

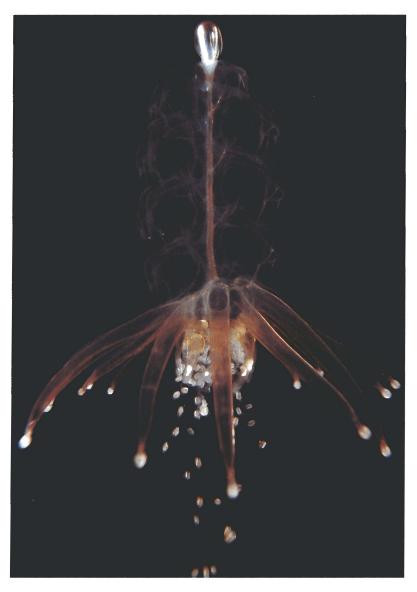
Ingo S. Wehrtmann Jorge Cortés *Editor*s





Part 8 Siphonophores

Karina Rodríguez-Sáenz and Rebeca Gasca



Physophora hydrostatica, a species from Pacific Costa Rica. (Photo: Steven H.D. Haddock)

Abstract Thirty-seven species of siphonophores are listed for the Costa Rican coastal and oceanic waters. All species were recorded from the Pacific coast, seven of them have been also collected off the Caribbean coast. These figures represent 10% of the 70 species known from the Caribbean Sea, and 42% of the 87 species known from the eastern tropical Pacific. Overall, the species recorded in Costa Rican waters represent close to 37% of the nearly 100 species known to be distributed in the tropical-equatorial belt. The relatively low biodiversity of siphonophores in the Caribbean coast of Costa Rica is clearly a result of the scarcity of research in this region. All the species currently known from Costa Rica are epipelagic forms living between the surface and 200 m, and have been recorded previously in the corresponding oceanographic regions of the Atlantic and Pacific oceans. Additional work in both oceanic and coastal waters is necessary, including coastal and estuarine ecosystems and also deep waters (>200 m), which most probably harbor a diverse siphonophore fauna.

Introduction

The Subclass Siphonophora is a peculiar group of marine cnidarians (Class: Hydrozoa); it is part of the commonly known "gelatinous zooplankton," which includes hydromedusae, scyphomedusae, ctenophores, salps, and appendicularians among other taxa. Most of the species of siphonophores are fully planktonic forms. The Portuguese Man-of-War (*Physalia physalis*). floating at the surface of the water, and a small group of benthic forms (Family: Rhodaliidae) (Pugh 1999b) are two noteworthy exceptions.

The siphonophores are efficient predators distributed in the entire world ocean from the surface to thousands of meters deep. Most of them inhabit oceanic waters, but a few species can live in neritic areas, and are often present in high densities in both neritic and oceanic waters. They are all carnivorous forms, capturing their prey using poisonous nematocysts, and most species are assignable to a passive feeding behavior. The use of luminescent lures to attract preys has been discovered recently (Haddock *et al.* 2005a). Because of their predating efficiency, siphonophores can cause major impacts on the structure and dynamics of the zooplankton communities

Habitat Crítico S.A., Environmental Consulting. P.O. Box 389-1300, San José, Costa Rica e-mails: karinarodriguez@racsa.co.cr; habitat@ice.co.cr

El Colegio de la Frontera Sur (ECOSUR)-Chetumal, A.P. 424. Chetumal, Quintana Roo 77000, México

e-mail: rgasca@ecosur.mx

K. Rodríguez-Sáenz

R. Gasca

8 Siphonophores 153

(Pagés *et al.* 1992). Recent studies have shown how pollution caused by human activities may have an impact on gelatinous zooplankton, including the population dynamics of siphonophores. These changes can eventually impact other components of the planktonic ecosystems (CIESM 2001).

We present a checklist of the siphonophores collected in coastal and oceanic environments of Costa Rica's Caribbean Sea and Pacific Ocean (Species Lists 8.1 and 8.2 are included on the CD-Rom). The lists include scientific results of international expeditions such as that of the "Albatross" in the eastern tropical Pacific during 1904 and 1905, and the "BONACCA" in the Western Caribbean Sea in 1963 (Alvariño 1972); they covered large areas of these regions, including a few stations in Costa Rican waters. Recent surveys of siphonophores in Costa Rican waters have been done and included basic aspects on the composition and distribution of this group (Ramírez 1988; Gasca & Suárez-Morales 1992; Nowaczyk 1998, Rodríguez 2001 unpublished data).

Taxonomic status/problems: The classic and best account on siphonophore taxonomy and morphology was written by Totton (1965); however, the systematics of the group has changed since then (Bouillon *et al.* 1992). Many species, mainly deepwater forms, have been described recently from different regions of the world (Pugh 1992a, b, c, 1995, 1999a, 2001, 2002, 2004; Pugh & Pagès 1995, 1997). Valuable publications on the biology and reproduction of the siphonophores are those by Mackie *et al.* (1987) and Carré & Carré (1993).

Siphonophores have been grouped into three orders, based on the presence of an apical pneumatophore (Cystonectae and Physonectae; absent in Calycophorae), and on the presence (Physonectae and Calycophorae) or absence (Cystonectae) of nectophores. The structure and life cycle of this group are complex. They have been designated as "colonies" formed by zooids, medusoids, and polypoids, each with different forms and functions. The complexity of siphonophores is not restricted to their morphological and physiological diversity; it extends also to the specific terminology used to define each of these variable structures. A recent revision of the morphological nomenclature in siphonophores is given in Haddock *et al.* (2005b). Nectophores and bracts are of particular importance for taxonomic identification.

Siphonophores are fragile, thus making it difficult to study them both taxonomically and ecologically. The use of traditional trawl nets may produce serious damages to the organisms and consequently complicate their identification and/or quantification. In recent years, blue water diving, submersibles, and remotely operated vehicles (ROVs) have solved most of this problem. It is now possible to obtain complete organisms and images of siphonophores, thus facilitating the identification of species and providing valuable information about their biology, ecology, and behavior. However, this kind of equipment is expensive (especially submersibles), and for developing countries it is almost impossible to acquire and maintain them. Due to this situation, most of the information on deepwater siphonophores is concentrated in a few sites around the world; hence, epipelagic trawls are, historically and currently, the most important source of the available information on the group. Moreover, the task of fixation-preservation of an intact organism is difficult. The slow addition of magnesium chloride to the seawater while the

organism is swimming helps to create a condition for relaxation previous to the fixation and preservation in formalin.

Their phylogenetic relations are still obscure and under investigation. There are few specialists working in the field, and some of them are listed below.

Species richness. The number of nominal species of siphonophores known to date is about 170 (worldwide), with many others discovered but yet undescribed. There are surveys on the general diversity of siphonophores in different regions of the Atlantic Ocean, such as the works by Pugh (1999b) in the southwestern Atlantic (96 species), by Stepanjants (1975) and by Suárez & Gasca (1991) in the Caribbean Sea and Gulf of Mexico (62 species), and by Pugh & Gasca (in press) for the Gulf of Mexico (80 species). In the Central Pacific Ocean, between 10°N and 20°S, Stepanjants (1977) recorded 55 species. Suárez & Gasca (1991) recorded 48 species from Mexican Pacific waters.

Overall, the diversity of siphonophores known in Costa Rica represents close to 20% of the known diversity of the group worldwide. In regional terms, the number of species estimated to inhabit the Pacific Ocean adjacent to the Costa Rican waters is ca. 90 species (Alvariño 1972, 1974, Stepanjants 1977, Gasca 2002), representing 42% of the species reported from this area. Roughly seven species have been reported from the Caribbean Sea of Costa Rica, which represents only 10% of the 70 species known from the Caribbean region (Alvariño 1972, 1974, Owre & Foyo 1972, Stepanjants 1975, Michel & Foyo 1976, Gasca 2002). These data show the need to increase surveys of the zooplankton community in both coasts of Costa Rica, but particularly in the Caribbean waters.

Specialists

PHIL R. PUGH: National Oceanography Centre, Southampton S014 3ZH, United Kingdom – Taxonomy. prp@noc.soton.ac.uk

CLAUDE CARRE and DANIELE CARRE: Station Zoologique, Faculté des Sciences de Paris, 06 Villefranche-sur-mer. France.

CASEY W. DUNN: Brown University Providence, 80 Waterman Street Box G-W, RI 02912 USA – casey_dunn@broun.edu

REBECA GASCA: El Colegio de la Frontera Sur (ECOSUR)-Chetumal. A.P. 424. Chetumal, Quintana Roo 77000. México – Taxonomy, geographic distribution. rgasca@ecosur.mx

Collections

Specimens of most of the species reported herein are deposited in a collection kept by K. Rodríguez as part of her Master thesis research. This material is held at CIMAR, Universidad de Costa Rica, San José. A few specimens, collected in previous projects, are deposited in the Museo de Zoología of the Universidad de Costa Rica.

8 Siphonophores 155

Conclusions

One of the reasons for the relatively low biodiversity found in the Caribbean coast of Costa Rica with respect to the Pacific coast is the scarcity of research in those waters. Additional collections of gelatinous zooplankton in both oceanic and coastal waters are necessary, including estuaries, but emphasizing the neritic and oceanic areas. The same recommendation is valid for the rest of Central America. Through understanding the ecology and distribution of siphonophores as an important group of predators, it will be easier to evaluate the processes determining the dynamics of the marine planktonic biota of Costa Rica, including the ecology and development of local fisheries.

References

- Alvariño A (1972) Zooplancton del Caribe, Golfo de México y regiones adyacentes del Pacífico. Mem. IV Congr. Nac. Oceanogr. México, pp 223–247
- Alvariño A (1974) Distribution of siphonophores in the regions adjacent to the Suez and Panama Canals. Fish Bull 22:41–76
- Bouillon J, Boero F Cicogna F Gili JM Hughes RH (1992) Non-siphonophoran Hydrozoa: what are we talking about? Sci Mar 56:279–284
- CIESM (2001) Gelatinous zooplankton outbreaks: theory and practice. CIESM workshop ser. No. 14, 112 p. Monaco. www.ciesm.org/publications/Naples01.pdf
- Carré C, Carré D (1993) Ordre des Siphonophores. In: Doumenc D (ed) Traité de Zoologie. Anatomie, Systématique, Biologie. Tome III. Fascicule 2. Cnidaires. Cténaires. Masson, Paris, pp. 523–596
- Cortés J (1996–1997) Biodiversidad marina de Costa Rica: Filo Cnidaria. Rev Biol Trop 44/45;323–334
- Gasca R (2002) Lista faunística y bibliografía comentada de los sifonóforos (Cnidaria) de México. An Inst Biol UNAM 73:123-143
- Gasca R Suárez E (1992) Sifonóforos (Cnidaria: Siphonophora) del Domo de Costa Rica. Rev Biol Trop 40:125–130
- Haddock SHD, Dunn CW, Pugh PR, Schnitzler CE (2005a) Bioluminescent and red-fluorescent lures in a deep-sea Siphonophore. Science 309:263.
- Haddock SHD, Dunn CW, Pugh PR (2005b) A re-examination of siphonophore terminology and morphology, applied to the description of two new prayinae species with remarkable bio-optical properties. J Mar Biol Ass U.K. 85:1–13.
- 4. Kirkpatrick P Pugh PR (1984) Siphonophores and Velellids: Keys and notes for the identification of the species. Linn Soc London Estuar Brack.-Water Sci Assoc, London, 154 p
- Mackie GO, Pugh PR, Purcell JE (1987) Siphonophore biology. Adv Mar Biol 24:97-262
- Michel HB, Foyo M (1976) Caribbean Zooplankton. Part. I. Siphonophora, Heteropoda, Copepoda, Euphausiacea, Chaetognatha and Salpidae. Off. Naval Res., Dept. Navy, Washington, DC, 549 p
- Nowaczyk J (1998) Gelatinöses Zooplankton im Golfo Dulce (Costa Rica), während des Überganges von der Regen zur Trockenzeit 1997- 1998. M.Sc. thesis, Univ. Bremen, Bremen, 82 p
- Owre HBM, Foyo M (1972) Studies on Caribbean zooplankton. Description of the program and results of the first cruise. Bull Mar Sci 22:483–521
- Pagès F, Gili JM Bouillon J (1992). Planktonic enidarians of the Benguela current. Sci Mar 56(Suppl 1):144

- Pugh PR (1992a) The status of the genus *Prayoides* (Siphonophora: Prayidae). J Mar Biol Assoc U.K. 72:895–909
- Pugh PR (1992b) Desmophyes haematogaster, a new species of prayine siphonophore (Calycophorae, Prayidae). Bull Mar Sci 50:89-96
- Pugh PR (1992c) A revision of the sub-family Nectopyramidinae (Siphonophora, Prayidae). Phil Trans Royal Soc Lond 335:281-322
- Pugh PR (1995) Clausophyes tropica (Siphonophorae, Calicophora) a new siphonophore species from the tropical Atlantic. Bull Mar Sci 57:453–459
- Pugh PR (1999a) A review of the genus *Bargmannia* Totton, 1954 (Siphonophorae, Physonectae, Pyrostephidae). Bull Nat Hist Mus Lond (Zool.) 65:51-72
- Pugh PR (1999b) Siphonophorae. *In*: Boltovskoy D (ed) South Atlantic Zooplankton. Backhuys, Leiden, The Netherlands, pp 467–511
- Pugh PR (2001) A review of the genus *Erenna* Bedot, 1904 (Siphonophora, Physonectae). Bull Nat Hist Mus Lond (Zool.) 67:169–182
- Pugh PR (2002) A new species of Rosacea (Siphonophora: Calycophorae: Prayidae) from the Gulf of Oman. J Mar Biol Ass U.K. 82:171-172
- Pugh PR (2004) A new species of *Physophora* (Siphonophora: Physonectae: Physophoridae) from the North Atlantic, with comments on related species. Syst Biodiv 2:251–270
- Pugh PR, Gasca R (2008) Siphonophora (Cnidaria: Hydrozoa) of the Gulf of Mexico. In: Felder DL, Camp DA (eds) Gulf of Mexico Origin, Waters and Biota. Biodivesity, vol 1 Texas A&M University Press, College Station, Texas (in press)
- Pugh PR Pagès F (1995) Is Lensia reticulata a diphyinae species (Siphonophorae, Calycophorae, Diphyidae)? A redescription. Sci Mar 52:181–192
- Pugh PR, Pagès F (1997) A re-description of *Lensia asymmetrica* Stepanjants, 1970 (Siphonophorae, Diphyidae). Sci Mar 61:153–161
- Ramírez AL (1988) Sistemática y distribución de los sifonóforos del Domo de Costa Rica. Thesis, Fac. Ciencias, UNAM, México, 207 p
- Rodríguez K (2001) Relación de eventos oceanográficos con la abundancia y composición del zooplancton, con énfasis en el zooplancton gelatinoso de la Isla del Coco. Consulting report for SINERGIA 66. Unpublished data, 11 p
- Rodríguez K (2005) Distribución espacial y temporal de la biomasa, composición y abundancia del zooplancton, con énfasis en hidromedusas de Bahía Culebra durante La Niña 1999 y el 2000. M.Sc. thesis, University of Costa Rica, San José, Costa Rica, 156 p
- Stepanjants S (1975) Species composition and distributional pattern of Siphonophora of the Caribbean, Gulf of Mexico and adjacent waters of the Atlantic. Trans P.P. Shirshov Inst Oceanol 100:96–126
- Stepanjants S (1977) Siphonophora of the central part of the Pacific Ocean. *In*: Explorations of the Fauna of the Seas XX (XXVIII). Marine Plankton. (Systematics and Faunistics), pp 54-81
- Suárez E, Gasca R (1991) Sifonóforos de México. Biología y Ecología. Centro de Investigaciones de Quintana Roo (CIQRO). Chetumal, Q. Roo, México. 178 p
- Totton AK (1965) A synopsis of the Siphonophora. Trust Brit Mus (Nat Hist), Lond, 230 p

Species List 8.1 Siphonophores collected in Caribbean of Costa Rica following the taxonomic arrangement and nomenclature proposed by Totton (1965) and Pagès et al. (1992)

	World			Oceanic		Habitat/	
Species	distribution.	distribution Central America Costa Rica	Costa Rica ^e		Depth(m)	distribution ^d Depth ^e (m) Community ^e References ^g	References ⁸
Order CYSTONECTAE							
Family PHYSALIIDAE							
Physalia physalis (Linnaeus, 1758)	ct-t	n.a.	೪	сb	0	ď	[1, 2, 6, 7]
Order CALYCOPHORAE							
Family DIPHYIDAE							
Abylopsis eschscholtzi (Huxley, 1859)	ct-t		ဗ	Бр	0200	wc	$[1, 2, 5, 6]^{h}$
Chelophyes appendiculata (Eschscholtz, 1829) ct-t	ct-t	H,N	ຽ	Ep	0-300	wc	[1, 2]
Chelophyes contorta (Lens and van Riemsdijk, 1908)	ī		11	də	0200	wc	[1, 2, 9] ^b
Enneagonum hyalinum Quoy and Gaimard, 1827	ct-s		22	də	300-1000	wc	[2, 4, 5]
Muggiaea atlantica Cunningham, 1892	EA,Med,WA CA	CA	5	Ep	0-100	wc	[1, 2, 4, 5] ^h
Sulculeolaria quadrivalvis Blainville, 1834	ct-s	G,P	CC	ф	0-100	wc	$[2, 3, 7, 9]^h$
*ct = circumtropical; ct-t = circumtropical-temp	erate; ct-s = cir	circumtropical-temperate; ct-s = circumtropical-subtropical; t-t = tropical-temperate; EA = eastern Atlantic; EP = eastern Pacific;	ical; t-t = tropi	al-temperate; E	A = eastern /	Atlantic; BP = e	astern Pacific;

"cf = circumtropical; cf-t = circumtropical-temperate; cf-s = circumtropical-subtropical; f-t = tropical Med = Mediterranean; WA = western Atlantic
bCA = all Central American countries; G = Guatemala; H = Honduras; N = Nicaragua; P = Panama
cC = Caribbean coast; IU = Isla Uvita

^dep = epipelagic
• Uppermost and lowermost occurrences (in meters)
^{f.wc} = water column; p = pleuston
^{f.wc} = weekerences are indicated by numbers according to the Reference list.

^{h.Rodríguez}, K. unpublished data

Species List 8.2 Siphonophores collected in Pacific waters of Costa Rica. The systematic arrangement and nomenclature used was that proposed by Totton (1965) and Pages et al. (1992)

	World	Central	Costa Rica	Oceanic		Habitat/	
Species	distribution ^a	Americab	Pacific ^c	distribution ^d	Depth ^e (m)	distribution ^d Depth ^e (m) Community ^f	References
Order CYSTONECTAE							
Family PHYSALIIDAE							
Physallia physalis Linnaeus, 1758	ct-t		GP,ICñ	surface	0	ď	[2, 6, 7, F. Pagès, 2000 personal communication]
Family RHIZOPHYSIDAE							
Rhysophysa eysenhardti Gegenbaur, 1859	ct		GP	ep,mp	n.a.	wc	[8, 9]
Order PHYSONECTAE							
Family AGALMIDAE							
Agalma elegans (Sars, 1846)	ct-t		D,GP	ep,mp	0-200	wc	[2, 6, 8]
Agalma okeni Eschscholtz, 1825	ct-t	CA	D,GP	eb	0-200	wc	[2, 6, 8]
Halistemma sp. Hexley, 1859	n.a.		IC	du	n.a.	wc	[8]
Nanomia bijuga (Chiaje, 1841)	ct-t		D,GP	ep,mp	0200	wc	[2, 8]
Family FORSKALIIDAE							
Forskalia edwarsi Kolliker, 1853	ct-t		GP	eb	0-250	wc	[8, 9]
Family PHYSOPHORIDAE							
Physophora hydrostatica Forskål, 1775	ct-t		Д	е́р	0200	wc	[2]
Order CALYCOPHORAE							
Family PRAYIDAE							
Amphicaryon ernesti Totton, 1954	ct-s		Д	eb	0-200	wc	[2]
Rosacea cymbiformis (Chiaje, 1822)	EA,Med,WA		GD	ep,mp	0-675	wc	[4, 5]
Rosacea flaccida Biggs, Pugh & Carré, 1978 n.a.	3 n.a.		GP	еb	0-50	wc	[8]

kål, 1776) ct-s D ep 0–200 wc 1853 ct-t D mp 300–500 wc 846) ct-s D ep 0–200 wc van ct-s D,GD,GP ep 0–200 wc lainville, 1834 ct-s D,GD,GP ep 0–200 wc lainville, 1835 ct-s D,GD,GP ep 0–200 wc schscholtz, ct-s ct-t D,GD,GP,IC ep 0–200 wc & van Riemsdijk, t-t G D,GD,GP,IC ep 0–200 wc Eysenhardt, ct-t N GD,GP,GP,IC ep 0–200 wc 1829) ct-t N GD,GP,GP,IC ep 0–200 wc Sys ct-t N GD,GP,GP,IC ep 0–200 wc 1925) ct-t N GD,GP,GP,IC ep 0–200 wc 1925) ct-t N GD,GP,GP,IC	raininy furrorounds							
ct-1 D mp 300-500 wc ct-s b,GD,GP ep 0-200 wc ct-s D,GD,GP ep 0-200 wc ct-s D ep 0-200 wc ct-t N GD,GP,GP,IC ep 0-500 wc ct-t N GD,GD,GP,IC ep 0-500 wc ct-t N GD,GP,GP,IC ep 0-500 wc ct-t N GD,GD,GP,IC ep 0-500 wc ct-t N GD,GP,GP,IC ep 0-500 wc ct-ts D,GD,GP,IC ep 0-500 wc ct-s D,GD,GP,IC ep 0-500 wc ct-s D,GD,GP,IC ep 0-500 wc		ct-s		D	еb	0-200	wc	[2, 9]
ct-s D ep 0-200 wc ct-s D,GB,GP ep 0-200 wc ct-s D,GB,GP ep 0-200 wc ct-t D ep 0-200 wc ct-t D ep 0-200 wc k, t-t D,GB,GP,IC ep 0-200 wc ct-t N GD,GP,GP,IC ep 0-500 wc ct-t N GD,GP,GP,IC ep 0-200 wc ct-t N GD,GP,GP,IC ep 0-200 wc ct-s D,GD,GP,IC ep 0-200 wc EA,EP,IO, WA,WP D,GD,GP,IC ep 0-200 wc wA,WW,WP D ep 0-200 wc ct-s D		ct-t		Q	du	300-500	wc	[2, 4]
ct-s D ep 0-200 wc ct-s D,GB,GP ep 0-200 wc ct-s D,GB,GP ep 0-200 wc ct-t D ep 0-200 wc k, t-t D,GB,GP,IC ep 0-500 wc ct-t N GB,GP,IC ep 0-500 wc ct-t N GB,GP,IC ep 0-200 wc ct-t N GB,GP,IC ep 0-200 wc ct-s D,GB ep 0-200 wc wA,wP D,GB ep 0-200 wc ct-s D,GB ep 0-200 wc ct-s D,GB ep 0-200 wc ct-s D ep,mp	Family DYPHIDAE							
ct-s D,GD,GP ep 0-200 wc ct-s D,GD,GP ep 0-200 wc ct-t D ep 0-200 wc ct-t D,GD,GP,IC ep 0-500 wc ct-t G D,GD,GP,IC ep 0-500 wc ct-t N GD,GP,GP,IC ep 0-500 wc ct-t N GD,GP,GP,IC ep 0-200 wc ct-s D,GD,GP,GP,IC ep 0-200 wc ct-s D,GD,GP,IC ep 0-200 wc wA,wP, P D,GD,GP,IC ep 0-200 wc wA,wP, WP D ep 0-200 wc ct-s D ep 0-300 wc ct-s D ep 0-200 wc ct-s D ep 0-200 wc ct-s D ep 0-200 wc ct-s D	1846)	ct-s		Д	ф	0-200	wc	[2, 4]
ct-s D,GD,GP ep 0-200 wc ct-s D ep 0-200 wc ct-t D ep 0-200 wc k, t-t D,GD,GP,IC ep 0-200 wc ct-t N GD,GP,IC ep 0-500 wc ct-t N GD,GP, GP,IC ep 0-200 wc ct-s D,GD,GP,IC ep 0-200 wc ct-s D,GD,GP,IC ep 0-200 wc ct-s D,GD,GP,IC ep 0-200 wc wA,wP D ep 0-200 wc ct-s D ep 0-200 wc ct-s D ep,mp 0-200 wc ct-s D ep,mp 0-200 wc		ct-s		D,GD,GP	də	0-200	wc	[2, 5, 8, 9]
ct-t. D ep 0–200 wc ct-t. D, GD, GP, C = 200 wc ct-t. D, GD, GP, IC ep 0–500 wc wc ct-t. D, GD, GP, IC ep 0–500 wc ct-t. N GD, GP ep 0–500 wc ct-t. N GD, GP, IC ep 0–500 wc ct-t. D, GD, GP, IC ep 0–500 wc ct-s. D, GD ep 0–200 wc wc wA, WP ep 0–500 wc ct-s. D, GD ep mp 0–200 wc ct-s. D ep mp 0–200 wc ct-s. D ep mp 0–200 wc ct-s. D ep mp 0–200 wc		ct-s		D,GD,GP	еb	0-200	wc	[2, 4, 5, 8]
ct-1 D ep 0–500 wc dijk, t-t D,GD,GP,IC ep 0–200 wc ct-t G D,GD,GP,IC ep 0–500 wc ct-t N GD,GP ep 0–500 wc ct-s D,GD,GP,IC ep 0–200 wc ct-s D,GD,GP,IC ep 0–200 wc ct-s D,GD,GP,IC ep 0–200 wc kWA,WP D ep 0–200 wc ct-s D ep,mp 0–200 wc ct-s D ep,mp 0–200 wc		ct-s.		Q	ер	0-200	wc	[2, 4]
Riemsdijk, t-t D,GD,GP,IC ep 0–200 wc ot-t G D,GD,GP,IC ep 0–500 wc ot-t N GD,GP ep 0–200 wc ct-t D,GD ep 0–200 wc ct-s D,GD ep 0–200 wc lers, 1860) temperate D ep 0–300 wc ct-s D ep,mp 0–200 wc ct-s D ep,mp 0–200 wc		ct-t		Q	də	0-500	wc	[2]
tert G D,GD,GP,IC ep 0–500 wc ct-t N GD,GP ep 0–500 wc ct-ts D,GD,GP,IC ep 0–200 wc ct-s D,GD ep 0–200 wc lers, 1860) temperate D ep 0–300 wc ct-s D ep,mp 0–300 wc ct-s D ep,mp 0–200 wc ct-s D ep,mp 0–200 wc	Chelophyes contorta (Lens & van Riemsdijk, 1908)	I		D,GD,GP,IC		0-200	wc	[2, 8, 9]
thardt, ct-t N GD,GP ep 0-500 wc ct-t D,GD,GP,IC ep 0-200 wc EA,EP,IO, D ep 0-200 wc WA,WP D ep 0-500 wc lers, 1860) temperate D ep 0-300 wc ct-s D ep,mp 0-200 wc ct-s D ep 0-200 wc		ct-t	g	D,GD,GP,IC	do :	0-500	wc	[2, 5, 8, 9]
ct-t D,GD,GP,IC ep 0–200 wc ct-s D,GD ep 0–200 wc EA,EP,IO, D ep 0–500 wc MA,WP D ep 0–300 wc ct-s D ep,mp 0–200 wc ct-s D ep o-200 wc	enhardt,	ct-t	Z	GD,GP	еb	0-500	wc	[4, 5, 8, 9]
ct-s D,GD ep 0-200 wc EA,EP,IO, D ep 0-500 wc WA,WP D ep 0-300 wc ct-s D ep,mp 0-200 wc ct-s D ep o-200 wc		ct-t		D,GD,GP,IC		0-200	wc	[2, 8]
EA,EP,10, WA,WP D ep 0-500 wc shlers, 1860) temperate D ep 0-300 wc ct-s D ep,mp 0-200 wc ct-s D ep 0-200 wc		ct-s		D,GD	ф	0200	wc	[2, 6, 9]
& Ehlers, 1860) temperate D ep 0-300 wc ct-s D ep,mp 0-200 wc ct-s D ep 0-200 wc		EA,EP,IO, WA,WP		Q	də	0-500	wc	[2, 9]
ct-s D ep,mp 0–200 wc ct-s D ep op,mp wc		temperate		Д	ф	0-300	wc	[2, 9]
ct-s D ep 0–200 wc		ct-s		Q	ep,mp	0-200	wc	[5, 9]
		ct-s		Q	сb	0-200	wc	[2, 4]
ct-s D,GD ep 0–200 wc	Lensia subtilis (Chun, 1886)	ct-s		D,GD	еb	0-200	wc	[2, 5, 9]

Species List 8.2 (continued)

	World	Central	Costa Rica	Oceanic		Habitat/	
Species	distribution*	Americab	Pacific ^e	distribution ^d	Depthe (m)	distribution ^d Depth ^e (m) Community ^f References ^s	References
Muggiaea atlantica Cunningham, 1892	EA,Med,WA		D,GD,GP,IC ep	də	0-100	WC	[2, 4, 5, 8]
Sphaeronectes gracilis (Claus, 1873)	n.a.		9	сb	0-100	WC	[5, 6]
Family ABYLIDAE							
8	8 ct-s		Д	сb	0200	WC	[2, 9]
Abylopsis eschscholtzi (Huxley, 1859)	ct-t		GD,GP,IC,D	ф	0-200	WC	[2, 5, 6, 8]
Abylopsis tetragona (Otto, 1823)	ct-t,Med	Q	පි	е́р	0-500	wc	[6, 8]
Bassia bassensis (Quoy & Gaimard, 1827)	ct-s	z	8	ф	0-500	wc	[4, 5]
Ceratocymba leuckarti (Huxley, 1859)	ct-s	-	Д	ф	0-700	wc	[2, 9]
Enneagonum hyalinum Quoy & Gaimard, 1827	ct-s		A	də	0200	wc	[2, 4, 5]

n.a. = information not available

*Ant = Antarctic; Arc = Arctic; cp = cosmopolitan; ct = circumtropical; ct-t = circumtropica-temperate; ct-s = circumtropical; st-t = tropical-temperate; BA = eastern Atlantic; BP = eastern Pacific; IO = Indian Ocean; IP = Indo-Pacific; Med = Mediterranean; WA = western Atlantic; WP = western Pacific

bCA = all Central American countries; G = Guatemala, N = Nicaragua; P = Panama
 b = Costa Rica Dome; GD = Golfo Dulce; GP = Golfo de Papagayo; IC = Isla del Coco; ICñ = Isla del Caño

dep = epipelagic; mp = mesopelagic; n = neritic

*Uppermost and lowermost occurrences (in meters)
'wc = water column; p = pleuston

References are indicated by numbers according to the reference list