

ChimeraLM: A genomic language model for detecting whole genome amplification artifacts in single-cell sequencing

Yangyang Li¹, Qingxiang Guo^{1†}, Rendong Yang^{1,2*}

¹Department of Urology, Northwestern University Feinberg School of Medicine, 303 E Superior St, Chicago, 60611, IL, USA.

²Robert H. Lurie Comprehensive Cancer Center, Northwestern University Feinberg School of Medicine, 675 N St Clair St, Chicago, 60611, IL, USA.

*Corresponding author(s). E-mail(s): rendong.yang@northwestern.edu;

Contributing authors: yangyang.li@northwestern.edu;

qingxiang.guo@northwestern.edu;

[†]These authors contributed equally to this work.

Abstract

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1 Main

Single-cell genomics has revolutionized our understanding of cellular heterogeneity and development by enabling the characterization of individual cells rather than bulk populations [1, 2]. This approach has proven instrumental in uncovering rare cell types, tracking developmental trajectories, and identifying somatic mutations that drive disease progression. However, the limited amount of DNA present in a single cell, typically only a few picograms, poses significant technical challenges for comprehensive genomic analysis [3, 4].

To overcome this fundamental limitation, [Whole Genome Amplification \(WGA\)](#) has become an essential preprocessing step in single-cell genomic studies [5, 6]. Various

WGA techniques, including Multiple Displacement Amplification (MDA), Multiple Annealing and Looping-based Amplification Cycles (MALBAC), and other emerging methods, can amplify the entire genome from a single cell by several orders of magnitude, generating sufficient DNA material for high-coverage sequencing [7–9]. This amplification enables researchers to achieve the depth and breadth of coverage necessary for reliable variant calling, copy number analysis, and structural variation detection.

Despite its critical role in single-cell genomics, WGA introduces systematic artifacts that can significantly impact downstream analyses [10, 11]. Among the most problematic are chimeric sequences—artificial DNA constructs formed when DNA fragments from different genomic loci are erroneously joined during the amplification process [10, 11]. These chimeric artifacts can manifest as false-positive structural variations that do not exist in the original cell [10]. The presence of such artifacts poses a substantial challenge for accurate Structural Variation (SV) detection, potentially leading to misinterpretation of genomic rearrangements and their biological significance.

Current computational approaches for identifying WGA-induced artifacts rely primarily on coverage-based metrics and read-pair orientation patterns [11, 12]. However, these methods often fail to distinguish between genuine structural variations and amplification artifacts, particularly when chimeric sequences exhibit complex rearrangement patterns or occur in repetitive genomic regions [13, 14]. The lack of robust artifact detection methods has limited the reliability of structural variant analysis in single-cell studies and hindered the full realization of single-cell genomics’ potential.

To address these challenges, we developed ChimeraLM, a genomic language model specifically designed to detect chimeric artifacts introduced by whole genome amplification. By leveraging deep learning approaches to capture sequence patterns and contextual information in genomic reads [15–17], ChimeraLM can effectively distinguish between genuine biological sequences and WGA-induced chimeric artifacts. This approach represents a significant advancement in single-cell genomic analysis, offering improved accuracy in artifact detection and enabling more reliable structural variant analysis in single-cell studies. This methodology represents a significant advancement in single-cell genomic analysis, offering a principled approach to improve the reliability of structural variant detection and enable more precise characterization of genomic alterations in individual cells.

In this study, we present ChimeraLM, demonstrate its superior performance compared to existing methods, and illustrate its practical applications in genomic studies.

2 Results

Topical subheadings are allowed. Authors must ensure that their Methods section includes adequate experimental and characterization data necessary for others in the field to reproduce their work. Authors are encouraged to include RIIDs where appropriate.

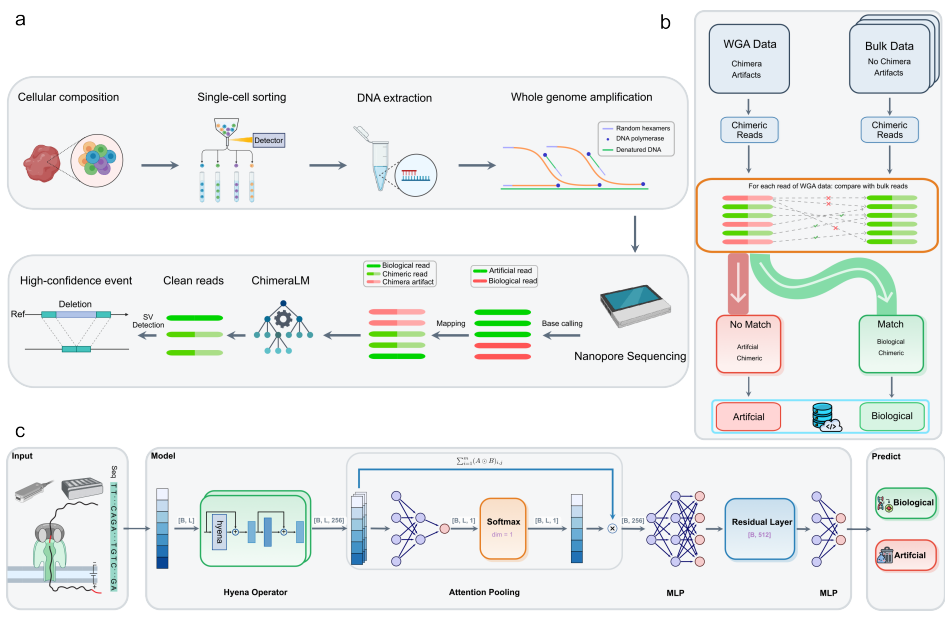


Fig. 1 Problem and Model

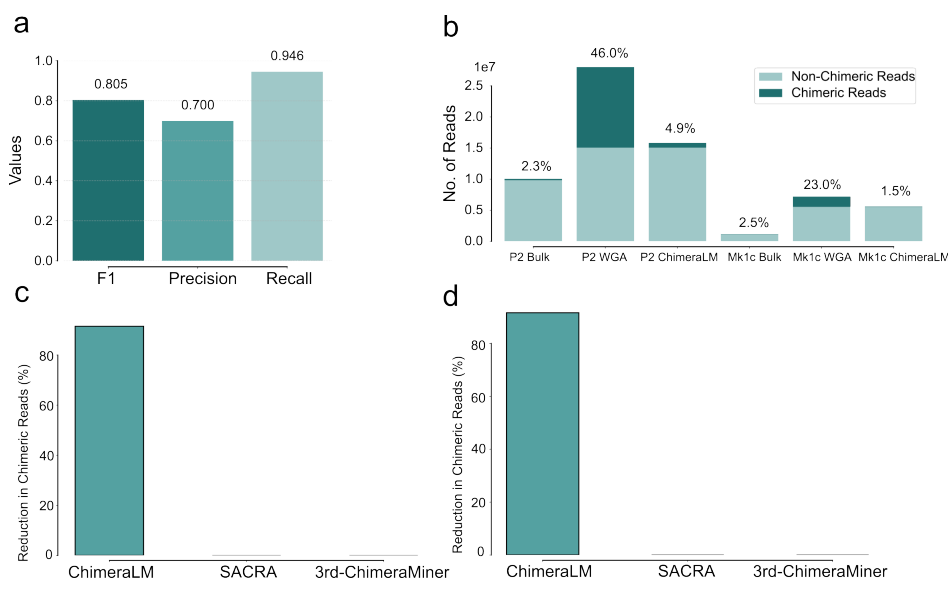


Fig. 2 Problem and Model

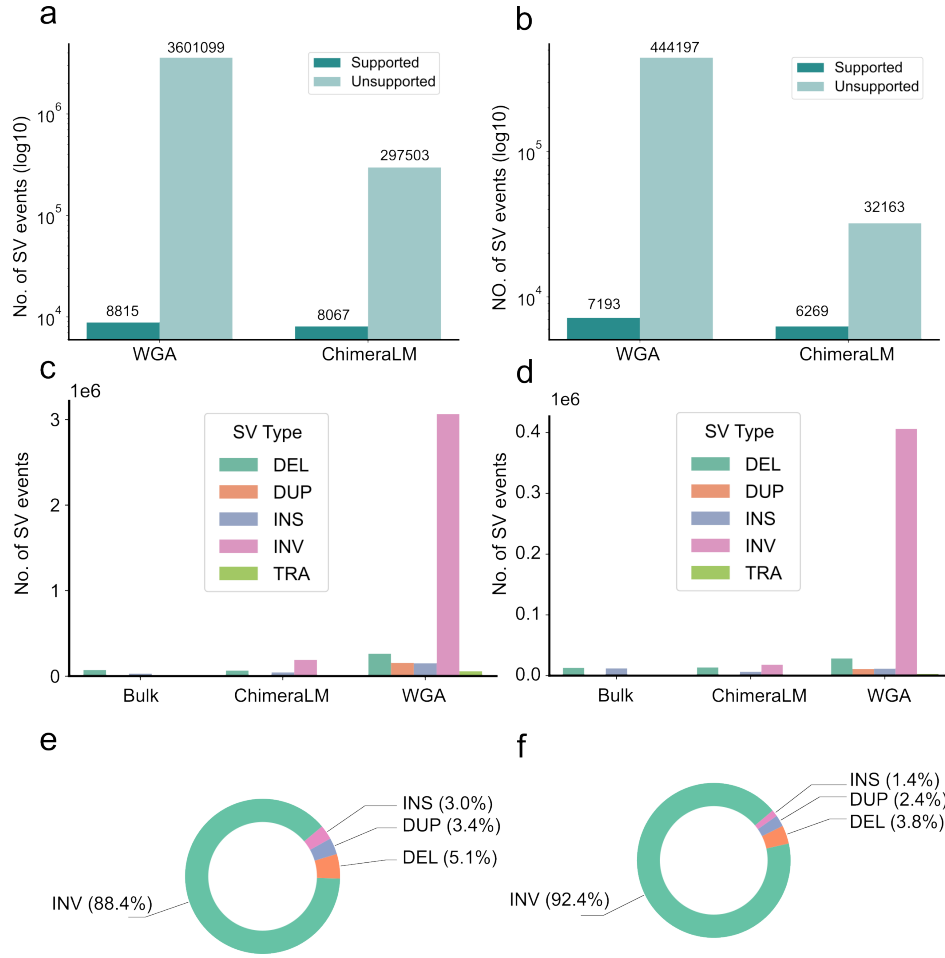


Fig. 3 Problem and Model

3 Methods

Supplementary information. This separation aligns with how many transcript assembly algorithms work:

1. First, chains of exons and splice junctions are identified from the data
2. Then, potential transcripts are derived by traversing the graph in different ways
3. Finally, relationships between different transcript graphs are established

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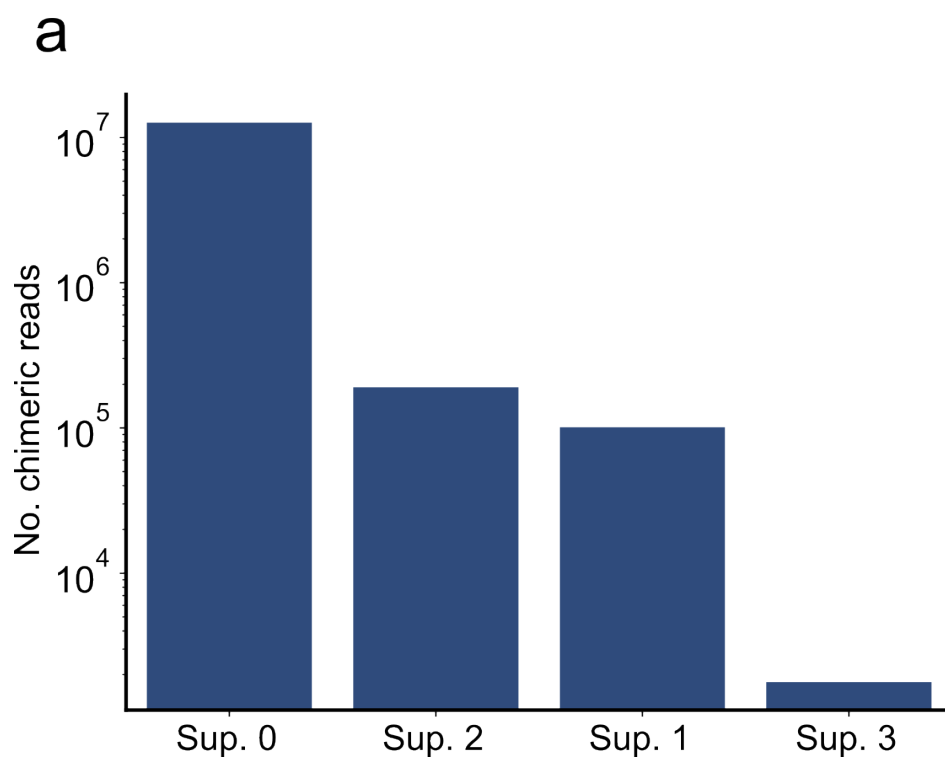


Fig. 4 Problem and Model

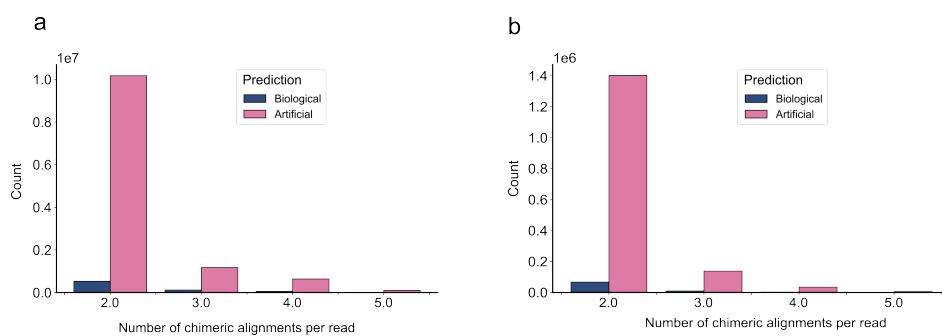


Fig. 5 Problem and Model

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Acronyms

MALBAC Multiple Annealing and Looping-based Amplification Cycles [2](#)

MDA Multiple Displacement Amplification [2](#)

SV Structural Variation [2](#)

WGA Whole Genome Amplification [1](#), [2](#)

Appendix A Section title of first appendix

An appendix contains supplementary information that is not an essential part of the text itself but which may be helpful in providing a more comprehensive understanding of the research problem or it is information that is too cumbersome to be included in the body of the paper.

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