



# PICI INTERNSHIP 2025

CATERINA PONTI

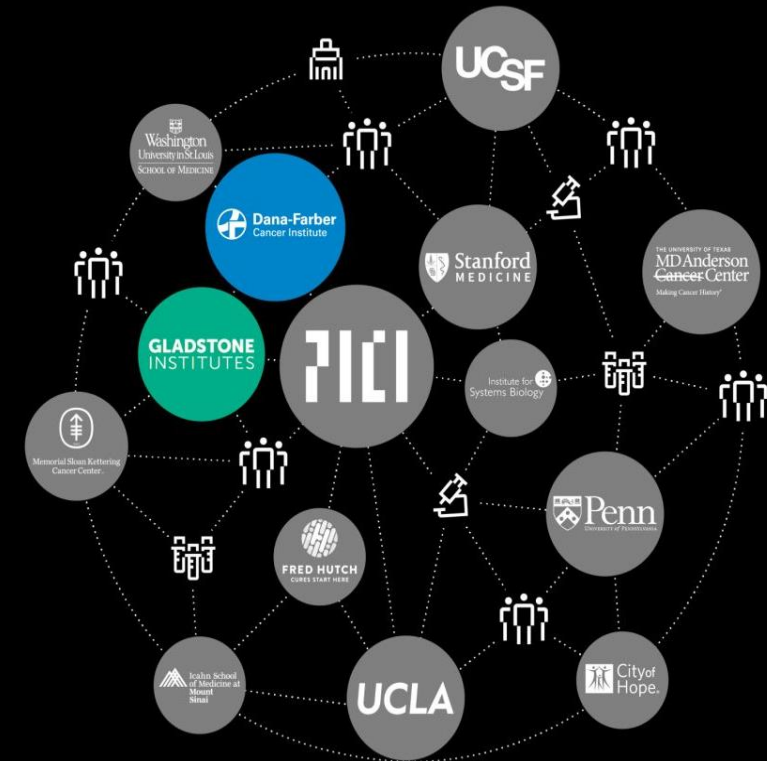
Data Science and Bioinformatics Intern

# PARKER INSTITUTE FOR CANCER IMMUNO THERAPY (PICI)

Our mission is to **accelerate the development of breakthrough immune therapies** to turn all cancers into curable diseases.

## WHAT IS PICI?

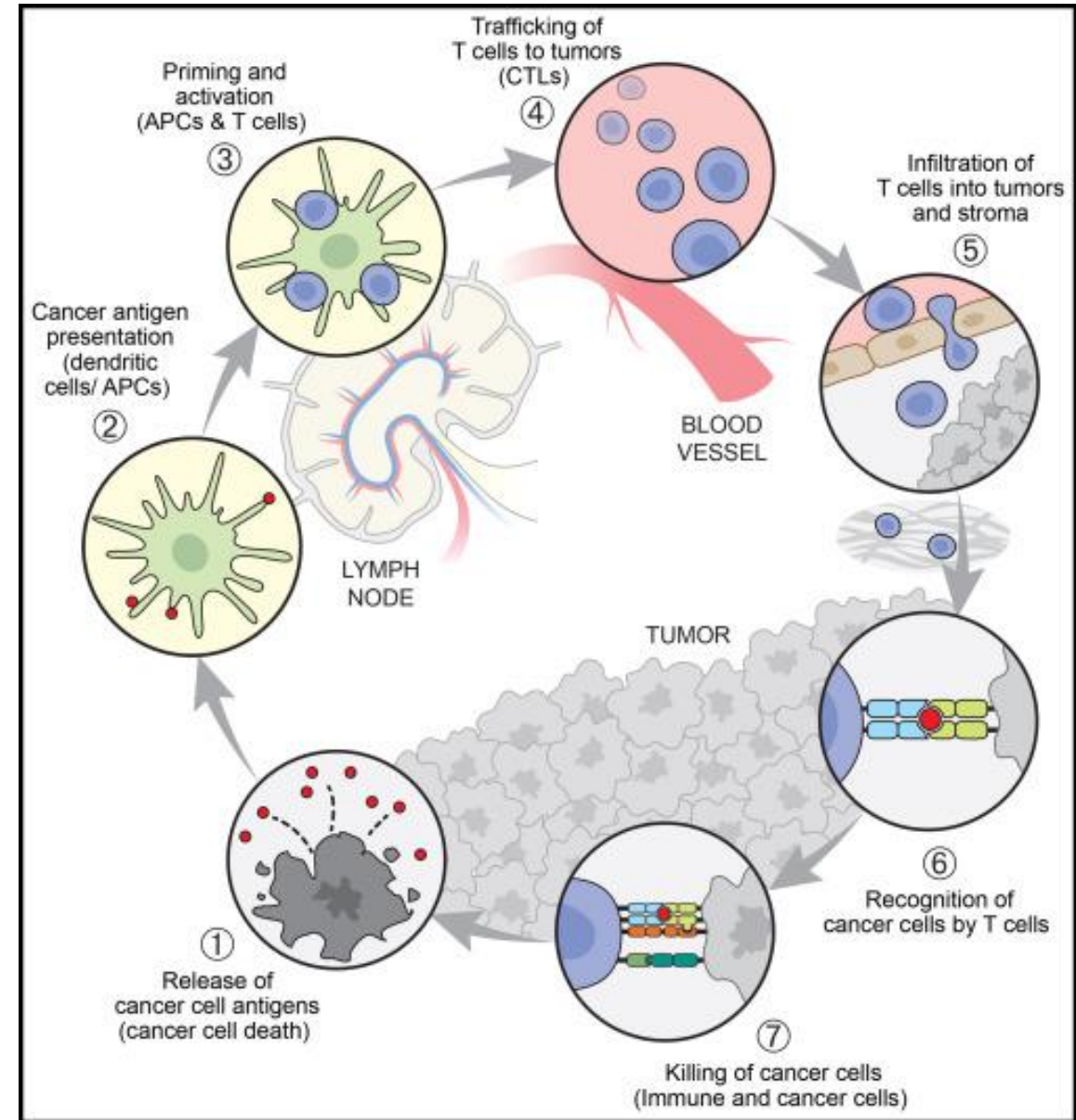
- Patient-centric, grant-giving non-profit organization
- Fund & accelerate high-impact research
- Drive collaboration (700+ Cross-Institutional PICI Investigators)
- Enable bold Innovations



# THE CANCER-IMMUNITY CYCLE

1. **Release:** Dying cancer cells release "markers" (Antigens).
2. **Presentation:** Specialized scout cells (Dendritic cells) capture these markers.
3. **Priming:** Scouts travel to lymph nodes to "teach" T-cells what the cancer looks like.
4. **Trafficking:** Newly trained T-cells enter the bloodstream to find the tumor.
5. **Infiltration:** T-cells exit the blood and "invade" the tumor tissue.
6. **Recognition:** T-cells identify the specific cancer cells using the markers from Step 1.
7. **Killing:** T-cells destroy the cancer cells, which releases *more* markers, restarting the cycle at Step 1.

**Where the System Breaks Down:** Cancer cells can **turn off** T cells by using "brakes" on the immune system called **immune checkpoints** -> this stops T cells from attacking



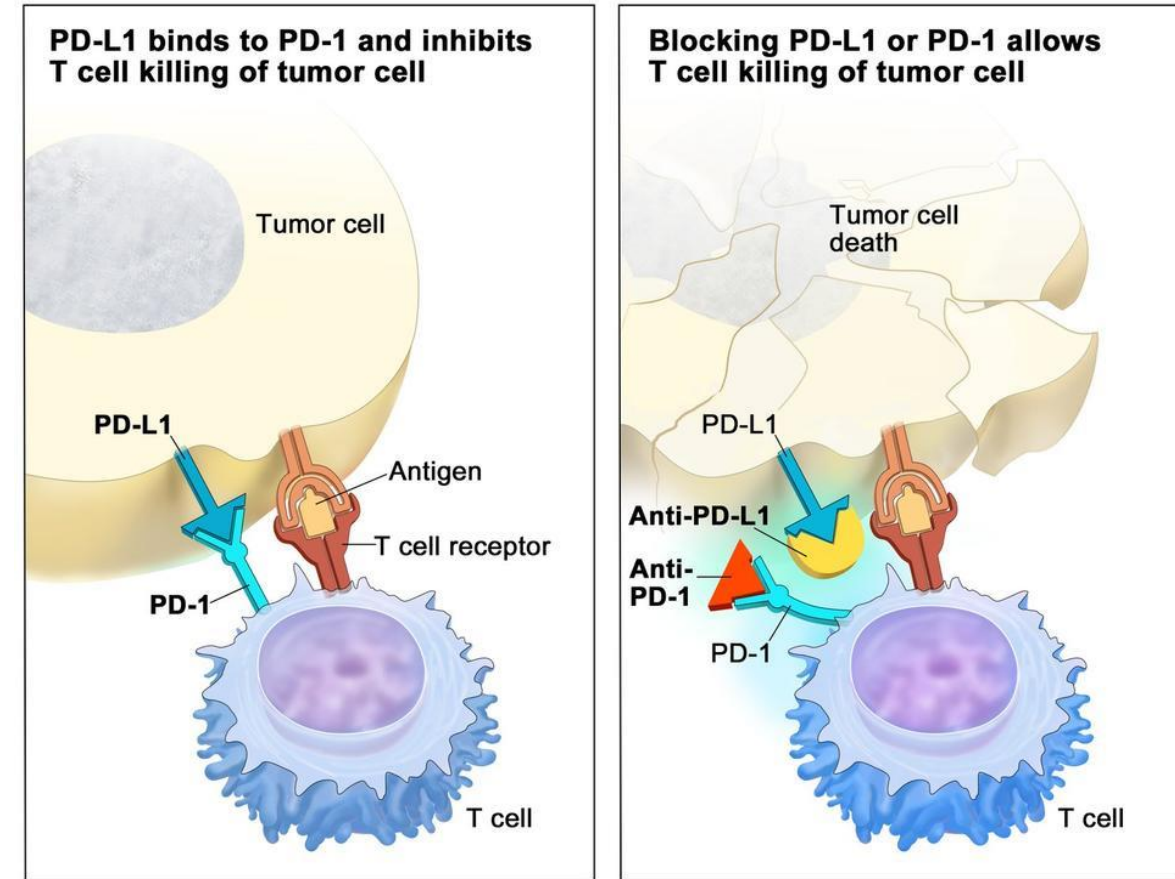
# IMMUNE CHECKPOINT INHIBITOR (ICI) THERAPY

Goal: To restart the Cancer-Immunity Cycle by removing the biochemical "brakes" (checkpoints) that cancer uses to survive.

- Anti-PD-1 / Anti-PD-L1 (e.g., pembrolizumab, atezolizumab)
- Anti-CTLA-4 (e.g., ipilimumab)

## Clinical Challenges

- Immune-related Adverse Events (irAEs) Because we are "releasing the brakes," the immune system can become overactive..
- It may mistakenly attack healthy organs (e.g., lungs, skin, or colon), leading to inflammation.



# RADIOHEAD Study Planning

## Study Support and Goals:

- Funded by the [Helmsley Charitable Trust](#), [JDRF](#), and [Bristol Myers Squibb](#).
- **Goal:** Identify **biomarkers** to prevent or intervene in:
  - Cancer immunotherapy-induced **Type 1 Diabetes**.
  - Severe Immune-Related Adverse Events (irAEs).

## Academic Guidance and Study Direction:

- [UCSF](#): data collection and trial objectives.
- [Huntsman Cancer Institute](#): data cleaning and curation work.

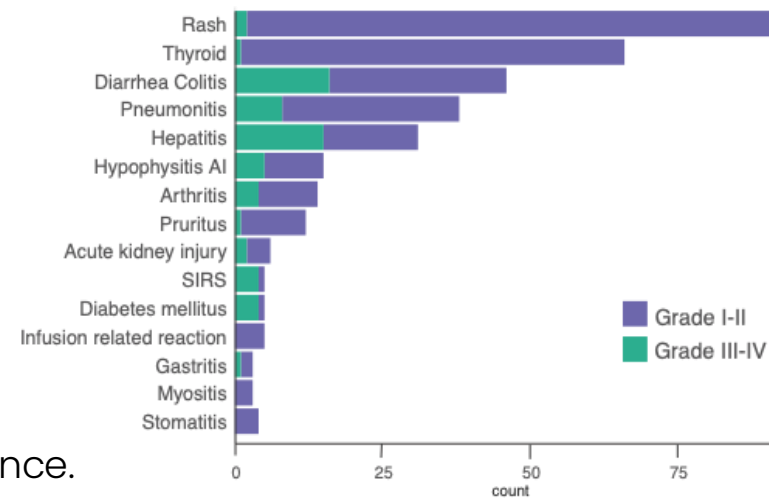
## Key Challenges:

- Recruitment for a Rare Event was difficult: 12 Expected, 3 Recruited.

## Root Cause Analysis:

- Patient Behavior
- COVID Impact
- Technical Issues
  - Difficulty in diagnosing T1D in community centers lacking specialized experience.

**New Focus:** Association between all **Immune-Related Adverse Events (irAEs)** and **Cancer Treatment Response**.

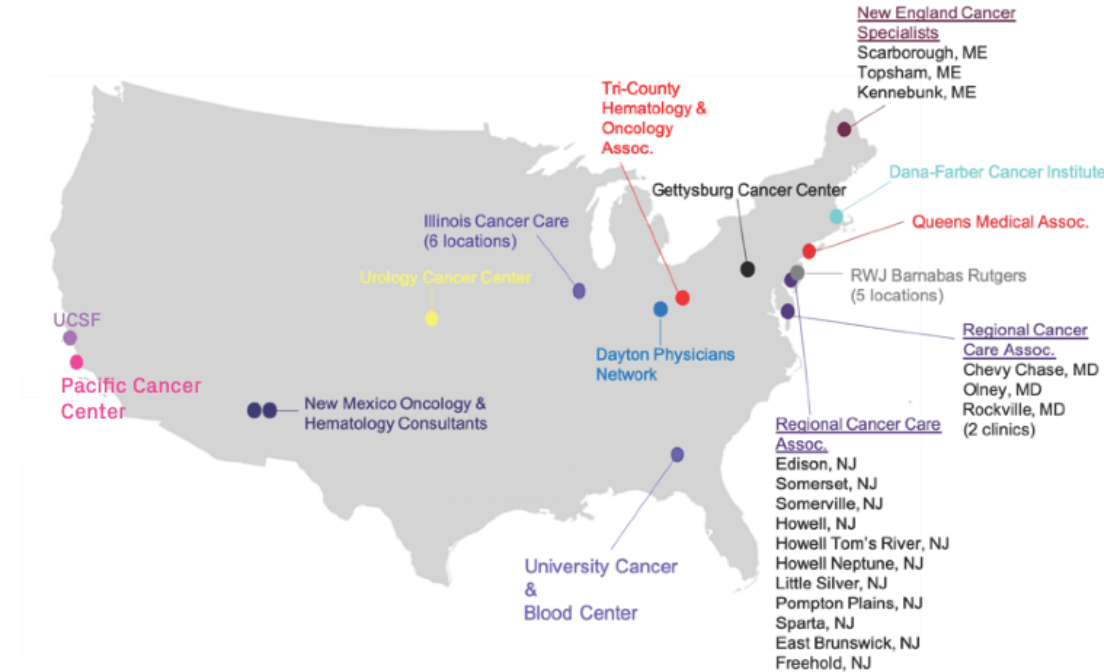


# RADIOHEAD Sample Collection

- **Standardized Processes:** All blood samples shipped overnight to ensure consistency across sites.
- **Central Laboratory:** Usage of a single central facility to eliminate processing variables.
- **Site Selection Strategy**
- **Batch Effect Analysis**

## Systemic Data Biases

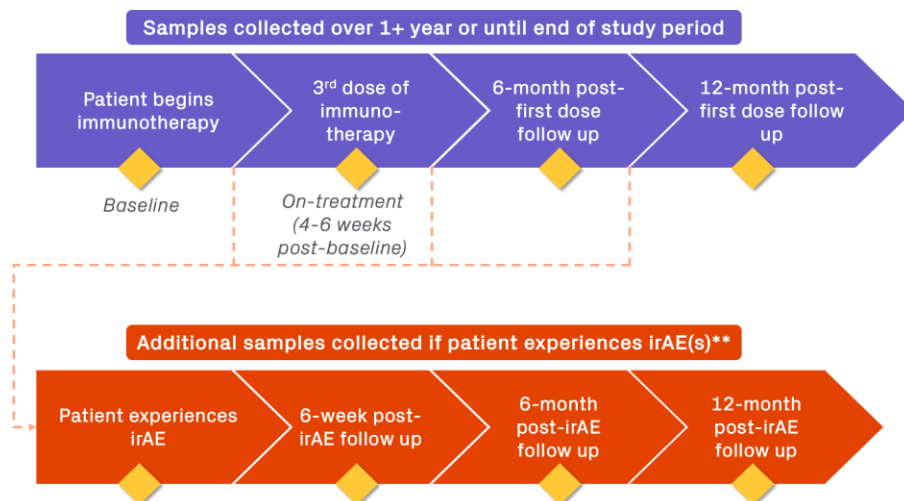
- Geographic Bias: study privileges expensive, resource-rich urban hubs
- Demographic Gap: underrepresentation of minority
- Economic Barriers: Cost of Access
- IRAE Potentially underreported
- Lack of Longitudinal Data (patient follow up is inconsistent because of pandemic)
- Clinical Limitations: additional disease information was often a free textbox





# What Makes RADIOHEAD Unique ?

- Scale and Data:
  - Data collected from **1,070** patients (~2.5 draws per patients).
  - Generated over **70,000** samples across **3,500+** combinations of timepoints.
  - Samples were collected over one year.
- Event-Driven Sampling
  - Additional samples collected in the event of an adverse event (irAE)



1070 Patients  
3500+ Timepoints

70K+ Sample  
Aliquots Banked

Patient Information

Whole Blood

Treatment Detail

Plasma

Outcomes Data

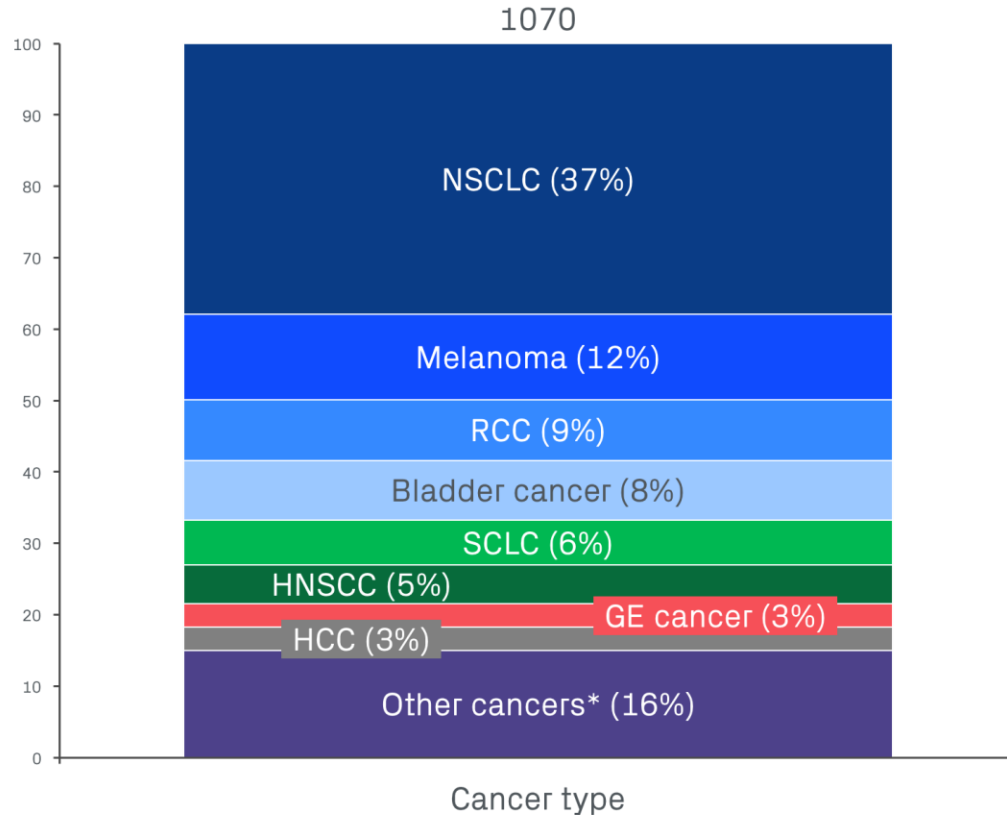
Serum

Additional Metadata

PBMC

# What Makes RADIOHEAD Unique ?

Distribution of patient tumor types in RADIOHEAD dataset  
Number of patients



## Comprehensive Multiomic Analyses

ctDNA

~750 genes, completed



Serum Proteomics

600+ proteins, completed



WES / SNP-panel

completed



Teiko.bio

HD Flow Cytometry

80 markers, in progress  
(complete by mid-2025)



Bulk-RNAseq

In progress (complete by mid-2025)

Single cell RNA-seq

Projected to start in April 2025



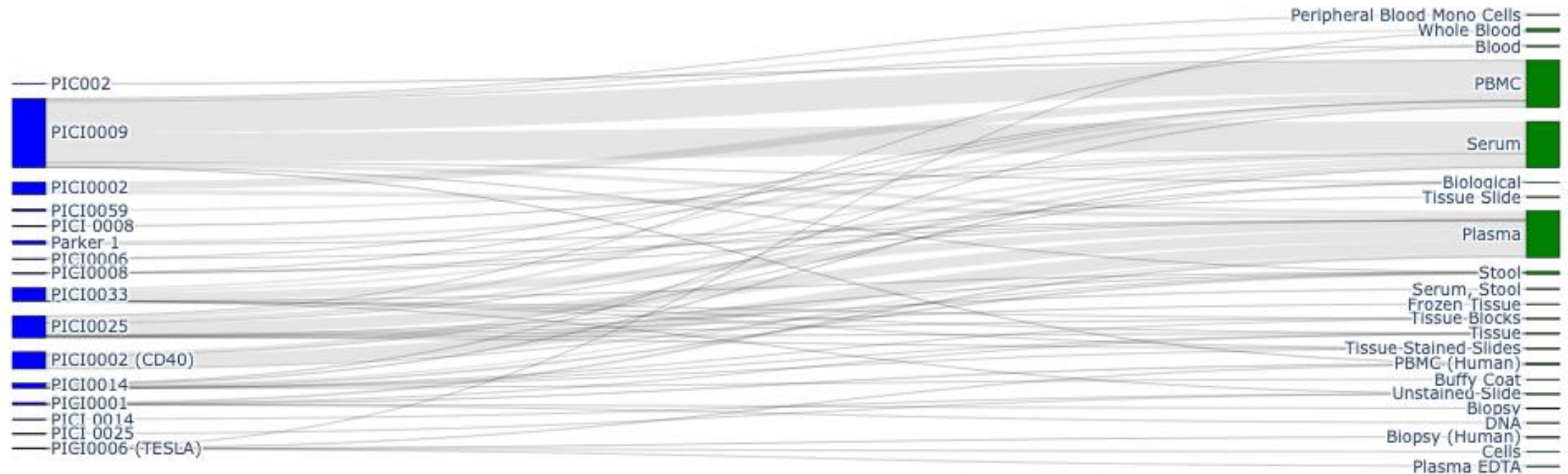
Metabolomics

Evaluating partners



# BIOREPOSITORY

## BIOREPOSITORY 2024 PULL



- The biorepository includes samples from 10 additional PICI-owned clinical trials.
- Limitations: inconsistent naming conventions and duplicate entries.
  - I developed a **data clean-up and standardization script** to support future data pulls.
- RADIOHEAD (PICI-009) is a high-priority study due to its external partnerships and multi-omics integration capabilities.

# RADIOHEAD DATASETS

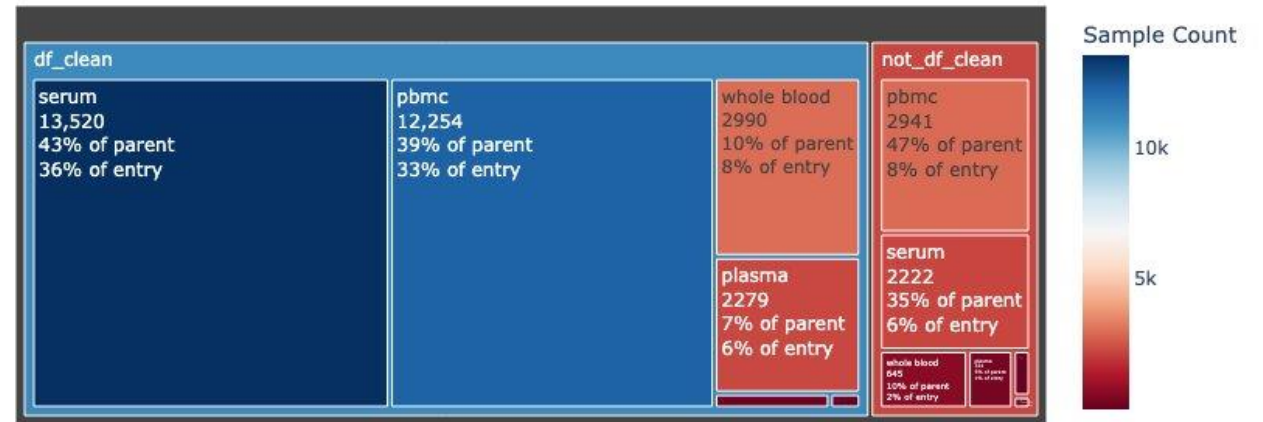
## Biorepository

Multiple pulls: keeping track of sample shipments

REDCap: clinical annotations + samples' barcodes of patients (1,300) collected during clinical trial

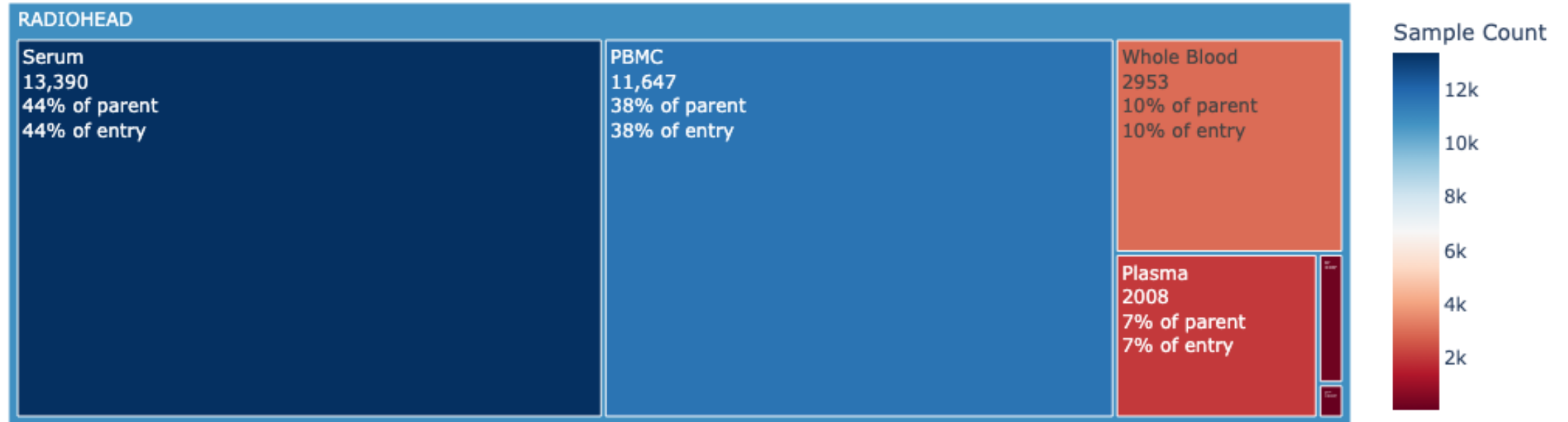
**df\_clean**: cleaned REDCap with 1,070 patients of all patients with a pretreatment sample

Sample Material Types in PICI 009 Biorepository  
(from df\_clean patients: 83.3% of all samples)

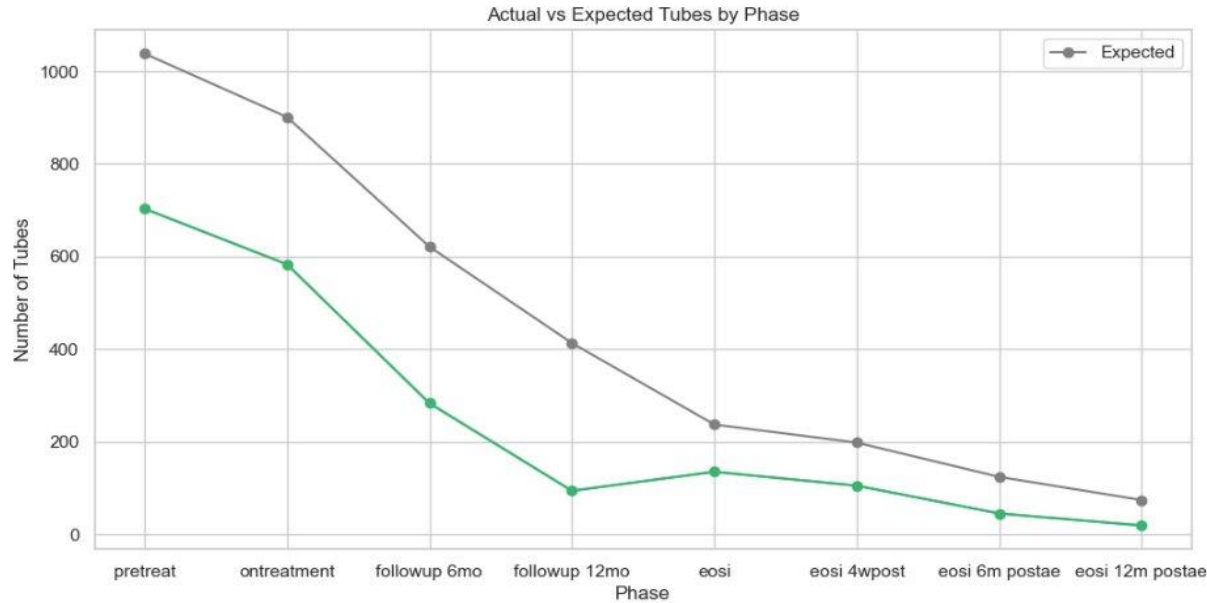


# BARCODES MATCHING AND OUTLIER DETECTION (RADIOHEAD)

Sample Material Types in Biorepository from the 1070 RADIOHEAD patients selected in df\_clean.

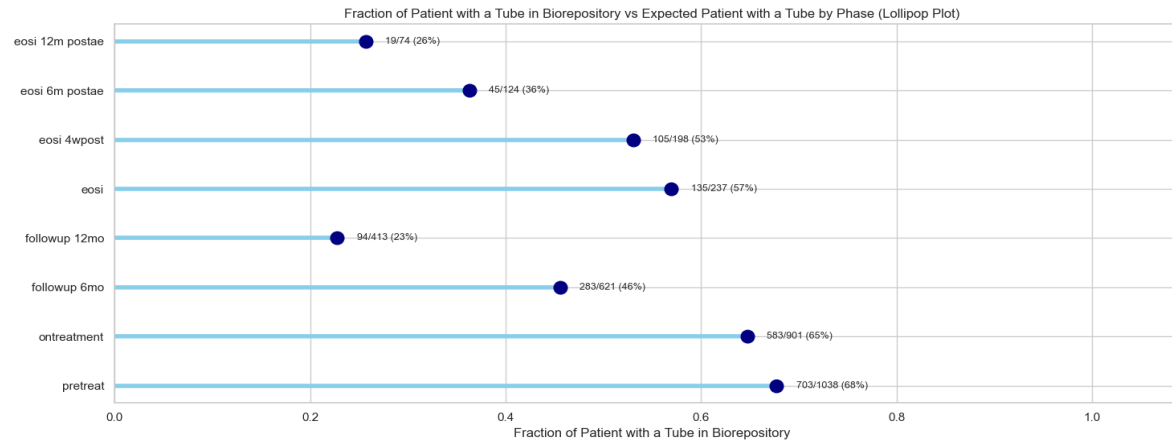


- Cleaned and standardized barcodes using custom parsing rules.
- Detected outliers (barcodes assigned to the wrong project name) using pairwise **Levenshtein distances** (via [RapidFuzz](#)) via [RapidFuzz](#) to compare alphanumeric barcodes.
- Matched barcodes between biorepository and REDCap using exact and substring matching.

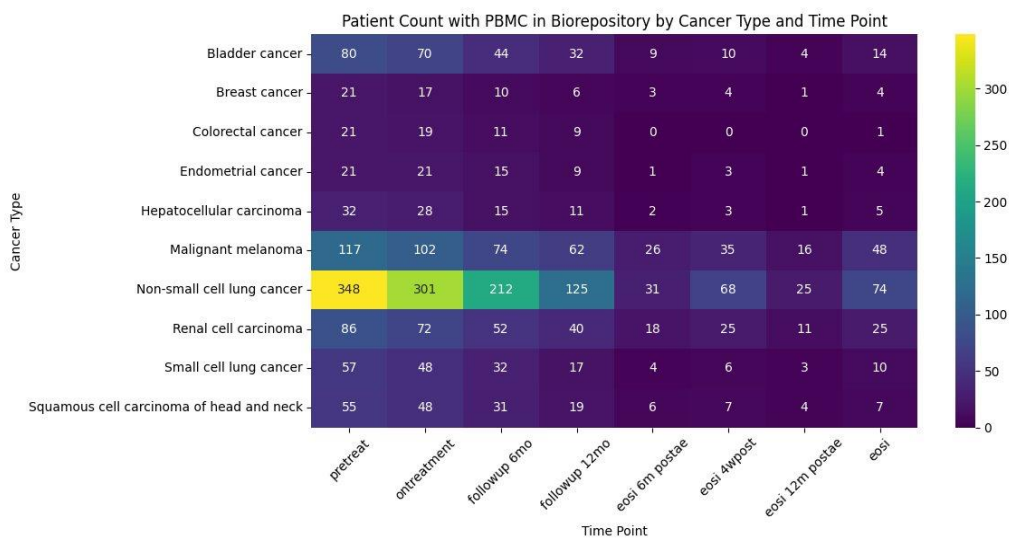
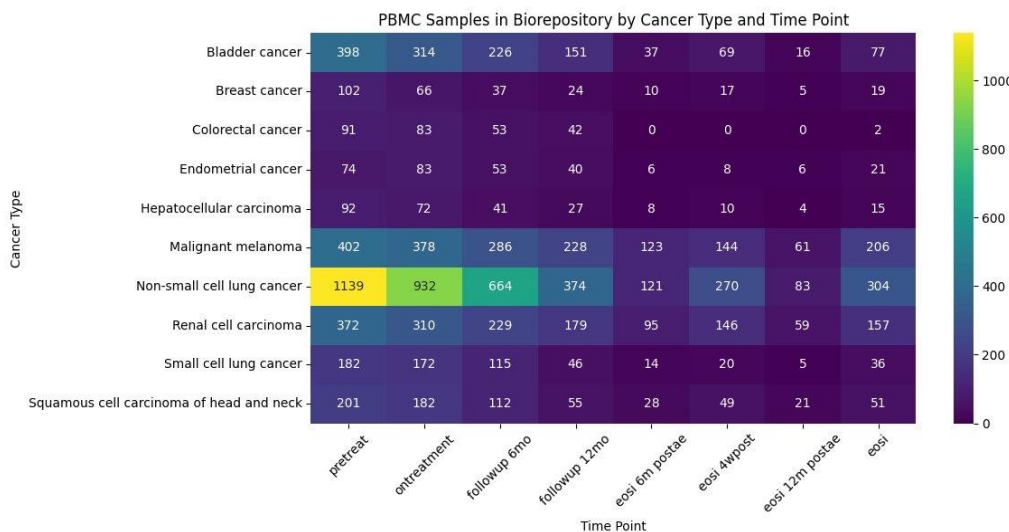


## SAMPLE COUNT AND PATIENT BY TIMEPOINT ACTUAL VS. EXPECTED (FROM REDCAP)

- Gap between the expected samples tracked in REDCap and the actual samples available in the biorepository as of 2025
- Percentage of expected tubes collected per patient during clinical trial vs samples available now in the biorepository



# SAMPLES AND PATIENT COUNT BY TIMEPOINT AND CANCER TYPE



1. Counts unique samples and patients remaining in the biorepository, grouped by cancer type and timepoint.
2. Provides a clear view of sample availability across disease groups and visits.
3. Helps scientists quickly identify gaps, trends, and well-represented cohorts when planning analyses or selecting samples.

## WHY MOVING FROM SCIRPTS TO A WEB PLATFORM?

- Internal Efficiency: Automated scripts to replace manual queries
- Partner Collaboration
- Real-Time Updates
- User Friendly

Use filters below to explore samples from the RADIOHEAD clinical trial.

Choose Biorepository

Biorepository: ☐ Biorepository 2024 ☒ Biorepository 2025

Group Samples by: Cancer Type v

Medical History: All

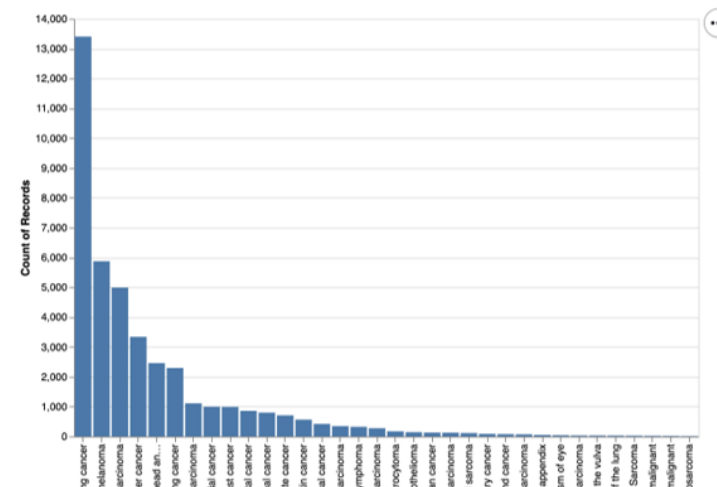
Pre-treatment: All

irAE: All

Color By: None

Apply Filters

### Sample Count Bar Chart



### Filtered Data Table

Participant ID	Barcode	BSI ID	Material Type	Barcode Timepoint
<a href="#">4-1010</a>	BK01750	PIC000001 1262	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 1266	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 2319	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 2323	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 2436	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 2486	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 2489	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 2538	Plasma	Pre-Treatment
<a href="#">4-1010</a>	BK01750	PIC000001 2541	Plasma	Pre-Treatment
<a href="#">3-1573</a>	10B86237-07	PIC000002 0343	Serum	Follow-Up
<a href="#">3-1573</a>	10B86237-07	PIC000002 0344	Serum	Follow-Up
<a href="#">3-1573</a>	10B86237-07	PIC000002 0345	Serum	Follow-Up

### Sample Distribution Donut Chart

Use filters below to explore samples from the RADIOHEAD clinical trial.

Biorepository: ☐ Biorepository 2024 ☒ Biorepository 2025

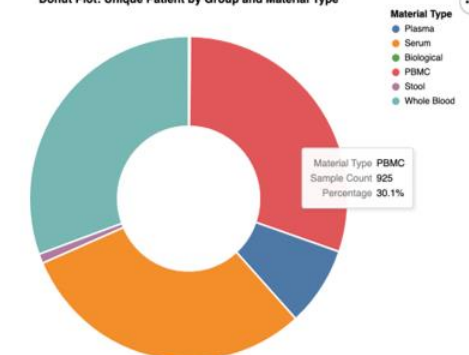
Cancer Type: All

Timepoint: All 

Group By: Patient Count

### Donut Plot: Unique Patient by Group and Material Type

Donut Plot: Unique Patient by Group and Material Type





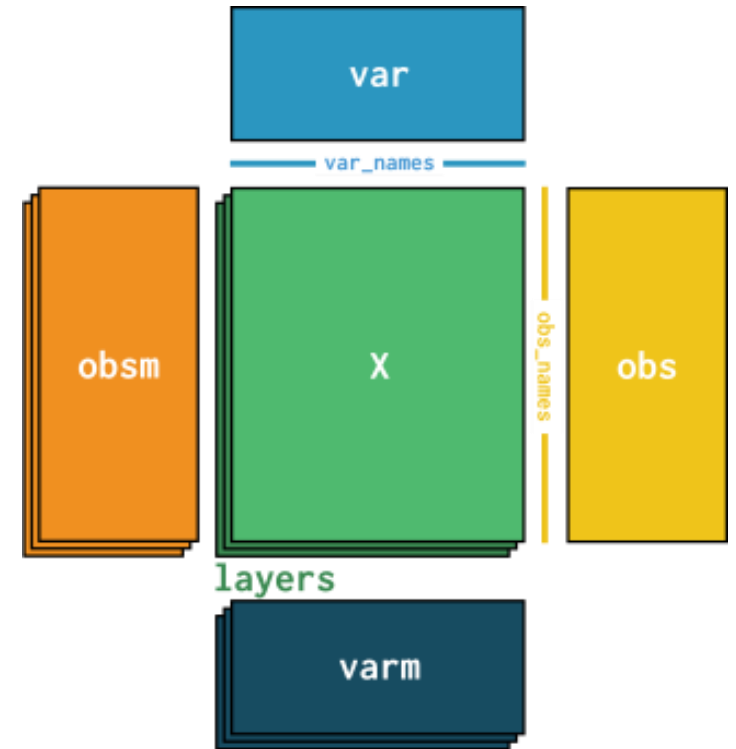
## Mass Cytometry Data (CyTOF) from Teiko

### Technology and Overview:

- Uses **antibodies** to quantify up to 50 single-cell biological markers.
- Partnership with **Teiko** completed data acquisition for all melanoma patients.

### Raw Data:

- Each sample ~10,000 cells
- High dimensionality: 50 markers x 10,000 cells x 500 samples
- **AnnData** (Annotated Data) object



Parse FCS into  
AnnData



Downsample cells per  
sample



Marker-  
Aware downsampling

# The Cloud Solution

## Challenges

- High-dimensional data → long runtimes & heavy compute
- Running on a laptop took ~8 hrs
- Local workflow limits collaboration

## The Cloud Solution

- What is the Cloud?
- Remote compute & storage instead of local machine

## Why the Cloud

- Scales analysis beyond local computing limits.
- Enables fast processing of large single-cell datasets.
- Supports reproducible, shareable pipelines.



# Cloud Workflow Architecture

## Cloud Environment Setup

- Linux Virtual Machine (Ubuntu)

## Data Access and Storage

- **GCS Bucket** mounted locally to the Virtual Machine for data and workflows access.

## Data Processing and Analysis

- Running pipelines for parsing (FCS to AnnData) and data analysis.

# Analytical Workflow

## 1. Data Processing

- Preprocessing and normalization of raw data using **Scanpy** and **Anndata** to ensure quality.

## 2. Dimensionality Reduction

- Application of **UMAP** to project high-dimensional marker data into interpretable 2D embeddings.

## 3. Unsupervised Clustering

- Utilizing **Leiden** (graph-based) and **FlowSOM** (SOM-based) algorithms to identify distinct cell populations.

## 4. Visualization & Output

- Generation of marker expression heatmaps, cluster comparison plots, and interactive visualizations.

# CLUSTER VALIDATION: SILHOUETTE ANALYSIS

**Clustering:** Cells were clustered using unsupervised methods (**Leiden** / **FlowSOM**), with K = 5 clusters selected for evaluation.

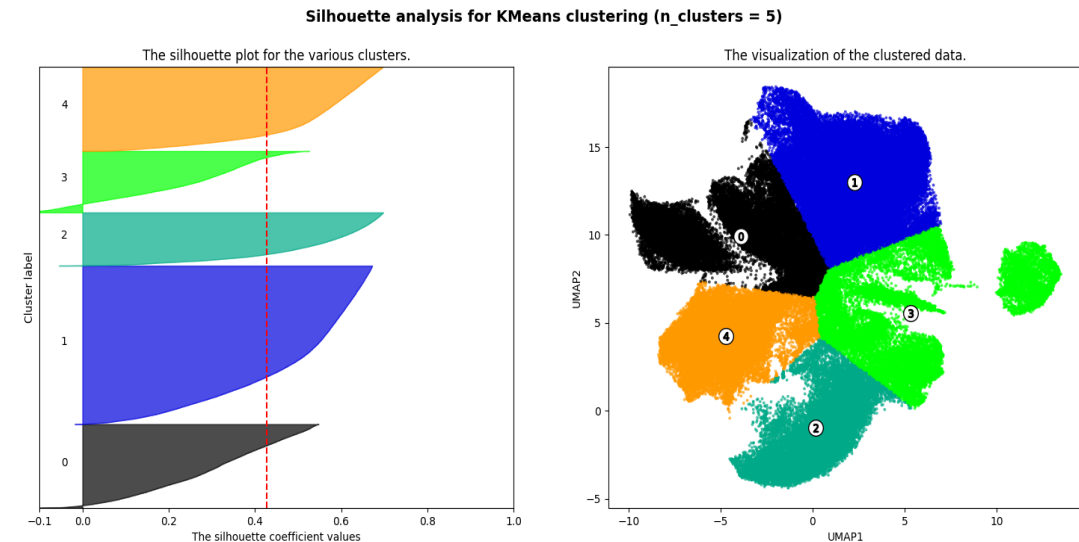
For every cell (data point)  $i$ , we calculate the **Silhouette Score** based on two distances:

- **Intra-cluster distance ( $a$ ):** Distance to cells in its own cluster (should be small).
- **Inter-cluster distance ( $b$ ):** Distance to cells in the nearest neighbor cluster (should be large).

Interpreting the Silhouette Plot

- **Each Line = One Cell:** The plot is made of horizontal bars; every single line represents an individual cell's score.
- **The "Knife" Shape:** You want thick, "knife-shaped" blocks. This indicates that most cells have high scores and are well-assigned.
- **The Threshold:** The vertical dashed line is the **average score**. You want most of your "knives" to extend past this line.

$$S(i) = \frac{b(i) - a(i)}{\max\{a(i), b(i)\}}$$

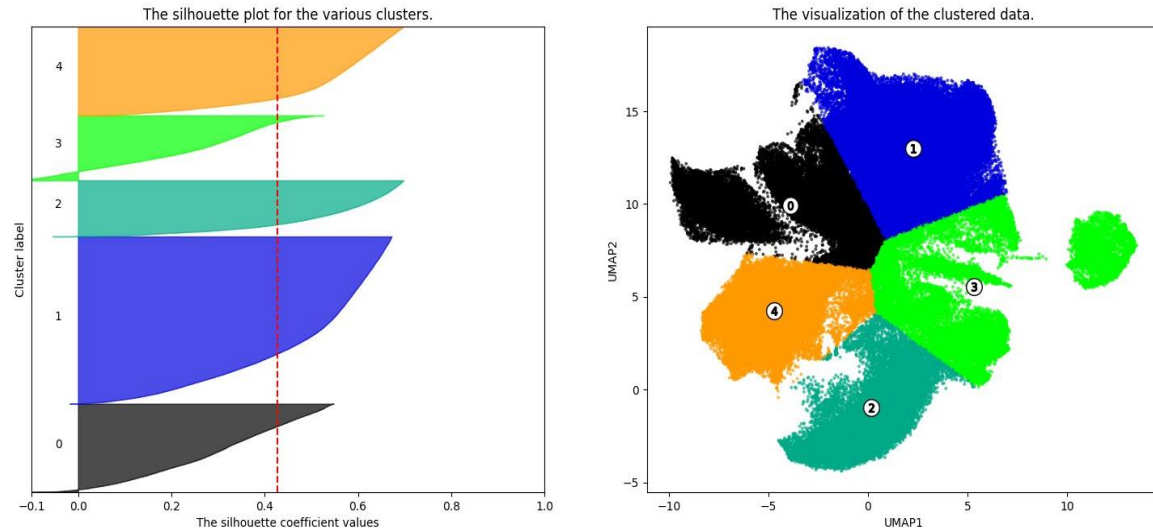




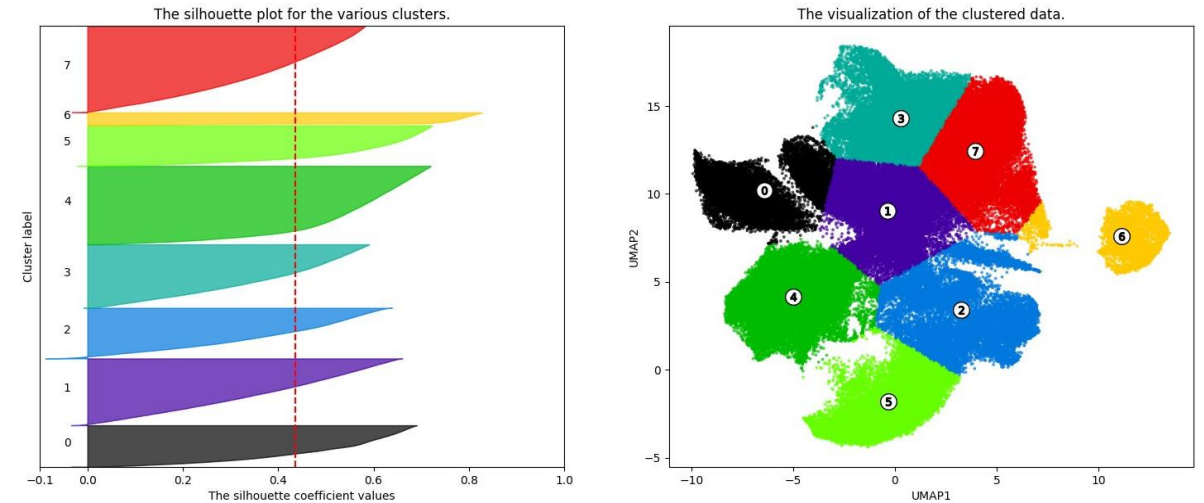
# HOW TO DETERMINE THE OPTIMAL NUMBER OF CLUSTERS (K)?

## SILHOUETTE PLOTS FOR K MEANS CLUSTERING ANALYSIS

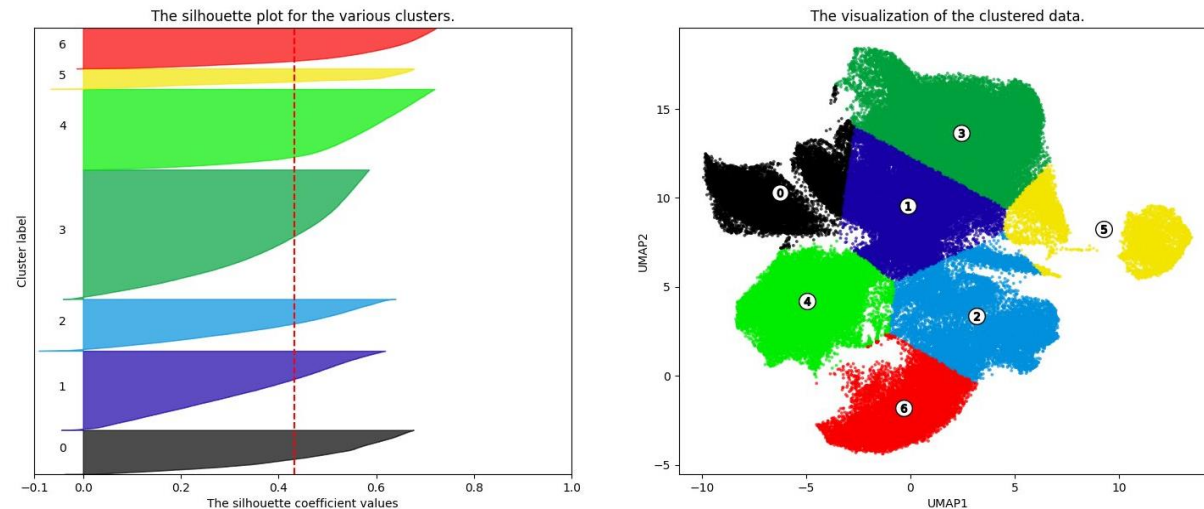
**Silhouette analysis for KMeans clustering (n\_clusters = 5)**



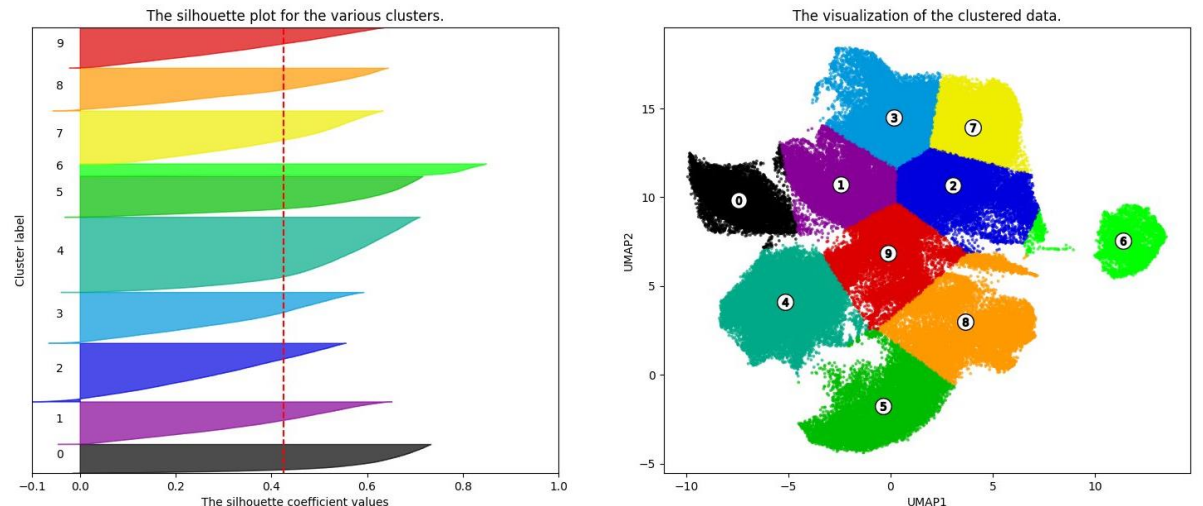
**Silhouette analysis for KMeans clustering (n\_clusters = 8)**



**Silhouette analysis for KMeans clustering (n\_clusters = 7)**



**Silhouette analysis for KMeans clustering (n\_clusters = 10)**





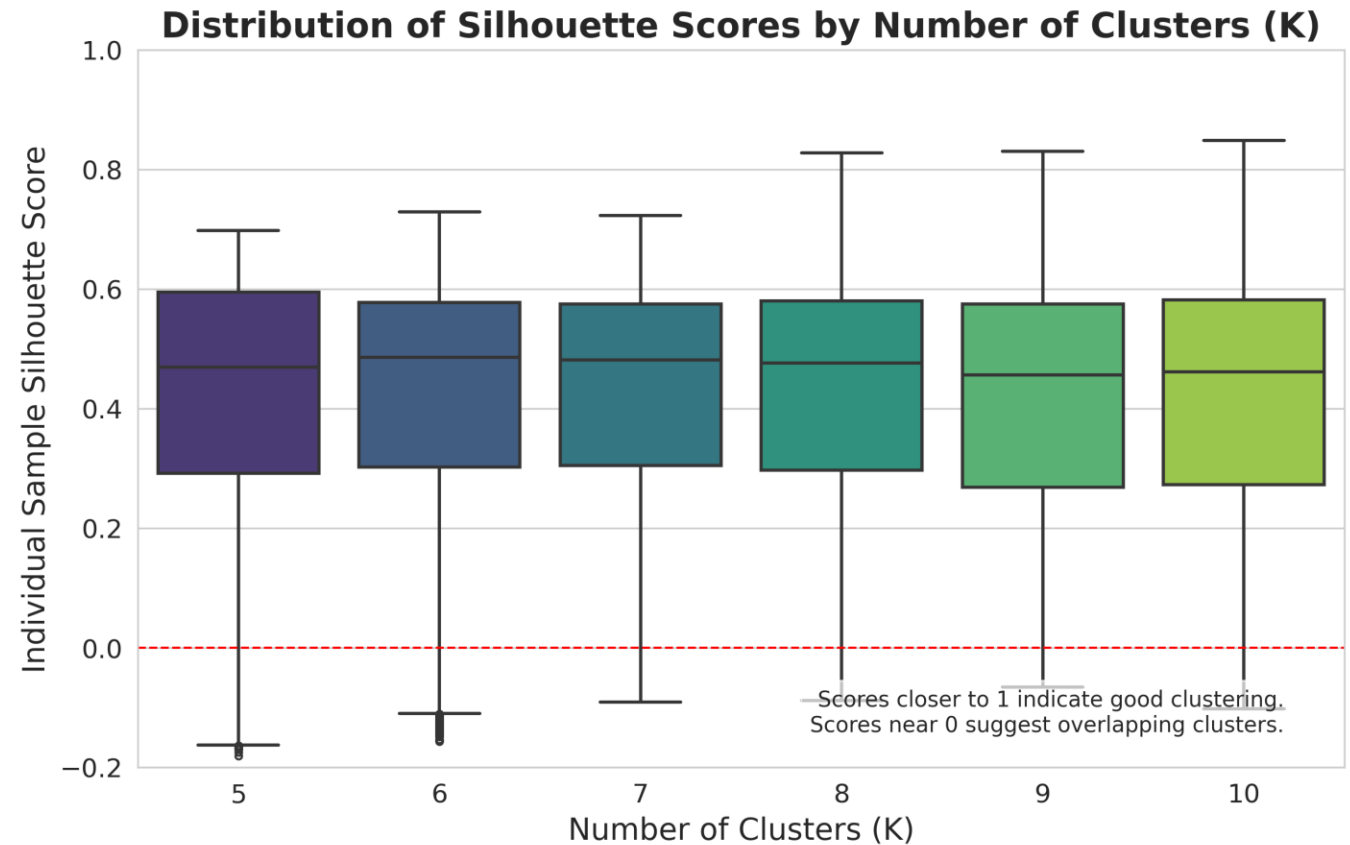
# HOW TO DETERMINE THE OPTIMAL NUMBER OF CLUSTERS (K)?

What is a **Silhouette Scores** Boxplot?

Represents the distribution of the silhouette scores for every data point across a range of cluster counts (K)

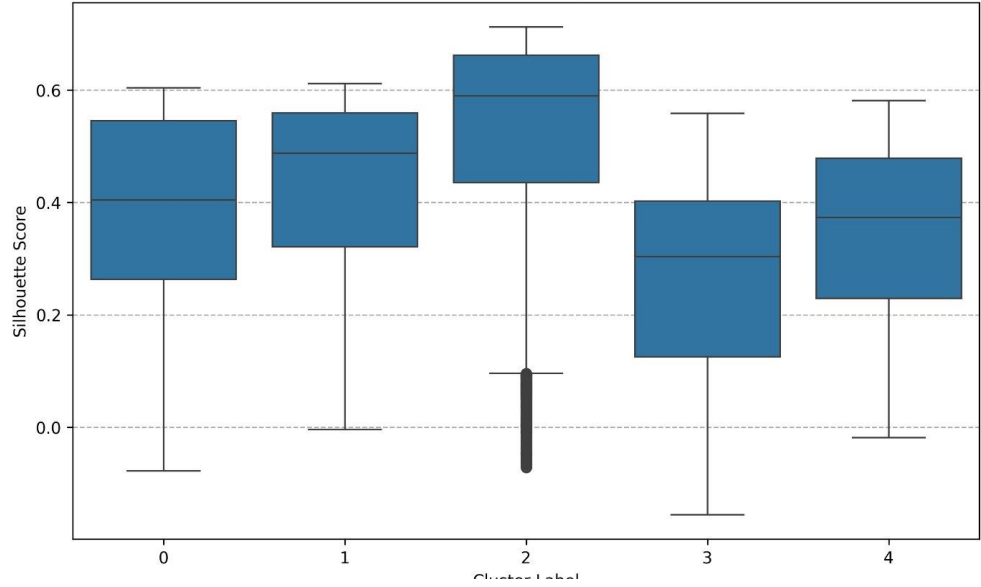
Optimal K:

- **Median of Each Silhouette Score:**  
higher indicates better separation on average
- **Interpreting Cluster Cohesion** (Variability/IQR): less spread suggests more consistent cluster assignments across samples

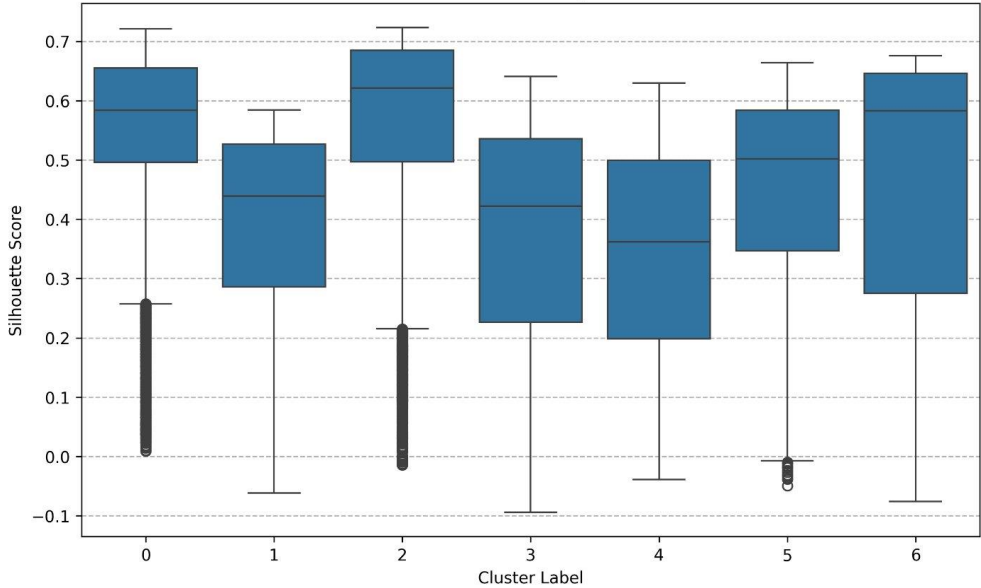


# HOW GOOD IN AVERAGE ARE MY CLUSTERS?

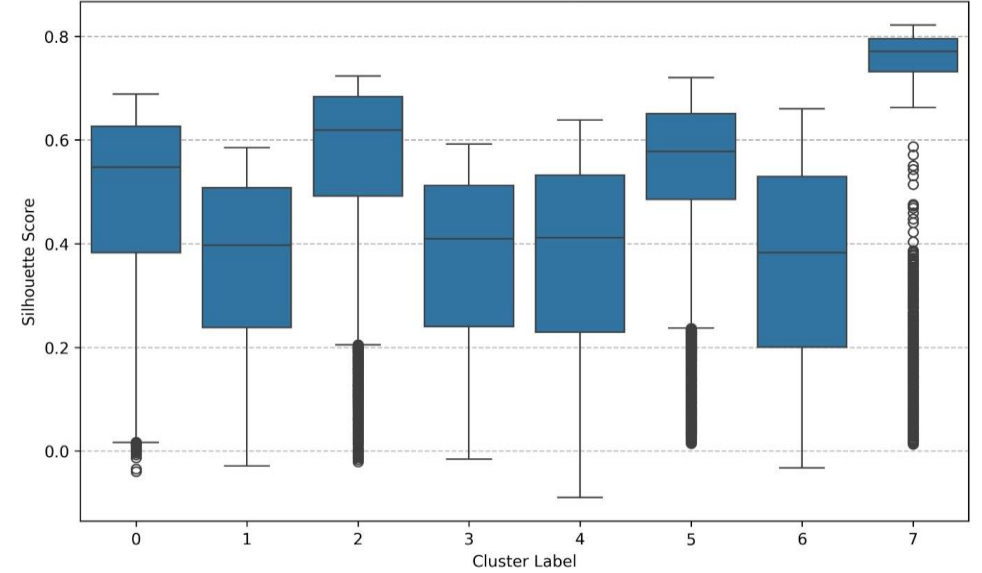
Silhouette Score Distribution per Cluster for K=5



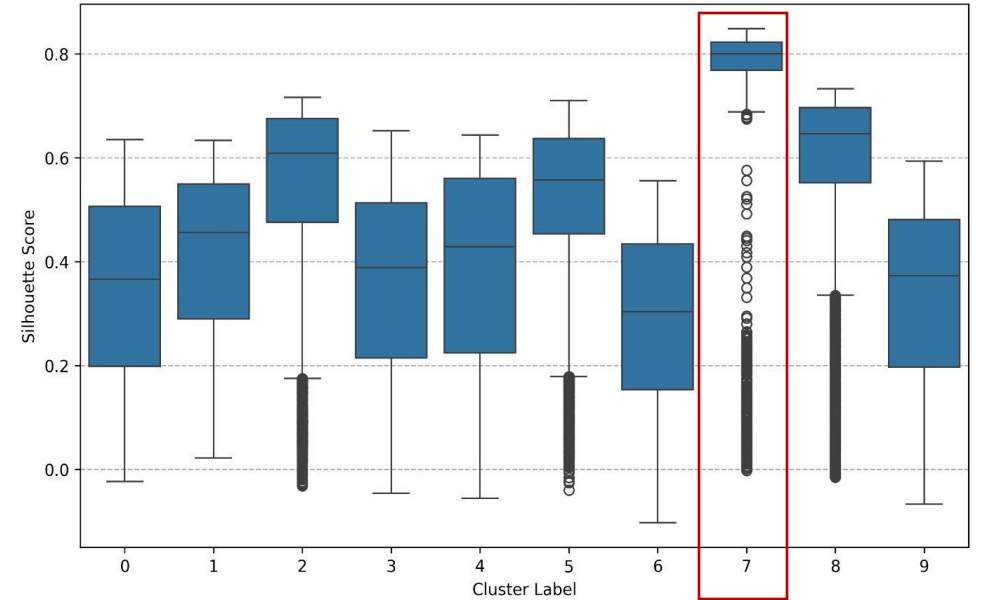
Silhouette Score Distribution per Cluster for K=7



Silhouette Score Distribution per Cluster for K=8

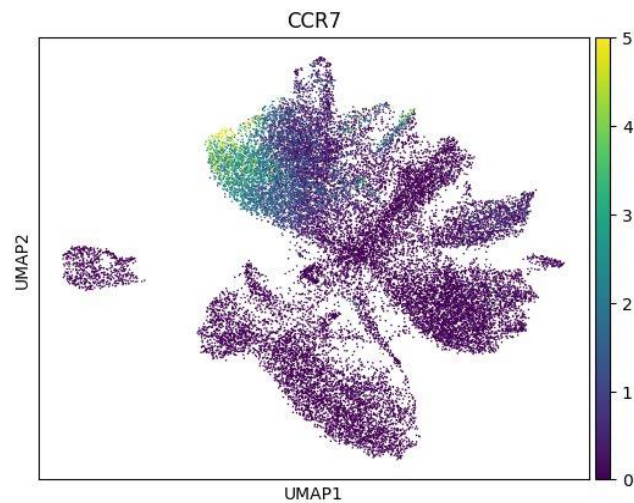
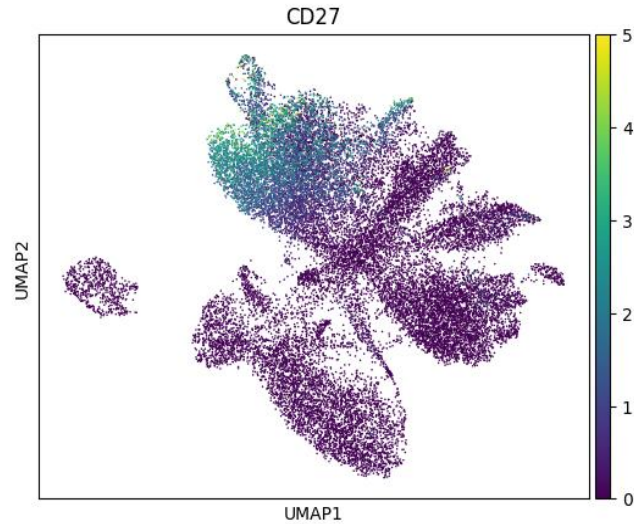


Silhouette Score Distribution per Cluster for K=10

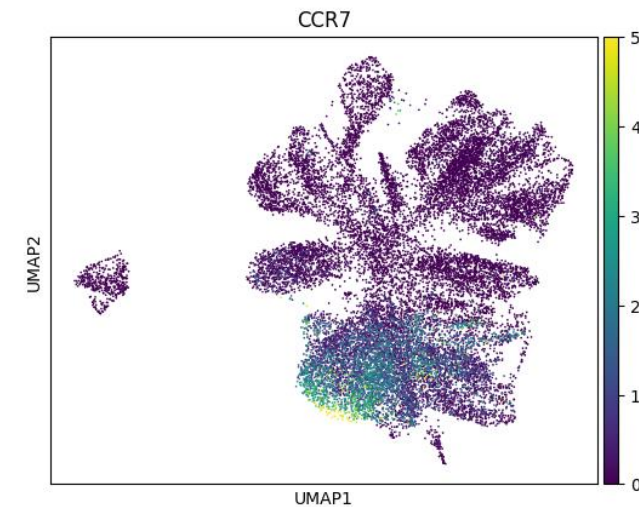
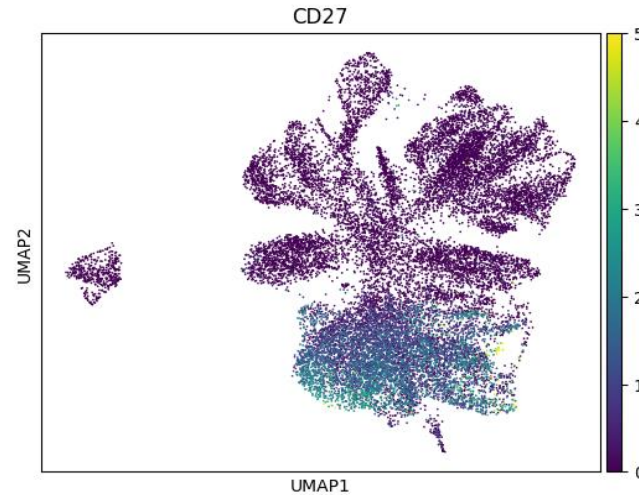


# Marker Expressions Across Treatment Stages

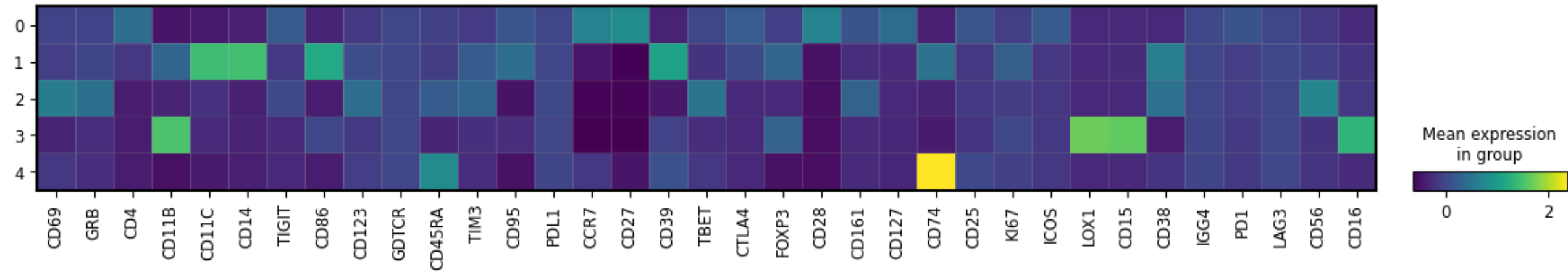
Pre Treatment



On Treatment



- **CD27:** co-stimulatory receptor on CD8<sup>+</sup> T cells, essential for their activation, clonal expansion, survival, and long-term memory formation in anti-tumor and anti-viral immunity
- **CCR7:** Chemokine receptor marking naive and central-memory CD8<sup>+</sup> T cells.



HOW CAN A  
COMPUTATIONAL  
APPROACH SUPPORT CELL  
MARKER CLUSTERING?  
HOW DATA ANALYSIS CAN  
SUPPORT BIOLOGY  
RESEARCH?

- Unbiased gating and clustering: can reveal new cell populations and relationships
- Validate biological gating
- Handling High Dimensionality and Automated Clustering

# Tools and Technologies

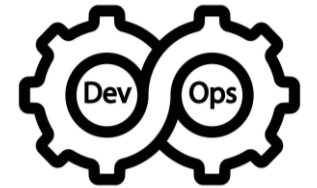
## Analysis & Parsing

- Pandas
- Anndata
- Scanpy
- Fcsparser
- pytometry



## DevOps & Environment

- Linux (Ubuntu)
- Miniconda
- Pyenv
- Git/GitHub
- Bash Scripting



## Web & Visualization

- Matplotlib
- Altair
- Flask
- SQLite3



## Next Steps

Apply mixed linear models to appropriately account for longitudinal data.

Integrate Circulating Tumor DNA (ctDNA) and mass cytometry together in analysis

Perform clustering analyses on additional cell types

Explore determinants of immune checkpoint responses and iRAEs

Run analyses in parallel across samples, donors, or batches.



# THANK YOU!

CHECK OUT MY CODE:

<https://github.com/caterinaponti/radiohead-pici-internship.git>



## CONTACT ME

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LinkedIn – <https://www.linkedin.com/in/caterina-ponti>

# SOURCES

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- *The Cancer-Immunity Cycle: Indication, Genotype, and Immunotype: Immunity*, [www.cell.com/immunity/fulltext/S1074-7613\(23\)00416-8](https://www.cell.com/immunity/fulltext/S1074-7613(23)00416-8).
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- Mellman, Ira, et al. "The cancer-immunity cycle: Indication, genotype, and immunotype." *Immunity*, vol. 56, no. 10, 2023, pp. 2188–2205, doi:10.1016/j.immuni.2023.09.011.
- Figure. Immune-related adverse events and impact on survival outcomes. Image from Zoe Quandt et al., Associations between immune checkpoint inhibitor response, immune-related adverse events, and steroid use in RADIOHEAD: a prospective pan-tumor cohort study, *Journal for Immunotherapy of Cancer*, vol. 13, no. 5, 12 May 2025, e011545, doi:10.1136/jitc-2025-011545. PubMed, <https://pubmed.ncbi.nlm.nih.gov/40355283/>
- Figure. Data center security layers, from One percent of Googlers get to visit a data center, but I did, Google Blog, 30 June 2020, [blog.google/inside-google/infrastructure/how-data-center-security-works/](https://blog.google/inside-google/infrastructure/how-data-center-security-works/)
- Figure. Mellman, Ira, et al. "The cancer-immunity cycle: Indication, genotype, and immunotype." *Immunity*, vol. 56, no. 10, 2023, pp. 2188–2205, doi:10.1016/j.immuni.2023.09.011. Figure of the cancer-immunity cycle.
- Figure: Immune checkpoint inhibitors illustrating PD-1/PD-L1 interaction and blockade (Credit: © Terese Winslow, National Cancer Institute, cancer.gov). Originally published on Cancer.gov in "Immune Checkpoint Inhibitors." <https://www.cancer.gov/about-cancer/treatment/types/immunotherapy/checkpoint-inhibitors>.