



Thesis Defence: Evaluation of Gene Finding Tools When Applied to *Trichoderma* Genomes

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Outline

- *Trichoderma* fungi.
- Novel *Trichoderma* genomes.
- Motivation.
- Research objectives.
- Workflow overview.
- Results.
- Conclusions and recommendations.
- Questions.



What is *Trichoderma*?

- *Trichoderma* is a genus of filamentous fungi that is ubiquitous in soil and plays a significant role in nutrient cycling.
- They are also used in biocontrol and as biofertilizers.
- Known for their production of plant cell wall degrading enzymes, and **secondary metabolites**.
- Further genomic studies can help in understanding their biology and potential applications.

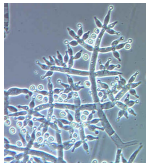


Figure: *T. harzianum*



Figure: *Trichoderma* colony

Novel *Trichoderma* Genomes

- *Trichoderma* species are diverse, with many species not yet fully characterized.
- DC1 and Tsth20, shown to improve drought and salt tolerance when applied to crops, and have been shown to breakdown hydrocarbons in soils.
- Recent advances in sequencing technology have made it possible to generate high-quality genomes for these species.



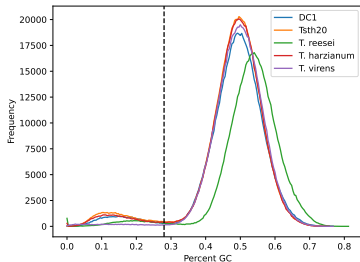
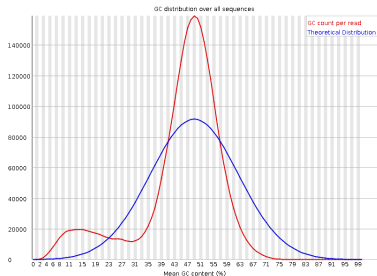
Motivation

- To understand *Trichoderma* biology, we must identify genes within their genomes.
- Gene prediction tools vary significantly in implementation and performance.
- **Few comparative studies exist for these tools in fungi, particularly in *Trichoderma*.**
- New high-quality *Trichoderma* genomes (DC1 and Tsth20) enable comparative evaluation of gene finding tools.



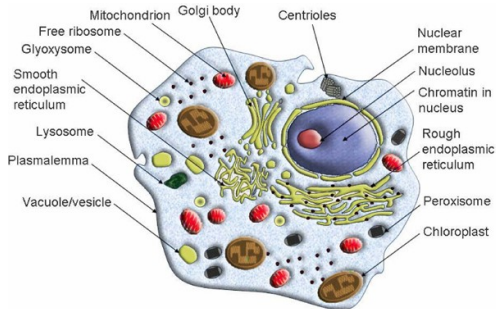
Gene Finding is Still Challenging

- We've been finding genes for decades. What's going on?
- Exons and introns as well as start and stop positions make gene finding tricky
- Inherent properties of genomes also complicate gene finding.

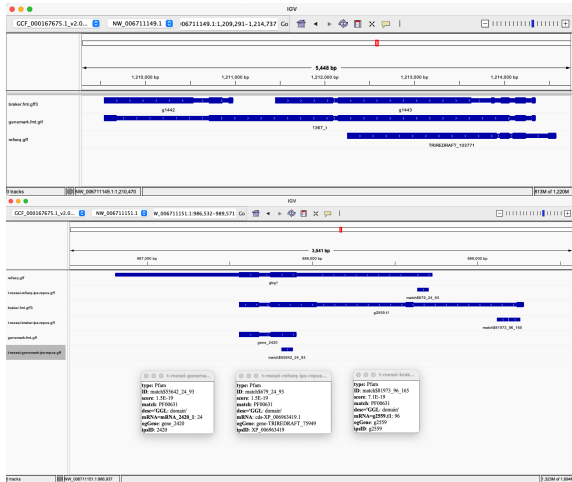


Genes are Sensitive

- Minor changes in start/stop positions can have major impacts on predicted protein sequences.
- Ex. The subcellular localization process relies heavily on N-terminal sequences.

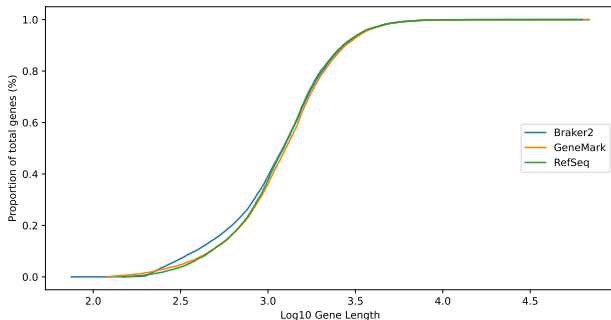


Uncertainty in Gene Predictions



Small but Significant Details

- Before diving too deep, how can we objectively confirm that gene finding tools predict different genes?
- Kolmogorov-Smirnov (KS) tests were performed on gene length distributions from preliminary gene predictions.

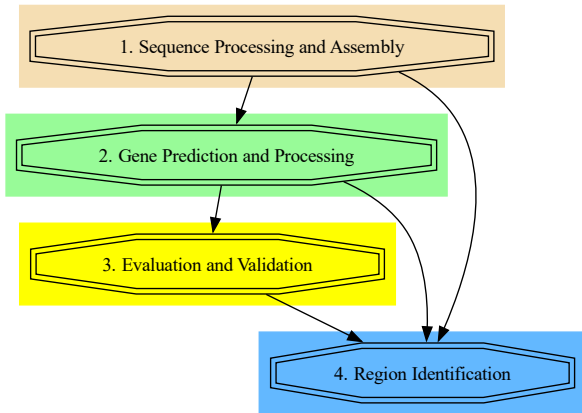


Research Objectives

- Assemble and evaluate novel assemblies of DC1 and Tsth20.
- Apply gene finders Braker2 and GeneMark in DC1, Tsth20, and three other RefSeq assemblies.
- Compare gene finding tools based on relevant criteria, including:
 - Number of genes and isoforms predicted.
 - Presence of conserved single-copy orthologs (BUSCOs).
 - Presence of functional domains and closely related protein sequences.
 - Presence of genes in AT-rich sequence.
 - Agreement of gene finders on start and stop positions of a gene.



Workflow Overview



Overview of Results

Category	Braker2	GeneMark	RefSeq
Availability	2	2	0
Ease of install	1	2	0
Ease of use	2	2	0
Number of genes predicted	1	3	3
Number of isoforms predicted	3	0	2
BUSCO Performance	3	2	1
Performance in AT-rich sequence	2	1	3
Predictions with InterProScan support	2	3	1
Cumulative Rank (Considering Availability)	16	15	N/A
Cumulative Rank (Ignoring Availability)	11	9	10



Key Findings

- Overall performance of gene finders are similar, but vary by specific criteria.
- Braker2 generally performs best, particularly when relevant training data is available.
- Candidate secondary metabolite gene clusters were found in both DC1 and Tsth20 assemblies.
- DC1 and Tsth20 assemblies are of high quality, providing a solid foundation for future genomic studies.

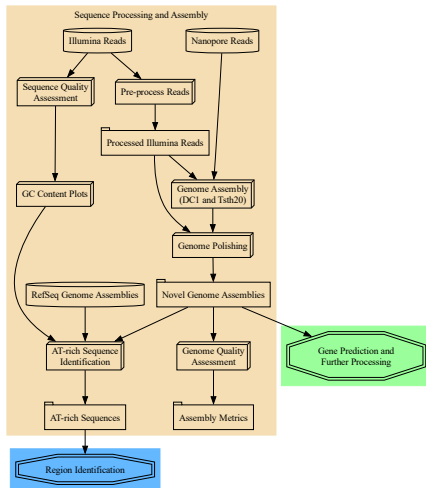


Acknowledgements

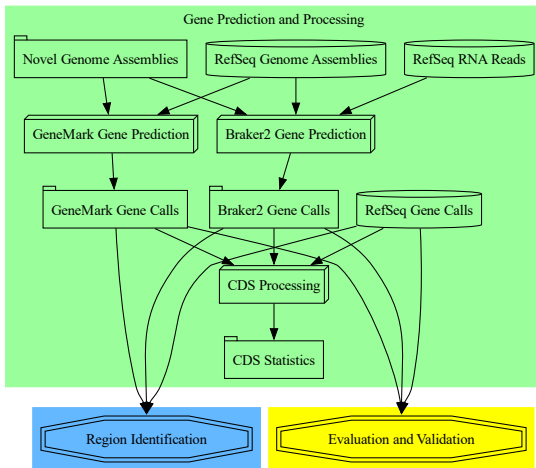
- My supervisors for their committed support through COVID and other challenges.
- Brendan Ashby and the Kaminskyj lab for providing the data for this project.
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- My committee members for their feedback and support.



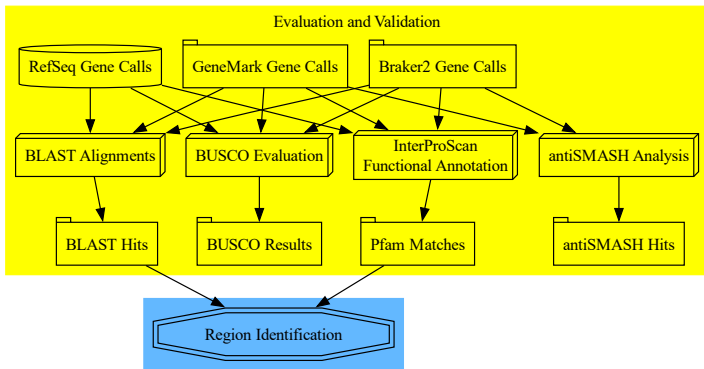
Workflow Details: Assembly



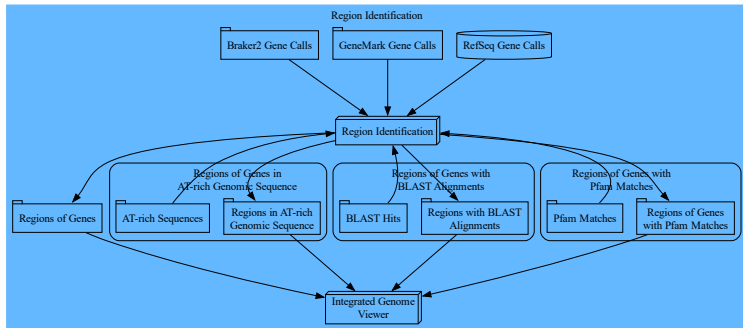
Workflow Details: Gene Finding



Workflow Details: Evaluation



Workflow Details: Regions



Datasets and Tools

- **Datasets:**

- Novel *Trichoderma* genomes: DC1 and Tsth20.
- Reference genomes and annotations: *T. reesei*, *T. harzianum*, *T. virens*.
- RNAseq training data from *T. reesei*.
- Benchmarking Universal Single-Copy Orthologs (BUSCO) fungal database.
- Protein sequence queries for tblastn from *T. atroviride*, *Fusarium graminearum*, and *Saccharomyces cerevisiae*.

- **Tools:**

- Sequence processing: FastQC, Trimmomatic, Hisat2.
- Genome assembly: NextDenovo and NextPolish.
- Gene finding tools: Braker2 and GeneMark-ES.
- Evaluation tools: BUSCO, tblastn, InterProScan, and custom scripts.





Regions



Image Credits

- *T. harzianum* image:
<https://en.wikipedia.org/wiki/Trichoderma>
- *Trichoderma colony* image:<https://biocontrol.entomology.cornell.edu/pathogens/trichoderma.php>

