NGS - quality control, alignment, visualisation

File types

File types

fasta	sequences
fastq	reads
sam/bam	alignments
bed	regions
gff	annotations
vcf	variants

fasta

- Plain sequence: *.fasta or *.fa
- Nucleotides or amino acids (proteins)
- Useful command:

  ```
  grep -c "^>" sequence.fasta
  ```

sequence.fasta

```
>sequence title1
ATCGTATCT
>sequence title2
ATGATGACGT
```

fastq files

reads.fastq

@D00283R:66:CC611ANXX:4:2311:2596:2330 1:N:0:TCCGGAG
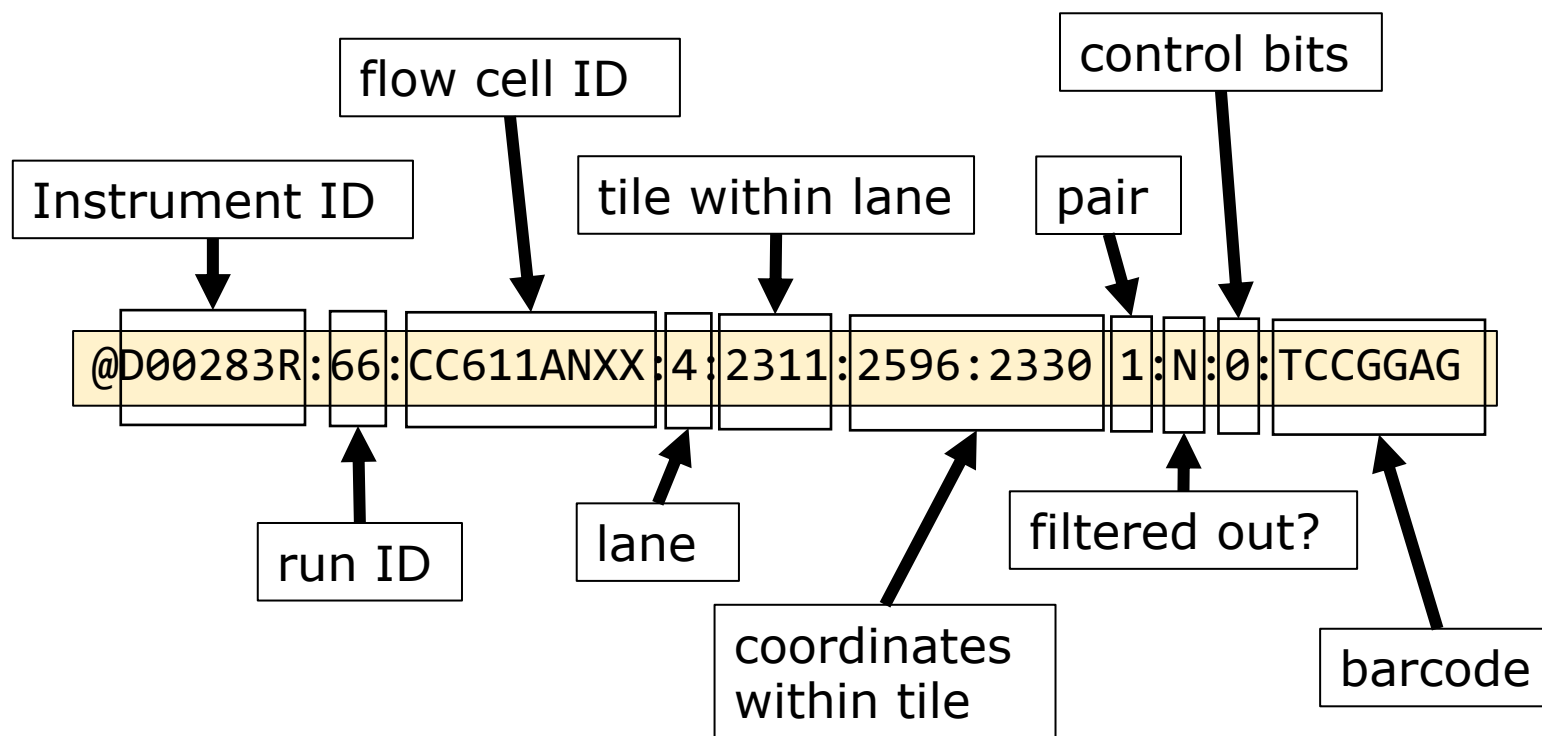ACTCTACGCTCAATAAAGATTTCTGATACGGCTCCTGAAATGCAGAATGAGT
+
B/<<<B<FFFFFFFFFFFBBFFFBFFFFBFFFF/FFFFFFFF/BFFFFFBFFF

title, starts with @

nucleotide sequence

optional description

base quality

```
!"#$%&'()*+,-./0123456789:;<=>?@ABCDEFGHI
|                              |   |        |
0.2........................26...31........41
```

fastq header

@D00283R:66:CC611ANXX:4:2311:2596:2330 1:N:0:TCCGGAG

- Instrument ID
- flow cell ID
- control bits
- tile within lane
- pair
- run ID
- lane
- coordinates within tile
- filtered out?
- barcode

5

Quiz Question 5

sam

sequence alignment format

Aim: alignments

sam header

```
@HD     VN:1.0  SO:coordinate
@SQ     SN:U00096.3     LN:4641652
@PG     ID:bowtie2      PN:bowtie2      VN:2.4.1
CL: "/opt/miniconda3/envs/ngs/bin/bowtie2-align-s \
--wrapper basic-0 \
-x /home/ubuntu/ecoli/ref_genome//ecoli-strK12-MG1655.fasta \
-1 /home/ubuntu/ecoli/trimmed_data/paired_trimmed_SRR519926_1.fastq \
-2 / home/ubuntu/ecoli/trimmed_data/paired_trimmed_SRR519926_2.fastq"
```

SAM column	example
read name	`@D00283R:66:CC611..`
flag	89
reference	`U00096.3`
start position	12513
mapping quality	42
CIGAR string	`5M2I7M3S`
reference name mate is mapped	`=`
start position mate	12928
fragment length	430
sequence	`CATCACCATTCCCAC`
base quality	`@>4:4C@89+&9CC@`
optional	`AS:i:-2`
optional	`XN:i:0`

Quiz Question 6

sam flags

Bit		Description
1	0x1	template having multiple segments in sequencing
2	0x2	each segment properly aligned according to the aligner
4	0x4	segment unmapped
8	0x8	next segment in the template unmapped
16	0x10	SEQ being reverse complemented
32	0x20	SEQ of the next segment in the template being reverse complemented
64	0x40	the first segment in the template
128	0x80	the last segment in the template
256	0x100	secondary alignment
512	0x200	not passing filters, such as platform/vendor quality controls
1024	0x400	PCR or optical duplicate
2048	0x800	supplementary alignment

	read paired?	properly aligned?	unmapped?	mate unmapped?	flag
read1	1	1	0	0	3
read2	1	0	0	1	9
read3	1	0	1	1	13
	1	2	4	8	

Quiz Question 7

bed

Browser Extensible Data

Aim: specify regions

regions.bed					
chr1	1701	1750	exon1	50	+
chr1	1780	1890	exon2	50	+

sequence title → start → end → name → score → strand
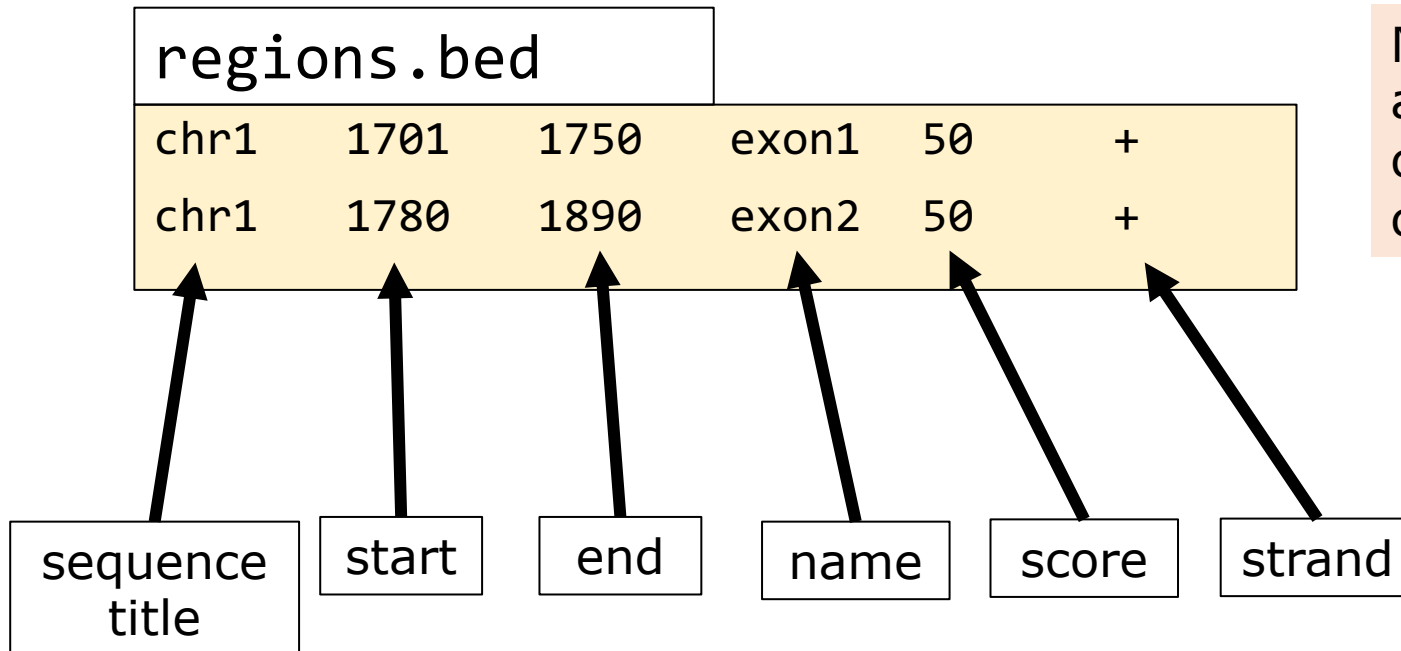
Numbering starts at 0!
chr1:1702-1750
chr1:1781-1890

gff

General Feature Format
Aim: annotation

seq name	source	feature	start	end	score	strand	frame	attributes
1	ensembl	mRNA	339070	346959	.	-	.	ID=…;
1	ensembl	exon	339070	339312	.	-	.	Parent=; …
1	ensembl	CDS	339070	339312	.	-	0	ID=;…

vcf

Variant Call Format

Aim: variants

```
##fileformat=VCFv4.2
##fileDate=20090805
##source=myImputationProgramV3.1
##reference=file:///seq/references/1000GenomesPilot-NCBI36.fasta
##contig=<ID=20,length=62435964,assembly=B36,md5=f126cdf8a6e0c7f379d618ff66beb2da,species="Homo sapiens",taxonomy=x>
##phasing=partial
##INFO=<ID=NS,Number=1,Type=Integer,Description="Number of Samples With Data">
##INFO=<ID=DP,Number=1,Type=Integer,Description="Total Depth">
##INFO=<ID=AF,Number=A,Type=Float,Description="Allele Frequency">
##INFO=<ID=AA,Number=1,Type=String,Description="Ancestral Allele">
##INFO=<ID=DB,Number=0,Type=Flag,Description="dbSNP membership, build 129">
##INFO=<ID=H2,Number=0,Type=Flag,Description="HapMap2 membership">
##FILTER=<ID=q10,Description="Quality below 10">
##FILTER=<ID=s50,Description="Less than 50% of samples have data">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Read Depth">
##FORMAT=<ID=HQ,Number=2,Type=Integer,Description="Haplotype Quality">
#CHROM POS     ID        REF   ALT    QUAL FILTER INFO                             FORMAT     NA00001        NA00002        NA00003
20     14370   rs6054257 G     A      29   PASS   NS=3;DP=14;AF=0.5;DB;H2          GT:GQ:DP:HQ 0|0:48:1:51,51 1|0:48:8:51,51 1/1:43:5:.,.
20     17330   .         T     A      3    q10    NS=3;DP=11;AF=0.017             GT:GQ:DP:HQ 0|0:49:3:58,50 0|1:3:5:65,3   0/0:41:3
20     1110696 rs6040355 A     G,T    67   PASS   NS=2;DP=10;AF=0.333,0.667;AA=T;DB GT:GQ:DP:HQ 1|2:21:6:23,27 2|1:2:0:18,2   2/2:35:4
20     1230237 .         T     .      47   PASS   NS=3;DP=13;AA=T                 GT:GQ:DP:HQ 0|0:54:7:56,60 0|0:48:4:51,51 0/0:61:2
20     1234567 microsat1 GTC   G,GTCT 50   PASS   NS=3;DP=9;AA=G                  GT:GQ:DP   0/1:35:4       0/2:17:2       1/1:40:3
```
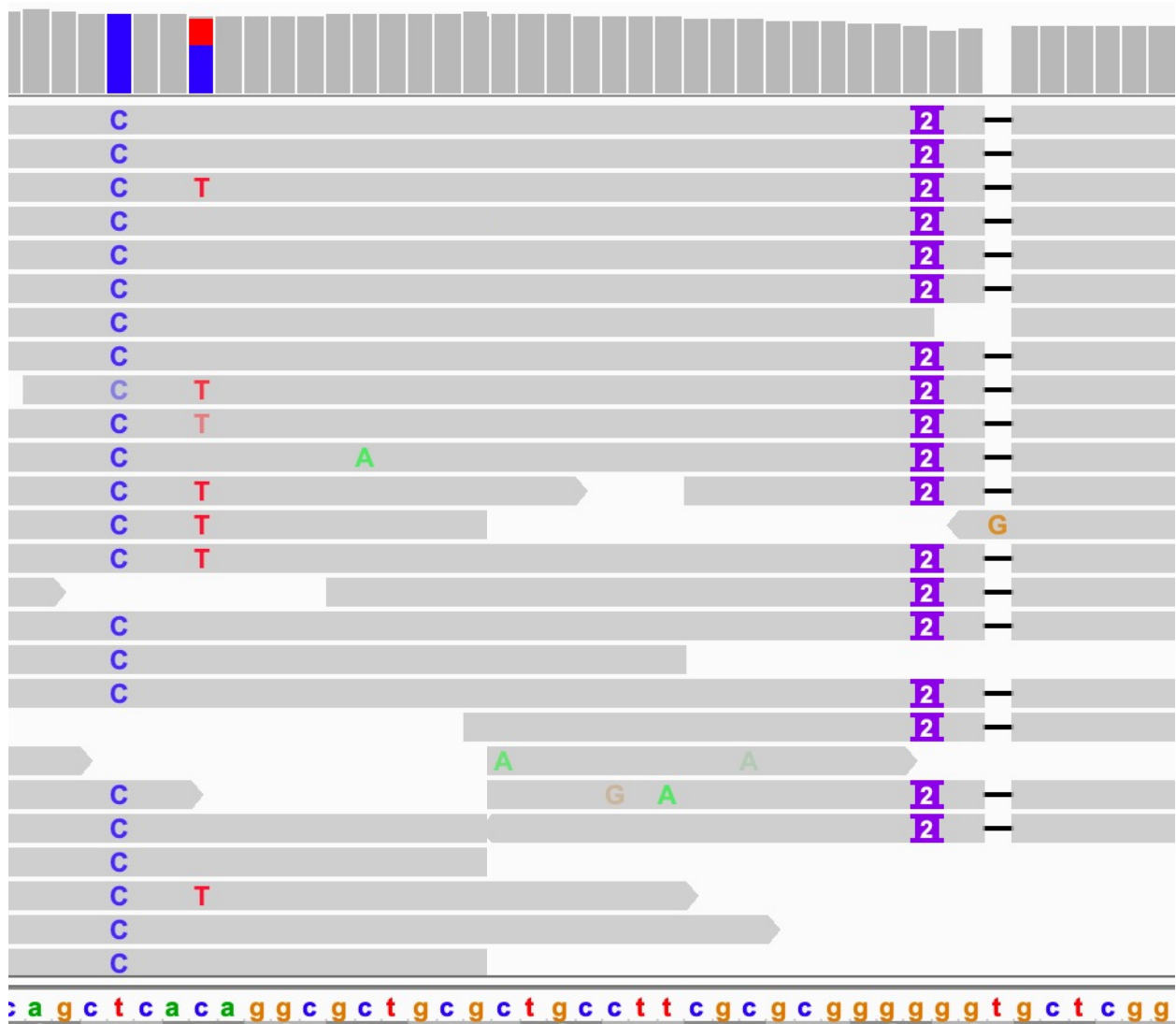
vcf

```
##fileformat=VCFv4.3
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##INFO=<ID=AA,Number=1,Type=String,Description="Ancestral Allele">
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##FORMAT=<ID=GQ,Number=1,Type=Integer,Description="Genotype Quality">
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##FORMAT=<ID=HQ,Number=2,Type=Integer,Description="Haplotype Quality">
```

#CHROM	POS	ID	REF	ALT	QUAL	FILTER	INFO	FORMAT	NA00001	NA00002
20	14370	rs6054257	G	A	29	PASS	NS=3;DP=14;AF=0.5;DB;H2	GT:GQ:DP:HQ	0|0:48:1:51,51	1|0:48:8:51,51
20	17330	.	T	A	3	q10	NS=3;DP=11;AF=0.017	GT:GQ:DP:HQ	0|0:49:3:58,50	0|1:3:5:65,3
20	1110696	rs6040355	A	G,T	67	PASS	NS=2;DP=10;AF=0.333,0.667;AA=T;DB	GT:GQ:DP:HQ	1|2:21:6:23,27	2|1:2:0:18,2
20	1230237	.	T	.	47	PASS	NS=3;DP=13;AA=T	GT:GQ:DP:HQ	0|0:54:7:56,60	0|0:48:4:51,51
20	1234567	microsat1	GTC	G,GTCT	50	PASS	NS=3;DP=9;AA=G	GT:GQ:DP	0/1:35:4	0/2:17:2

samples