

# Biological Data Formats

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## 1 Sources of data

The major sources of data are sequences databases:

1. NCBI: <http://www.ncbi.nlm.nih.gov>
2. EBI: <http://www.ebi.ac.uk/>
3. ENSEMBL: <http://www.ensembl.org/index.html>
4. UCSC Genome Browser: <https://genome.ucsc.edu/>

## 2 Human Reference Genome

1. Genome Reference Consortium. <http://www.ncbi.nlm.nih.gov/projects/genome/assembly/grc/>
2. Gencode (ENSEMBL). <http://www.encodegenes.org/>

## 3 FASTA format

The most common file format for sequence files.

```
>gi|5524211|gb|AAD44166.1| cytochrome b [Elephas maximus maximus]
LCLYTHIGRNIYYGSYLYSETWNTGIMLLITMATAFMGYVLPWGQMSFWGATVITNLFSAIPYIGTNLV
EWIWGGFSVDKATLNRFFAFHFILPFTMVALAGVHLTFLHETGSNNPLGLTSDSDKIPFHPYYTIKDFLG
LLILILLILLILLALLSPDMLGDPDNHMPADPLNTPLHIKPEWYFLFAYAILRSVPNKLGGVLALFLSIVIL
GLMPFLHTSKHRSMMLRPLSQALFWTLTMDLLTLTWIGSQPVEYPYTIIGQMASILYFSIILAFLPIAGX
IENY
```

## 4 GenBank format

Sample GenBank record: <http://www.ncbi.nlm.nih.gov/Sitemap/samplerecord.html>

## 5 Annotation

Annotation is a way to provide extra information over the raw sequence. Some sequence file formats by design have the annotation built into the formation. An example is GenBank. Most commonly, annotation comes in separate files. The files generally of two types:

### 1. GTF

```
381 Twinscan    CDS 380 401 .   +   0   gene_id "001"; transcript_id "001.1";
381 Twinscan    CDS 501 650 .   +   2   gene_id "001"; transcript_id "001.1";
381 Twinscan    CDS 700 707 .   +   2   gene_id "001"; transcript_id "001.1";
```

### 2. GFF

The file format specification can be found here: <http://useast.ensembl.org/info/website/upload/gff.html?redirect=no>

## 6 Problem 1

Bert Vogelstein in a Science paper published in 2013 (PMID: 23539594) reported a list of Tumor Suppressor genes and Oncogenes. The list is available in the `data` folder as `vogelstein_tsg.txt`. Use the UNIPROT REST API get the protein IDs for the genes.

[https://rest.uniprot.org/uniprotkb/search?query=reviewed:true+AND+organism\\_id:9606+AND+gene:BRCA1&format=tsv&fields=accession,reviewed](https://rest.uniprot.org/uniprotkb/search?query=reviewed:true+AND+organism_id:9606+AND+gene:BRCA1&format=tsv&fields=accession,reviewed)

Not working: “<http://www.uniprot.org/uniprot/?query=organism:9606+AND+gene:BRCA1&format=tab&columns=id,reviewed>”

```
cat 'vogelstein_tsg.txt' | head
```

```
## ACVR1B
## APC
## ARID1A
## ARID1B
## ARID2
## ASXL1
## ATM
## ATRX
## AXIN1
## B2M
```

We will not loop through the gene list and get the ids from Uniprot.

```
for i in `cat vogelstein_tsg.txt`;do wget -q -O - "https://rest.uniprot.org/uniprotkb/search?query=reviewed:true+AND+organism_id:9
```

## 7 Problem 2

Download the Swissprot FASTA file from the UNIPROT website ([ftp://ftp.uniprot.org/pub/databases/uniprot/current\\_release/knowledgebase/complete/uniprot\\_sprot.fasta.gz](ftp://ftp.uniprot.org/pub/databases/uniprot/current_release/knowledgebase/complete/uniprot_sprot.fasta.gz)). Write a script to extract the sequences corresponding to the IDs created in Problem 1 from this file.

```
library(seqinr)
library(stringr)
fasta <- read.fasta("data/uniprot_sprot.fasta.gz", seqtype = "AA", as.string = T)
ids <- names(fasta)
ex.ids <- str_match(ids, "\\S+\\|(?\\S+)\\|\\S+")
ex.ids <- ex.ids[,2]
required_ids <- read.table("data/reviewed.txt")[,1]
subset_fasta <- fasta[ ex.ids %in% required_ids ]
length(required_ids)
write.fasta(subset_fasta, names=names(subset_fasta), file.out = "output_file.faa" )
```