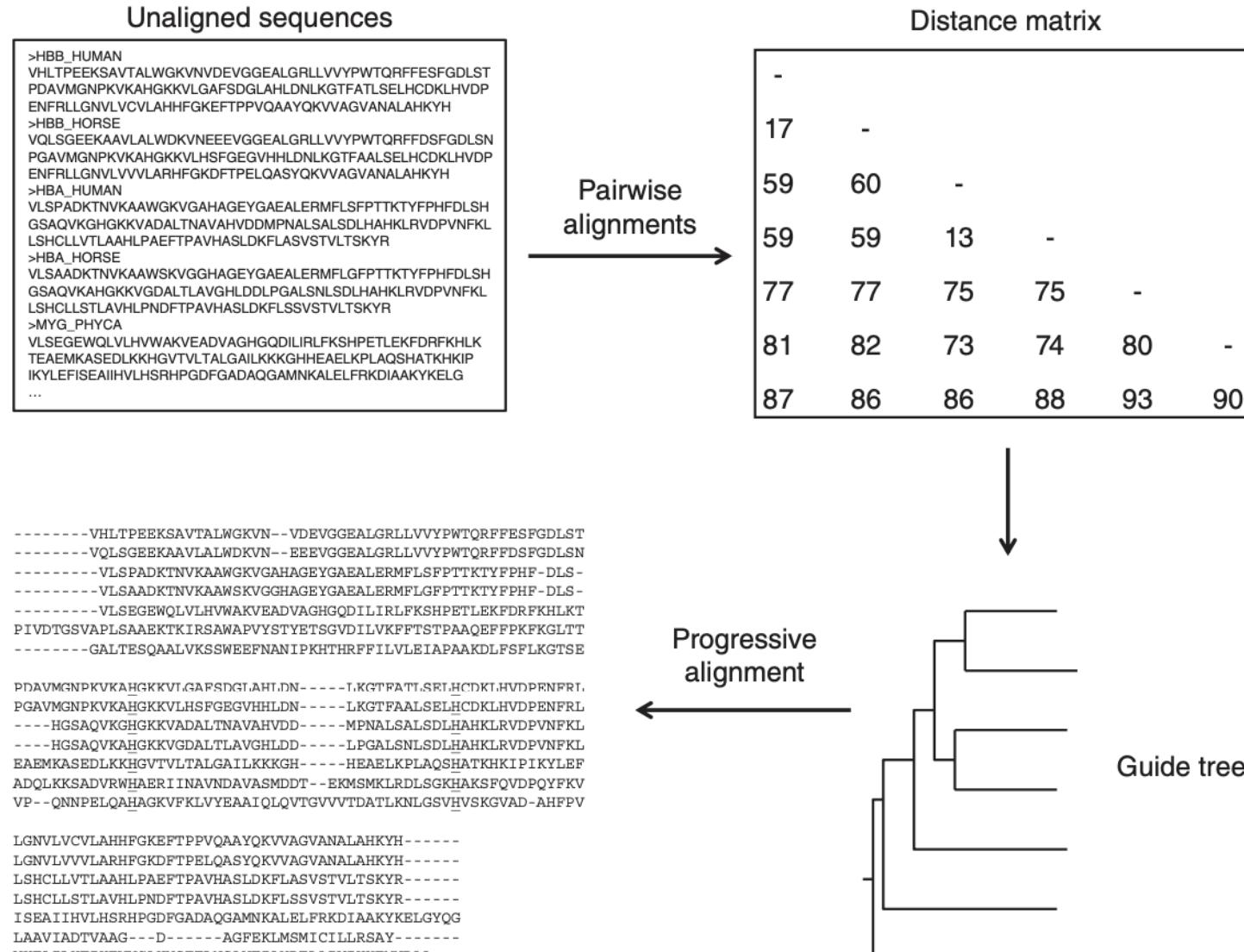
**Figure 8.1** An example multiple sequence alignment of seven globin protein sequences. One position is highlighted.

When constructing an MSA, one must also consider insertions and deletions from diverged sequences.

This means that MSA packages must be able to find an arrangement of null characters or “gaps” that maximizes the alignment of homologous residues.

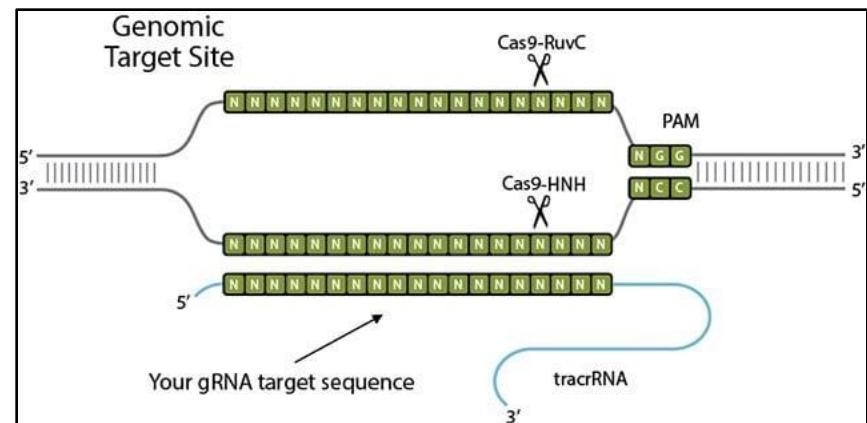
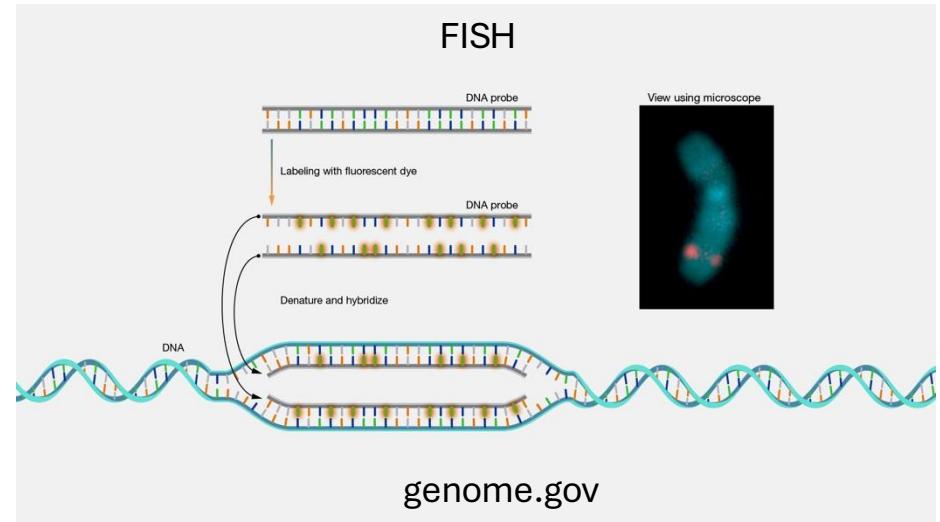
Given a scoring scheme for residue matches and scores for gaps, one can attempt to find an MSA that produces the best overall score (and, thereby, the best overall alignment).

# Progressive alignment process



# Multiple sequence alignments - Applications

- DNA/Protein: look for functional homology (consensus sequences)
- DNA: highlight conserved regions in the genome
  - ✓ Transcription factor binding sites
  - ✓ Regions to design probes/oligos (fluorescent in situ hybridization - FISH, PCR)
- Proteins: conserved sequence regions (~functionality)
  - ✓ Active sites (eg. cleave site of an endonuclease)
  - ✓ Predicting 3D structure (eg. [AlphaFold](#))
  - ✓ Identify protein family members
  - ✓ Experiment design (CRISPR/Cas modifications)



CRISPR/Cas system  
Source: Sigma-Aldrich

# Alignments algorithms

## - Lecture02 & Lecture03 -

- Pairwise Alignment
  - Global
  - Local
  - Scoring system
- Multiple Sequence Alignment (MSA)
- Heuristic – Database search

# Pairwise alignments & Databases (DBs)



- It all started in the early 1980's:
  - European Molecular Biology Laboratory - EMBL (1980)
  - GenBank (1982).
    - to establish a central database of DNA sequences rather than have scientists submit sequences to journals.
- By the late 1980's there were too many sequences.
  - Transition to digital format
  - But the question was: How to efficiently find related (homologous) sequences in the DBs??
- Development of tools to allow access to sequence databases (FASTP 1984, BLAST 1990, ENTREZ 1992).

# Pairwise alignments & Databases (DBs)



WHY not to run pairwise alignments against the DB?

|         |          | GenBank  |           | WGS   |           |
|---------|----------|----------|-----------|-------|-----------|
| Release | Date     | Bases    | Sequences | Bases | Sequences |
| 3       | Dec 1982 | 680338   | 606       |       |           |
| 62      | Dec 1989 | 37183950 | 31229     |       |           |

... against over 250 million sequences??

|     |          |               |           |                |            |
|-----|----------|---------------|-----------|----------------|------------|
| 264 | Dec 2024 | 5085904976338 | 254365075 | 32983029087303 | 3957195833 |
|-----|----------|---------------|-----------|----------------|------------|

# Database searches

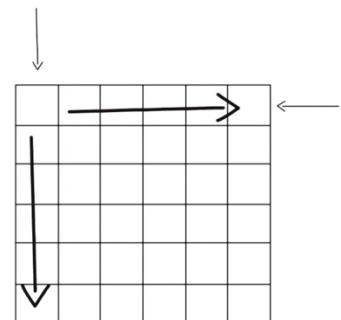


- Pairwise alignments are not adequate
- FASTP (Lipman and Pearson 1985)
- FASTA (Pearson and Lipman 1998)
- BLAST (Altschul et al. 1990)
- And others: heuristic methods
  - Fast approximation to dynamic programming
  - Contributes to a higher speed execution
  - But we have to make some assumptions...

# Database searches



- Let's say we want to search a query against the UniProtKB/Swiss-Prot protein knowledgebase (2024):
  - 57,1864 sequence entries
  - 207,016,062 amino acids
  - Query: 50 amino acids
  - How would you do it via dynamic programming?

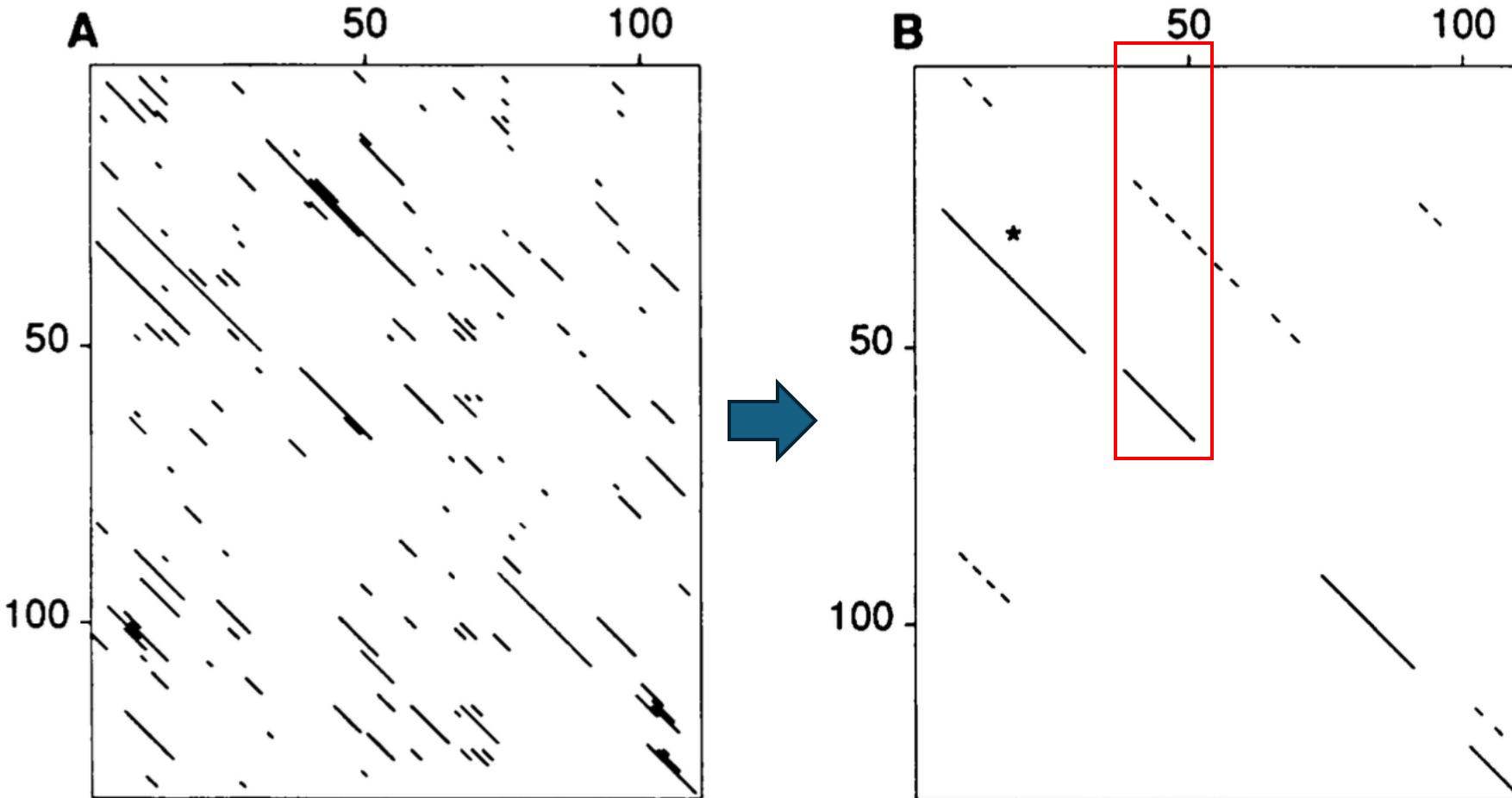


# FAST algorithms

- FASTP (Lipman and Pearson 1985) was the first widely used program designed for database similarity searching.
- FAST (FASTP, FASTA) **assumes that small local areas of similarity are more common in two related sequences than in two unrelated sequences.**
- The FAST algorithms can be divided into four major steps:
  - Identify regions of identity (words). The word length parameter or k-tuple is called ***ktup*** (equivalent to word size ***W*** in BLAST).
  - Scan the regions using a scoring matrix and save the best initial regions (using a Threshold)
  - Join initial regions with scores > Threshold (T)
  - Optimized initial score

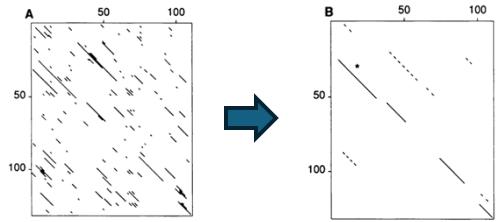
# FASTA algorithm

Find all regions of local identity

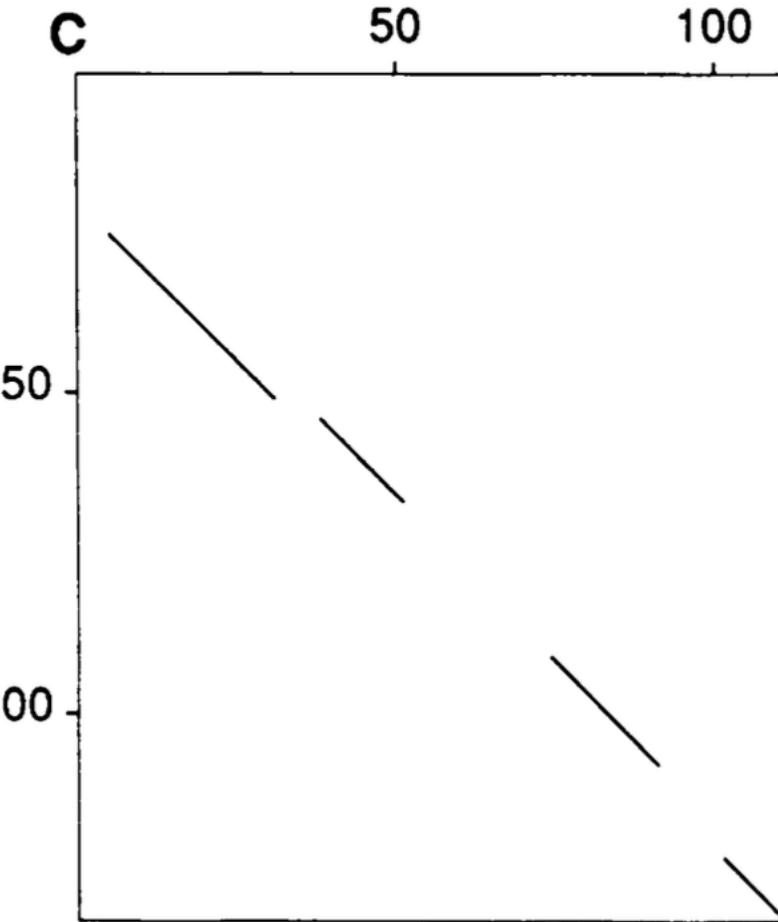


Select the best regions of local identity using a threshold score  $*$  ( $T$ ).

Note that some regions are mutually exclusive (red rectangle).



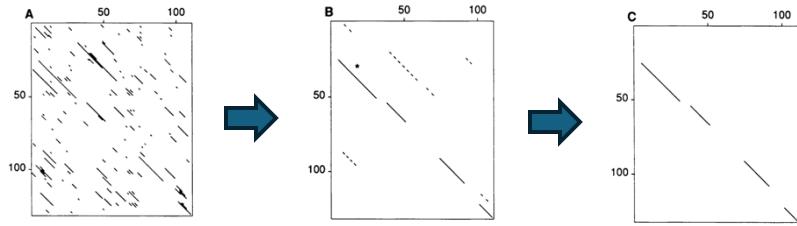
# FASTA algorithm



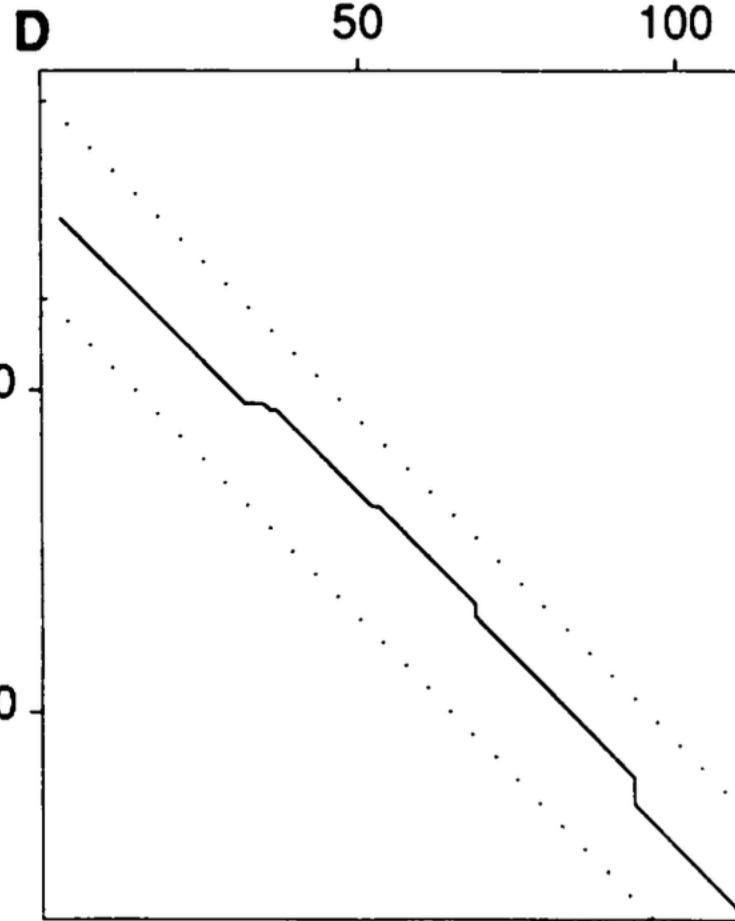
After the initial round of scoring, the top diagonal are selected.

Dashed lines (mutually exclusive) fell off. They were mutually exclusive with the best score.

Now, join the regions of the highest similarity.



# FASTA algorithm



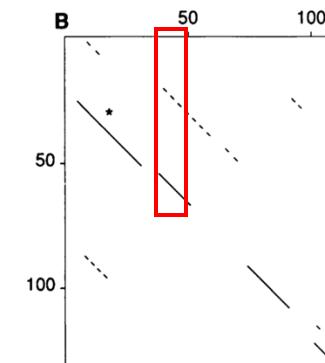
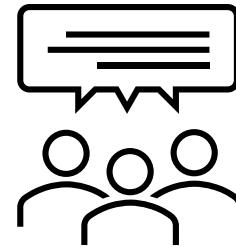
Recalculate an optimized alignment centered around the highest-scoring initial region.

Run dynamic programming along the diagonal to optimize the alignment.

The Smith-Waterman algorithm is applied to yield the optimal pairwise alignments between the two sequences.

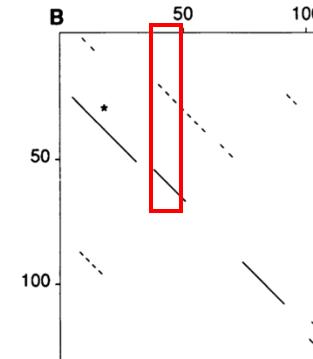
# FASTA algorithm

- FAST assumption: small areas of local similarity with the highest scores are most likely to be evolutionary conserved regions.
- Do you see any problem with this assumption?



# FASTA algorithm

- FAST assumption: small areas of local similarity with the highest scores are most likely to be evolutionary conserved regions.
- But we also assume **duplications** and **gene conversions** are not present (exclusive areas in the dot blot).
  - Initially, not many sequences deposited carried **duplications** (single copy human genes, mitochondrial genes).
  - More organisms being sequenced: alignments can show tricked scenarios where the FAST algorithm is far from perfect.



# FASTA algorithm

- Instead of doing a full dynamic programming query vs. database.
- First, check if there is enough k-tuple (*ktup*) matches to be worthy.
- Then, extend and merge the k-tuples.
- It is similar to dynamic programming....but faster.

**FASTA Sequence Comparison at the U. of Virginia**

New: Annotation features available for Uniprot/SwissProt/PIR1 library searches.

**UVa FASTA Server**

**About**

- Getting started
- [fasta\\_guide.pdf](#)

**Other FASTA Servers**

- EMBL-EBI
- KEGG (Japan)

**References**

- FASTA
- FASTX/FASTY
- Statistics
- FASTS/FASTF

**Software**

- FASTA v36
- ChangeLog
- Downloads
- Sequence Libraries

**Other resources**

- PSI-Search2
- BLASTP with annotations
- CHAPS - Convert HTML and Profiles
- FASTA Exercises
- NCBI BLAST server
- EMBL-EBI Server

The **FASTA** programs find regions of local or global similarity between Protein or DNA sequences, either by searching Protein or DNA databases, or by identifying local duplications within a sequence. Other programs provide information on the statistical significance of an alignment. Like **BLAST**, **FASTA** can be used to infer functional and evolutionary relationships between sequences as well as help identify members of gene families.

|   |   |
|---|---|
| <b>Protein</b>  | <b>Nucleotide</b>   |
| <ul style="list-style-type: none"><li>• Protein-protein <b>FASTA</b></li><li>• Protein-protein Smith-Waterman (<b>ssearch</b>)</li><li>• Global Protein-protein (Needleman-Wunsch) (<b>gsearch</b>)</li><li>• Global/Local protein-protein (<b>glsearch</b>)</li><li>• Protein-protein with unordered peptides (<b>fasts</b>)</li><li>• Protein-protein with mixed peptide sequences (<b>fastf</b>)</li></ul> | <ul style="list-style-type: none"><li>• Nucleotide-Nucleotide (DNA/RNA <b>fasta</b>)</li><li>• Ordered Nucleotides vs Nucleotide (<b>fastm</b>)</li><li>• Un-ordered Nucleotides vs Nucleotide (<b>fasts</b>)</li></ul> |
| <b>Translated</b>   | <b>Statistical Significance</b>   |
| <ul style="list-style-type: none"><li>• Translated DNA (with frameshifts, e.g. ESTs) vs Proteins (<b>fastx/fasta</b>)</li><li>• Protein vs Translated DNA (with frameshifts) (<b>tfastx/tfasta</b>)</li><li>• Peptides vs Translated DNA (<b>tfasts</b>)</li></ul>  | <ul style="list-style-type: none"><li>• Protein vs Protein shuffle (<b>prss</b>)</li><li>• DNA vs DNA shuffle (<b>prss</b>)</li><li>• Translated DNA vs Protein shuffle (<b>prfx</b>)</li></ul>                         |
| <b>Local Duplications</b>   |   |
| <ul style="list-style-type: none"><li>• Local Protein alignments (<b>lalign</b>)</li><li>• Plot Protein alignment "dot-plot" (<b>palign</b>)</li><li>• Local DNA alignments (<b>lalign</b>)</li><li>• Plot DNA alignment "dot-plot" (<b>palign</b>)</li></ul>   |   |

[https://fasta.bioch.virginia.edu/fasta\\_www2/fasta\\_list2.shtml](https://fasta.bioch.virginia.edu/fasta_www2/fasta_list2.shtml)

## Other search options:

Scoring matrix: **open:** **ext:** **Ktup:** Statistical estimates

Blosum50 (25%) ▾ -10 ▾ -2 ▾ ktup = 2 ▾ Default ▾

Output limits: Max  Show Histogram

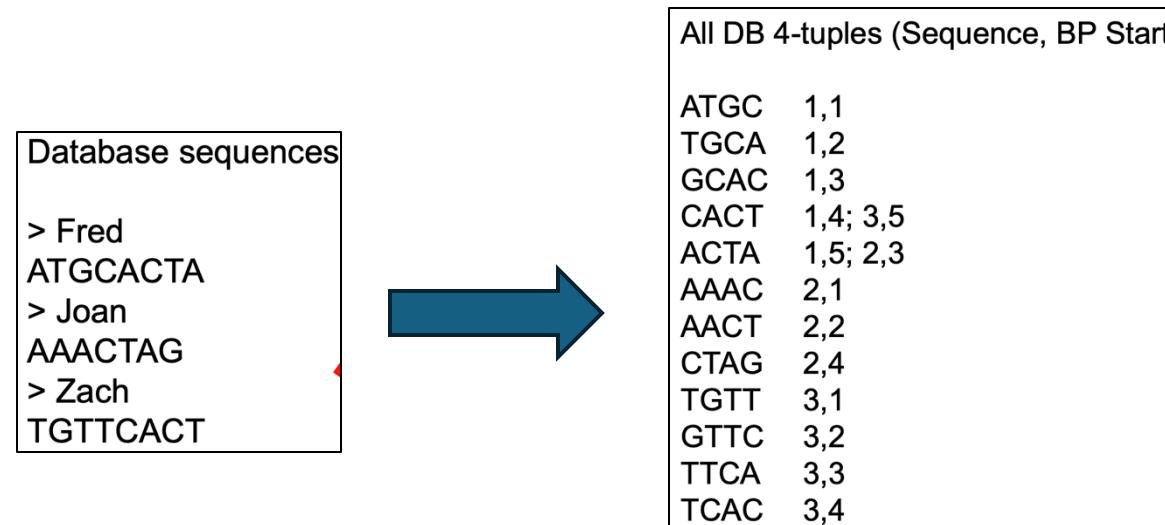
E(): Best E(): aligns:

5.0 ▾    Hide Alignments

# Database search

WHAT IF....

- database sequences are preprocessed with a list of all seed of size k.
- Then, we only need to search the query against the indexed database



# BLAST

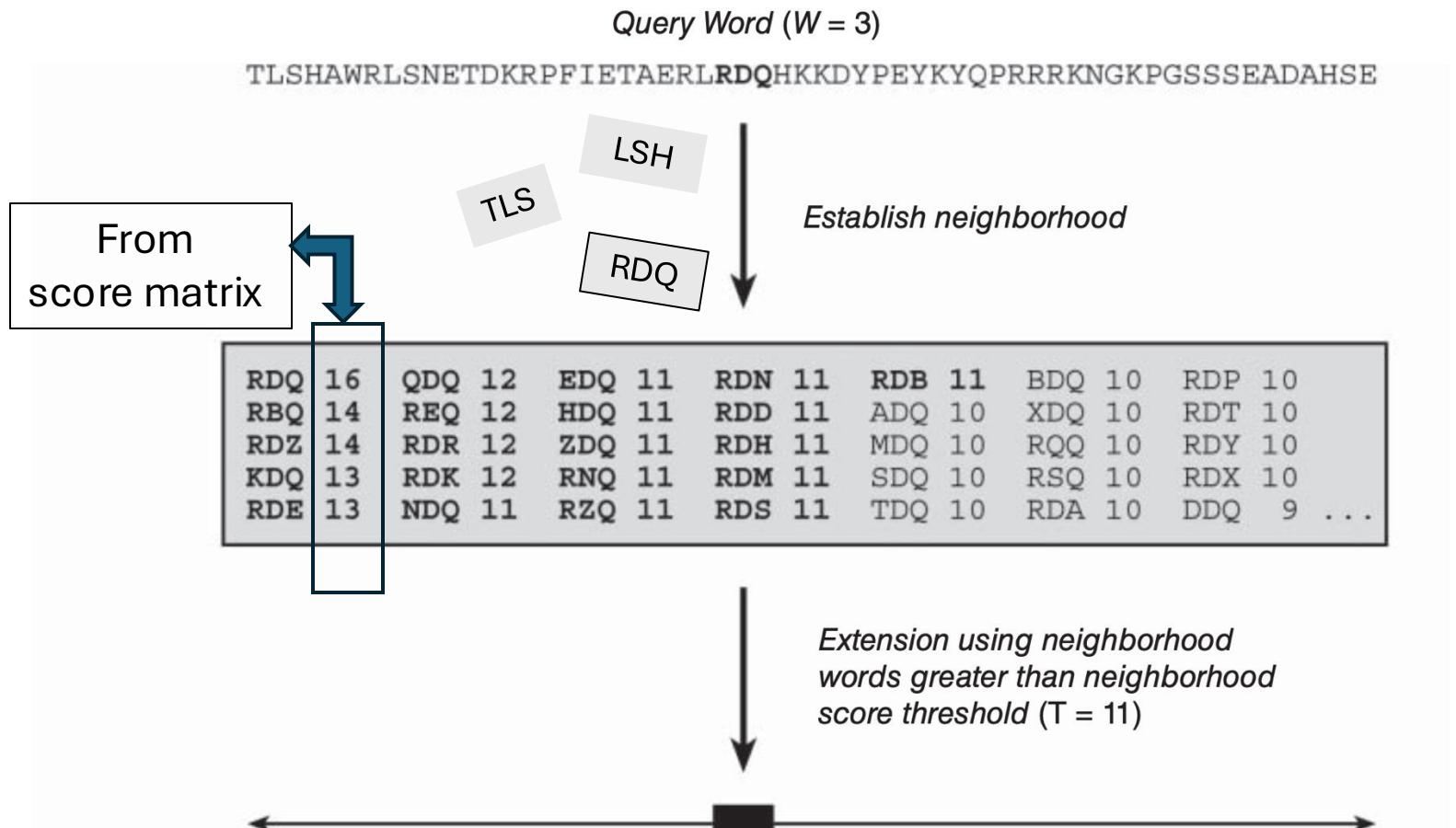
- Basic Local Alignment Search Tool (Altschul et al. 1990) in Perusal.
- BLAST heuristically finds ***high-scoring segment pairs (HSPs)***:
  - Identical length segments from 2 sequences with statistically significant match scores
- Not only the best region of local alignment, but also whether there are other plausible alignments.
- BLAST method begins by seeding the search with a small subset of letters (query **word**).

## BLAST algorithms

---

| Program | Query                             | Database                          |
|---------|-----------------------------------|-----------------------------------|
| BLASTN  | Nucleotide                        | Nucleotide                        |
| BLASTP  | Protein                           | Protein                           |
| BLASTX  | Nucleotide, six-frame translation | Protein                           |
| TBLASTN | Protein                           | Nucleotide, six-frame translation |
| TBLASTX | Nucleotide, six-frame translation | Nucleotide, six-frame translation |

## The initiation of a BLAST search



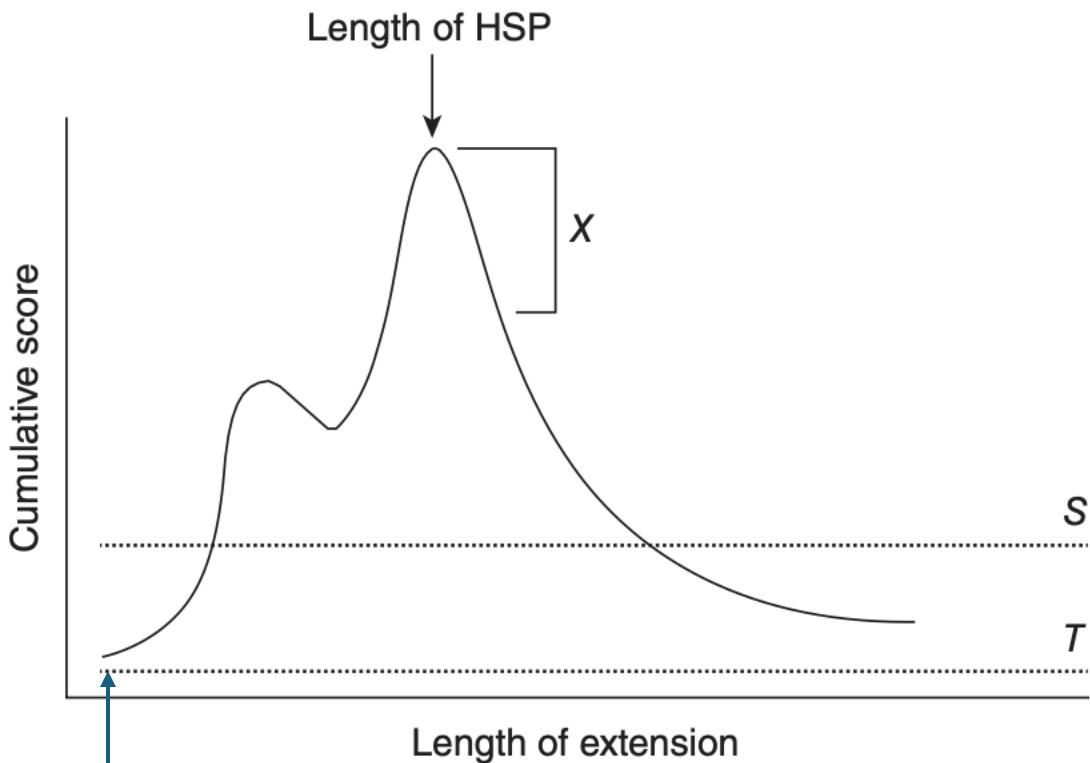
- **Query word** ( $w=3$ ) is RDQ
- TLS, LSH, ...RDQ

- BLAST search RDQ in the DB, but also related words (with conservative substitutions)
- **Neighborhood:** related words using scoring matrix

- We use a cut-off in the neighborhood: **score threshold** ( $T$ )

- Once the query (RDQ) word is aligned with another word (REQ) from DB (with  $T > 11$ ).
- Now BLAST attempt to extend the alignment in both directions.

## How to determine the maximal length of extension



- The number of residues (**extension**) is plotted vs. the **cumulative score** from the alignment.
- As the alignment gets extended:
  - matches with a positive score (conservative substitutions): score increase
  - Mismatches and gaps: score decrease
- As soon as the cumulative score breaks the **score threshold S**, the alignment is reported in the BLAST output.
- The resulting alignment is called a high-scoring segment pair, or HSP.

T: neighborhood score threshold  
S: minimum score to return a BLAST hit  
X: significance decay

RDQ  
|||  
REQ

## After HSP identification

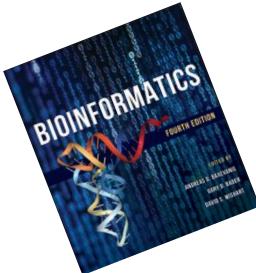
| Sequences producing significant alignments     |                        |             |                 |             |             |         |            | Download | Select columns | Show 100 | ? |
|--|------------------------|-------------|-----------------|-------------|-------------|---------|------------|----------|----------------|----------|---|
|  |                        | Description | Scientific Name | Common Name | Query Cover | E value | Per. Ident | Acc. Len | Accession      |          |   |
| <input checked="" type="checkbox"/> select all | 100 sequences selected |             |                 |             |             |         |            |          |                |          |   |

### Box 3.2 The Karlin-Altschul Equation

As one might imagine, assessing the putative biological significance of any given BLAST hit based simply on raw scores is difficult, since the scores are dependent on the composition of the query and target sequences, the length of the sequences, the scoring matrix used to compute the raw scores, and numerous other factors. In one of the most important papers on the theory of local sequence alignment statistics, Karlin and Altschul (1990) presented a formula which directly addresses this problem. The formula, which has come to be known as the Karlin-Altschul equation, uses search-specific parameters to calculate an expectation value (*E*). This value represents the number of HSPs that would be expected purely by chance. The equation and the parameters used to calculate *E* are as follows:

$$E = kmNe^{-\lambda s}$$

where *k* is a minor constant, *m* is the number of letters in the query, *N* is the total number of letters in the target database,  $\lambda$  is a constant used to normalize the raw score of the high-scoring segment pair, with the value of  $\lambda$  varying depending on the scoring matrix used; and *S* is the score of the high-scoring segment pair.



- How to know if the alignment is significant?
- The cumulative score is used, along other parameters, to get a new value: ***E*** value :
  - ***E*** gives the number of expected HSPs having a score of *S* or more that BLAST would find purely by chance.
  - ***E*** provides a measure of whether an HSP is a false positive.
  - Lower ***E*** values: greater biological significance.
  - Higher ***E*** values: more likely to get a hit like that by chance.

## After HSP identification

| Sequences producing significant alignments     |                        |             |                 |             |             |         |            |          |
|--|------------------------|-------------|-----------------|-------------|-------------|---------|------------|----------|
|  |                        | Description | Scientific Name | Common Name | Query Cover | E value | Per. Ident | Acc. Len |
| <input checked="" type="checkbox"/> select all | 100 sequences selected |             |                 |             |             |         |            |          |

***E*** (HSP with Score > S), also called ***E* value**.  
The expected value (***E***) is a parameter that describes the number of hits one can expect to see by chance when searching in a particular database.

$$E = kmne^{-\lambda S}$$

Expected number of alignments (by chance) →  $E = kmne^{-\lambda S}$

Labels and components:

- A minor constant
- Scaling factor
- Raw score
- Normalized score
- Length of query
- Length of database
- Search space

- How to know if the alignment is significant?
- The cumulative score is used, along other parameters, to get a new value: ***E* value** :
  - ***E*** gives the number of expected HSPs having a score of S or more that BLAST would find purely by chance.
  - ***E*** provides a measure of whether an HSP is a false positive.
  - Lower ***E*** values: greater biological significance.
  - Higher ***E*** values: more likely to get a hit like that by chance.

## After HSP identification

*E* value: 1

Expect to get 1 **false positive** in a DB  
of this size given the alignment score

*E* value: 0.01

Expect to get 0.01 **false positive** in a  
DB of this size given the alignment  
score

- How to know if the alignment is significant?
- The cumulative score is used, along other parameters, to get a new value: ***E* value** :
  - ***E*** gives the number of expected HSPs having a score of S or more that BLAST would find purely by chance.
  - ***E*** provides a measure of whether an HSP is a false positive.
  - Lower ***E*** values: greater biological significance.
  - Higher ***E*** values: more likely to get a hit like that by chance.

# BLASTN

- Nucleotide query
- Nucleotide database
- BLASTN uses larger word size than protein searches
  - Default word size is ..... nucleotides?
  - Increases the likelihood of unique strings
  - Reduces the DB sequences considered

— Algorithm parameters

**General Parameters**

|                              |   |   |
|------------------------------|---|---|
| Max target sequences         | 100 <input type="button" value="▼"/>  | Select the maximum number of aligned sequences to display <a href="#">?</a> |
| Short queries                | <input checked="" type="checkbox"/> Automatically adjust parameters for short input sequences <a href="#">?</a> |   |
| Expect threshold             | 0.05 <input type="button" value="?"/>   |   |
| Word size                    | <input type="button" value="?"/>  |   |
| Max matches in a query range | 0 <input type="button" value="?"/>  |   |

**Scoring Parameters**

|                       |   |
|-----------------------|---|
| Match/Mismatch Scores | 1,-2 <input type="button" value="▼"/> <a href="#">?</a>   |
| Gap Costs             | Linear <input type="button" value="▼"/> <a href="#">?</a> |

**Filters and Masking**

|        |   |
|--------|---|
| Filter | <input checked="" type="checkbox"/> Low complexity regions <a href="#">?</a>                  |
|        | <input type="checkbox"/> Species-specific repeats for: Homo sapiens (Human) <a href="#">?</a> |
| Mask   | <input checked="" type="checkbox"/> Mask for lookup table only <a href="#">?</a>              |
|        | <input type="checkbox"/> Mask lower case letters <a href="#">?</a>                            |

# BLASTP

- Protein query/database
- Smaller word size:
  - The default word-size is 5
  - Amino acid sequences are more likely to be unique than nucleotide sequences(20 vs. 4 residues)
  - Uses substitution matrices (PAM, BLOSUM) to identify more unique w-size strings (neighborhood table).
  - If word-size decreases: **increase sensitivity / decrease selectivity**
    - ✓ More probable to find distant matches
    - ✓ More time to compute (more hits)
  - If word-size increase: **increase selectivity / decrease sensitivity**
    - ✓ More probable to find close matches only
    - ✓ Increase compute speed

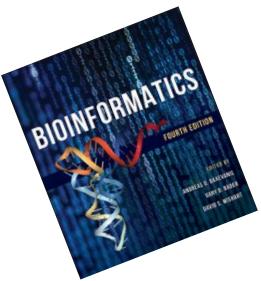
# PSI-BLAST

- Position-Specific Iterated BLAST is particularly well suited for identifying **distantly related proteins** – proteins that may not have been found using the traditional BLASTP method.
- Use results from BLAST (above specific *E* value) to construct a consensus (align BLAST hits into a multiple sequence alignment - MSA).
- PSI-BLAST relies on the use of position-specific scoring matrices (PSSMs)
- Scoring in PSSMs depends on the frequencies of residues in a specific column of the MSA (not from scores of PAM/BLOSUM)
- Search database with PSSMs matrix (instead of the original query)

# PSI-BLAST

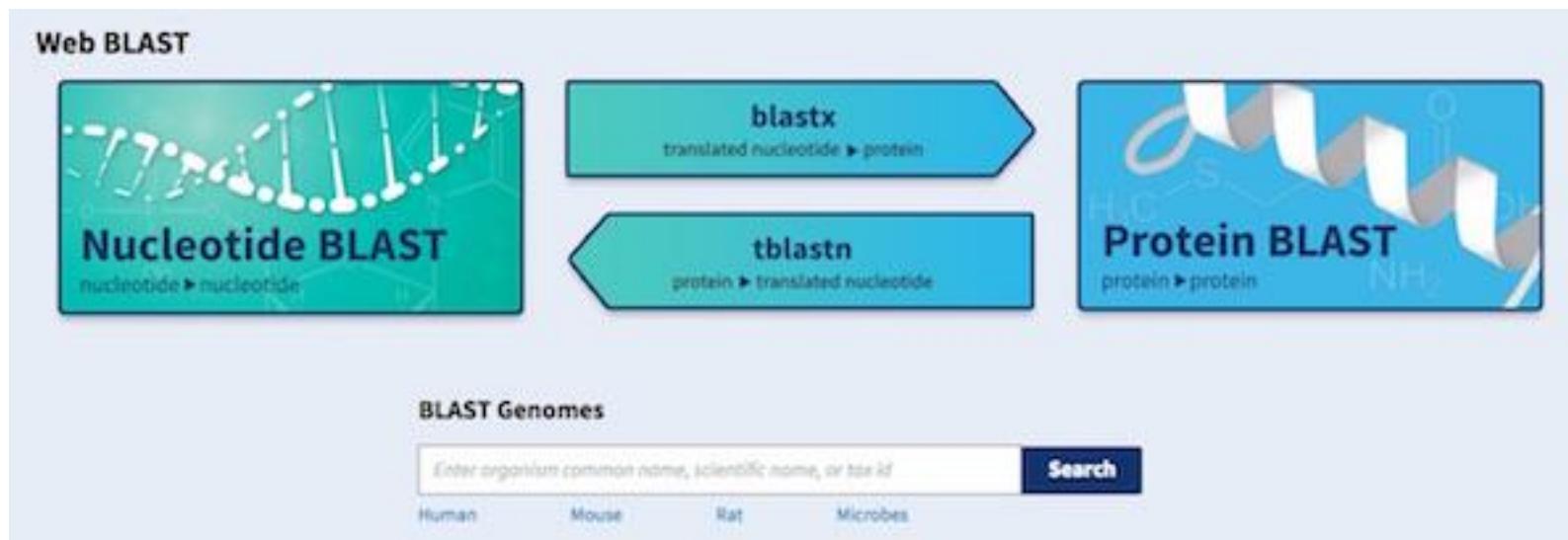


**Figure 3.13** Constructing a position-specific scoring matrix (PSSM). In the upper portion of the figure is a multiple sequence alignment of length 10. Using the criteria described in the text, the PSSM corresponding to this multiple sequence alignment is shown in the lower portion of the figure. Each row of the PSSM corresponds to a column in the multiple sequence alignment. Note that position 8 of the alignment always contains a threonine residue (T), whereas position 10 always contains a glycine (G). Looking at the corresponding scores in the matrix, in row 8, the threonine scores 150 points; in row 10, the glycine also scores 150 points. These are the highest values in the row, corresponding to the fact that the multiple sequence alignment shows absolute conservation at those positions. Now, consider position 9, where most of the sequences have a proline (P) at that position. In row 9 of the PSSM, the proline scores 89 points – still the highest value in the row, but not as high a score as would have been conferred if the proline residue was absolutely conserved across all sequences. The first column of the PSSM provides the deduced consensus sequence.



# Heuristic methods

- FASTA, BLAST are approximations. Remember, we are trying to find a needle in a haystack.
- Based on heuristics
- If search don't "seed" well, it can miss evolutionary close sequences
- But they are useful for practical purposes



# Heuristic methods

- FASTA begins the search by looking for exact matches of words, while BLAST allows for conservative substitutions in the first step.
- BLAST allows for automatic masking of sequences, while FASTA does not.
- FASTA will return one and only one alignment for a sequence in the hit list, while BLAST can return multiple results for the same sequence, each result representing a distinct HSP.
- Since FASTA uses a version of the more rigorous Smith–Waterman alignment method, it generally produces better final alignments and is more apt to find distantly related sequences than BLAST. For highly similar sequences, their performance is fairly similar.
- BLAST runs faster than FASTA, since FASTA is more computationally intensive.

What database to search?

Search the smallest comprehensive database likely to contain your protein

# Quick Discussion!

BLASTP

Query ID [BAJ99511.1](#) (Barley)



Source: Wikipedia

Protein  Advanced  Help

GenPept ▾  Change region shown

**predicted protein [Hordeum vulgare subsp. vulgare]**

GenBank: BAJ99511.1

Go to:

LOCUS BAJ99511 406 aa linear PLN 20-MAY-2011

DEFINITION predicted protein [Hordeum vulgare subsp. vulgare].

ACCESSION BAJ99511

VERSION BAJ99511.1

DBSOURCE accession [AK368308.1](#)

KEYWORDS .

SOURCE Hordeum vulgare subsp. vulgare (domesticated barley)

ORGANISM [Hordeum vulgare subsp. vulgare](#)  
Eukaryota; Viridiplantae; Streptophyta; Embryophyta; Tracheophyta;  
Spermatophyta; Magnoliopsida; Liliopsida; Poales; Poaceae; BOP  
clade; Pooideae; Triticodae; Triticeae; Hordeinae; Hordeum.

Analyze this    
Identify Conserved Domains  
Highlight Sequence Features  
Find in this Sequence

Related Information   
Nucleotide  
PubMed

- What hits do you get?
- Taxonomy?
- Are they biologically significant?