

Gen 711 Final Project – Bacterial Genome Assembly

By, Caylin Grove, Ethan Morgan, and Graham Collinsworth

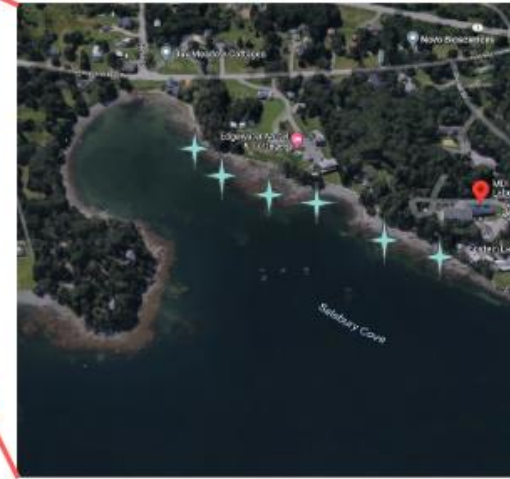
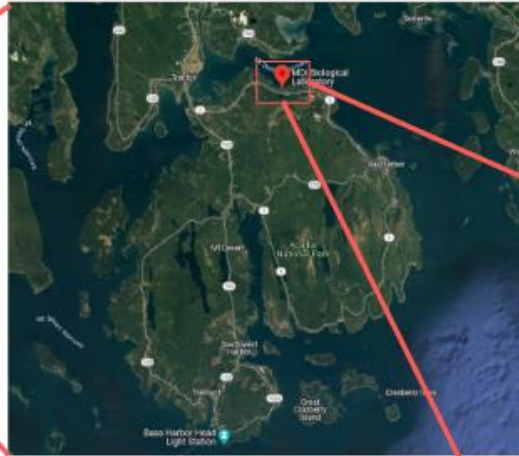
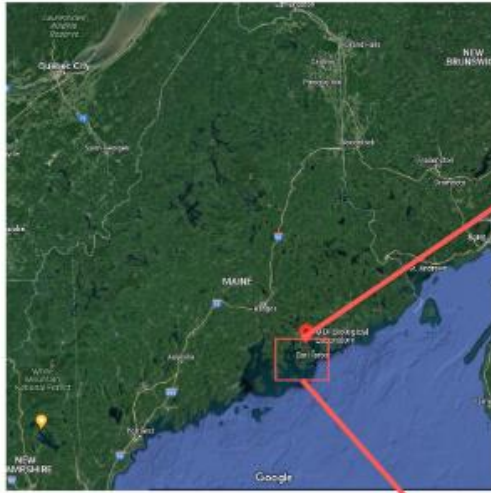




Background Information

- Data was collected by Anthony Hay, Steven Weicksel, Dana-Lynn Koomoa-Lange, Leah Elliot, Melissa Chisholm, Princess Rodriguez
- Pathway was created by Joseph Sevigny on Github
- Seaweed eating microbes were collected in MDIBL, Acadia National Park
- Their DNA was extracted and Illumina libraries were generated
- Illumina sequencing of the samples occurred

Sampling Location: MDIBL, Acadia National Park



Some General Information

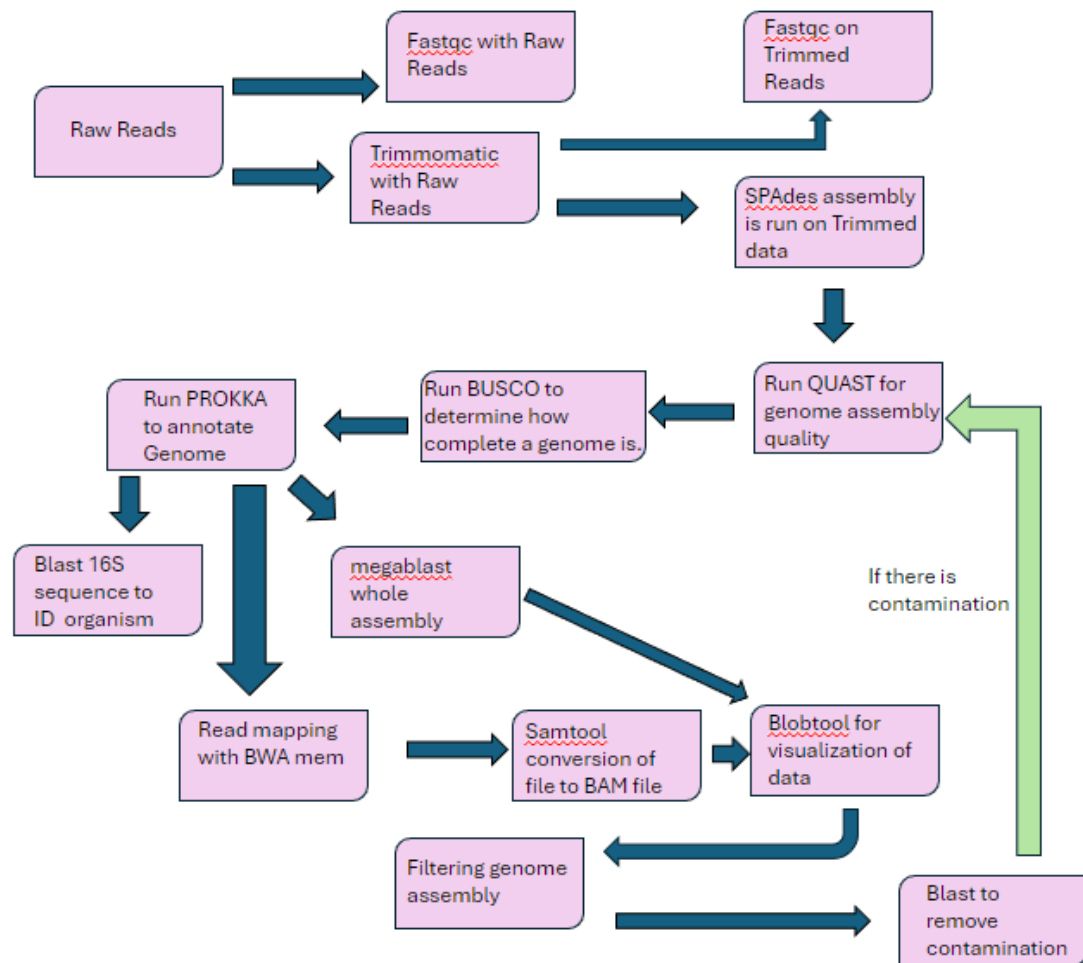
- We each ran one set of samples

Ethan Morgan: 69_S8_L001_R*_001.fastq.gz

Graham Collinsworth: DC1_S41_L001_R*_001.fastq.gz

Caylin Grove: 2_S26_L001_R*_001.fastq.gz

- We worked through each step together so our method was combined into one .sh file

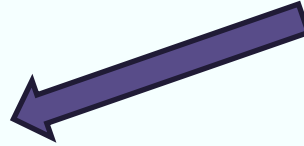


Methods Overview

Fastqc: Summarizes Read Quality and Base Pair composition



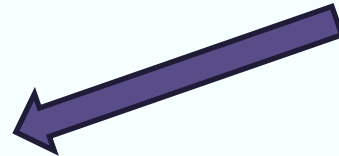
Trimmomatic: Removes Adapters and “Trims” off Low quality BP scores



SPAdes: Takes Trimmed Fastqc reads and assembles a genome



QUAST: Determines How well a genome was assembled



BUSCO: Finds portions of Single copy orthologs to determine how complete a genome is.



Methods Overview

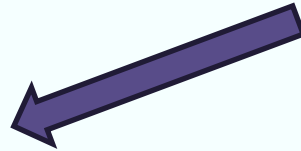


PROKKA: “Rapid prokaryote
genome annotation
pipeline”

BWA MEM Read Mapping:
Aligns reads to a reference
sequence and produces a
sequence alignment map
“SAM” file



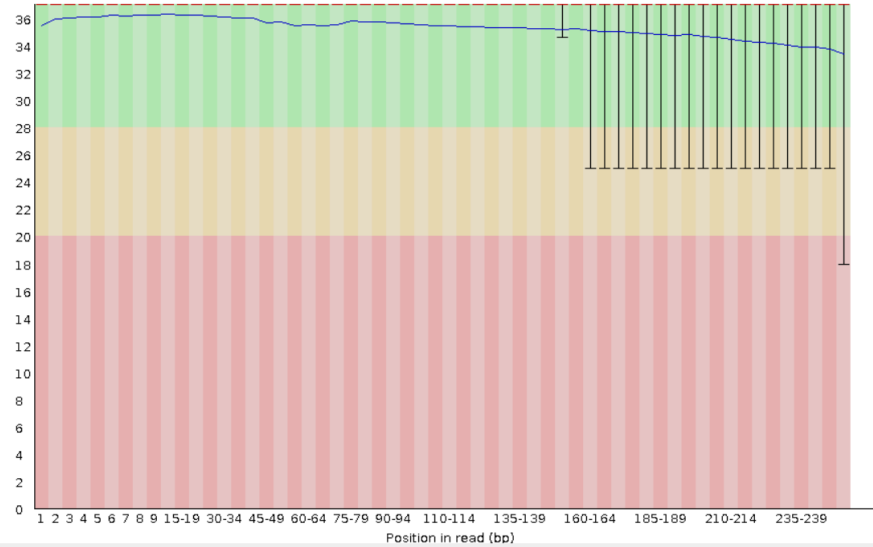
BLAST: Uses 16S sequence in
order to relay potential
taxonomic identities



Blobtools: visualizes genome
assembly from SAM and
BAM files.

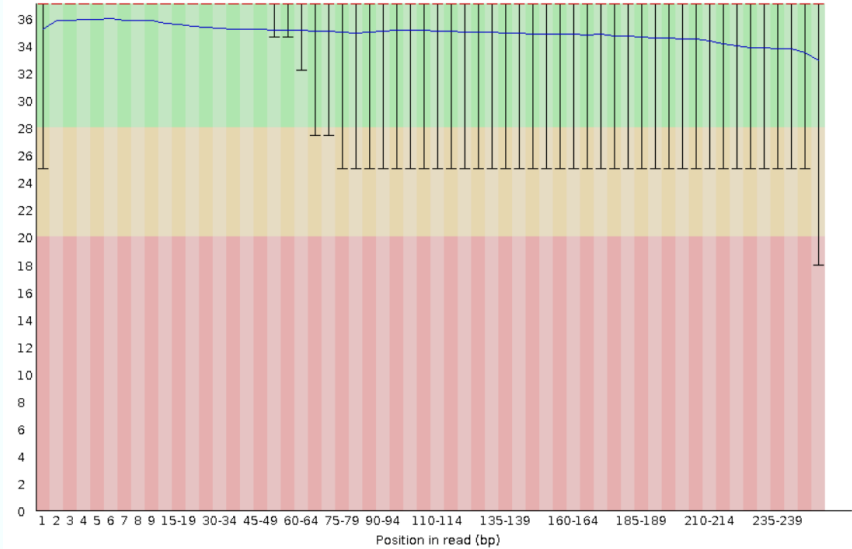
Fastqc run on samples -Caylin

Quality scores across all bases (Sanger / Illumina 1.9 encoding)



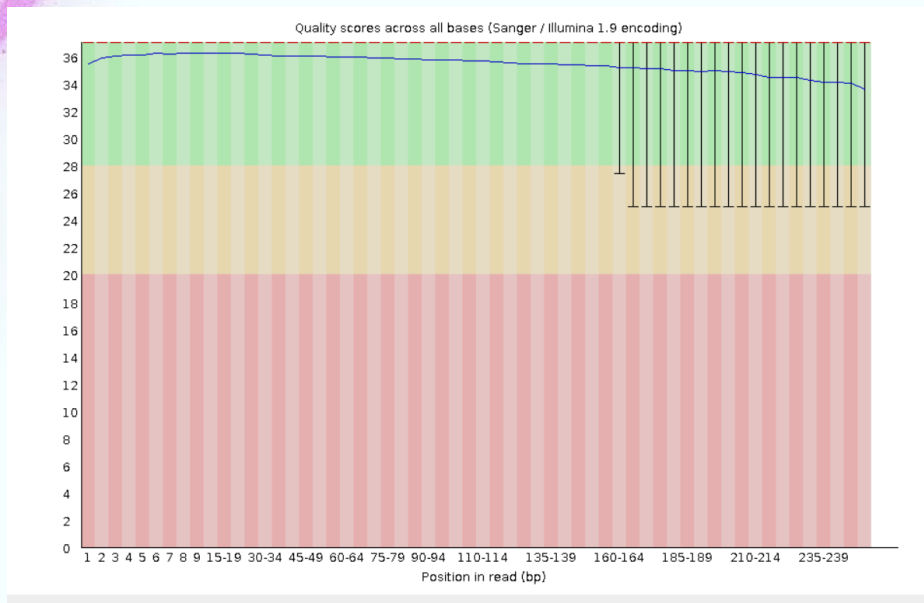
R1

Quality scores across all bases (Sanger / Illumina 1.9 encoding)

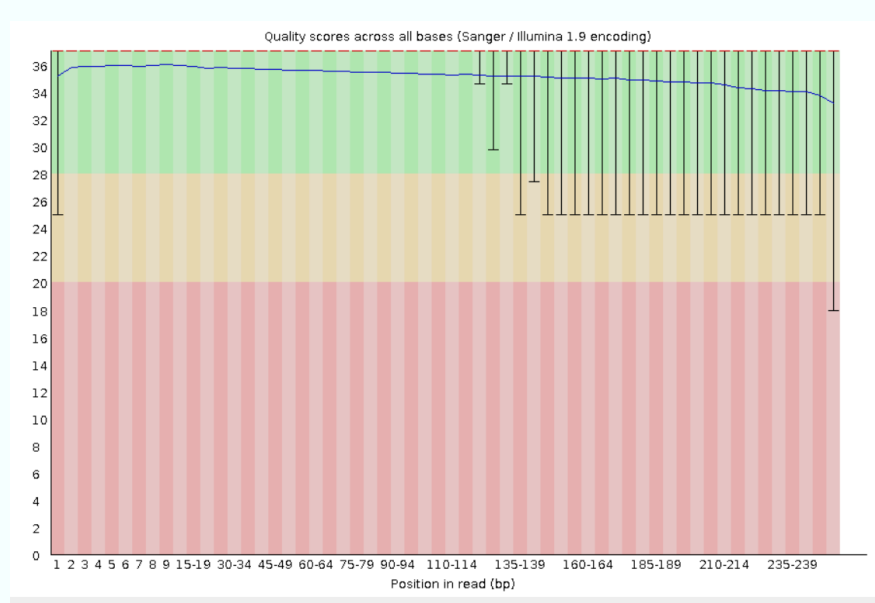


R2

Trimmomatic - Caylin



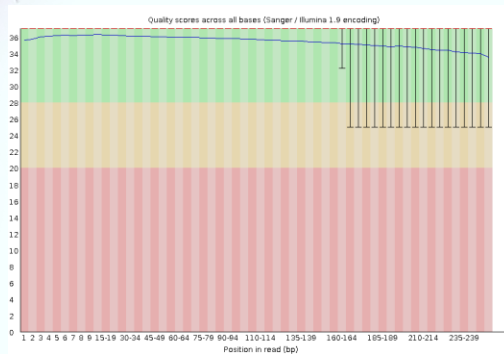
R1



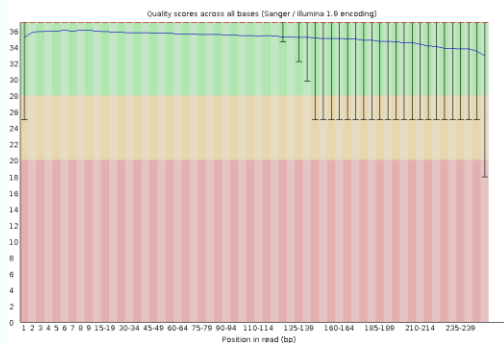
R2

Fastqc run on samples -Ethan

Raw Data

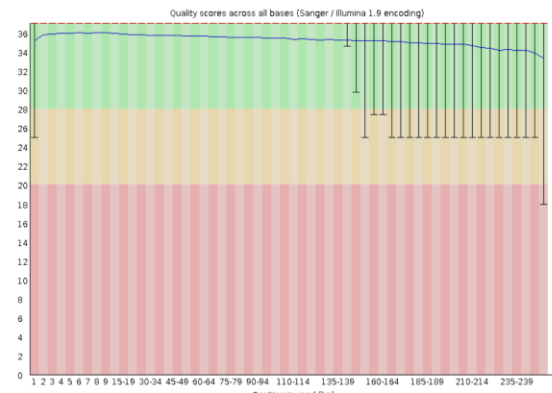
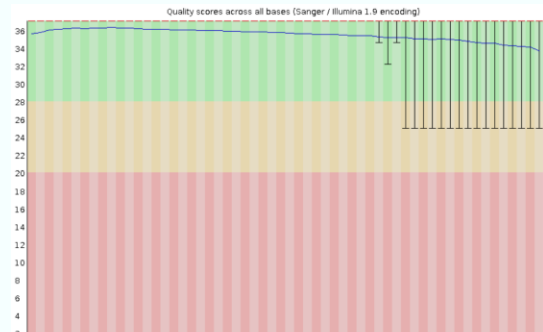


R1



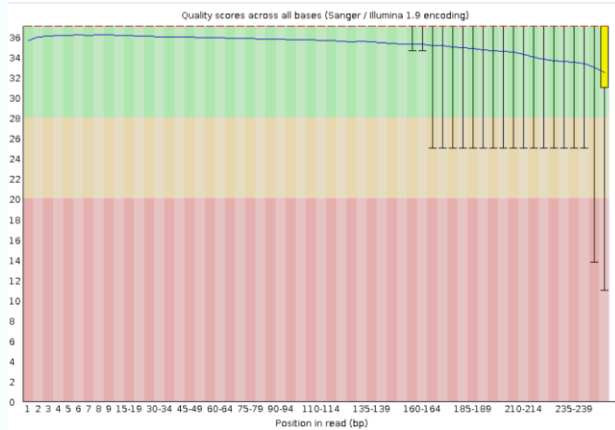
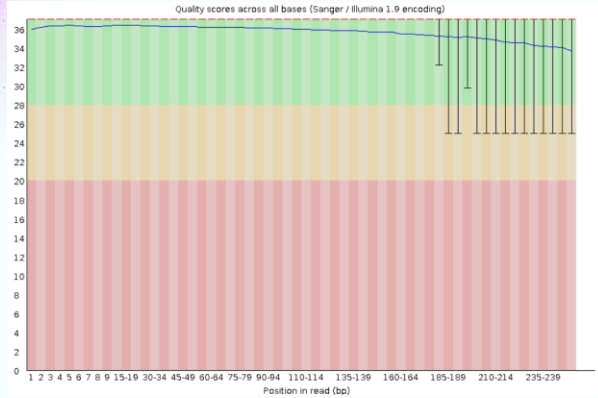
R2

Trimmomatic Data

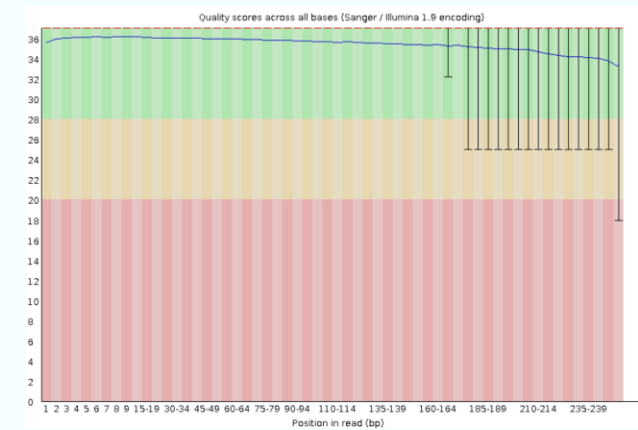
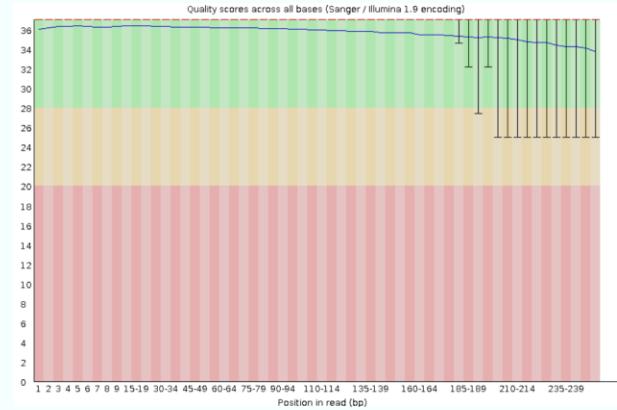


Fastqc - Graham

Raw Data



Trimmomatic Data



Quast – Caylin

```
cmg1164@ron: ~  
GNU nano 4.8 report.txt  
All statistics are based on contigs of size >= 500 bp, u  
  
Assembly contigs  
# contigs (>= 0 bp) 2399  
# contigs (>= 1000 bp) 233  
# contigs (>= 5000 bp) 172  
# contigs (>= 10000 bp) 149  
# contigs (>= 25000 bp) 114  
# contigs (>= 50000 bp) 63  
Total length (>= 0 bp) 9850381  
Total length (>= 1000 bp) 8854820  
Total length (>= 5000 bp) 8715164  
Total length (>= 10000 bp) 8563183  
Total length (>= 25000 bp) 8006791  
Total length (>= 50000 bp) 6113044  
# contigs 649  
Largest contig 204992  
Total length 9114677  
GC (%) 72.34  
N50 75477  
N75 40575  
L50 37  
L75 80  
# N's per 100 kbp 0.00
```

Before filtering

```
GNU nano 4.8 report.txt  
All statistics are based on contigs of size >= 500 bp, u  
  
Assembly Streptomyces_A1277_filtered  
# contigs (>= 0 bp) 250  
# contigs (>= 1000 bp) 221  
# contigs (>= 5000 bp) 170  
# contigs (>= 10000 bp) 147  
# contigs (>= 25000 bp) 112  
# contigs (>= 50000 bp) 61  
Total length (>= 0 bp) 8590294  
Total length (>= 1000 bp) 8570309  
Total length (>= 5000 bp) 8444055  
Total length (>= 10000 bp) 8292074  
Total length (>= 25000 bp) 7735682  
Total length (>= 50000 bp) 5841935  
# contigs 250  
Largest contig 204992  
Total length 8590294  
GC (%) 72.41  
N50 75477  
N75 41882  
L50 35  
L75 75  
# N's per 100 kbp 0.00
```

After filtering

QUAST Results: Ethan

N50: 36704

Total Length: 7,914,737

contigs: 373

```
Assembly          contigs
# contigs (>= 0 bp)      509
# contigs (>= 1000 bp)   353
# contigs (>= 5000 bp)   265
# contigs (>= 10000 bp)  212
# contigs (>= 25000 bp)  106
# contigs (>= 50000 bp)  36
Total length (>= 0 bp)   7914737
Total length (>= 1000 bp) 7852311
Total length (>= 5000 bp) 7612487
Total length (>= 10000 bp) 7228826
Total length (>= 25000 bp) 5404723
Total length (>= 50000 bp) 2990284
# contigs              373
Largest contig         176229
Total length           7865799
GC (%)                 72.13
N50                     36704
N75                     22132
L50                      59
L75                      128
# N's per 100 kbp      0.00
```

After Filtering:


N50: 36636

Total Length: 7,758,289



contigs: 366

```
Assembly          Streptomyces_A1277_filtered
# contigs (>= 0 bp)      366
# contigs (>= 1000 bp)   349
# contigs (>= 5000 bp)   262
# contigs (>= 10000 bp)  209
# contigs (>= 25000 bp)  105
# contigs (>= 50000 bp)  35
Total length (>= 0 bp)   7758289
Total length (>= 1000 bp) 7746526
Total length (>= 5000 bp) 7510519
Total length (>= 10000 bp) 7126858
Total length (>= 25000 bp) 5333177
Total length (>= 50000 bp) 2918738
# contigs              366
Largest contig         176229
Total length           7758289
GC (%)                 72.13
N50                     36636
N75                     22296
L50                      59
L75                      126
# N's per 100 kbp      0.00
```

Quast – Graham



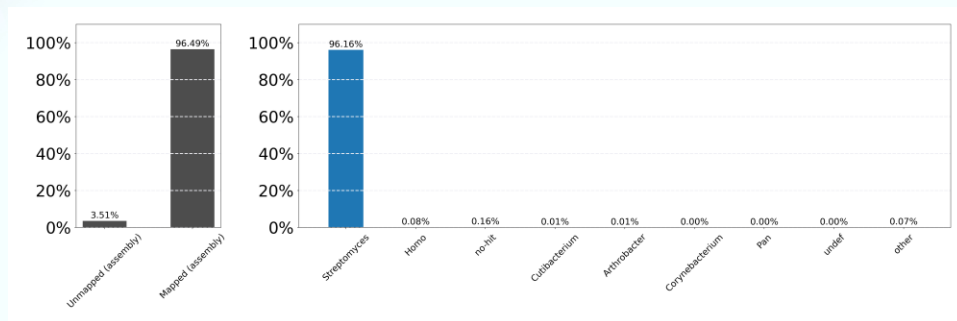
Assembly	contigs
# contigs (≥ 0 bp)	2619
# contigs (≥ 1000 bp)	1953
# contigs (≥ 5000 bp)	1400
# contigs (≥ 10000 bp)	1110
# contigs (≥ 25000 bp)	633
# contigs (≥ 50000 bp)	278
Total length (≥ 0 bp)	48882566
Total length (≥ 1000 bp)	48572897
Total length (≥ 5000 bp)	47137826
Total length (≥ 10000 bp)	45064031
Total length (≥ 25000 bp)	37091509
Total length (≥ 50000 bp)	24476204
# contigs	2162
Largest contig	490780
Total length	48712931
GC (%)	53.70
N50	50144
N75	26414
L50	276
L75	612
# N's per 100 kbp	0.00



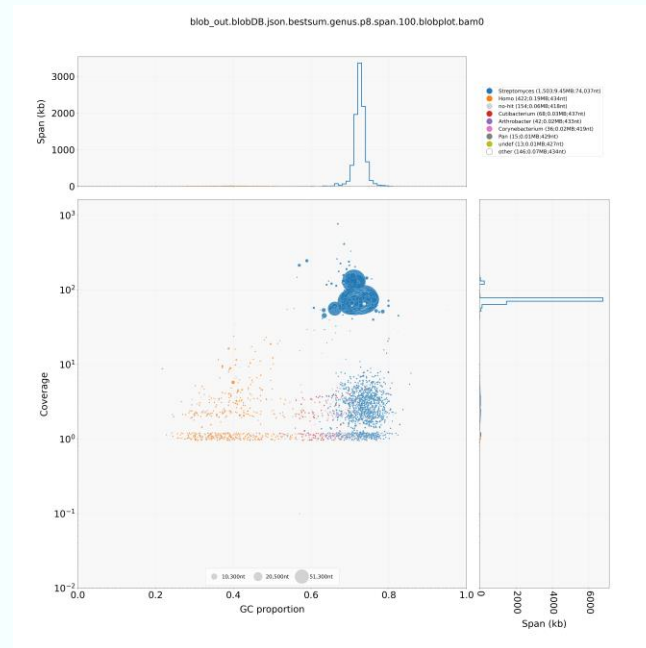
After Filtering

Assembly	Final_filtered
# contigs (≥ 0 bp)	187
# contigs (≥ 1000 bp)	157
# contigs (≥ 5000 bp)	115
# contigs (≥ 10000 bp)	102
# contigs (≥ 25000 bp)	79
# contigs (≥ 50000 bp)	55
Total length (≥ 0 bp)	8717754
Total length (≥ 1000 bp)	8697586
Total length (≥ 5000 bp)	8611512
Total length (≥ 10000 bp)	8517060
Total length (≥ 25000 bp)	8135470
Total length (≥ 50000 bp)	7281407
# contigs	187
Largest contig	490780
Total length	8717754
GC (%)	62.52
N50	140893
N75	61137
L50	20
L75	42
# N's per 100 kbp	0.00

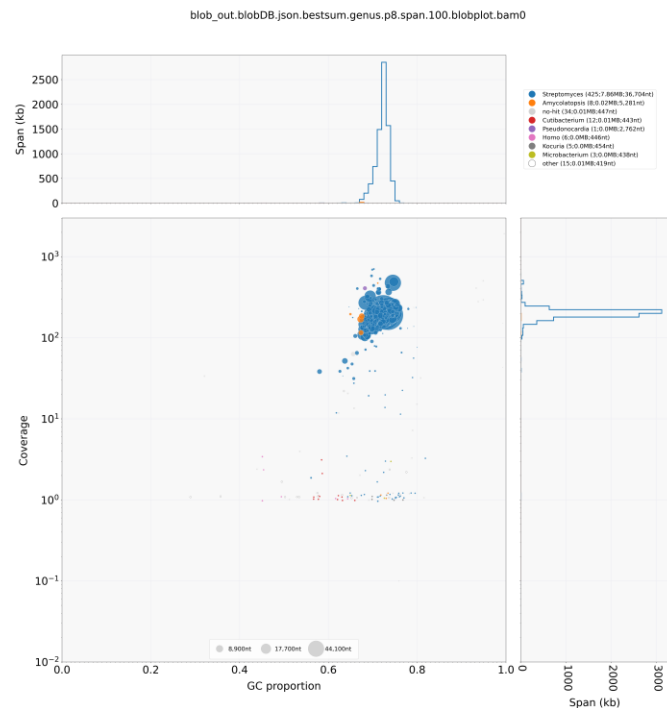
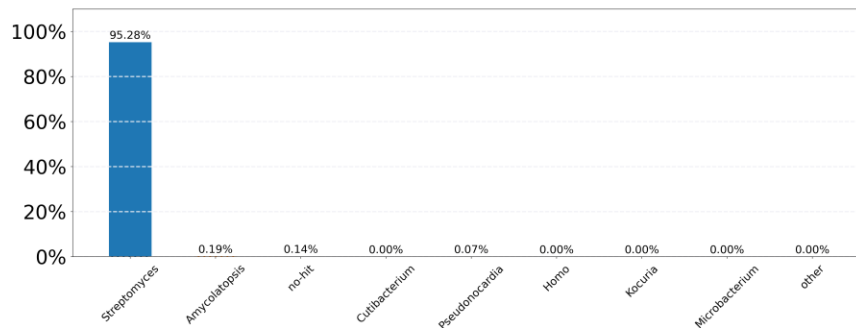
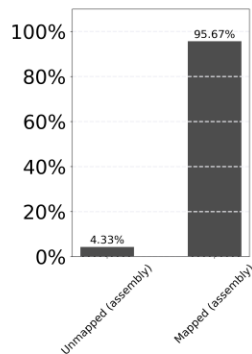
Blobtool (visualize genome assembly) - Caylin



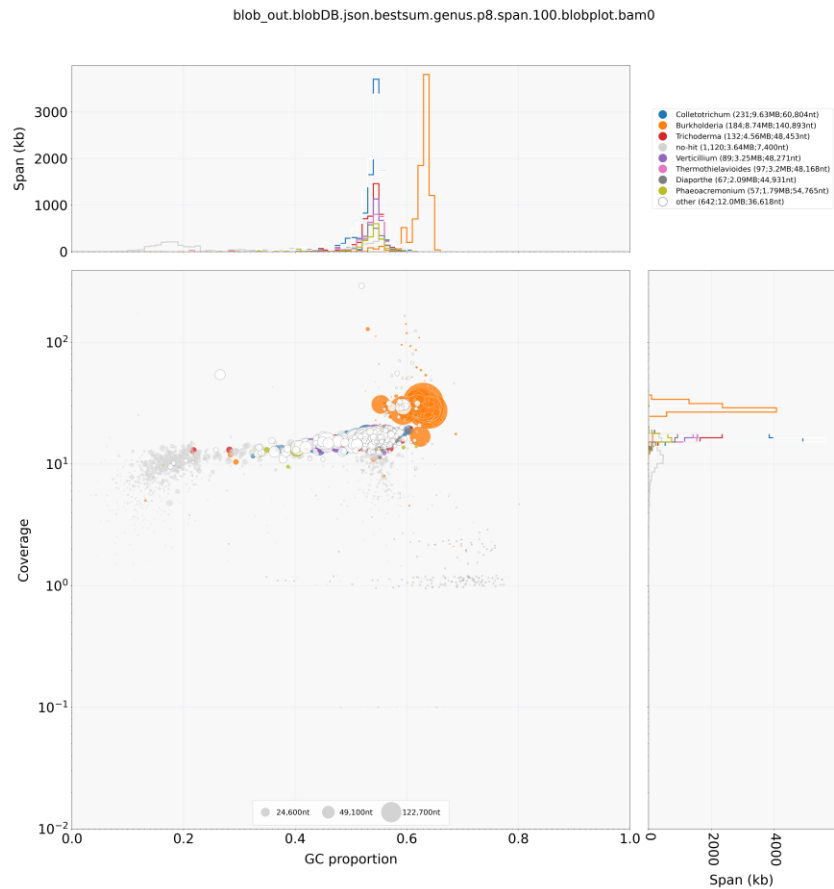
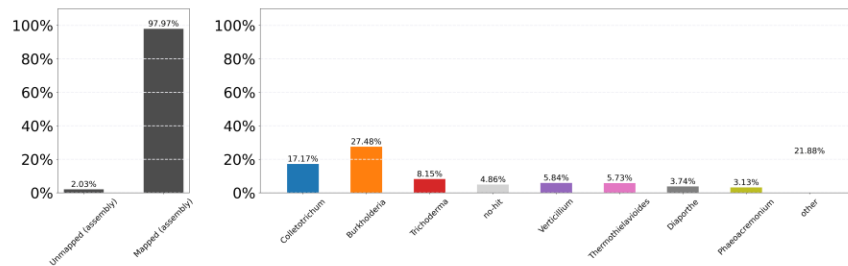
Before filtering



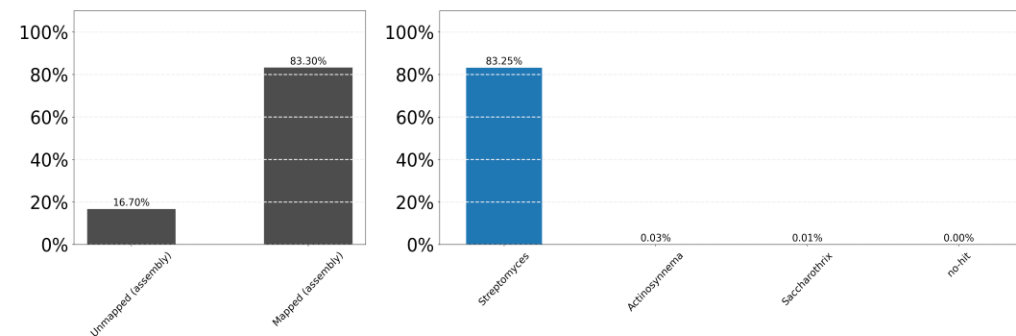
Blobtools - Ethan



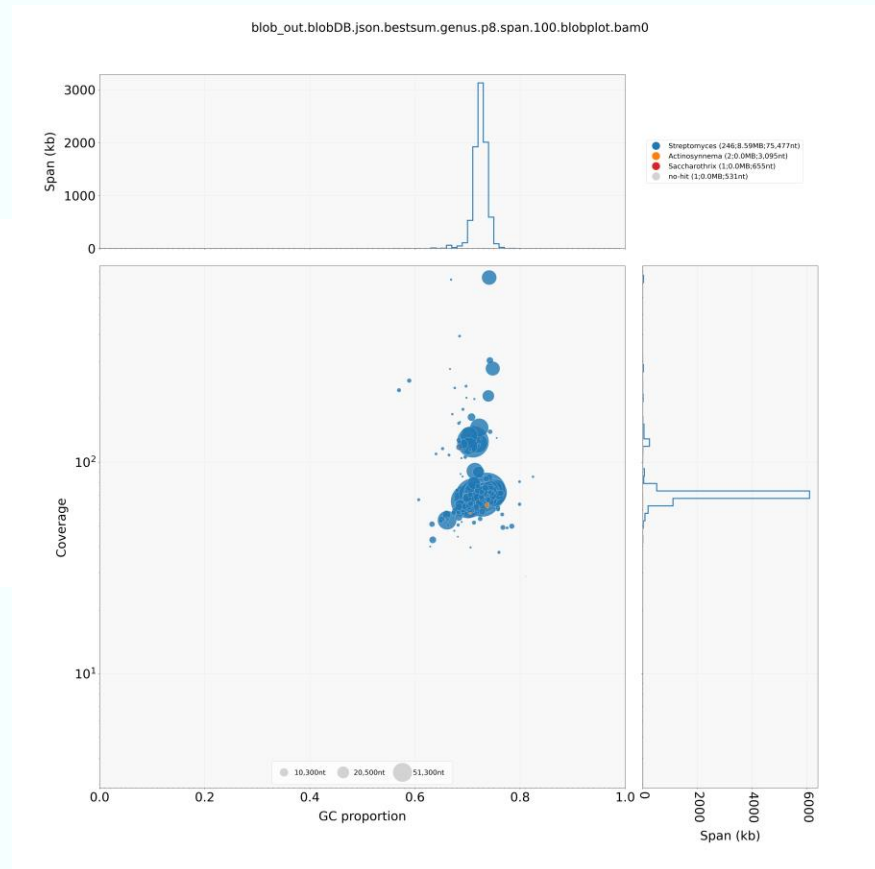
Blobtools – Graham



Blobtools ReRun with Filtered Data -Caylin



After filtering



Identification: Streptomyces

The figure displays two bar charts and a bubble plot related to the identification of *Streptomyces*.

Assembly Mapping:

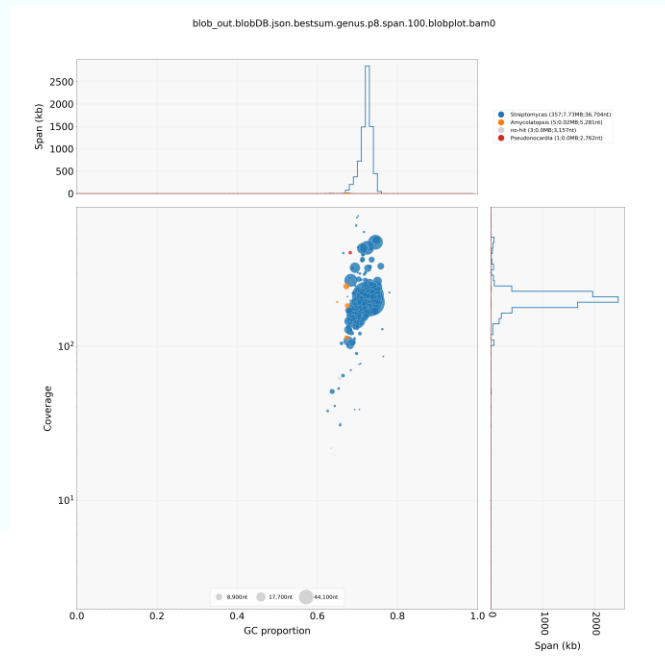
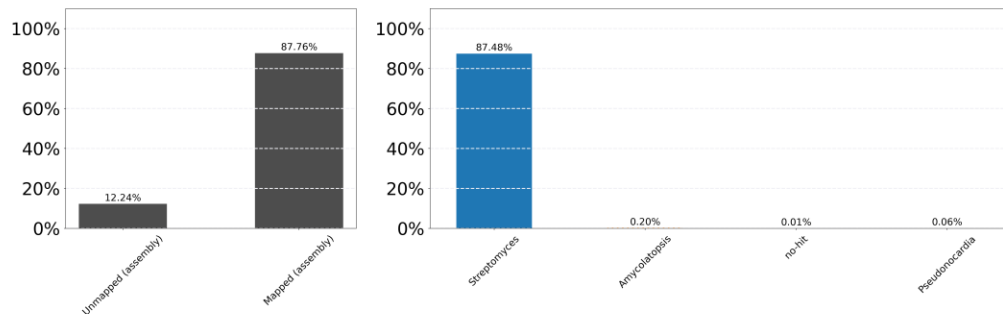
Category	Percentage
Unmapped (assembly)	12.24%
Mapped (assembly)	87.76%

Taxonomic Identification:

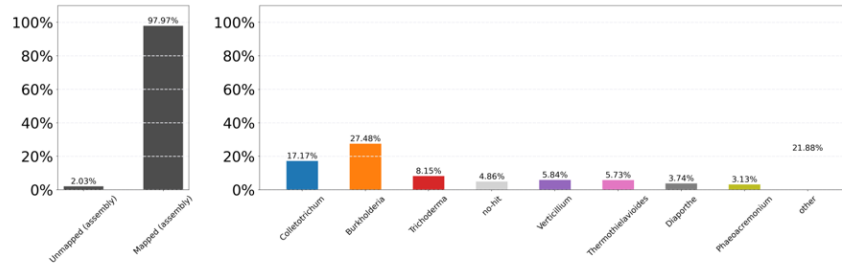
Species	Percentage
<i>Streptomyces</i>	87.48%
<i>Amycolatopsis</i>	0.20%
no-hit	0.01%
<i>Pseudonocardia</i>	0.06%

Bubble Plot:

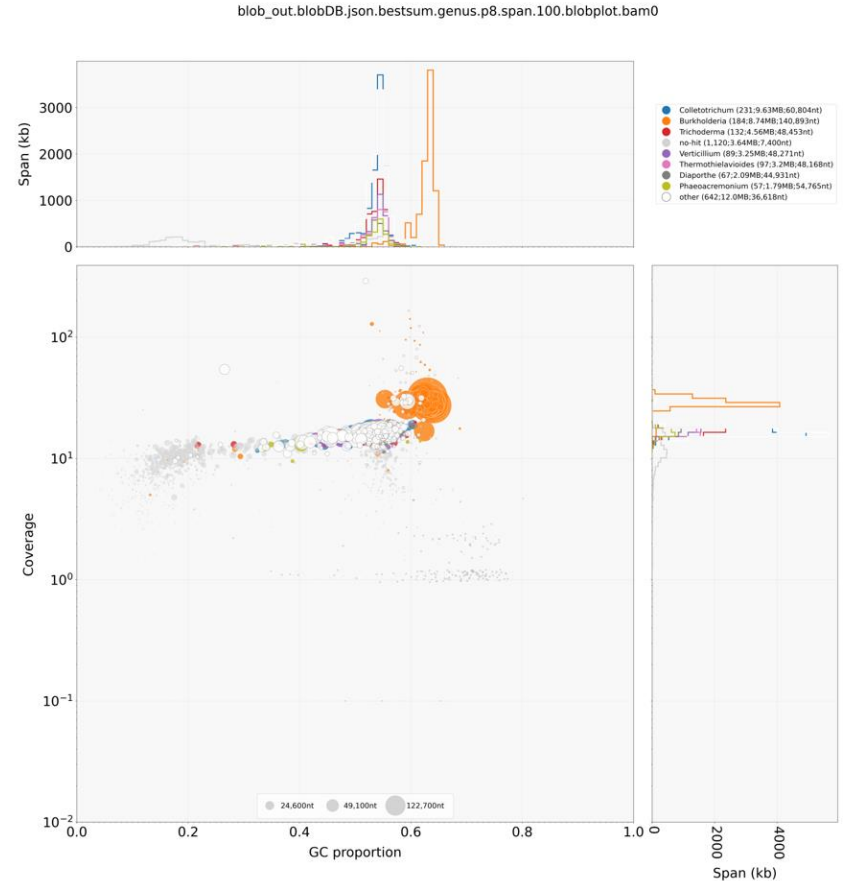
The bubble plot shows Coverage (Y-axis, log scale from 10^1 to 10^2) versus GC proportion (X-axis, 0.0 to 1.0) and Span (kb) (X-axis, 0 to 2000). The legend indicates three size ranges: 6,300bp (small grey circle), 17,700bp (medium grey circle), and 44,100bp (large grey circle). The plot shows a dense cluster of blue bubbles around a GC proportion of 0.6 and a coverage of 10^1 to 10^2 . A red line indicates the coverage threshold at approximately 500.



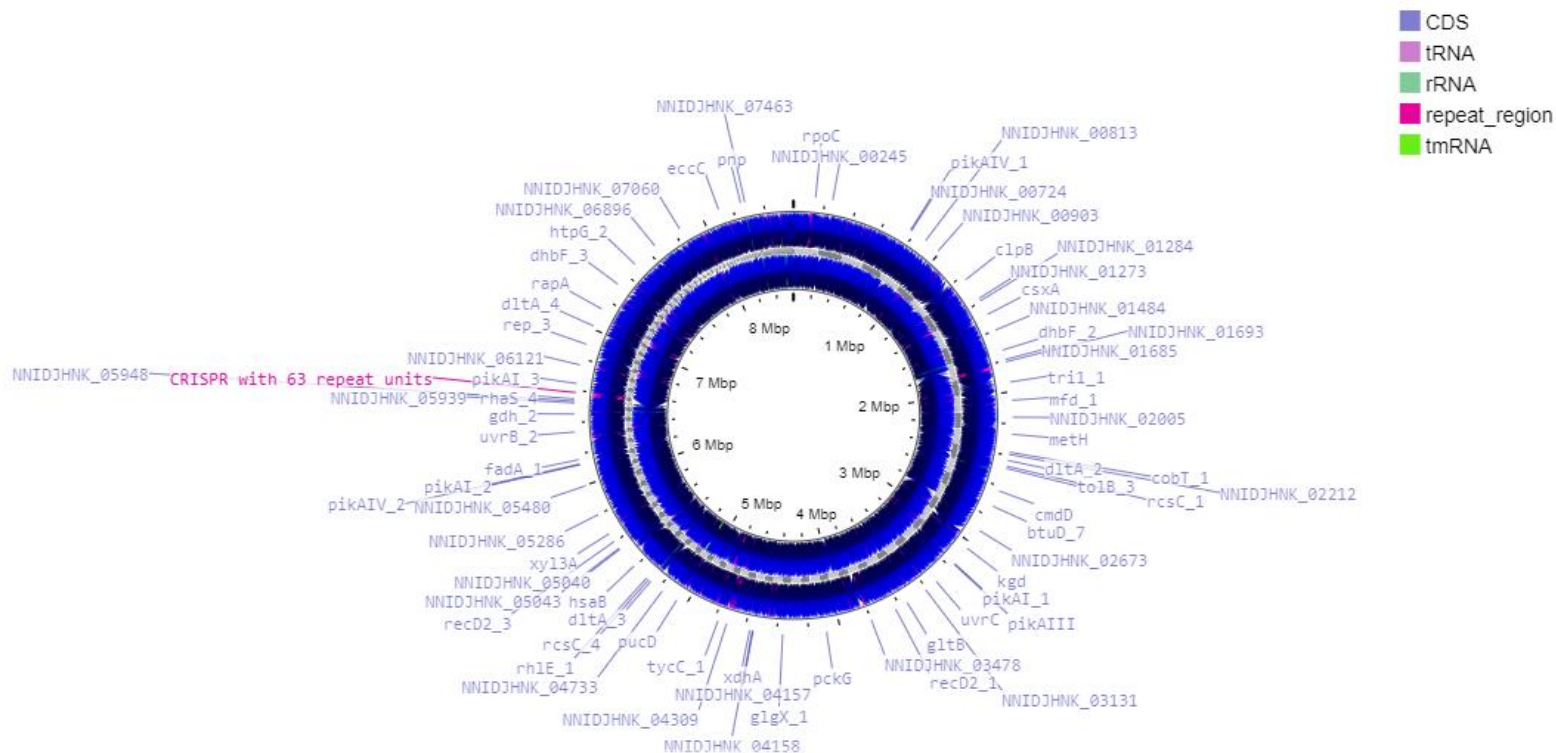
Blobtools ReRun with Filtered Data -Graham



Burkholderia Species

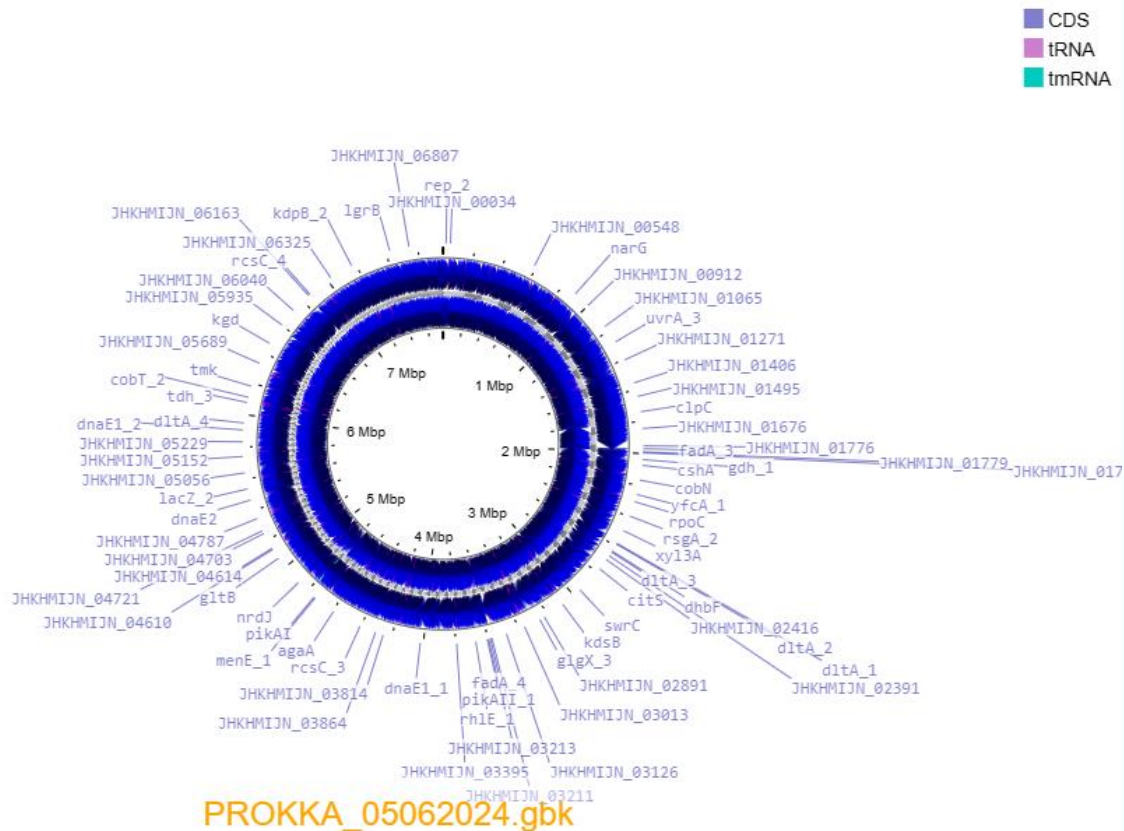


Genome Visualization - Caylin



PROKKA_05032024.gbk

Genome Visualization – Ethan








●



Concluding Remarks



- Our experience showed us that coding takes time and patience
 - Some Conda environments did not contain all necessary commands and we needed to download new condas
 - BWA mem failed multiple times due to files not being in the proper format we had to find the correct format to run BWA mem on
- 
- 
- 

Bibliography

Grant JR, Enns E, Marinier E, Mandal A, Herman EK, Chen C, Graham M, Van Domselaar G, and Stothard P
Proksee: in-depth characterization and visualization of

bacterial genomes

Nucleic Acids Research, 2023, gkad326,
<https://doi.org/10.1093/nar/gkad326>

Sevigny, Joseph. (2024). MDIBL-T3-WGS-Tutorial. <https://github.com/Joseph7e/MDIBL-T3-WGS-Tutorial>

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THANKS

DO YOU HAVE ANY QUESTIONS?

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