Evaluation of Gene Finding Tools on *Trichoderma* Genomes - Update

July, 2025

Connor Burbridge

Outline

- Progress update with results
- Progress assessment
- Timeline for completion
- Discussion of next steps

Background

- Gene finding is a critical step in genome annotation.

- *Trichoderma* species are important for agriculture and biotechnology.

- Various tools exist for gene prediction, but their implementations and performance can vary significantly.

- Few studies have compared these tools in fungi, and even fewer in *Trichoderma*.

- In addition, high-quality reference genomes are not available for all species, which makes comparisons difficult.

Why *Trichoderma*?

- *Trichoderma* species are ubiquitous in soil and play a significant role in nutrient cycling.
- They are also used in biocontrol and as biofertilizers.
- Known for their production of plant cell wall degrading enzymes, and secondary metabolites.
- Further genomic studies can help in understanding their biology and potential applications.
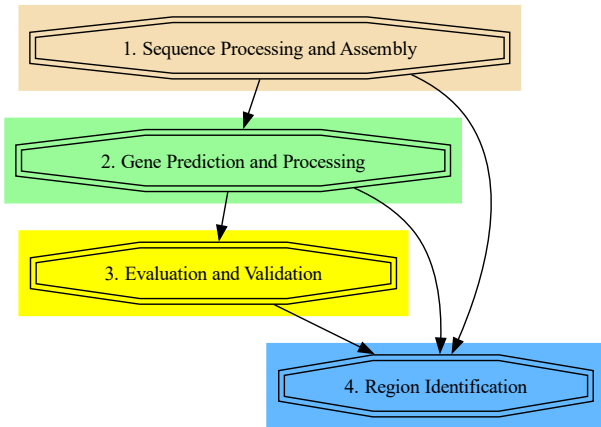
Novel *Trichoderma* Genomes

- *Trichoderma* species are diverse, with many species not yet fully characterized.

- Recent advances in sequencing technology have made it possible to generate high-quality genomes for these species.

- Two novel genomes of *Trichoderma* species have been sequenced at the Global Institute for food Security

- DC1 and Tsth20, shown to improve drought and salt tolerance when applied to crops.

- These genomes provide an opportunity to evaluate gene finding tools in a comparative context, and contribute to the understanding of *Trichoderma* biology.

Workflow Overview

Datasets and Tools

- **Datasets:**
  - *Trichoderma* genomes: DC1 and Tsth20.
  - Reference genomes and annotations: *T. reesei, T. harzianum, T.virens.*
  - RNAseq training data from *T. reesei*.
  - Benchmarking Universal Single-Copy Orthologs (BUSCO) fungal database.
  - Protein sequence queries for tblastn from *T. atroviride, Fusarium graminearum, and Saccharomyces cerevisiae*.
- **Tools:**
  - Sequence processing: FastQC, Trimmomatic, Hisat2.
  - Genome assembly: NextDenovo and NextPolish.
  - Gene finding tools: Braker2 and GeneMark-ES.
  - Evaluation tools: BUSCO, tblastn, InterProScan, and custom scripts.

Assembly Results

Name	Total Contigs	Total Length	Largest Contig	GC%	N50	L50
DC1	8	38.6 Mb	11.49 Mb	47.97	5.69 Mb	3
Tsth20	7	41.58 Mb	8.02 Mb	47.33	6.52 Mb	3
T. harzianum	532	40.98 Mb	4.08 Mb	47.61	2.41 Mb	7
T. virens	93	39.02 Mb	3.45 Mb	49.25	1.83 Mb	8
T. reesei	77	33.39 Mb	3.75 Mb	52.82	1.21 Mb	9

- DC1 and Tsth20 assemblies are high quality with few contigs in comparison to other *Trichoderma* assemblies.

- Input sequences and assemblies show a bimodal distribution of GC content.

- AT-rich sequence content may be related to transposable elements and repeat-induced mutations, which may be of interest in secondary metabolite production.

Gene Finding Results

Assembly	Braker2		GeneMark		RefSeq	
	Genes	CDS	Genes	CDS	Genes	CDS
DC1	8546	8637	11353	11353	N/A	N/A
Tsth20	8784	8858	12362	12362	N/A	N/A
T. reesei	9659	10175	9196	9196	9109	9118
T. harzianum	8314	8385	12164	12164	14269	14090
T. virens	7801	7863	11866	11866	12405	12406

- Braker2 predicts more genes and coding sequences than GeneMark and RefSeq in *T. ressei*, but fewer in other assemblies.

- GeneMark and RefSeq predictions are similar, except in *T. harzianum*, where RefSeq predicts 17% more genes.

- GeneMark only predicts one coding sequence per gene, while Braker2 and RefSeq predict multiple coding sequences.

Examining Coding Sequence Lengths

Genome	Tool #1	Tool #2	P-value
DC1	Braker2	GeneMark	0.999
Tsth20	Braker2	GeneMark	0.965
T. reesei	Braker2	GeneMark	$9.481 * 10^{-07}$
T. reesei	GeneMark	RefSeq	0.002
T. reesei	Braker2	RefSeq	$1.340 * 10^{-07}$
T. harzianum	Braker2	GeneMark	0.863
T. harzianum	GeneMark	RefSeq	$4.313 * 10^{-52}$
T. harzianum	Braker2	RefSeq	$4.674 * 10^{-55}$
T. virens	Braker2	GeneMark	0.635
T. virens	GeneMark	RefSeq	$7.352 * 10^{-12}$
T. virens	Braker2	RefSeq	$1.794 * 10^{-09}$

BUSCO Results

Strain	Complete	Single	Duplicated	Fragmented	Missing
DC1	99.5	80.2	19.3	0.1	0.4
Tsth20	99.9	81.7	18.2	0.0	0.1
T. harzianum	99.7	80.2	19.5	0.0	0.3
T. virens	99.8	79.0	20.8	0.1	0.1
T. reesei	99.9	85.5	14.4	0.1	0.0

(a) Braker2

Strain	Complete	Single	Duplicated	Fragmented	Missing
DC1	99.2	98.8	0.4	0.3	0.5
Tsth20	99.8	99.1	0.7	0.0	0.2
T. harzianum	99.6	98.9	0.7	0.0	0.4
T. virens	99.7	99.2	0.5	0.1	0.2
T. reesei	99.6	99.5	0.1	0.0	0.4

(b) GeneMark

Strain	Complete	Single	Duplicated	Fragmented	Missing
T. harzianum	99.9	99.2	0.7	0.0	0.1
T. virens	99.5	98.8	0.7	0.3	0.2
T. reesei	99.8	99.5	0.3	0.0	0.2

(c) RefSeq

Agreement of Gene Predictions

- In DC1 and Tsth20, BRaker2 and GeneMark tend to agree on theh start and stop positions of genes when they predict the same gene. Both gene finders have a large portion of singleton predictions.

Regions: 11269

Full Support: 8482

Singletons: 2787

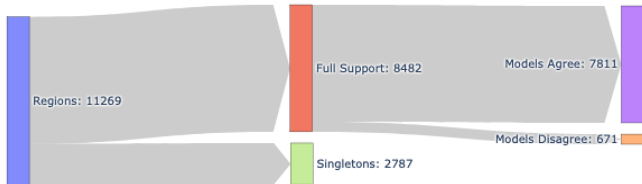Models Agree: 7811

Models Disagree: 671

Figure: DC1

- In *T. reesei*, Braker2, GeneMark and RefSeq tend to disagree more on the start and stop positions of genes when they predict the same gene.
- Disagreement is more pronounced in *T. harzianum* and *T. virens*, where genes with partial support from more than one gene finder are common.
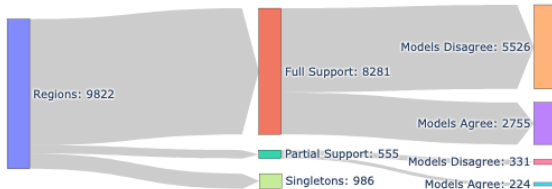

Figure: *T. reesei*

InterProScan Results

- k

Assembly	Braker2	GeneMark	RefSeq
DC1	$\frac{10676}{14479}$	$\frac{8416}{11354}$	N/A
Tsth20	$\frac{11389}{15546}$	$\frac{9168}{12373}$	N/A
T. reesei	$\frac{8471}{11704}$	$\frac{6990}{9196}$	$\frac{6964}{9111}$
T. harzianum	$\frac{11370}{15408}$	$\frac{9061}{12164}$	$\frac{9293}{14065}$
T. virens	$\frac{11249}{15062}$	$\frac{8871}{11866}$	$\frac{9062}{12383}$