

Assignment 3: Data Exploration

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OVERVIEW

This exercise accompanies the lessons in Environmental Data Analytics on Data Exploration.

Directions

1. Rename this file `<FirstLast>_A03_DataExploration.Rmd` (replacing `<FirstLast>` with your first and last name).
2. Change “Student Name” on line 3 (above) with your name.
3. Work through the steps, **creating code and output** that fulfill each instruction.
4. Assign a useful **name to each code chunk** and include ample **comments** with your code.
5. Be sure to **answer the questions** in this assignment document.
6. When you have completed the assignment, **Knit** the text and code into a single PDF file.
7. After Knitting, submit the completed exercise (PDF file) to the dropbox in Sakai.

TIP: If your code extends past the page when knit, tidy your code by manually inserting line breaks.

TIP: If your code fails to knit, check that no `install.packages()` or `View()` commands exist in your code.

Set up your R session

1. Check your working directory, load necessary packages (tidyverse, lubridate), and upload two data sets: the ECOTOX neonicotinoid data set (ECOTOX_Neonicotinoids_Insects_raw.csv) and the Niwot Ridge NEON data set for litter and woody debris (NEON_NIWO_Litter_massdata_2018-08_raw.csv). Name these data sets “Neonics” and “Litter”, respectively. Be sure to include the subcommand to read strings in as factors.

```
getwd() #checking working directory
```

```
## [1] "Z:/EDE_Fall2023"
```

```
library(tidyverse) #loading necessary package
```

```
## -- Attaching core tidyverse packages ----- tidyverse 2.0.0 --  
## v dplyr      1.1.3      v readr      2.1.4  
## v forcats    1.0.0      v stringr    1.5.0
```

```
## v ggplot2 3.4.3      v tibble 3.2.1
## v lubridate 1.9.2    v tidyr 1.3.0
## v purrr 1.0.2
## -- Conflicts ----- tidyverse_conflicts() --
## x dplyr::filter() masks stats::filter()
## x dplyr::lag() masks stats::lag()
## i Use the conflicted package (<http://conflicted.r-lib.org/>) to force all conflicts to become errors

library(lubridate) #loading necessary package
Neonics <- read.csv("../Data/Raw/ECOTOX_Neonicotinoids_Insects_raw.csv") #import
#data with a relative path, naming it
Litter <- read.csv("../Data/Raw/NEON_NIWO_Litter_massdata_2018-08_raw.csv") #import
#data with a relative path, naming it
```

Learn about your system

2. The neonicotinoid data set was collected from the Environmental Protection Agency's ECOTOX Knowledgebase, a database for ecotoxicology research. Neonicotinoids are a class of insecticides used widely in agriculture. The data set that has been pulled includes all studies published on insects. Why might we be interested in the ecotoxicology of neonicotinoids on insects? Feel free to do a brief internet search if you feel you need more background information.

Answer: Neonicotinoids impact non-target insects, such as bees, and aquatic invertebrate communities. Bees are very important pollinators, and the toxicity they have been experiencing for decades because of insecticides (like Neonicotinoids also others) has led to a significant decrease in bee population, which can be disastrous for ecosystems. Aquatic ecosystems are also impacted by neonicotinoids and it's difficult to remove pollutants from waterways.

3. The Niwot Ridge litter and woody debris data set was collected from the National Ecological Observatory Network, which collectively includes 81 aquatic and terrestrial sites across 20 ecoclimatic domains. 32 of these sites sample forest litter and woody debris, and we will focus on the Niwot Ridge long-term ecological research (LTER) station in Colorado. Why might we be interested in studying litter and woody debris that falls to the ground in forests? Feel free to do a brief internet search if you feel you need more background information.

Answer: Litter and woody debris are important in forest and stream ecosystems because they are a part of carbon budgets and nutrient cycling, provide habitats for terrestrial and aquatic organisms, and influence water flows and sediment transport.

4. How is litter and woody debris sampled as part of the NEON network? Read the NEON_Litterfall_UserGuide.pdf document to learn more. List three pieces of salient information about the sampling methods here:

Answer: 1. Sampling sites range from a size of 20m x 20 m (low-statured vegetation over the tower airsheds) to 40 m x 40 m (forested tower airsheds). 2. Litter trap placement within plots may be targeted or randomized, depending on the vegetation. If the site has 50% aerial cover of woody vegetation >2m in height, litter traps are randomly placed. Otherwise, there's a described method on how to place the traps. 3. Group traps are sampled once annually, while sampling for elevated traps varies by the site's vegetation; deciduous forest sites should have frequent sampling (once every 2 weeks) during senescence, while evergreen sites need infrequent year-round sampling (once every 1-2 months).

Obtain basic summaries of your data (Neonics)

5. What are the dimensions of the data set?

```
dim(Neonics) # 188 rows, 19 columns
```

```
## [1] 4623 30
```

```
dim(Litter) # 4623 rows, 30 columns
```

```
## [1] 188 19
```

6. Using the `summary` function on the “Effect” column, determine the most common effects that are studied. Why might these effects specifically be of interest?

```
Neonics$Effect <- as.factor(Neonics$Effect) #convert the character values in the
#column into factors. This allows us to quantify character values, particularly
#when they have a fixed and known set of possible values
summary(Neonics$Effect) #summarizes the column to find the most common values
```

```
##      Accumulation      Avoidance      Behavior      Biochemistry
##           12           102           360           11
##      Cell(s)      Development      Enzyme(s) Feeding behavior
##           9           136           62           255
##      Genetics      Growth      Histology      Hormone(s)
##          82           38           5           1
##      Immunological      Intoxication      Morphology      Mortality
##          16           12           22           1493
##      Physiology      Population      Reproduction
##           7           1803           197
```

```
sort(summary(Neonics$Effect), decreasing = TRUE)
```

```
##      Population      Mortality      Behavior Feeding behavior
##          1803          1493           360           255
##      Reproduction      Development      Avoidance      Genetics
##          197           136           102           82
##      Enzyme(s)      Growth      Morphology      Immunological
##          62           38           22           16
##      Accumulation      Intoxication      Biochemistry      Cell(s)
##          12           12           11           9
##      Physiology      Histology      Hormone(s)
##           7           5           1
```

Answer: The two most common studied effects are Population (1803 examples) and Mortality (1493). Mortality is likely studied to measure when the cause of death is by direct action of the chemical. Population measures the group of organisms of the same species in the same area at a given time. This would allow for a measure of biodiversity and, depending on how it is measured, may allow for a calculation of mortality rate.

7. Using the `summary` function, determine the six most commonly studied species in the data set (common name). What do these species have in common, and why might they be of interest over other insects? Feel free to do a brief internet search for more information if needed. [TIP: The `sort()` command can sort the output of the summary command...]

```
Neonics$Species.Common.Name <- as.factor(Neonics$Species.Common.Name) #convert
#the character values in the column into factors. This allows us to quantify
#character values, particularly when they have a fixed and known set of possible
#values
summary(Neonics$Species.Common.Name) #summarizes the column to find the most
```

##	Honey Bee	Parasitic Wasp
##	667	285
##	Buff Tailed Bumblebee	Carniolan Honey Bee
##	183	152
##	Bumble Bee	Italian Honeybee
##	140	113
##	Japanese Beetle	Asian Lady Beetle
##	94	76
##	Euonymus Scale	Wireworm
##	75	69
##	European Dark Bee	Minute Pirate Bug
##	66	62
##	Asian Citrus Psyllid	Parastic Wasp
##	60	58
##	Colorado Potato Beetle	Parasitoid Wasp
##	57	51
##	Erythrina Gall Wasp	Beetle Order
##	49	47
##	Snout Beetle Family, Weevil	Sevenspotted Lady Beetle
##	47	46
##	True Bug Order	Buff-tailed Bumblebee
##	45	39
##	Aphid Family	Cabbage Looper
##	38	38
##	Sweetpotato Whitefly	Braconid Wasp
##	37	33
##	Cotton Aphid	Predatory Mite
##	33	33
##	Ladybird Beetle Family	Parasitoid
##	30	30
##	Scarab Beetle	Spring Tiphia
##	29	29
##	Thrip Order	Ground Beetle Family
##	29	27
##	Rove Beetle Family	Tobacco Aphid
##	27	27
##	Chalcid Wasp	Convergent Lady Beetle
##	25	25
##	Stingless Bee	Spider/Mite Class
##	25	24
##	Tobacco Flea Beetle	Citrus Leafminer
##	24	23
##	Ladybird Beetle	Mason Bee

##	23	22
##	Mosquito	Argentine Ant
##	22	21
##	Beetle	Flatheaded Appletree Borer
##	21	20
##	Horned Oak Gall Wasp	Leaf Beetle Family
##	20	20
##	Potato Leafhopper	Tooth-necked Fungus Beetle
##	20	20
##	Codling Moth	Black-spotted Lady Beetle
##	19	18
##	Calico Scale	Fairyfly Parasitoid
##	18	18
##	Lady Beetle	Minute Parasitic Wasps
##	18	18
##	Mirid Bug	Mulberry Pyralid
##	18	18
##	Silkworm	Vedalia Beetle
##	18	18
##	Araneoid Spider Order	Bee Order
##	17	17
##	Egg Parasitoid	Insect Class
##	17	17
##	Moth And Butterfly Order	Oystershell Scale Parasitoid
##	17	17
##	Hemlock Woolly Adelgid Lady Beetle	Hemlock Woolly Adelgid
##	16	16
##	Mite	Onion Thrip
##	16	16
##	Western Flower Thrips	Corn Earworm
##	15	14
##	Green Peach Aphid	House Fly
##	14	14
##	Ox Beetle	Red Scale Parasite
##	14	14
##	Spined Soldier Bug	Armoured Scale Family
##	14	13
##	Diamondback Moth	Eulophid Wasp
##	13	13
##	Monarch Butterfly	Predatory Bug
##	13	13
##	Yellow Fever Mosquito	Braconid Parasitoid
##	13	12
##	Common Thrip	Eastern Subterranean Termite
##	12	12
##	Jassid	Mite Order
##	12	12
##	Pea Aphid	Pond Wolf Spider
##	12	12
##	Spotless Ladybird Beetle	Glasshouse Potato Wasp
##	11	10
##	Lacewing	Southern House Mosquito
##	10	10
##	Two Spotted Lady Beetle	Ant Family

```
##                10                9
##                Apple Maggot        (Other)
##                9                670
```

#common values

```
sort(summary(Neonics$Species.Common.Name), decreasing = TRUE)
```

```
##                (Other)                Honey Bee
##                670                667
##                Parasitic Wasp        Buff Tailed Bumblebee
##                285                183
##                Carniolan Honey Bee        Bumble Bee
##                152                140
##                Italian Honeybee        Japanese Beetle
##                113                94
##                Asian Lady Beetle        Euonymus Scale
##                76                75
##                Wireworm        European Dark Bee
##                69                66
##                Minute Pirate Bug        Asian Citrus Psyllid
##                62                60
##                Parastic Wasp        Colorado Potato Beetle
##                58                57
##                Parasitoid Wasp        Erythrina Gall Wasp
##                51                49
##                Beetle Order        Snout Beetle Family, Weevil
##                47                47
##                Sevenspotted Lady Beetle        True Bug Order
##                46                45
##                Buff-tailed Bumblebee        Aphid Family
##                39                38
##                Cabbage Looper        Sweetpotato Whitefly
##                38                37
##                Braconid Wasp        Cotton Aphid
##                33                33
##                Predatory Mite        Ladybird Beetle Family
##                33                30
##                Parasitoid        Scarab Beetle
##                30                29
##                Spring Tiphia        Thrip Order
##                29                29
##                Ground Beetle Family        Rove Beetle Family
##                27                27
##                Tobacco Aphid        Chalcid Wasp
##                27                25
##                Convergent Lady Beetle        Stingless Bee
##                25                25
##                Spider/Mite Class        Tobacco Flea Beetle
##                24                24
##                Citrus Leafminer        Ladybird Beetle
##                23                23
##                Mason Bee        Mosquito
##                22                22
##                Argentine Ant        Beetle
```

##	21	21
##	Flatheaded Appletree Borer	Horned Oak Gall Wasp
##	20	20
##	Leaf Beetle Family	Potato Leafhopper
##	20	20
##	Tooth-necked Fungus Beetle	Codling Moth
##	20	19
##	Black-spotted Lady Beetle	Calico Scale
##	18	18
##	Fairyfly Parasitoid	Lady Beetle
##	18	18
##	Minute Parasitic Wasps	Mirid Bug
##	18	18
##	Mulberry Pyralid	Silkworm
##	18	18
##	Vedalia Beetle	Araneoid Spider Order
##	18	17
##	Bee Order	Egg Parasitoid
##	17	17
##	Insect Class	Moth And Butterfly Order
##	17	17
##	Oystershell Scale Parasitoid	Hemlock Woolly Adelgid Lady Beetle
##	17	16
##	Hemlock Wooly Adelgid	Mite
##	16	16
##	Onion Thrip	Western Flower Thrips
##	16	15
##	Corn Earworm	Green Peach Aphid
##	14	14
##	House Fly	Ox Beetle
##	14	14
##	Red Scale Parasite	Spined Soldier Bug
##	14	14
##	Armoured Scale Family	Diamondback Moth
##	13	13
##	Eulophid Wasp	Monarch Butterfly
##	13	13
##	Predatory Bug	Yellow Fever Mosquito
##	13	13
##	Braconid Parasitoid	Common Thrip
##	12	12
##	Eastern Subterranean Termite	Jassid
##	12	12
##	Mite Order	Pea Aphid
##	12	12
##	Pond Wolf Spider	Spotless Ladybird Beetle
##	12	11
##	Glasshouse Potato Wasp	Lacewing
##	10	10
##	Southern House Mosquito	Two Spotted Lady Beetle
##	10	10
##	Ant Family	Apple Maggot
##	9	9

Answer: The six most commonly studied species are the Honey Bee (667), Parasitic Wasp (285), Buff Tailed Bumblebee (183), Carniolan Honey Bee (152), Bumble Bee (140), and Italian Honey Bee (113). All of these species are pollinators are not target insects of the insecticide. They are likely studied most because of the aforementioned broad decrease of pollinator (specially bees) populations because of insecticide use.

8. Concentrations are always a numeric value. What is the class of `Conc.1..Author.` column in the data set, and why is it not numeric?

```
class(Neonics$Conc.1..Author.)
```

```
## [1] "character"
```

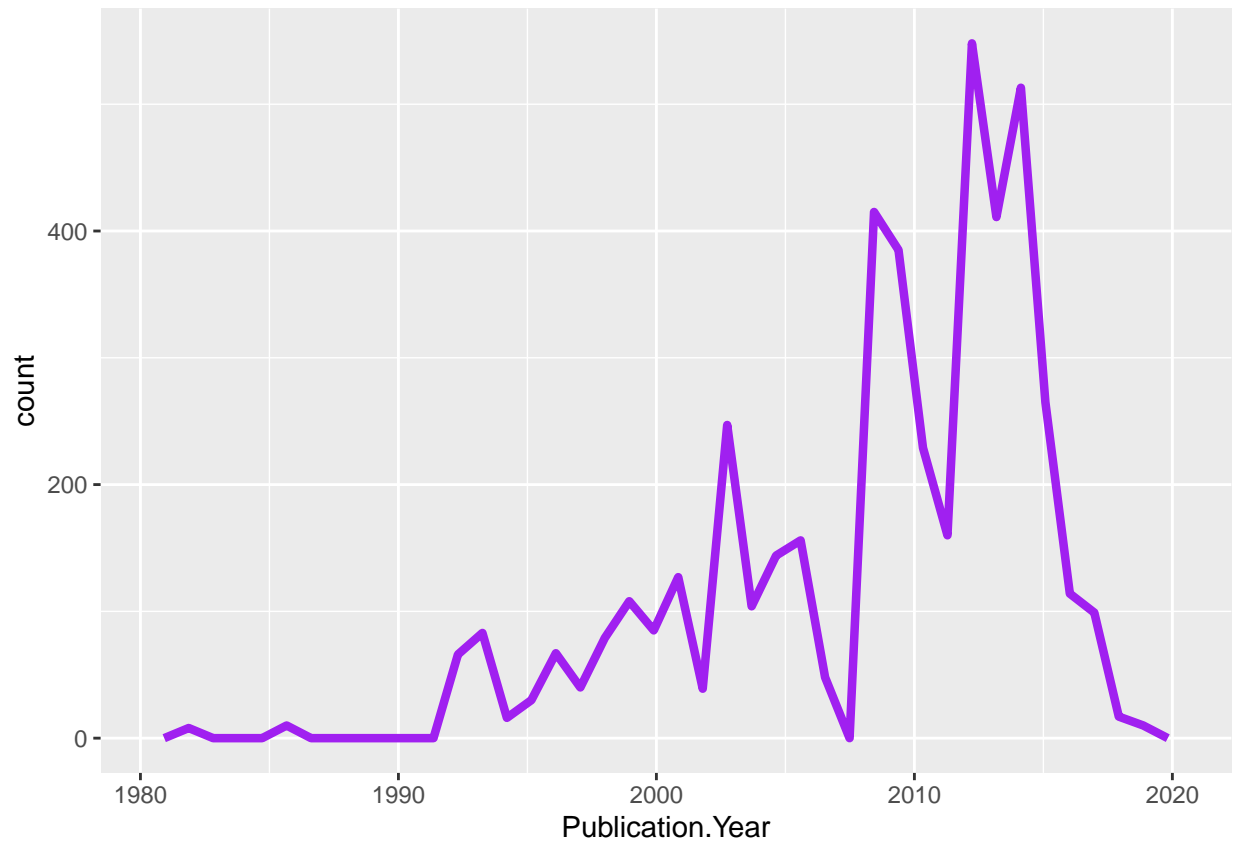
Answer: The values in the `Conc.1..Author.` column are characters. The numeric concentrations were reported as characters because not all elements are reported on the same unit and they are not measuring the same thing. `Conc.1.Type..Author.` outlines what the row is measuring (such as active ingredient), while `Conc.1.Units..Author.` determines what unit is to be used. This also allows for non-numeric characters, such as `~` to denote approximation and `/`. By listing the values as characters, the dataset authors discourage summarizations of a column that is not standardized.

Explore your data graphically (Neonics)

9. Using `geom_freqpoly`, generate a plot of the number of studies conducted by publication year.

```
ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year), bins = 40, color = "purple", size = 1.5)
```

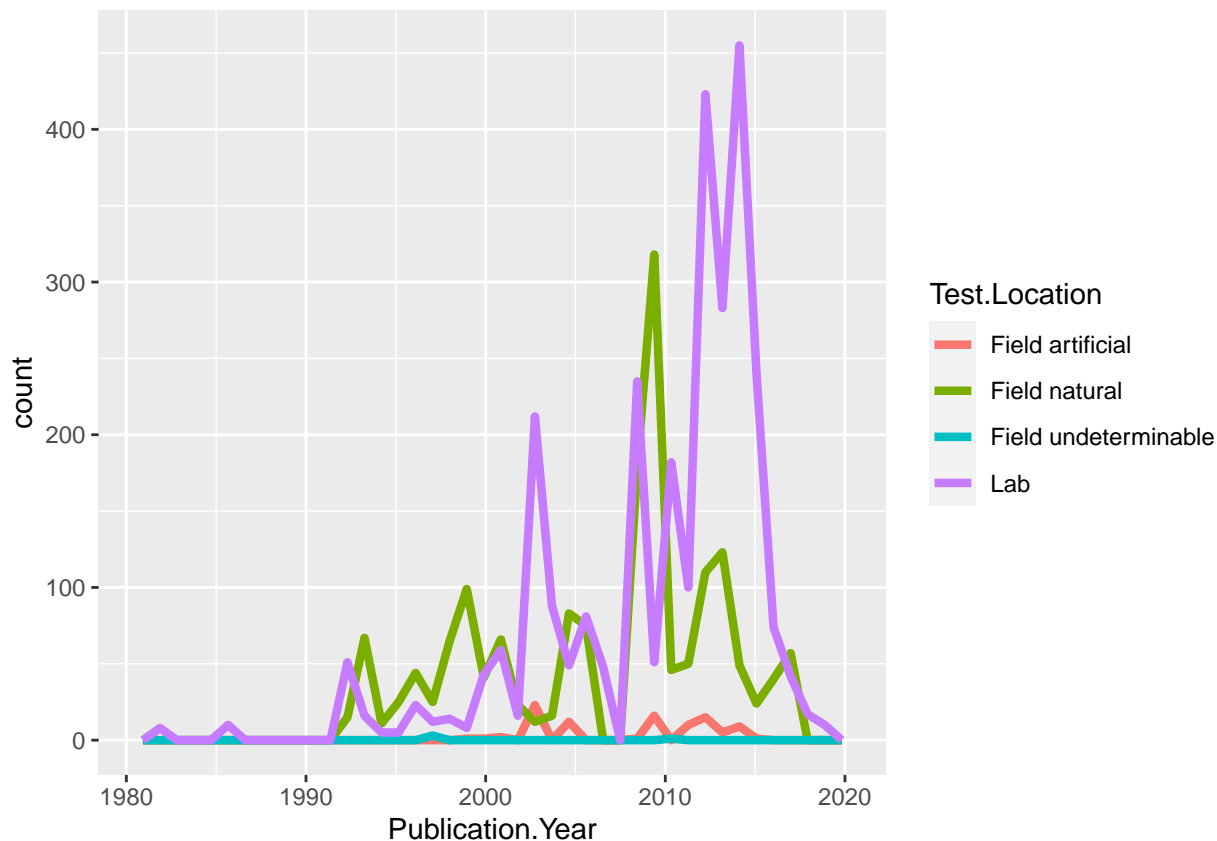
```
## Warning: Using 'size' aesthetic for lines was deprecated in ggplot2 3.4.0.  
## i Please use 'linewidth' instead.  
## This warning is displayed once every 8 hours.  
## Call 'lifecycle::last_lifecycle_warnings()' to see where this warning was  
## generated.
```

*#I graphed the specific column, increased the number of bins to see subtler
#trends over the years (40 is the difference between 2020 and 1980), and changed
#the line color and size to make it more visible.*

10. Reproduce the same graph but now add a color aesthetic so that different Test.Location are displayed as different colors.

```
ggplot(Neonics) +  
  geom_freqpoly(aes(x = Publication.Year, color = Test.Location), bins = 40, size = 1.5)
```



*#I kept the same basic graph but added the color determinant to be the elements
#in TestLocation to create 4 separate lines*

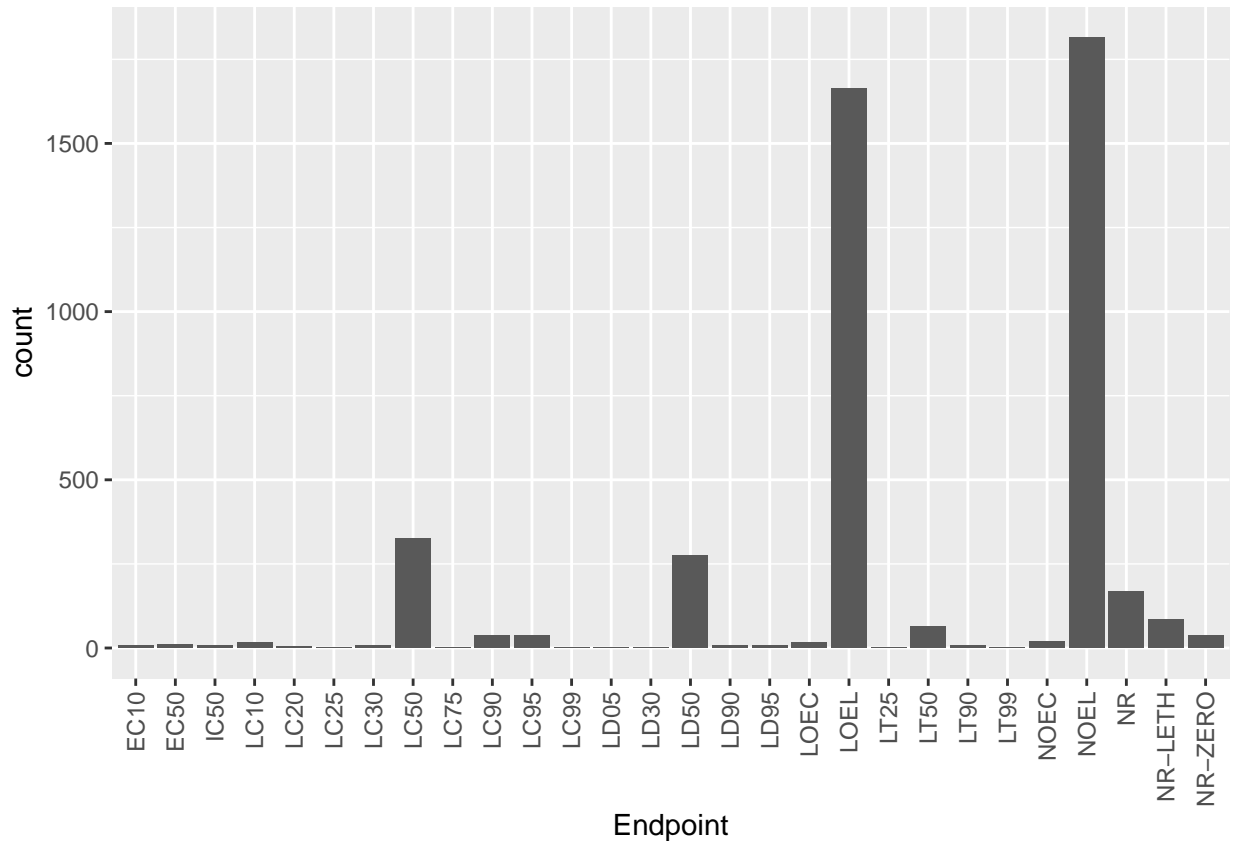
Interpret this graph. What are the most common test locations, and do they differ over time?

Answer: By having the elements in Test.Location be the determining factor for the coloring, it creates a line graph for each related element; in this case, a graph of the studies published by year, by test location. The four types of location are Field Artificial, Field Natural, Field Undeterminable, and Lab. Lab is by far the most frequent, though there are times (such as in the 90s and right before 2010) when Field Natural (the second most popular type, is the most used test location for the studies published that year. Field artificial was used sometimes, but not every year and not many published studies use it. Field Undeterminable is an unused Test Location type in published studies.

11. Create a bar graph of Endpoint counts. What are the two most common end points, and how are they defined? Consult the ECOTOX_CodeAppendix for more information.

[**TIP:** Add `theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1))` to the end of your plot command to rotate and align the X-axis labels...]

```
ggplot(Neonics) +
  geom_bar(aes(x = Endpoint)) +
  theme(axis.text.x = element_text(angle = 90, vjust = 0.5, hjust=1)) #rotates
```



#and aligns the x-axis labels

Answer: The two most common end points are NOEL and LOEL. NOEL is No-Observable-Effect-Level, whereused to note the highest concentration producing effects that aren't significantly different from control responses. LOEL is Lowest-Observable-Effect-Level, used to note the lowest concentration producing effects that are significantly different from controls.

Explore your data (Litter)

- Determine the class of collectDate. Is it a date? If not, change to a date and confirm the new class of the variable. Using the `unique` function, determine which dates litter was sampled in August 2018.

```
class(Litter$collectDate) #check the class of the column; it is currently character
```

```
## [1] "character"
```

```
Litter$collectDate <- as.Date(Litter$collectDate) #Transform column to Date format
class(Litter$collectDate) #confirm the new class of the Collect Date column
```

```
## [1] "Date"
```

```
unique(Litter$collectDate) #determine when litter was sampled in Aug 2018
```

```
## [1] "2018-08-02" "2018-08-30"
```

13. Using the `unique` function, determine how many plots were sampled at Niwot Ridge. How is the information obtained from `unique` different from that obtained from `summary`?

```
unique(Litter$siteID)
```

```
## [1] "NIWO"
```

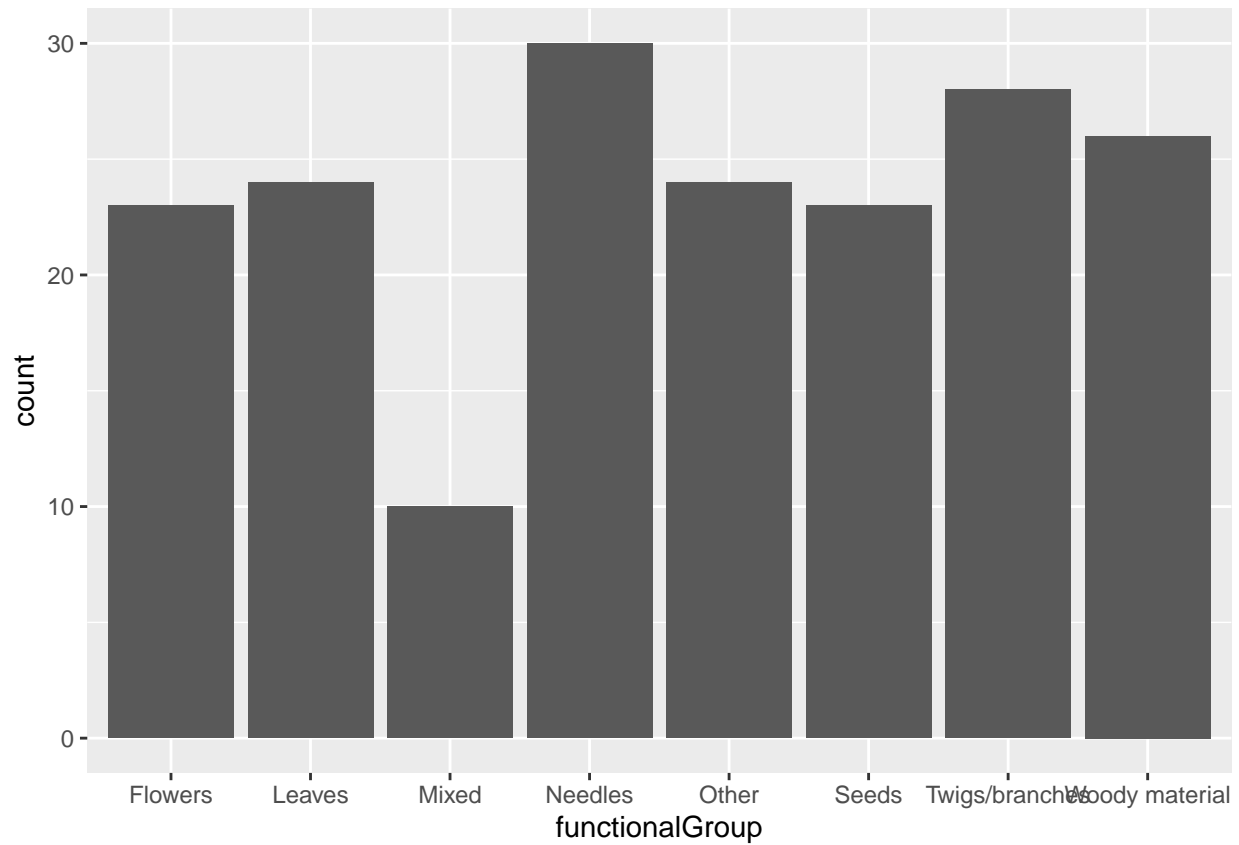
```
Litter$siteID <- as.factor(Litter$siteID) #change it to factor as before to make  
#it possible to count them in summary  
summary(Litter$siteID)
```

```
## NIWO  
## 188
```

Answer: All 188 plots were sampled at Niwot Ridge. The information is the same, just shown differently. Unique tells you the amount of unique values there are, so if there's only one value throughout the column and there are no NA/empty cells, you can assume its the length of your dataset. Summary, on the other hand, tells you the value in the column. If you turn it into an element, it can tell you the number of occurrences of every value listed, but since there's only one here, it lists the length of the dataset.

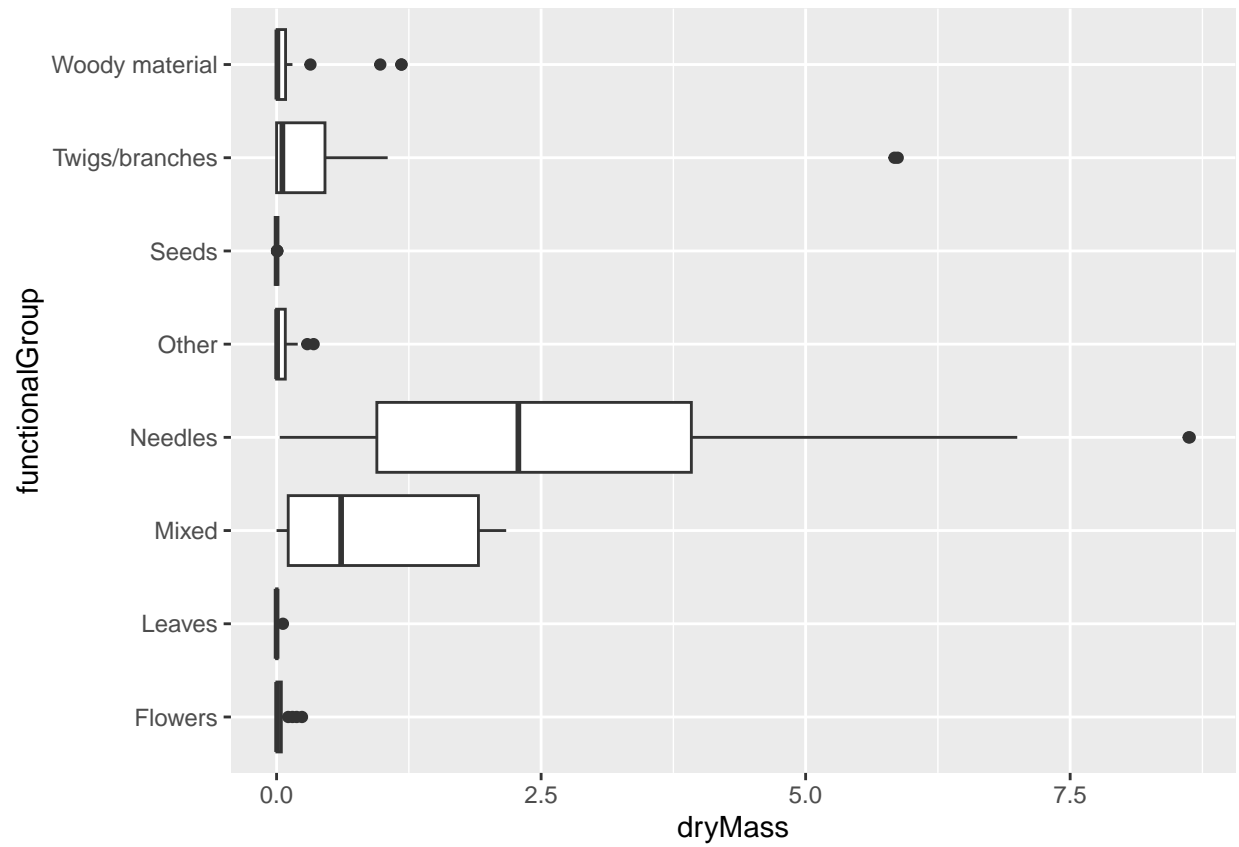
14. Create a bar graph of functionalGroup counts. This shows you what type of litter is collected at the Niwot Ridge sites. Notice that litter types are fairly equally distributed across the Niwot Ridge sites.

```
ggplot(Litter) +  
  geom_bar(aes(x = functionalGroup))
```

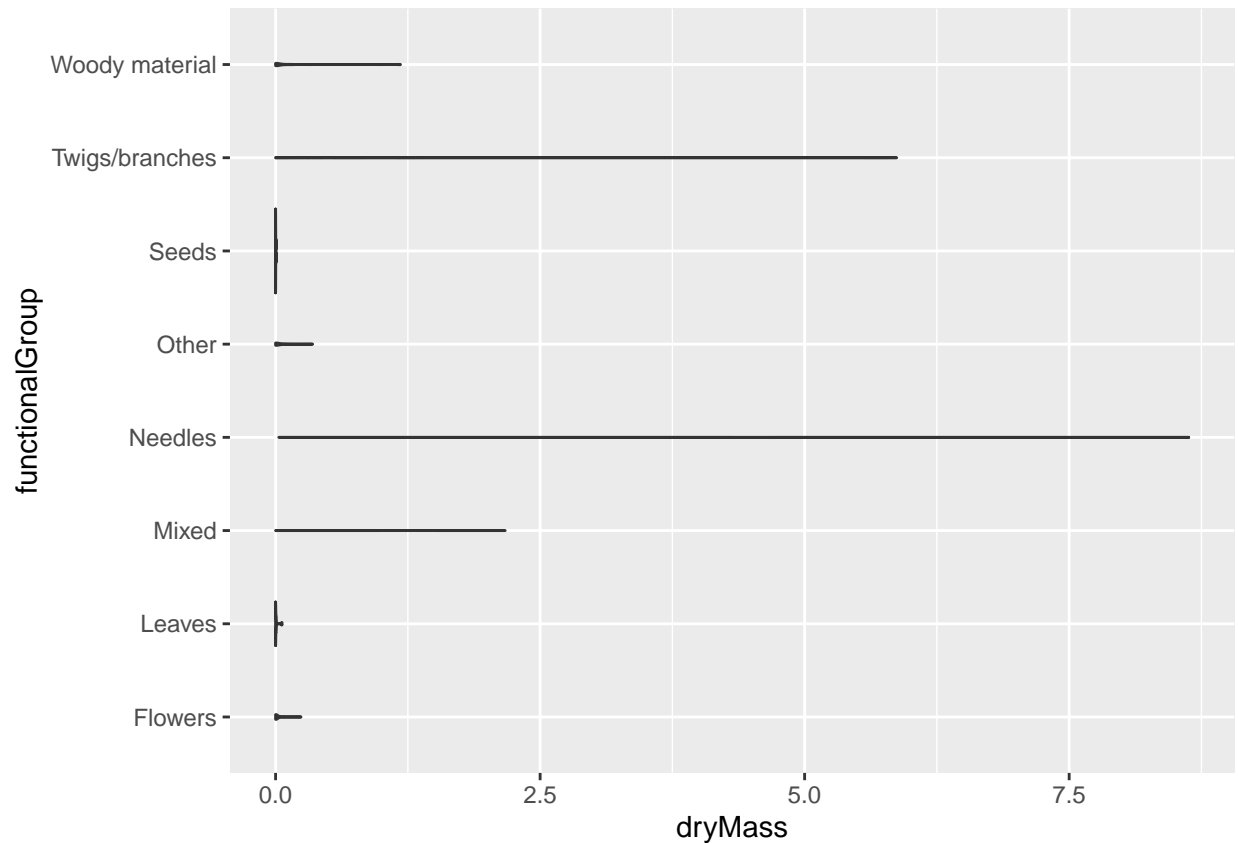


15. Using `geom_boxplot` and `geom_violin`, create a box plot and a violin plot of `dryMass` by `functionalGroup`.

```
ggplot(Litter) +  
  geom_boxplot(aes(x = dryMass, y = functionalGroup))
```



```
ggplot(Litter) +  
  geom_violin(aes(x = dryMass, y = functionalGroup))
```



Why is the box plot a more effective visualization option than the violin plot in this case?

Answer: The violin plot is eant to show the distribution and probability density at different values, but this dataset is too disperse to easily visualize any trend with this plot. The boxplot, on the other hand, always hs a visible box to visualize the median, quartiles, and outliers.

What type(s) of litter tend to have the highest biomass at these sites?

Answer: Needles.