Four new records of family Diphyidae (Hydrozoa: Siphonophorae) in Korean waters

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Siphonophores are unique, gelatinous zooplankton, which many individuals gather and live like one "Superorganism". The role of individuals in the colony differs greatly depending on their morphological difference, making them more unique. In this study, we report four species belonging to Diphyidae Quoy and Gaimard, 1827 sampled from the South Sea and off Jeju Island, Korea. Two *Chelophyes* Totton, 1932 (*C. appendiculata* (Eschscholtz, 1829); *C. contorta* (Lens and van Riemsdijk, 1908)) and two *Eudoxoides* Huxley, 1859 (*E. mitra* (Huxley, 1859); *E. spiralis* (Bigelow, 1911)) species are described with multi-focus stacked digital images. Our findings update the confirmed order Siphonophorae Eschscholtz, 1829 in Korea to be three suborders, five families, eight genera, and 13 species. In addition, we summarize the synonyms and global distributions of these four newly recorded species in Korean waters.

Keywords: Calycophorae, Cnidaria, eudoxid, nectophore, northwestern Pacific, polygastric

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Introduction

Siphonophores are gelatinous zooplankton belonging to the phylum Cnidaria Hatschek, 1888 class Hydrozoa Owen, 1843. They are exclusively marine and mostly holoplanktonic organisms (Totton, 1965). Currently there are 187 described species in Siphonophorae, although it is expected that there will be more species (Munro *et al.*, 2018). Siphonophores are complex, polymorphic hydrozoans and have a unique colony stage in their life cycle (Pugh, 1974). Although siphonophores appear as one giant single creature, actually it is a colony composed of many individuals. For this unique colony, Mackie (1963) named it "Superorganism". Individual in the colony has significant morphological difference depending on the function (movement, feeding, defense, and reproduction).

Siphonophorae has been traditionally divided into three suborders (Totton, 1965; Pugh, 1999; Mapstone, 2014): Cystonectae Haeckel, 1887 (characterized by the presence of a pneumatophore and absence of nectophores), Physonectae Haeckel, 1888 (characterized by the presence of both a pneumatophore and nectophores), and Calycophorae Leuckart, 1854 (characterized by the absence of a pneumatophore and presence of nectophores).

Diphyidae, well known for bullet-shape, is the most diverse and representative group among seven families

belonging to Calycophorae (Mapstone, 2009). Over 60% of known siphonophores are small, bullet-shaped colonies (Grossmann et al., 2014). Diphyidae has two distinct phases, polygastric (creating eudoxid through asexual reproduction), and eudoxid (creating polygastric through sexual reproduction), as an adult (Dunn and Wagner, 2006). Diphyidae is typically composed of two dissimilar nectophores with mouth-plate and longitudinal ridges. Their somatocyst only exists in the anterior nectophore, not reaching the anterior end (Mapstone, 2009). Diphyidae is clearly distinguished from other families based on these unique characters. However, most species belonging to the Diphyidae share these characters within the family, so they were initially grouped into a single genus Diphyes Cuvier, 1817. Later on, they were divided 45 species in eight genera, based on detailed features such as the depth of hydroecium, the length of somatocyst, and the ostial teeth. In the past, Diphyes was the most diverse genus, however, currently Lensia Totton, 1932 is the most diverse one within Siphonophorae with 26 valid species (Grossmann et al., 2014). In contrast, the smallest genus is Dimophyes Moser, 1925 with only one reported species. Chelophyes and Eudoxoides, addressed in this study, are the second smallest genera, including two species for each genus. Both of these genera were originally classified as *Diphyes*, and were reestablished in 1932 by Totton.

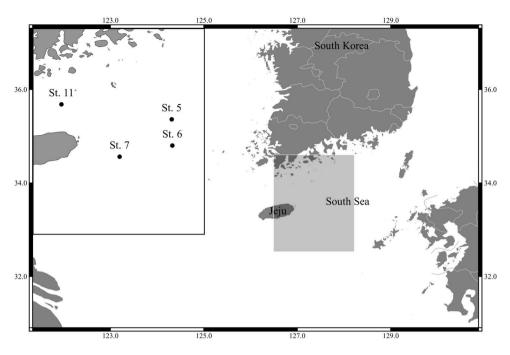


Fig. 1. The map of Korean waters and sampling stations marked with spot. The upper left box magnifies the stations on the map (gray area).

Only two genera (*Diphyes* Cuvier, 1817; *Muggiaea* Busch, 1851) and four species of Diphyidae have been recorded in Korea (NIBR, 2019). *Diphyes* and *Muggiaea* have round and deep hydroecium. The mouth-plate is undivided in *Diphyes* and divided in *Muggiaea*. On the other hand, *Chelophyes* and *Eudoxoides* have a medium depth hydroecium and divided mouth-plate, so they are clearly distinguished from other genera. However, the distinction between *Chelophyes* and *Eudoxoides* has been controversial, they are only discriminated from each other by fine discrepancies in the shape of the ventral notch, the ridges reaching the apex, and the shape of hydroecium cavity.

This study reports four unrecorded species belonging to Diphyidae found in the South Sea and off Jeju Island, Korea. Both genera, *Chelophyes* and *Eudoxoides*, are also reported for the first time in Korea.

MATERIALS AND METHODS

Sample collections

Zooplankton samples were collected in April and August 2018 during an oceanographic vessel, R/V Dongbaek, in the South Sea and off Jeju Island of South Korea (Fig. 1). We towed a plankton net (mesh size: 200 µm, Ø: 60 cm) vertically from the bottom to surface. The net mouth was equipped with a flowmeter (Hydro-Bios, Germany) to determine the volume of filtered water at

each tow. Temperature and salinity of surface layer of seawater were recorded using a CTD (CastAway-CTD, Sontek, USA) at each station (Table 1). Zooplankton samples were immediately split into two aliquots using a Folsom plankton splitter. One aliquot was fixed in 5 or 10% neutralized formalin solution for morphological observation and stored at room temperature. The other aliquot was fixed in 99.9% ethanol and stored in 4°C for further molecular study. Siphonophores were sorted out from zooplankton samples using a Live Insect Forceps (26029-10, F.S.T, Germany) under a stereomicroscope (Olympus SZX7, Japan) and stored in 20 mL glass vials filled with 5% neutralized formalin at room temperature. All materials of four newly recorded species were deposited in the invertebrate collection of the National Institute of Biological Resources (NIBR), Korea.

Morphological analysis

Siphonophore individuals were observed on a petri dish (Ø: 5 cm) filled with 5% neutralized formalin solution under a stereomicroscope (Olympus SZX7, Japan). All specimens were identified using descriptions and illustrations from literature including Totton (1954; 1965), Kirkpatrick and Pugh (1984), and Mapstone (2009). We used the terminology proposed by Mapstone (2009) to describe the specimens. Digital photographs of specimens were taken using a digital camera (Olympus PEN Lite E-PL3, Japan) connected to the stereomicroscope (Olympus SZX7, Japan), and side lights on dark field. Photographs were

| Station | Latitude | Longitude | Date | Depth (m) | Temperature (°C) | Salinity (PSS) |
|---------|-----------------|------------------|--------------|-----------|------------------|----------------|
| | | | 2018. 04. 27 | 107 | 17.87 | 34.47 |
| 5 | 33°41′17.9376″N | 127°53′22.6860″E | 2018.09.14 | 108 | 26.77 | 33.88 |
| | | | 2018. 04. 27 | 111 | 18.02 | 34.49 |
| 6 | 33°25′27.6924″N | 127°53′45.6252″E | 2018.09.14 | 117 | 26.99 | 33.71 |
| | | | 2018. 04. 26 | 133 | 18.35 | 34.32 |
| 7 | 33°18′43.6968″N | 127°21′52.8552″E | 2018.09.13 | 127 | 26.71 | 33.58 |
| | | | 2018. 04. 26 | 95 | 15.86 | 34.43 |
| 11 | 33°50′21.2388″N | 126°46′43.7880″E | 2018.09.12 | 88 | 23.20 | 33.48 |

Table 1. CTD (Conductivity, Temperature, and Depth) data and sample fixation methods of each station.

taken at various focus distances and multi-focus stacking was performed using Helicon Focus 7 (version: 7.5.1, Helicon Soft, Ukraine). The objects of the photographs were cropped and moved to a black background with a scale bar by using Adobe Photoshop CS6 (Version: 13.0 × 64, Adobe, USA). Size measurements of the right lateral and dorsal views (up to 15 individuals) were performed using Axiovision Rel. 4.8 (Version: AxioVs40 V 4.8.1.0, Carl Zeiss, Germany). The measurement points were determined by reference to Nishiyama *et al.* (2016).

Systematics

Class Hydrozoa Owen, 1843 Subclass Hydroidolina Collins, 2000 Order Siphonophorae Eschscholtz, 1829 Suborder Calycophorae Leuckart, 1854 Family Diphyidae Quoy and Gaimard, 1827 Subfamily Diphyinae Quoy and Gaimard, 1827 Genus *Chelophyes* Totton, 1932

Diagnosis

Bullet-shaped anterior nectophore with pointed apex. Five serrated ridges. Five ridges that only partly reaching the apex. A short hydroecium compared to *Eudoxoides*. Claw-shaped hydroecium. Small serrated hydroecium and mouth-plate. Pentagonal cross section. Anterior nectophore and posterior nectophore, both existing. Divided mouth-plate with two trapezoidal serrated wings equal. A club-shaped somatocyst with long peduncle. Somatocyst that does not reach nectosac apex. Canal that follows the shape of a nectosac. No baso-dorsal or lateral ostial teeth at the level of the ostial. The end of the mouth-plate with the point. Hydroecium cavity with made of slanted arches from the lateral view. U-shaped shallow notch from the ventral view.

1. Chelophyes appendiculata (Eschscholtz, 1829) (Fig. 2)

Synonymy

Diphyes appendiculata Eschscholtz, 1829: 138–139, pl. 12, fig. 7; Huxley, 1859: 34, pl. 1, figs. 2a–c; Bigelow, 1911: 248–249, pl. 7, figs. 5–6, pl. 8, figs. 7–8, pl. 9, fig. 6, pl. 10, fig. 6, pl. 11, fig. 1.

Diphyes gracilis Gegenbaur, 1853: 309–315, pl. 16, figs. 5–7.

Chelophyes appendiculata Totton, 1932: 354; 1954: 127, pl. 4, figs. 1, 3; 1965: 185–187; pl. 32, fig. 4, pl. 33, fig. 6; Stepanjants, 1967: 191, figs. 131–132a; Kirkpatrick and Pugh, 1984: 108, fig. 48; Daniel, 1985: 263, figs. 72a–e; Mackie *et al.*, 1987: fig. 36; Pagès and Gili, 1992: 95, fig. 40; Gamulin and Krsinić, 2000: 103, figs. 59a–c; Mapstone, 2009: 201, fig. 55.

Material examined

Five anterior nectophores (NIBRIV0000862385, one vial), St. 6 (33°25′27.6924″N, 127°53′45.6252″E), South Sea, Korea, 14 September 2018, collected by Jaehyeon Kim and Jisu Yeom.

Descriptions

Polygastric phase (Fig. 2)

Mean length and width, 10.50 mm and 3.52 mm, respectively (Table 2). Long bullet-shaped anterior nectophore with pointed apex (Fig. 2A–D). Sharp tip of nectosac facing to apex (Fig. 2E₁; Subscripts indicate arrow number). Canal passing along surface of nectosac (Fig. 2E₂). Five serrated ridges. Pentagonal cross section. Right lateral, right, and left lower ridges reaching anterior nectophore apex (Fig. 2E₃). Claw-shaped short hydroecium (Fig. 2F). Small serrated margin and tip of hydroecium (Fig. 2F_{1,2}). Horizontal-patterned surface of nectosac (Fig.

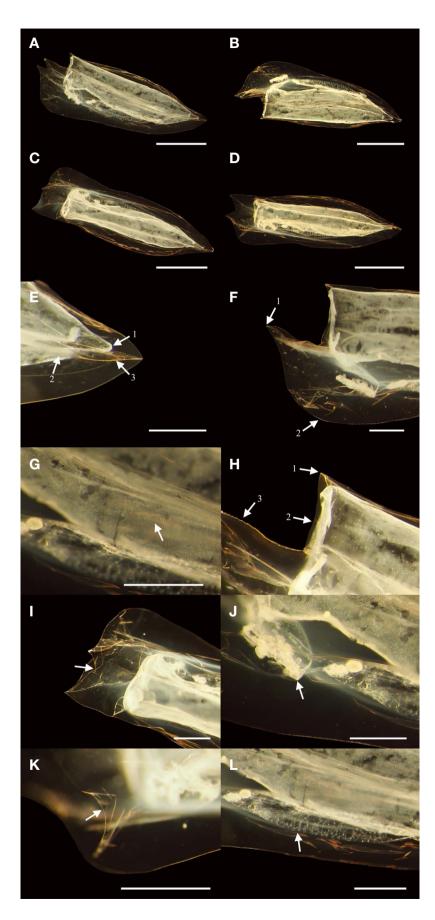


Fig. 2. Chelophyes appendiculata anterior nectophore. (A) Right lateral view, (B) Left lateral view, (C) Dorsal view, (D) Ventral view, (E) Apex; 1: Tip of Apex; 2: Canal; 3: Ridge, (F) Hydroecium; 1: Tip of hydroecium; 2: Serrated right ventral ridge, (G) Surface of nectosac; Horizontal-pattern of nectosac, (H) Ostial teeth; 1: No baso-dorsal ostial teeth; 2: No baso-lateral ostial teeth, 3: Serrated wing, (I) Dorsal view of mouth-plate; Divided mouth-plate, (J) Right lateral view of stem; Stem attachment point, (K) Ventral view of hydroecium; U-notch, (L) Right lateral view of somatocyst; somatocyst with oil droplets. Scale bars: A-D (3 mm); E-L (1 mm).

2G). No baso-dorsal or lateral ostial teeth at level of ostial (Fig. 2H_{1, 2}). Serrated ridge of wing (Fig. 2H₃). Mouthplate divided with two trapezoidal wings; equal on both sides, or larger on right one (Fig. 2I). Sharp pointed end of mouth-plate. Hydroecium cavity with slanted arches (Fig. 2J). Gastrozooid originating at end of hydroecium cavity arch. Stem originating at apex of hydroecium cavity. Stem attachment point with broad rootlike-shape. U-shaped shallow notch in ventral view (Fig. 2K). Oval-shaped somatocyst with small circular materials (Fig. 2L). Round oil droplets at end of somatocyst. Somatocyst reaching 3/4 nectosac length (Fig. 2A–D).

Remarks

Chelophyes appendiculata has a claw-shaped hydroecium, no conspicuous ostial teeth, and the long cylindrical somatocyst as the original description (Eschscholtz, 1829). Most of the morphological characters are confirmed from our materials, although there are minor discrepancies compared to the original description; in the original description the somatocyst narrows more sharply towards the end and no oil droplets, while in present specimens it has oval shaped somatocyst with oil droplets. We regard these differences as variation within the range of individual because the shape of the overall somatocyst was the similar, and the oil droplets are easy to damage.

Gegenbaur (1853) described *C. appendiculata* as *Diphyes gracilis*; the illustrations are almost perfectly consistent with the present materials, although there is no report on the serrated ridges being clearly visible in the present specimen (Fig. 2F_{1,2}). Totton (1932) established the genus *Chelophyes* characterizing with no baso-dorsal and lateral teeth from anterior and posterior nectophores; the apical wall is oblique and runs into the dorsal wall without the marked angle. Based on those synapomorphic characters, Totton (1932) reallocated *D. appendiculata* into the genus *Chelophyes*.

2. Chelophyes contorta (Lens and van Riemsdijk, 1908) (Fig. 3)

Synonymy

Diphyes contorta Lens and van Riemsdijk, 1908: 39–41, pl. 6, figs. 48–50; Bigelow, 1911: 254–255, pl. 7, figs. 7–8, pl. 8, fig. 3, pl. 11, fig. 2.

Chelophyes contorta Totton, 1932: 357–358, fig. 27; 1954: 130, fig. 65; 1965: 187–188, figs. 125–126, pl. 32, figs. 7–8.

Material Examined

Ten anterior nectophores (NIBRIV0000862386, one

vial), St. 6 (33°25′27.6924″N, 127°53′45.6252″E), South Sea, Korea, 14 September 2018, collected by Jaehyeon Kim and Jisu Yeom.

Descriptions

Polygastric phase (Fig. 3)

Mean length and width, 4.94 mm and 1.87 mm, respectively (Table 2). Bullet-shaped anterior nectophore with pointed apex (Figs. 3A-D). Bluntly pointed anterior nectophore compared with Chelophyes appendiculata. Sharp tip of nectosac facing to anterior nectophore apex (Fig. 3E₁). Small circular material covering tip of nectosac. Canal passing along surface of nectosac. Five serrated ridges (Fig. 3E₂). Pentagonal cross section. Right ventral ridge not reaching anterior nectophore apex (Fig. 3A). Horizontal-patterned surface of nectosac (Fig. 3F). Claw-shaped hydroecium (Fig. 3G). Short hydroecium compared with C. appendiculata. Serrated tip of hydroecium (Fig. 3G₁). No baso-dorsal or lateral teeth at level of ostial (Fig. 3G_{2,3}). Smooth ostial regions. Narrow hydroecium cavity with slanted arches. Gradual apex of hydroecium cavity compared with C. appendiculata. Stem originating at apex of hydroecium cavity (Fig. 3H₁). Gastrozooid originating at end of hydroecium cavity arch. Stem attachment point with broad rootlike-shape. Club-shaped somatocyst with long peduncle (Fig. 3H₂). Right bending head of club. Opaque white somatocyst. Oil droplets at various positions of somatocyst. Somatocyst reaching 1/2-2/3 nectosac length (Fig. 3A-D). Mouth-plate divided with two trapezoidal wings; equal on both sides, or larger on right one (Fig. 3I). Small serrated mouth-plate. Sharp pointed end of mouth-plate (Fig. 3J₁). U-shaped shallow notch in ventral view (Fig. 3J₂).

Remarks

Chelophyes contorta is characterized by the absolute contortion of the somatocyst, and the related facets (Lens and van Riemsdijk, 1908). In addition, the club-shaped somatocyst curved to the right is unique character within the family. The key characters are confirmed from our present materials, except for few minor characters; the incomplete median ridge beginning at the base of facet near the velum is visible in the original description, while it is not present in our materials; the peduncle of somatocyst is short (Lens and van Riemsdijk, 1908, fig. 48 in pl. 6), but present materials have long somatocyst similar to those in the fig. 49 in pl. 6 (Lens and van Riemsdijk, 1908). As in the case of C. appendiculata, Totton (1932) reallocated C. contorta from Diphyes to Chelophyes based on the characteristic of no baso-dorsal and lateral teeth at the level of ostial.

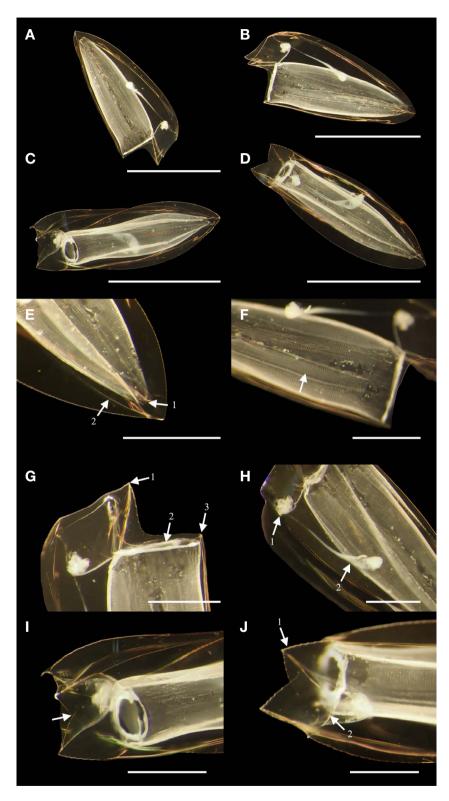


Fig. 3. Chelophyes contorta anterior nectophore. (A) Right lateral view, (B) Left lateral view, (C) Dorsal view, (D) Ventral view, (E) Apex; 1: Tip of Apex; 2: Serrated ridge, (F) Surface of nectosac; Horizontal-pattern of nectosac, (G) Hydroecium and ostial teeth; 1: Tip of hydroecium; 2: No baso-lateral ostial teeth; 3: No baso-dorsal ostial teeth, (H) Right lateral view of somatocyst and stem; 1: Stem attachment point; 2: Somatocyst, (I) Dorsal view of mouth-plate; Divided mouth-plate, (J) Ventral view of hydroecium; 1: Serrated wing; 2: U-notch. Scale bars: A-D (3 mm); E-J (1 mm).

 Table 2. Size measurement of unrecorded four Diphyidae species.

| | | | | | | | Average | Average (Min-Max) (mm) | (mu | | | | | |
|-------------------------------------|----|---------------------|------------------|--|---------------------|---------------------|---------------------|------------------------|------------------|------------------|---|------------------|-----------------------|--------------------------------------|
| Specimen | z | TL | ANH | ANW | HN | MO | SH | SW | HH | HW | ГМН | LMW | RMH | RMW |
| Chelophyes appendiculata | 5 | 10.50 (9.76–11.15) | 9.11 (8.59–9.57) | 10.50 9.11 3.52 8.51 1.89 5.53 0.35 1.48 2.37 1.19 0.95 (9.76–11.15) (8.59–9.57) (3.14–3.87) (7.67–9.15) (1.46–2.11) (4.94–5.94) (0.15–0.68) (1.23–1.75) (1.71–2.91) (0.92–1.55) (0.74–1.17) | 8.51 (7.67–9.15) | 1.89 (1.46–2.11) | 5.53 (4.94–5.94) | 0.35 (0.15-0.68) | 1.48 (1.23–1.75) | 2.37 (1.71–2.91) | 1.19 (0.92–1.55) | 0.95 (0.74–1.17) | 1.32 (1.14–1.48) | 1.32 1.18 (1.14-1.48) (0.98-1.44) |
| Chelophyes contota | 10 | 4.94 (3.10–5.45) | 4.13 (2.58-4.58) | 4.94 4.13 1.87 3.86 1.03 (3.10–5.45) (2.58–4.58) (0.99–2.21) (2.37–4.39) (0.46–1.28) | 3.86 (2.37–4.39) | 1.03 (0.46–1.28) | 2.00 (1.25–2.43) | 0.33 (0.13-0.47) | 0.80 (0.55-0.91) | 1.12 (0.68-1.31) | 2.00 0.33 0.80 1.12 0.72 0.47 (1.25-2.43) (0.13-0.47) (0.55-0.91) (0.68-1.31) (0.40-0.84) (0.25-0.57) | 0.47 | 0.75 (0.47–0.91) | 0.75 0.54 (0.47-0.91) (0.30-0.67) |
| Eudoxoides mitra | 15 | 9.42 (8.21–10.41) | 7.84 (6.92–8.70) | 9.42 7.84 2.64 7.07 1.55 2.32 0.47 1.69 1.50 1.57 0.63 (8.21-10.41) (6.92-8.70) (2.32-3.10) (6.32-7.71) (1.14-1.91) (1.86-2.99) (0.17-0.88) (1.35-1.94) (1.24-1.71) (1.14-1.84) (0.41-0.83) | 7.07 (6.32–7.71) | 1.55 (1.14-1.91) | 2.32 (1.86–2.99) | 0.47 (0.17–0.88) | 1.69 (1.35–1.94) | 1.50 (1.24-1.71) | 1.57 (1.14-1.84) | 0.63 (0.41–0.83) | 1.48 (1.23–1.70) | 1.48 0.79 (1.23–1.70) (0.42–1.05) |
| Eudoxoides spiralis | 15 | 4.48 (3.88–5.41) | 3.38 (2.98–3.97) | $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | 3.07 (2.69–3.61) | 0.77 (0.43–1.13) | 1.33 (0.88–1.58) | 0.29 (0.15-0.49) | 1.09 (0.78–1.54) | 0.89 (0.67–1.02) | 1.02 $(0.88-1.21)$ | 0.26 (0.18-0.35) | $1.10 \\ (0.98-1.28)$ | 1.10 0.46 (0.98-1.28) (0.35-0.61) |
| | | TL | ML | ВН | BW | НS | GW | | | | | | | |
| Eudoxoides spiralis (Eudoxid) | 1 | 4.08 | 0.93 | 1.97 | 0.93 | 2.76 | 0.98 | | | | | | | |

HW. Hydroecium width, LMH: Left mouth-plate height, LMW: Left mouth-plate width, RMH: Right mouth-plate height, RMW: Right mouth-plate width, TW: Total width, BH: Bract height, BW: Bract width, GH: N: Numbers, TL: Total length, ANH: Anterior nectophore height, ANW: Anterior nectophore width, NH: Nectosac height, OW: Ostium width, SH: Somatocyst height, SW: Somatocyst width, HH: Hydroecium height Gonophore height, GW: Gonophore width Genus Eudoxoides Huxley, 1859

Diagnosis

Bullet-shaped anterior nectophore with pointed apex. Small serrated 5 longitudinal ridges. Pentagonal cross section. Complete dorsal ridge. Claw-shaped hydroecium. Divided mouth-plate. Somatocyst not reaching apex of nectosac. Lancet-shaped and serrated wings. Small serrated hydroecium and mouth-plate. Posterior nectophore which is present or absent.

3. Eudoxoides mitra (Huxley, 1859) (Fig. 4)

Synonymy

Diphyes mitra Huxley, 1859: 36–37, pl. 1, fig. 4. Diphyes gracilis Bedot, 1896: 370–372, pl. 12, figs. 4, 8. Eudoxoides mitra Totton, 1932: 358–360, figs. 28–29; 1936: 234; 1965: 188–189, pl.33, figs. 4–5; Leloup, 1934: 28; Moore, 1949: 17, figs. 30–36; Sears, 1950: 3.

Material Examined

Six anterior nectophores (NIBRIV0000862387, one vial), St. 5 (33°41′17.9376″N, 127°53′22.6860″E), South Sea, Korea, 27 April 2018, collected by Eunha Choi and Nayeon Park; nine anterior nectophores (NIBRIV 0000862388), St. 6 (33°25′27.6924″N, 127°53′45.6252″E), South Sea, Korea, 14 September 2018, collected by Jaehyeon Kim and Jisu Yeom.

Descriptions

Polygastric phase (Fig. 4)

Mean length and width, 9.42 mm and 2.64 mm, respectively (Table 2). Bullet-shaped with pointed apex (Fig. 4A-D). Not spirally twisted. Small circular material covering tip of nectosac facing to apex (Fig. 4E₁). Canal passing along surface of nectosac (Fig. 4E2). Small serrated five longitudinal ridges reaching apex (Fig. 4E₃). Hydroecium with pointed tip (Fig. 4F1). Long hydroecium part below ostial level compared with part above. Gastrozooid originating at end of hydroecium cavity arch (Fig. 4F₂). No lateral ostial teeth (Fig. 4G₁). Ostial teeth originating from junction of baso-dorsal ridge and ostial (Fig. 4G₂). Acute outer angles of mouth-plate wing. Concave distal edges of wing. Small serrated mouth-plate (Fig. 4F, H). Mouth-plate divided with two lanceolate wings; equal in both sides, or larger in left one (Fig. 4H). Oval-shaped somatocyst with short peduncle (Fig. 4I₁). Round oil droplets at end of somatocyst (Fig. 4I₂). Somatocyst with small circular materials. Somatocyst reaching 1/2-1/3 nectosac length (Fig. 4A-C). Stem attachment point with broad rootlike-shape (Fig. 4I₃). Horizontal-patterned surface of nectosac (Fig. 4J).

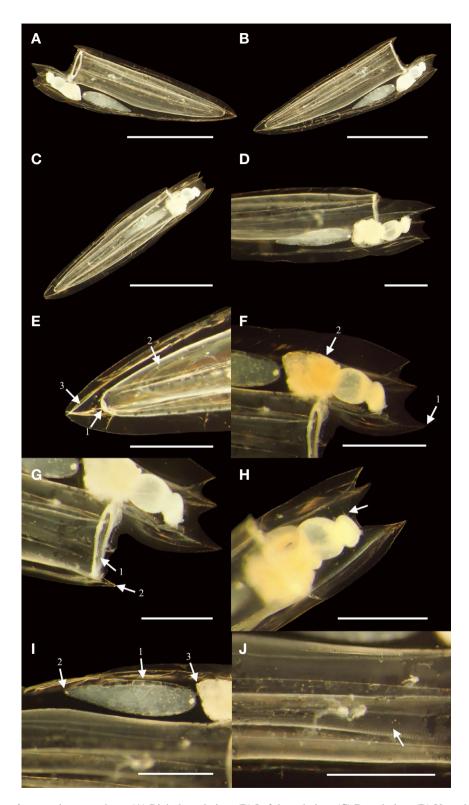


Fig. 4. Eudoxoides mitra anterior nectophore. (A) Right lateral view, (B) Left lateral view, (C) Dorsal view, (D) Ventral view, (E) Apex; 1: Tip of Apex; 2: Canal; 3: Ridge, (F) Hydroecium; 1: Tip of hydroecium; 2: Gastrozooid, (G) Ostial teeth; 1: No baso-lateral ostial teeth, 2: Baso-dorsal ostial teeth, (H) Dorsal view of mouth-plate; Divided mouth-plate, (I) Somatocyst; 1: Somatocyst; 2: Oil droplets; 3: Stem attachment point, (J) Surface of nectosac; Horizontal-pattern of nectosac. Scale bars: A-C (3 mm); D-J (1 mm).

Remarks

Based on the original description (Huxley, 1859), Eudoxoides mitra has an interesting combination of characters including (1) anterior nectophore has obtusely pointed at its apex, (2) hydroecium attains hardly more than one fourth of the length, (3) the narrow neck of the somatocyst is obtusely conical with a slightly recurved apex, (4) the anterior wall of hydroecium is formed below by two triangular plates. The same characters are detected from our present materials although there are minor discrepancies including (1) nectophore is a bit blunter. (2) the somatocyst is slightly shorter, (3) the hydroecium is round and the serrate of ridge is larger. Present specimens also can be regarded as Diphyes gracilis originally reported by Bedot (1896), however Totton (1932) synonymized the species to E. mitra. Totton (1965) provides detailed illustrations of anterior nectophores, which are almost perfectly consistent with the present specimen, except that the somatocyst is relatively shorter in the earlier report.

4. Eudoxoides spiralis (Bigelow, 1911) (Fig. 5)

Synonymy

Diphyes spiralis Bigelow, 1911: 249–251, pl. 7, fig. 4, pl. 8, figs. 1–2, pl. 9, fig. 3, pl. 11, fig. 4.

Eudoxoides spiralis Totton, 1932: 360–363, fig. 30; 1936: 234; 1965: 189–191, pl. 32, figs. 5–6; Moore, 1949: 16, figs. 23–29; Sears, 1950: 3; Totton and Fraser, 1955: 55, figs. 2–3, 6; Kirkpatrick and Pugh, 1984: 110–111, fig. 49; Pugh, 1999: 154, pl. 60, fig. F.

Material Examined

Five anterior nectophores (NIBRIV0000862389, one vial), St. 6 (33°25′27.6924″N, 127°53′45.6252″E), South Sea, Korea, 14 September 2018, collected by Jaehyeon Kim and Jisu Yeom; six anterior nectophores (NIBRIV 0000862390, one vial) and one eudoxid (NIBRIV0000 862391, one vial), St. 5 (33°41′17.9376″N, 127°53′22.6860″E), South Sea, Korea, 14 September 2018, collected by Jaehyeon Kim and Jisu Yeom; three anterior nectophores (NIBRIV0000862382, one vial), St. 7 (33°18′43.6968″N, 127°21′52.8552″E), South Sea, Korea, 13 September 2018, collected by Jaehyeon Kim and Jisu Yeom; one anterior nectophores (NIBRIV0000862383, one vial), St. 11 (33°50′21.2388″N, 126°46′43.7880″E), South Sea, Korea, 26 April 2018, collected by Eunha Choi and Nayeon Park.

Descriptions

Polygastric phase (Fig. 5A-J)

Mean length and width, 4.48 mm and 1.36 mm, re-

spectively (Table 2). Overall twisted bullet-shaped with pointed apex (Fig. 5A-D). Small circular material covering tip of nectosac facing to apex (Fig. 5E₁). Nectosac with same twist-shaped as nectophore. Five twisted and serrated longitudinal ridges. Left ventral ridge combining to right before apex (Fig. 5E₂). Serrated and pointed tip of hydroecium (Fig. 5F₁). Hook-shape of hydroecium in lateral view (Fig. 5F₂). Small serrated hydroecium (Fig. 5F₃). No baso-dorsal or lateral ostial teeth at level of ostial (Fig. 5F_{4,5}). Stem attachment point with broad rootlike-shape (Fig. 5G₁). Oval-shaped somatocyst with short peduncle (Fig. 5G₂). Somatocyst with small circular and column-shaped materials. Round oil droplets at end of somatocyst (Fig. 5G₃). Somatocyst reaching 1/2-1/3 nectosac length (Fig. 5A-D). Oblique somatocyst to right of main axis (Fig. 5D). Slanted fish-scaly surface of nectosac (Fig. 5H). Deep V-shaped notch located in ventral wall (Fig. 5I). Right ventral ridges faced to notch. Curved basal end of left ventral ridge reaching the mid-ventral line before level of ostium. Dissimilar basal ends of two ventral ridges. Triangular space consisting of hydroecium converged to dorsal and opened to ventral. Small divided mouth-plate with two lanceolate wings (Fig. 5J). Larger right lanceolate wing than left one.

Eudoxid phase Eudoxid (Fig. 5K-L)

Conical shape with one longer downside (Fig. 5K). Sharp apex of ridges. Two serrated ridges. Long plump oval-shaped phyllocyst. Small circular materials in phyllocyst. Round oil droplets at end of phyllocyst. Tip of phyllocyst facing to bract apex (Fig. 5K₁). Basal end of phyllocyst point with broad rootlike-shape (Fig. 5K₂). About 2.76 mm in gonophore length and about 0.98 mm in gonophore width (Table 2). Junction of bract and gonophore (Fig. 5L). About 4.08 mm in eudoxid total length about 1.97 mm in bract length and about 0.93 mm in bract width (Table 2). Twisted column-shaped gonophore (Fig. 5L). Smooth attachment point (Fig. 5L₁). Slanted fishscaly surface same as anterior nectophore. Ovum attached inside of eudoxid (Fig. 5L₂). Serrated ridges. Short and pointed hydroecium. No conspicuous baso-ostial teeth in ostium (Fig. 5L₃).

Remarks

Based on the original description (Bigelow, 1911), the most remarkable feature of *Eudoxoides spiralis* is that the entire nectophore is spirally twisted in the same clockwise direction. This species has four ridges at the apex, the larger right wing, and neither baso-lateral nor baso-dorsal teeth as in our present specimens. There is a slight difference, with the original description showing plumper nectophore than the present one. Similar to the case of *E. mi*-

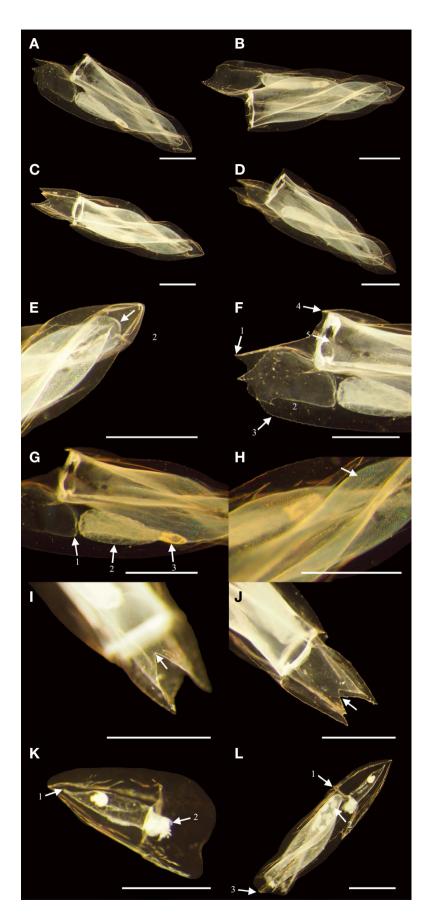


Fig. 5. Eudoxoides spiralis anterior nectophore and Eudoxid. (A) Right lateral view, (B) Left lateral view, (C) Dorsal view, (D) Ventral view, (E) Apex; 1: Tip of Apex; 2: Ridge, (F) Hydroecium; 1: Tip of hydroecium; 2: Hook of hydroecium; 3: Serrated right ventral ridge; 4: No baso-dorsal ostial teeth; 5: No baso-lateral ostial teeth, (G) Somatocyst; 1: Stem attachment point; 2: Somatocyst; 3: Oil droplets, (H) Surface of nectosac; Slanted fish-scaly pattern of nectosac, (I) Ventral view of hydroecium; V-notch, (J) Dorsal view of mouth-plate; Divided mouth-plate, (K) Bract of Eudoxid; 1: Apex of phyllocyst; 2: Gastrozooid, (L) Gonophore of Eudoxid; 1: Attachment point; 2: Ovum; 3: Ostium. Scale bars: 1 mm.

Table 3. Global distribution of four Diphyidae species described in this paper. Records after the original descriptions were integrated. St. 5–7, 11 can be found in the fig. 1.

| | | | Newly record | led species | | |
|-------------------|---|-----------------------------|------------------------|---------------------|------------------------|---|
| Ocean | Regions | Chelophyes appendiculata | Chelophyes contorta | Eudoxoides mitra | Eudoxoides spiralis | References |
| North Pacific | Canada, Canadian Pacific water | • | | | | Mapstone, 2009 |
| | China, East China Sea | | | • | • | Gao <i>et al.</i> , 2002; Tao <i>et al.</i> , 2005 Xu <i>et al.</i> , 2008 |
| | China, South China Sea | • | • | • | • | Li et al, 2012; Lo <i>et al.</i> , 2012; 2014; Tao <i>et al.</i> , 2005; Zhang and Xu, 1980; Zhang <i>et al.</i> , 2005 |
| | Costa Rica | | • | • | | Gasca and Suárez, 1992 |
| | Japan, Sagami Bay | • | • | • | • | Grossmann and Lindsay, 2013 |
| | Korea, St. 5 | | | • | • | This study |
| | Korea, St. 6 | • | • | • | • | This study |
| | Korea, St. 7 | | | | • | This study |
| | Korea, St. 11 | | | | • | This study |
| | Philippines, Celebes Sea | | • | • | • | Bigelow, 1928; Grossmann <i>et al</i> . 2015 |
| | Philippines, Sulu Sea | | • | | • | Grossmann et al., 2015 |
| | Taiwan | • | • | • | • | Hsieh <i>et al.</i> , 2013; Lo <i>et al.</i> , 2012; 2014; Zhang <i>et al.</i> , 2005 |
| | USA, California | • | | | • | Alvariño, 1991; Lluch-Cota <i>et al.</i> , 2007; Longhurst, 1967 |
| South Pacific | Australia, Coral Sea: Great Barrier Reef | | | • | | Totton, 1932 |
| | Chile | • | • | • | • | Palma and Silva, 2006 |
| | Chile, Easter Island | • | • | • | • | Palma and Silva, 2006 |
| | New Zealand, Exclusive Economic Zone (EEZ) | • | | • | • | Cairns et al., 2009 |
| | Papua New Guinea | | • | • | • | Pagès et al., 1989 |
| | Peru | | | • | • | Ayón et al., 2008 |
| North Atlantic | Adriatic Sea | • | | | • | Gamulin and Krsinić, 2000; Hure <i>et al.</i> , 2018 |
| | British Isles, North and West of the British Isles | | | | • | Fraser, 1967 |
| | Caribbean Sea | • | | | • | Alvariño, 1974; Michel and Foyo, 1977 |
| | Colombia | | • | • | • | Palomino et al., 2019 |
| | France, Villefranche- Sur-Mer | • | | | • | Leloup, 1935 |
| | Italy, Tyrrhenian Sea | | | | • | Zagami et al., 1996 |
| | Mediterranean Sea | • | • | • | • | Andersen <i>et al.</i> , 2001; Bouillon <i>et al.</i> , 2004; Mapstone, 2001; Sardou and Andersen, 1993 |
| | Mexico, Gulf of Mexico | • | • | • | • | Felder and Camp, 2009; Pugh and Gasca, 2009 |
| | Porcupine Seabight | • | | | | Kirkpatrick and Pugh, 1984 |
| | Sargasso Sea | • | | | | Purcell, 1981 |
| | Spain, Canary Island | | | • | • | Owre and Foyo, 1972; Soldevilla and Hernández, 1991 |
| | United Kingdom, Bermuda | • | | • | • | Hela <i>et al.</i> , 1953; Lo and Biggs, 1996; Moore, 1949 |
| | USA, Florida Current | | | _ | | Moore and Corwin, 1956 |

Table 3. Continued.

| | | | Newly record | ed species | | |
|-------------------|------------------------------|-----------------------------|------------------------|---------------------|------------------------|---|
| Ocean | Regions | Chelophyes appendiculata | Chelophyes contorta | Eudoxoides mitra | Eudoxoides spiralis | References |
| South Atlantic | Angola, Benguela Current | • | • | • | • | Pagès and Gili, 1992; Pagès <i>et al.</i> , 1991 |
| | Brazil | | | • | • | Gusmão <i>et al.</i> , 2015; Lang da Silveira <i>et al.</i> , 2011 |
| Indian | Arab emirates | | • | | | Sharaf and Al-Ghais, 1997 |
| Ocean | Arabian Sea | | | • | | Peter et al., 2018 |
| | Australia, Western Australia | | | | • | McCosker, 2016 |
| | India, Bay of Bengal | | | | • | Li et al., 2017 |
| | Indonesia | • | • | | | Wang et al., 2018 |
| | Jordan, Gulf of Aqaba | | • | • | | Mańko <i>et al</i> ., 2017 |
| | Madagascar | | | • | | Patriti, 1970 |
| | Pakistan | • | • | | | Morandini et al., 2015 |

tra, Totton (1932) reallocated *E. spiralis* from *Diphyes* to *Eudoxoides*. The eudoxid phase of *E. spiralis* is identified by Totton (1932) with the conical bract as in other eudoxid. It is easily identified as *E. spiralis* due to the twisted gonophore.

DISCUSSIONS

We conducted a morphological study of four unrecorded species of Diphyidae found in the South Sea and off Jeju Island, Korea. Two species belong to *Chelophy*es and another two species to *Eudoxoides*.

Chelophyes appendiculata is the type species of Chelophyes, and the most abundant species in the family Diphyidae (Totton, 1965). Up to 20 mm in length, it is larger than the other species in the same family (Kirkpatrick and Pugh, 1984). Specimens observed in this study were about 10 mm in length, twice as large as C. contorta (Table 2). Chelophyes appendiculata has a big difference in the shape of somatocyst compared with C. contorta, so the distinction between them is clear. Interestingly, E. mitra has a similar shape of somatocyst and size of anterior nectophore compared with C. appendiculata, so they are difficult to distinguish especially in lateral view. According to the previous reports (C. appendiculata: Gegenbaur, 1853; E. mitra: Bedot, 1896), these two species were identified as a single species, Diphyes gracilis. For clear identification, we need to check the apex view, whether there are three ridges reaching the apex (C. appendiculata) or five ridges (E. mitra), as suggested in Totton (1965).

Chelophyes contorta is about 7 mm in length (Totton, 1965). The mean length of present specimens is 5 mm, and about half size of *C. appendiculata* (Table 2). *Chelo-*

phyes contorta has a unique club-shaped somatocyst curved to the right. The number of ridges reaching the apex is four, which is the same as *E. spiralis*. However, the right-ventral ridge does not reach the apex in *C. contorta*, while the left-ventral ridge does not reach the apex in *E. spiralis* (Totton, 1965).

Eudoxoides mitra is the type species of Eudoxoides. It has been recorded up to 12 mm in length (Totton, 1965). Specimens observed in this study were about 10 mm in length. It is a large species like C. appendiculata (Table 2). Eudoxoides mitra has a straight body shape, and also has a posterior nectophore. Therefore, the distinction between E. mitra and E. spiralis is quite clear based on their body form.

Eudoxoides spiralis is 2–6 mm in length in the original description (Bigelow, 1911) and up to 11–12 mm in subsequent studies (Totton, 1965; Kirkpatrick and Pugh, 1984). The present specimens were about 5 mm in length, which fits well into the size range of the original report. Eudoxoides spiralis has a unique anterior nectophore which is completely twisted. In addition, E. spiralis has four ridges reaching the apex, while E. mitra has five ridges reaching the apex (Totton, 1965).

The genus *Eudoxoides* has a debatable history. Huxley (1859) established the genus, however it was Totton (1932) who defined and allocated *E. mitra* to *Eudoxoides*. This genus has a medium depth hydroecium and divided mouth-plate, and their anterior nectophore has a complete dorsal ridge. Since *Chelophyes* shares these characters with *Eudoxoides*, and their differences are not clear, it is necessary to check the generic identity of *Eudoxoides* in further molecular analysis.

Chelophyes appendiculata and E. mitra are the most abundant species in the family Diphyidae (Totton, 1965)

and they occur mainly in warm waters (Kirkpatric and Pugh, 1984). Chelophyes contorta and E. spiralis are rarer compared to other species belonging to Diphyidae (Totton, 1965). However, in the tropical western Indian Ocean, C. contorta is more abundant than C. appendiculata (Totton, 1954). The type localities of these four species are the Mediterranean Sea (C. appendiculata, Eschscholtz, 1829), Malay Archipelago (C. contorta, Lens and van Riemsdijk, 1908), southeast of Mauritius (E. mitra, Huxley, 1859) and tropical eastern Pacific Ocean (E. spiralis, Bigelow, 1911). Since their original reports, their distributions have been limited to warm waters (Mapstone, 2009). However, high temperature does not necessarily cause their high abundance. In the case of the Gulf of Mexico, where siphonophores appear frequently, colonies decrease at conditions of over 28.1°C (Sanvicente-Añorve et al., 2009) and extreme salinity (>36.5 or <34 PSU, Sanvicente-Añorve et al., 2007). According to Pakhomov et al. (1994), C. appendiculata and E. spiralis appear in the subtropical convergence regions but are absent in the Antarctic Polar Fronts showing limited distribution in low temperature.

These four species have been reported in the northwestern Pacific Ocean (Table 3). During the surveyed period, temperature and salinity were about 15.3°C and 34.6 PSU in Sagami Bay, Japan (Grossmann and Lindsay, 2013). In the Taiwan Strait, temperature ranged 14.7–24.4°C in winter and 25.18–30.08°C in summer, while salinity ranged 32.3–34.7 PSU in winter and 32.1–34.3 PSU in summer (Hsieh *et al.*, 2013). In the present study, conditions ranged 15.86–26.99°C in temperature and 33.48–34.49 PSU in salinity (Table 1). Those values are within the ranges of temperature and salinity in previous reports.

These four species are distributed globally except for the Antarctic and Arctic Oceans (Table 3). They are epipelagic (Mapstone, 2009), eurythermic, and euryhaline showing cosmopolitan distribution (Hsieh et al., 2013). However, morphological identification of siphonophores can be confusing due to their phenotypic plasticity. Their gelatinous bodies are easily damaged, and many records have been described based on dissociated specimens (Totton, 1965; Dunn et al., 2005). In addition, siphonophores have intraspecific size variation. For example, E. spiralis has a difference up to six times in adult size (2-12 mm). This size difference is quite large although gelatinous zooplanktons normally have variability in size depending on the habitat (Bouillon and Boero, 2000). This suggests the presence of cryptic species, although there are many difficulties in determining the cryptic species due to the lack of data (Moura et al., 2008; Pontin and Cruickshank, 2012). It would be useful if we can compare molecular markers of Siphonophorae in further study.

In conclusion, our findings update the confirmed Siphonophorae in Korea to be three suborders, five families, eight genera, and 13 species. These data suggest basic information on the biodiversity of siphonophores in Korean waters.

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REFERENCES

Alvariño, A. 1974. Distribution of siphonophores in the regions adjacent to the Suez and Panama Canals. Fishery Bulletin 72(2):527-546.

Alvariño, A. 1991. Abundancia y distribución batimétrica diurna y nocturna de los sifonóforos durante las cuatro estaciones del año 1969, en aguas de California y Baja California. Investigaciones marinas CICIMAR, 6:1-37.

Andersen, V., A. Gubanova, P. Nival and T. Ruellet. 2001. Zooplankton community during the transition from spring bloom to oligotrophy in the open NW Mediterranean and effects of wind events. 2. Vertical distributions and migrations. Journal of Plankton Research 23(3):243-261.

Ayón, P., M.I. Criales-Hernandez, R. Schwamborn and H.J. Hirche. 2008. Zooplankton research off Peru: a review. Progress in Oceanography 79(2-4):238-255.

Bedot, M. 1896. Les Siphonophores de la Baie d'Amboine: Étude Suivie d'une Revision de la Famille des Agalmidae. Revue suisse de zoologie 3(3):367-414.

Bigelow, H.B. 1911. The Siphonophorae. Reports on the scientific research expedition to the tropical Pacific. Albatross XXIII. Memoirs of the Museum of Comparative Zoology at Harvard College 38(2):173-401.

Bigelow, H.B. 1928. Hydromedusae, siphonophores and ctenophores of the "Albatross" Philippine Expedition. In: Contributions to the biology of the Philippine Archipelago and adjacent region. Bulletin United States National Museum 100(1):279-362.

Bouillon, J. and F. Boero. 2000. The Hydrozoa: A new clas-

- sification in the light of old knowledge. Thalassia Salentina 24:1-45.
- Bouillon, J., M.D. Medel, F. Pagès, J.M. Gili, F. Boero and C. Gravili. 2004. Fauna of the Mediterranean hydrozoa. Scientia Marina 68:5-438.
- Cairns, S.D., L.A. Gerhswin, F. Brook, P.R. Pugh, E.W. Dawson, V.O. Ocaña, W. Vervoort, G. Williams, J. Watson, D.M. Opresko, P. Schuchert, P.M. Hine, D.P. Gordon, H.J. Campbell, A.J. Wright, J. Sanchez and D.G. Fautin. 2009. Phylum cnidaria; corals, medusae, hydroids, myxozoa. New Zealand Inventory of Biodiversity. Volume 1. Kingdom Animalia: Radiata, Lophotrochozoa, Deuterostomia. pp. 59-101.
- Daniel, R. 1985. The fauna of India and the adjacent countries. Coelenterata: Hydrozoa, Siphonophora. Zoological Survey of India Publication, Calcutta, India.
- Dunn, C.W. and G.P. Wagner. 2006. The evolution of colony-level development in the Siphonophora (Cnidaria: Hydrozoa). Development Genes and Evolution 216(12):743-754.
- Dunn, C.W., P.R. Pugh and S.H.D. Haddock. 2005. Molecular phylogenetics of the Siphonophorae (Cnidaria), with implications for the evolution of functional specialization. Systematic Biology 54:916-935.
- Eschscholtz, F. 1829. System der Acalephen. Eine ausführliche Beschreibung aller medusenartigen Strahltiere. Ferdinand Dümmler, Berlin. pp. 1-190.
- Felder, D.L. and D.K. Camp (Eds.). 2009. Gulf of Mexico origin, waters, and biota: Biodiversity. Texas A and M University Press, Texas.
- Fraser, J. 1967. I. Siphonophora in the Plankton to the North and West of the British Isles. Proceedings of the Royal Society of Edinburgh. Section B. Biology 70(1):1-30.
- Gamulin, T. and F. Krsinić. 2000. Kalikofore (Siphonophora, Calycophorae) Jadranskog i Sredozemnog Mora. Calycophores (Siphonophora, Calycophorae) of the Adriatic Sea and Mediterranean Seas. Natura Croatica 9(2):1-198.
- Gao, S., H. Hong and S. Zhang. 2002. Phylum Cnidaria class Hydrozoa subclass Siphonophorae. Invertebrata 27 (Phylum Cnidaria). Fauna Sinica. pp. 1-275.
- Gasca, R. and E. Suárez. 1992. Sifonóforos (Cnidaria: Sipbonopbora) del Domo de Costa Ric. Revista de biología tropical 40(1):125-130.
- Gegenbaur, C. 1853. Beiträge zur näheren Kenntniss der Schwimmpolypen (Siphonophoren). Zeitschrift für wissenschaftliche Zoologie 5:285-344.
- Grossmann, M.M. and D.J. Lindsay. 2013. Diversity and distribution of the Siphonophora (Cnidaria) in Sagami Bay, Japan, and their association with tropical and subarctic water masses. Journal of Oceanography 69(4):395-411.
- Grossmann, M.M., A.G. Collins and D.J. Lindsay. 2014. Description of the eudoxid stages of *Lensia havock* and *Lensia leloupi* (Cnidaria: Siphonophora: Calycophorae), with a review of all known Lensia eudoxid bracts. Systematics and Biodiversity 12(2):163-180.

- Grossmann, M.M., J. Nishikawa and D.J. Lindsay. 2015. Diversity and community structure of pelagic enidarians in the Celebes and Sulu Seas, Southeast Asian tropical marginal seas. Deep Sea Research Part I: Oceanographic Research Papers 100:54-63.
- Gusmão, L.M.O., X.F.G. Diaz, M. de Melo Jr, R. Schwamborn and S. Neumann-Leitão. 2015. Jellyfish diversity and distribution patterns in the tropical Southwestern Atlantic. Marine Ecology 36(1):93-103.
- Hela, I., H.B. Moore and H. Owre. 1953. Seasonal changes in the surface water masses and in their plankton in the Bermuda area. Bulletin of Marine Science 3(3):157-167.
- Hsieh, H.Y., S.F. Yu and W.T. Lo. 2013. Influence of monsoon-driven hydrographic features on siphonophore assemblages in the Taiwan Strait, western North Pacific Ocean. Marine and Freshwater Research 64(4):348-358.
- Hure, M., H. Mihanović, D. Lučić, Z. Ljubešić and P. Kružić. 2018. Mesozooplankton spatial distribution and community structure in the South Adriatic Sea during two winters (2015, 2016). Marine Ecology 39(1).
- Huxley, T.H. 1859. The Oceanic Hydrozoa: A Description of the Calycophoridae and Physophoridae Observed During the Voyage of H.M.S. Rattlesnake in the Years 1846-1850. Ray society.
- Kirkpatrick, P.A. and P.R. Pugh. (Eds.). 1984. Siphonophores and velellids: keys and notes for the identification of the species. Synopses of the British fauna 29:154.
- Lang da Silveira, F. and A. Carrara Morandini. 2011. Checklist dos Cnidaria do estado de São Paulo, Brasil. Biota Neotropica 11(1):445-454.
- Leloup, E. 1934. Siphonophores calycophorides de l'Ocean Atlantique tropical et austral. Bulletin du Musée Royal d'Histoire Naturelle de Belgique 10(6):1-87.
- Leloup, E. 1935. Les siphonophores de la rade de Villefranche-sur-Mer (Alpes Maritimes, France). Bulletin de Musée Royal d'Histoire Naturelle de Belgique 11(31):1-12.
- Lens, A.D. and T. van Riemsdijk. 1908. The Siphonophora of the "Siboga" Expedition. Siboga-Expeditie 9:1-130.
- Li, K.Z., J.Q. Yin, L.M. Huang and X.Y. Song. 2012. Comparison of siphonophore distributions during the southwest and northeast monsoons on the northwest continental shelf of the South China Sea. Journal of Plankton Research 34(7):636-641.
- Li, K., J. Yin, L. Huang, Y. Tan and Q. Lin. 2017. A comparison of the zooplankton community in the Bay of Bengal and South China Sea during April-May, 2010. Journal of Ocean University of China 16(6):1206-1212.
- Lluch-Cota, S.E., E.A. Aragon-Noriega, F. Arreguín-Sánchez, D. Aurioles-Gamboa, J.J. Bautista-Romero, R.C. Brusca, R. Cervantes-Duarte, R. Cortés-Altamirano, P. Del-Monte-Luna, A. Esquivel-Herrera, G. Fernández, M.E. Hendrickx, S. Hernández-Vázquez, H. Herrera-Cervantes, M. Kahru, M. Lavín, D. Lluch-Belda, D.B. Lluch-Cota, J. López-Martínez, S.G. Marinone, M.O. Nevárez-Martínez,

- S. Ortega-García, E. Palacios-Castro, A. Parés-Sierra, G. Ponce-Díaz, M. Ramírez-Rodríguez, C.A. Salinas-Zavala, R.A. Schwartzlose and A.P. Sierra-Beltrán. 2007. The Gulf of California: review of ecosystem status and sustainability challenges. Progress in Oceanography 73(1):1-26
- Lo, W.T. and D.C. Biggs. 1996. Temporal variability in the night-time distribution of epipelagic siphonophores in the North Atlantic Ocean at Bermuda. Journal of Plankton Research 18(6):923-939.
- Lo, W.T., P.R. Kang and H.Y. Hsieh. 2012. Siphonophores from a transect off southern Taiwan between Kuroshio Current and South China Sea. Zoological Studies 51(8): 1354-1366.
- Lo, W.T., S.F. Yu and H.Y. Hsieh. 2014. Hydrographic processes driven by seasonal monsoon system affect siphonophore assemblages in tropical-subtropical waters (Western North Pacific Ocean). PIOS One 9(6):e100085.
- Longhurst, A.R. 1967. Diversity and trophic structure of zooplankton communities in the California Current. Deep Sea Research and Oceanographic Abstracts 14(4):393-402.
- Mackie, G.O. 1963. Siphonophores, bud colonies, and superorganisms. The Lower Metazoa (E. Dougherty, ed.). University of California Press, Berkeley, pp. 329-337.
- Mackie, G.O., P.R. Pugh and J.E. Purcell. 1987. Siphonophore Biology. Advances in Marine Biology 24:97-262.
- Mańko, M.K., A.W. Słomska and K. Jażdżewski. 2017. Siphonophora of the Gulf of Aqaba (Red Sea) and their associations with crustaceans. Marine Biology Research 13(5):480-485.
- Mapstone, G.M. 2001. Siphonophora, In: M.J. Costello *et al*. (eds.), European register of marine species: a checklist of the marine species in Europe and a bibliography of guides to their identification. Collection Patrimoines Naturels 50:120-122.
- Mapstone, G.M. 2009. Siphonophora (Cindaria: Hydrozoa) of Canadian Pacific waters. NRC Research Press, Ottawa, Ontario, Canada. pp. 1-302.
- Mapstone, G.M. 2014. Global diversity and review of the Siphonophorae (Cnidaria: Hydrozoa). PLOS One 9(2): e87737.
- McCosker, E. 2016. Influence of oceanographic conditions on coastal zooplankton assemblages at three IMOS National Reference Stations in Western Australia. Masters by Coursework thesis, Murdoch University.
- Michel, H.B. and M. Foyo. 1977. Studies on Caribbean zooplankton. In: Cooperative investigations of the Caribbean and adjacent regions, II. FAO Fisheries Reports 200:275-289.
- Moore, H.B. 1949. The zooplankton of the upper wqaters of the Bermuda area of the North Atlantic. Bulletin of the Bingham Oceanographic Collection, Yale University 12(2):1-97.
- Moore, H.B. and E.G. Corwin. 1956. The Effects of Tempera-

- ture, Illumination and Pressure on the Vertical Distribution of Zooplankton. Bulletin of Marine Science 6(4):273-287.
- Morandini, A.C., S. Gul, V. Haussermann and U. Pörschmann. 2015. Checklist of cnidarians from Pakistani waters. Check List 11:1.
- Moura, C.J., D.J. Harris, M.R. Cunha and A.D. Rogers. 2008. DNA barcoding reveals cryptic diversity in marine hydroids (Cnidaria, Hydrozoa) from coastal and deep-sea environments. Zoologica Scripta 37:93-108.
- Munro, C., S. Siebert, F. Zapata, M. Howison, A. Damian-Serrano, S.H. Church, F.E. Goetz, P.R. Pugh, S.H. Haddock and C.W. Dunn. 2018. Improved phylogenetic resolution within Siphonophora (Cnidaria) with implications for trait evolution. Molecular Phylogenetics and Evolution 127:823-833.
- National Institute of Biological Resources (NIBR). 2019. National List of Species of Korea (2018) [Available from: http://kbr.go.kr, accessed 25 September 2019].
- Nishiyama, E.Y., E.M. Araujo and O.M.P. Oliveira. 2016. Species of *Lensia* (Cnidaria: Hydrozoa: Siphonophorae) from southeastern Brazilian waters. Zoologia 33(6):1-14.
- Owre, H.B. and M. Foyo. 1972. Studies on Caribbean Zooplankton. Description of the Program and Results of the First Cruise. Bulletin of Marine Science 22(2):483-521.
- Pagès, F. and J.M. Gili. 1992. Siphonophores (Cnidaria, Hydrozoa) of the Benguela Current (southeastern Atlantic). Scientia Marina 56(1):65-112.
- Pagès, F., J.M. Gili and J. Bouillon. 1989. The Siphonophores (Cnidaria, Hydrozoa) of Hansa Bay, Papua New Guinea, Indo-Malayan Zoology 6:133-140.
- Pagès, F., H.M. Verheye, J.M. Gili and J. Flos. 1991. Short-term effects of coastal upwelling and wind reversals on epiplanktonic cnidarians in the southern Benguela ecosystem. South African Journal of Marine Science 10(1):203-211
- Pakhomov, E.A., R. Perissinotto and C.D. McQuaid. 1994. Comparative structure of the macrozooplankton/micronekton communities of the Subtropical and Antarctic Polar Fronts. Marine ecology progress series. Oldendorf 111(1):155-169.
- Palma, S. and N. Silva. 2006. Epipelagic siphonophore assemblages associated with water masses along a transect between Chile and Easter Island (eastern South Pacific Ocean). Journal of Plankton Research 28(12):1143-1151.
- Palomino, U.J., R. López, M.J. Gibbons, F. Gusmão and A.J. Richardson. 2019. Siphonophores from surface waters of the Colombian Pacific Ocean. Journal of the Marine Biological Association of the United Kingdom 99(1):67-80.
- Patriti, G. 1970. Aperçu systématique de la faune de siphonophores des zones superficielles et subsuperficielles des eaux du large de Tuléar (S.W. de l'Océan Indien, Madagascar). Recueil des Travaux de la Station marine d'Endoume. Fascicule hors série suppl 10:285-303.
- Peter, S., B. Manojkumar, D. Pillai, A. Velusamy, B. Kamarudeen, P. Sreeparvathy and F. Agnes. 2018. Distribution

- and Diversity of Gelatinous Zooplankton in the South Eastern Arabian Sea, Kanyakumari to off Kollam. Vestnik Zoologii 52(5):379-388.
- Pontin, D.R. and R.H. Cruickshank. 2012. Molecular phylogenetics of the genus *Physalia* (Cnidaria: Siphonophora) in New Zealand coastal waters reveals cryptic diversity. Hydrobiologia 686:91-105.
- Pugh, P.R. 1974. The vertical distribution of the siphonophores collected during the SOND cruise, 1965. Journal of the Marine Biological Association of the United Kingdom 54(1):25-90.
- Pugh, P.R. 1999. Siphonophorae, In: Boltvoskoy D (Ed.) Zooplankton. Backhuys Publishers, Leiden 1:467-513.
- Pugh, P.R. and R. Gasca. 2009. Siphonophorae (Cnidaria) of the Gulf of Mexico. Gulf of Mexico origins, waters, and biota 1:395-402.
- Purcell, J.E. 1981. Dietary composition and diel feeding patterns of epipelagic siphonophores. Marine Biology 65(1):83-90.
- Sanvicente-Añorve, L., C. Alba, M.A. Alatorre and C. FloresCoto. 2007. Cross-shelf and vertical distribution of siphonophore assemblages under the influence of freshwater outflows in the southern Gulf of Mexico. Hydrobiologia 586:69-78.
- Sanvicente-Añorve, L., C. Alba, C. Flores-Coto and M. CastilloRivera. 2009. Siphonophores off a riverine system in the southern Gulf of Mexico: factors affecting their distribution and spatial niche breadth and overlap. Aquatic Ecology 43:423-435.
- Sardou, J. and V. Andersen. 1993. Micronekton and macroplankton in the Ligurian Sea (Mediterranean Sea): Diel migrations and vertical distributions. Oceanologica Acta, Paris 16(4):381-392.
- Sears, M. 1950. Notes on siphonophores. I. Siphonophores from the Marshall Islands. Journal of Marine Research 9(1):1-16.
- Sharaf, G.M. and S.M. Al-Ghais. 1997. Distribution of zooplankton in offshore waters of the west coast of the united arab emirates. Kuwait Journal of Science and Engineering 24(1):131-144.
- Soldevilla, F.L. and P.H. Hernández. 1991. Preliminary list of zooplankton of the Canary Islands. II. Siphonophora, Pteropoda, Heteropoda, Ostracoda, Amphipoda and Decapoda. Boletim do Museu Municipal do Funchal 43(230):149-158.
- Stepanjants, S.D. 1967. Siphonophores of the seas of the USSR and the northern part of the Pacific Ocean. Opred Faune SSSR 96:1-216.

- Tao, Z., W. Rong and C. Yaqu. 2005. Net macrozooplankton community classification on the shelf area of the East China Sea and the Yellow Sea in spring and autumn. Acta Ecologica Sinica 25(7):1531-1540.
- Totton, A.K. 1932. Siphonophora. Scientific Reports, Great Barrier Reef Expedition 1928-29, 4:317-374.
- Totton, A.K. 1936. Plankton of the Bermuda Oceanographic Expeditions. VII. Siphonophora taken during the Year 1931. Zoologica: scientific contributions of the New York Zoological Society 21(4):231-240.
- Totton, A.K. 1954. Siphonophora of the Indian Ocean together with systematic and biological notes on related specimens from other oceans. Discovery Reports 27:1-162.
- Totton, A.K. 1965. A synopsis of the Siphonophora. British Museum (Natural History), The Thanet Press, Margate. 314.
- Totton, A.K. and J.H. Fraser. 1955. Siphonophora. Sub-Order: Calycophorae. Family: Diphyidae. Genera: Dimophyes, Muggiaea, Sulculeolaria, Chelophyes, Eudoxoides. Conseil International pour L'Exploration de la Mer Zooplankton Sheet 55:1-4.
- Wang, Y., X. Chen, B. Xing, R. Sun, N. Fitria, P. Xiang, C. Wang and M. Lin. 2018. Zooplankton composition and distribution in the Lembeh Strait of North Sulawesi, Indonesia. Acta Oceanologica Sinica 37(12):35-44.
- Xu, Z., M. Lin and Q. Gao. 2008. Causal analysis of the diversity of medusae in East China Sea. Frontiers of Biology in China 3(3):300-307.
- Zagami, G., F. Badalamenti, L. Guglielmo and A. Manganaro. 1996. Short-term variations of the zooplankton community near the Straits of Messina (North-eastern Sicily): relationships with the hydrodynamic regime. Estuarine, Coastal and Shelf Science 42(5):667-681.
- Zhang, J. and Z. Xu. 1980. On the geographical distribution of the Siphonophores in the China Sea. Acta Scientiarum Naturalium 19(3):100-108.
- Zhang, J., J. Hwang, G. Lian and S. Tang. 2005. Species diversity and abundance distribution of pelagic siphonophores from Nanwan Bay of Taiwan Island in late autumn and early winter. Journal of Tropical Oceanography 24(1):40-49.

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