

## MEMORY RESEARCH

# Competition between engrams influences fear memory formation and recall

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Collections of cells called engrams are thought to represent memories. Although there has been progress in identifying and manipulating single engrams, little is known about how multiple engrams interact to influence memory. In lateral amygdala (LA), neurons with increased excitability during training outcompete their neighbors for allocation to an engram. We examined whether competition based on neuronal excitability also governs the interaction between engrams. Mice received two distinct fear conditioning events separated by different intervals. LA neuron excitability was optogenetically manipulated and revealed a transient competitive process that integrates memories for events occurring closely in time (coallocating overlapping populations of neurons to both engrams) and separates memories for events occurring at distal times (disallocating nonoverlapping populations to each engram).

Memory of an event is thought to be represented by an ensemble of neurons, referred to as its memory trace or engram (1, 2). Despite recent advances in localizing and manipulating single engrams, little is known about how multiple engrams interact to influence memory function. Two engrams may engage nonoverlapping neuronal populations, thus minimizing interference between distinct memory representations. Alternatively, engrams may engage overlapping neuronal populations to functionally link those memories. Here, we examined the rules governing engram interaction.

In lateral amygdala (LA), a region critical for conditioned fear memory (3–5), eligible neurons compete for engram allocation. Neurons with relatively higher function of transcription factor CREB or increased excitability at training preferentially win this competition and are allocated to an engram (6–10). Silencing these neurons prevents memory expression, whereas their activation artificially elicits memory expression, indicating that these allocated neurons are both necessary and sufficient for expression of that memory (7–10). Similar competition governs engram allocation in mice without experimental manipulation of CREB or excitability (11).

We asked if two fear-conditioning events that occur closely in time are coallocated to overlapping populations of neurons, thereby func-

tionally linking these memories. Mice received two events (event 1, event 2) featuring distinct auditory conditioned stimuli (CS1 or CS2, Fig. 1A; see supplementary materials) separated by varying intertraining intervals (ITIs; 1.5, 3, 6, 18, or 24 hours). Event 2 (CS2+footshock) was the same across groups, but event 1 content and timing differed. Memory for event 1 was stable across ITIs (fig. S1B). However, event 2 memory was enhanced if event 1 (CS1+footshock) occurred shortly before (1.5 to 6 hours, but not 18 to 24 hours; Fig. 1B and fig. S2, B and C), even if event 1 consisted of light-footshock pairing (fig. S2A). Event 2 memory enhancement depended on previous fear conditioning (not sensitization); mice failed to show enhanced event 2 memory if event 1 consisted of CS1 alone or immediate footshock.

To examine if neural representations of events separated by short (not long) intervals are coallocated to overlapping neuronal populations, we used fluorescent in situ hybridization (FISH) for two activity-dependent genes. Nuclear *arc* mRNA labels neurons that were active in the preceding 5 min, whereas nuclear *homer1a* (*h1a*) mRNA labels neurons that were active in the preceding 30 to 40 min (12). Mice received event 1 and event 2 separated by 6 or 24 hours. During the memory test, CS1 was presented; then, 25 min later, CS2 was presented. Brains were removed 5 min later. *arc*<sup>+</sup> neurons were activated by CS2, *h1a*<sup>+</sup> neurons were activated by CS1, and *arc*<sup>+</sup>/*h1a*<sup>+</sup> neurons were activated by both CSs (part of both engrams). Two control groups trained on event 1 only and tested under two conditions provided upper and lower overlap boundaries. During the test, the CS1-CS1 group was reexposed to CS1 twice (*h1a*<sup>+</sup> = first CS1 reexposure, *arc*<sup>+</sup> = second CS1 reexposure, “overlap ceiling”), whereas the CS1-CS3 group was reexposed to CS1 and novel CS3 (“floor”). The overlap observed in mice trained on event 1 and

event 2 separated by 24 hours was not different from floor conditions, whereas the overlap observed in mice similarly trained with a 6-hour interval was higher, indicating coallocation (Fig. 1C and fig. S3, A to C).

To test whether coallocation functionally links memories, we extinguished event 2 memory (repeatedly presented CS2 without footshock) and examined the effect on event 1 memory. After CS2 extinction, all groups froze less to CS2. However, mice trained with short (6 hours), but not long (24 hours), ITIs also froze less in response to CS1, even though CS1 was not explicitly extinguished (Fig. 1D).

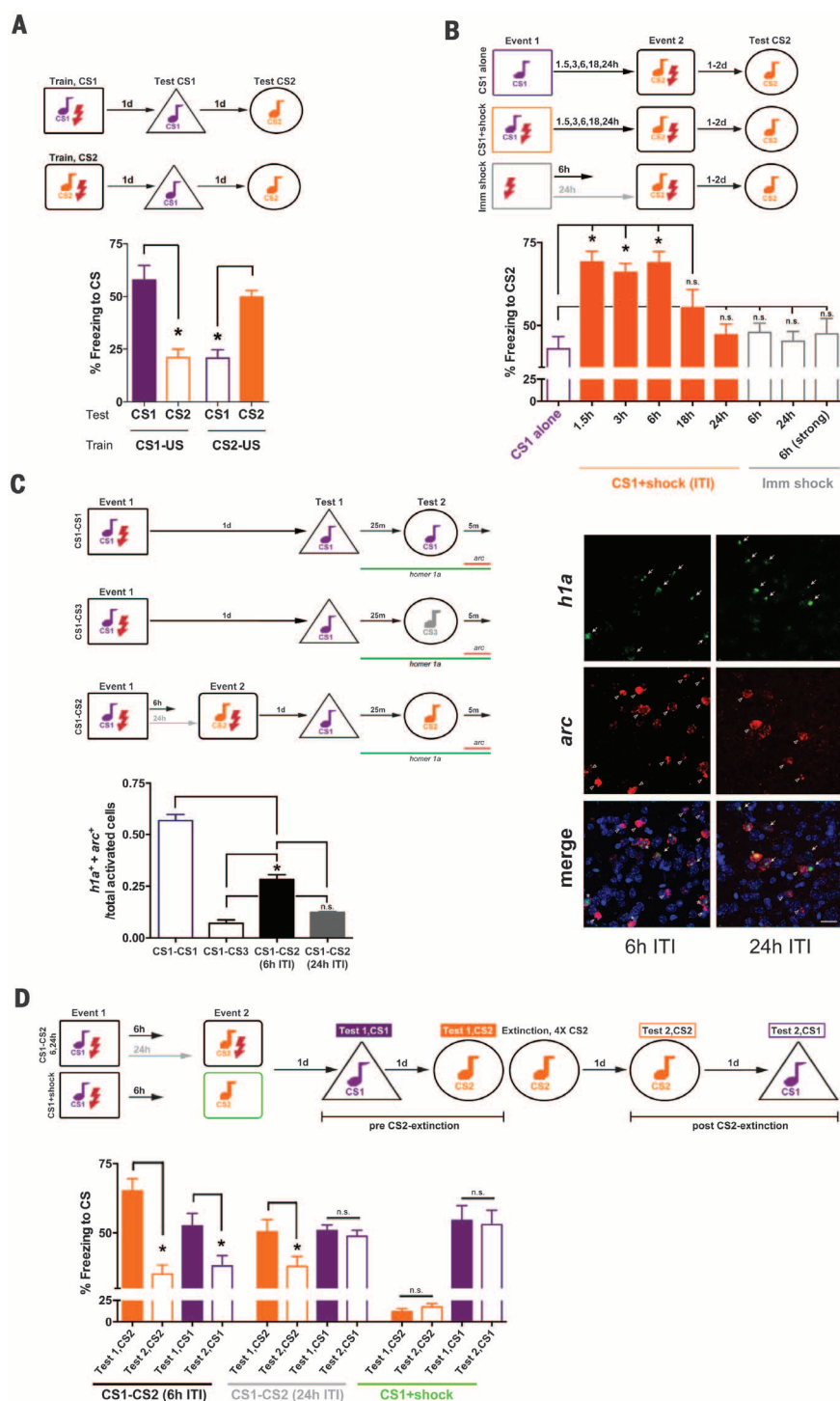
We next examined potential molecular and local circuit mechanisms governing engram interaction. Fear conditioning transiently (1.5 to 6 hours, but not 18 to 24 hours) increased the number of LA principal neurons with activated CREB (Fig. 2A), in agreement with reports showing that learning transiently increases neuronal excitability (13, 14). CREB function cycles over time, in part, due to autoregulatory feedback (15) and, consistent with this, we observed that a depolarizing stimulus in cultured neurons first activated CREB, then increased an inhibitory CREB isoform (fig. S4).

The oscillation in CREB function and neuronal excitability provides a potential mechanism by which engrams supporting events occurring closely in time are coallocated, whereas events occurring at more remote time points are disallocated (Fig. 2B) (16). Although the size of an LA engram supporting a fear memory remains stable, a competitive process determines which neurons comprise each engram (7–10). If event 1 transiently increases CREB function and neuronal excitability in a population of LA neurons and event 2 occurs while these neurons are more excitable than their neighbors (within 6 hours), then overlapping neurons would be coallocated to both events. However, if event 2 occurs when event 1 neurons are no longer more excitable (e.g., 24 hours after event 1), the engram underlying event 2 is disallocated, thus functionally disambiguating these memories. We tested these predictions by manipulating LA neuronal excitability to control allocation. We sought to force coallocation to artificially link normally separated memories (24-hour interval) and force disallocation to artificially separate normally linked memories (6-hour interval).

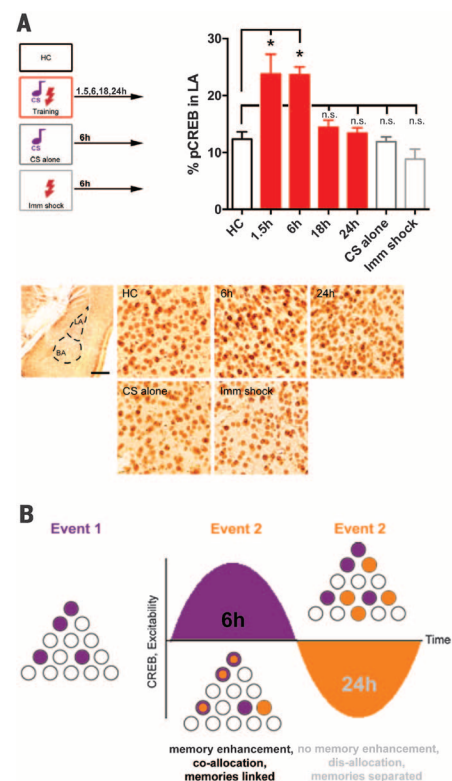
To bidirectionally manipulate excitability in the same neuron, we constructed a replication-defective herpes simplex viral vector (HSV) expressing both channelrhodopsin-2 (ChR2, excitatory opsin) and halorhodopsin (NpHR3.0, inhibitory opsin) [HSV-NpACY (17, 18)]. Blue light (via ChR2) increases excitability, whereas red light (via NpHR3.0) inhibits activity (Fig. 3A and figs. S5 and S7). HSV infects ~15% of LA principal neurons (10) (Fig. 3A and fig. S6). We verified this approach using one fear-conditioning event. To allocate HSV-infected neurons, we excited them (blue light, BL+) before training (10). To confirm their allocation, we tested mice under basal conditions (no red light, RL-) and while inhibiting infected neurons (red light, RL+). Mice trained immediately after activation of infected neurons froze less when these

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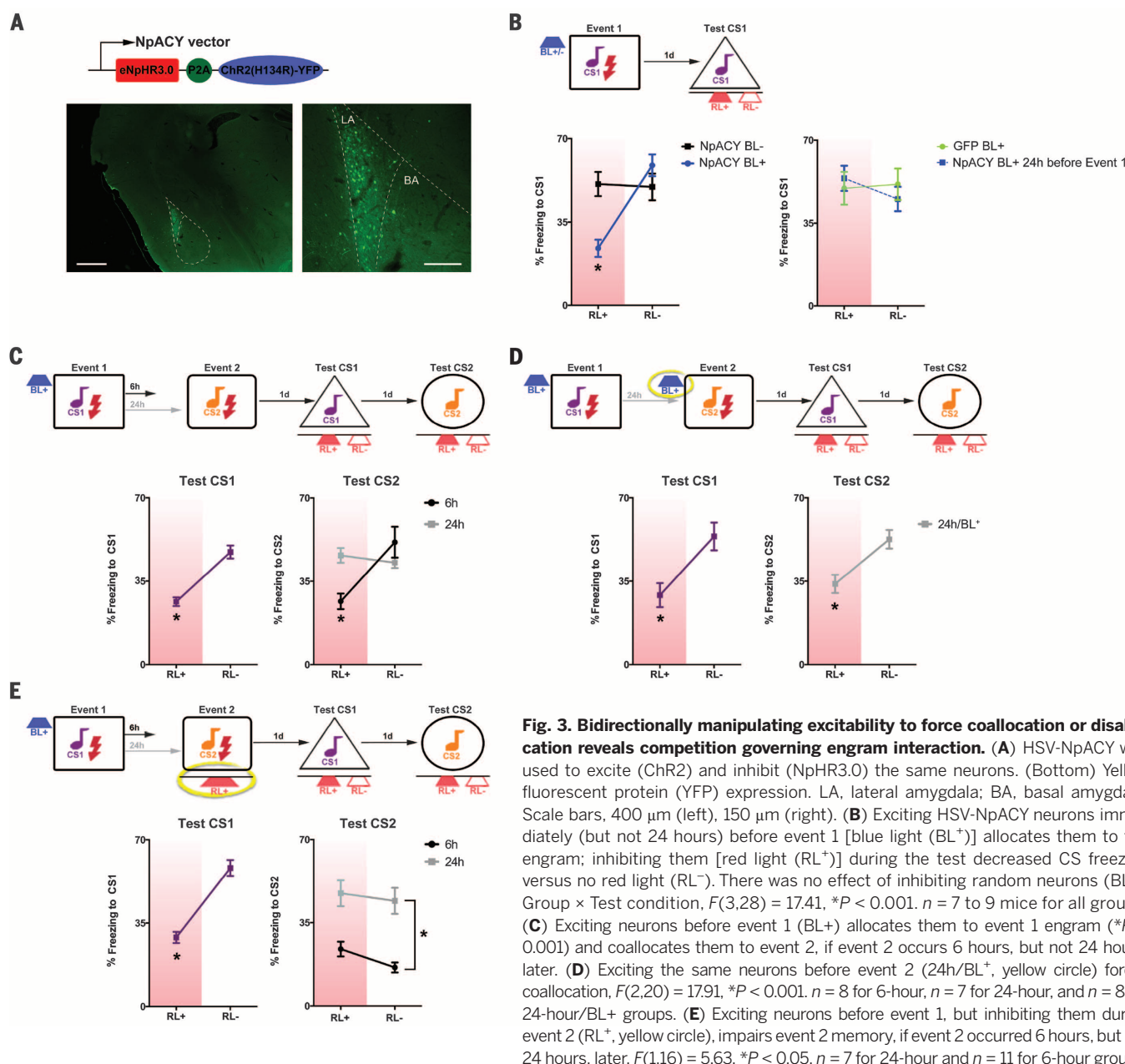
**Fig. 1. Events occurring closely in time are coallocated to overlapping engrams, and memories become linked.** (A) Mice distinguish conditioned stimuli (CS1, CS2), freezing more in response to trained, than untrained, auditory CS. ANOVA: Train-CS × Test-CS,  $F(1,12) = 44.29$ ,  $*P < 0.001$ . Means ± SEM.  $n = 7$  mice per group.  $*P < 0.05$ . (B) Event 2 memory was enhanced if event 1 (CS1+shock) occurred within a short intertraining interval (ITI). Enhancement was not due to CS1 alone or immediate shock (Imm shock) during event 1;  $F(8,142) = 17.03$ ,  $*P < 0.001$ .  $n = 56$  for CS1 alone (see fig. S1A),  $n = 5$  to 16 for other groups. n.s., not statistically different. (C) Engrams for events with a 6-hour (but not 24-hour) ITI coallocated to overlapping LA neurons;  $F(3,8) = 119.01$ ,  $*P < 0.001$ . (Right) Coallocation in 6-hour, but not 24-hour, ITI.  $arc^+$  (CS2 reexposure, red),  $homer1a^+$  ( $h1a^+$ , CS1 reexposure, green) neurons, 4',6-diamidino-2-phenylindole (DAPI; blue, nuclear stain). Scale bar, 20  $\mu$ m.  $n = 3$  mice per group. (D) Extinguishing event 2 memory also decreased event 1 memory if a 6-hour (but not 24-hour) ITI was used. Group × CS-Freezing,  $F(2,50) = 12.53$ ,  $*P < 0.001$ .  $n = 12$  mice, 6-hour, 24-hour groups;  $n = 4$  mice, control group.



**Fig. 2. Engram interaction may be governed by neuronal CREB function and excitability.** (A) (Top) Fear conditioning transiently increases the percentage of LA neurons with activated CREB (pCREB) relative to home-cage (HC), CS alone (6h), or immediate shock (6h, Imm shock);  $F(6,21) = 12.53$ ,  $*P < 0.001$ . (Bottom) Left panel: LA. Scale bar, 400  $\mu$ m. Right panels: pCREB staining. Scale bar, 25  $\mu$ m.  $n = 3$  to 5 mice per group. (B) Schematic of hypothesized engram interaction. Event 1 transiently increases CREB function and excitability in a population of LA neurons (purple). If event 2 occurs when these neurons have elevated CREB and excitability (6h), then engrams are coallocated, memories are linked (purple+orange neurons), and event 2 memory is enhanced. If event 2 occurs later (24h), neurons activated by event 1 are no longer more excitable (perhaps they are in a “refractory-like period”), engrams are disallocated to nonoverlapping neurons, and memories are distinct.

neurons were inhibited (Fig. 3B), indicating that optogenetically exciting neurons before training was sufficient for allocation. Inhibiting a similar number of neurons in control groups (BL-, BL+ 24 hours before training, BL+ but no opsin) did not affect memory expression, nor did BL+ affect training variables or function as a cue during the test (fig. S8B).

We confirmed time-limited coallocation of two events. Principal neurons expressing HSV-NpACV were excited immediately before event 1. Infected neurons were coallocated to event 2, if event 2 occurred 6 hours, but not 24 hours, later (Fig. 3C). To artificially link normally separated memories (24-hour interval), we excited the same population of HSV-infected neurons before event 1 and event 2, forcing coallocation. Inhibiting infected neurons



**Fig. 3. Bidirectionally manipulating excitability to force coallocation or disallocation reveals competition governing engram interaction.** (A) HSV-NpACY was used to excite (ChR2) and inhibit (NpHR3.0) the same neurons. (Bottom) Yellow fluorescent protein (YFP) expression. LA, lateral amygdala; BA, basal amygdala. Scale bars, 400  $\mu$ m (left), 150  $\mu$ m (right). (B) Exciting HSV-NpACY neurons immediately (but not 24 hours) before event 1 [blue light (BL<sup>+</sup>)] allocates them to the engram; inhibiting them [red light (RL<sup>+</sup>)] during the test decreased CS freezing versus no red light (RL<sup>-</sup>). There was no effect of inhibiting random neurons (BL<sup>-</sup>). Group  $\times$  Test condition,  $F(3,28) = 17.41$ ,  $*P < 0.001$ .  $n = 7$  to 9 mice for all groups. (C) Exciting neurons before event 1 (BL<sup>+</sup>) allocates them to event 1 engram ( $*P < 0.001$ ) and coallocates them to event 2, if event 2 occurs 6 hours, but not 24 hours, later. (D) Exciting the same neurons before event 2 (24h/BL<sup>+</sup>, yellow circle) forces coallocation,  $F(2,20) = 17.91$ ,  $*P < 0.001$ .  $n = 8$  for 6-hour,  $n = 7$  for 24-hour, and  $n = 8$  for 24-hour/BL<sup>+</sup> groups. (E) Exciting neurons before event 1, but inhibiting them during event 2 (RL<sup>+</sup>, yellow circle), impairs event 2 memory, if event 2 occurred 6 hours, but not 24 hours, later.  $F(1,16) = 5.63$ ,  $*P < 0.05$ .  $n = 7$  for 24-hour and  $n = 11$  for 6-hour groups.

impaired both event 1 and event 2 memory expression (Fig. 3D), indicating overlapping engrams of infected neurons. Similarly, we artificially linked normally separated memories (24-hour interval) by virally overexpressing CREB (vCREB) to excite the same neurons during event 1 and event 2. Silencing vCREB-infected neurons with the inhibitory DREADD receptor hM4Di [designer receptors exclusively activated by designer drug (19)] disrupted expression of both memories (fig. S9, A to D).

To artificially separate normally linked memories (6-hour interval), we excited infected neurons before event 1, then inhibited (RL<sup>+</sup>) these neurons during event 2. Surprisingly, event 2 memory was impaired, even when mice were tested without red light (Fig. 3E). Event 2 memory was also disrupted if we used vCREB to allocate neurons before event 1 and DREADDs

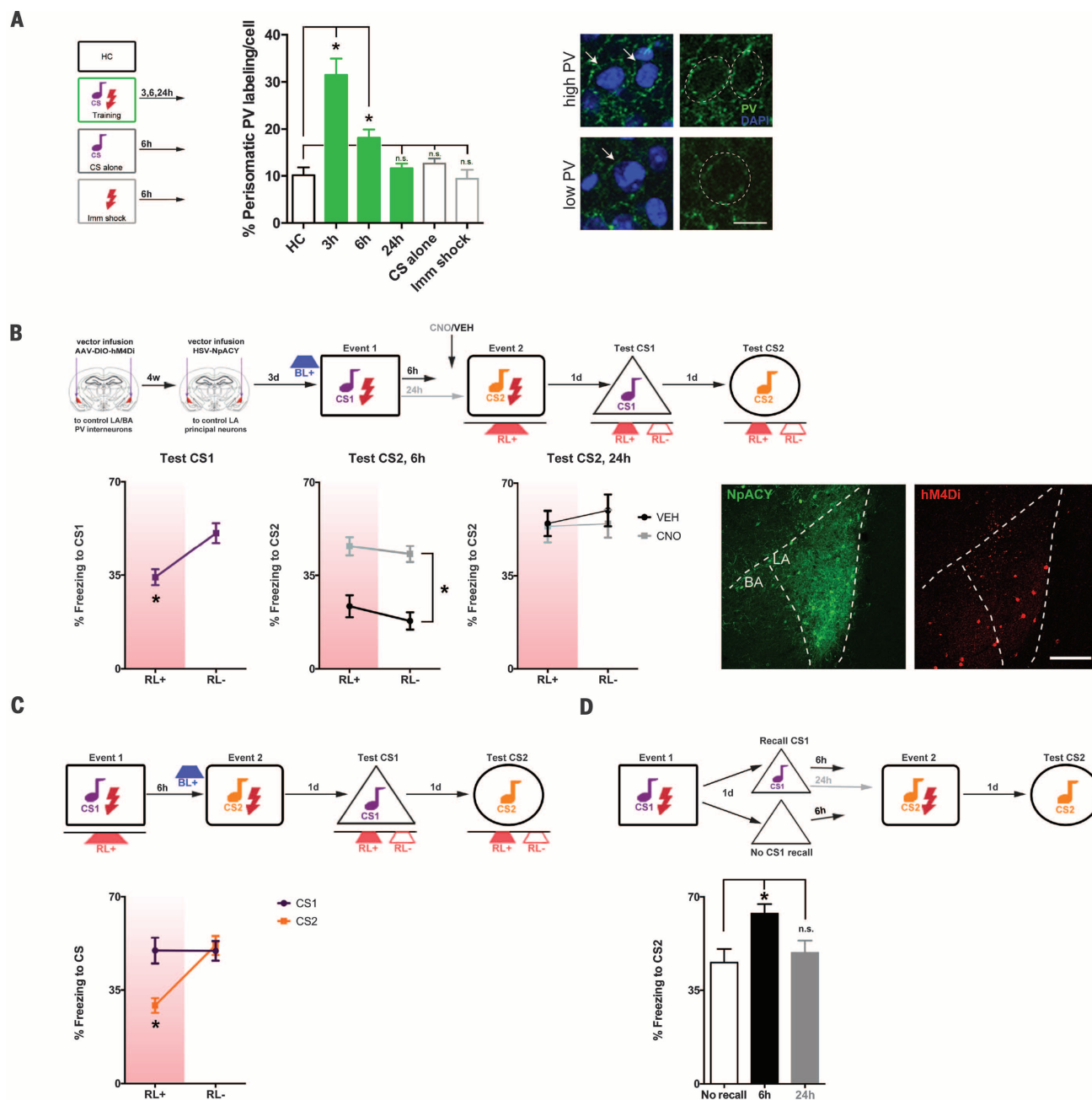
to inhibit these neurons during event 2 (fig. S9E). In contrast, event 2 memory was intact in optogenetic experiments if the interval between events was increased to 24 hours (Fig. 3E). These results indicate that initially allocated neurons transiently prevent nonallocated neurons from being allocated to another engram (winning neurons inhibit losing neurons).

In neural networks, winner-take-all competition maintains sparse coding and is regulated by excitation and inhibition (20). Local LA inhibition is mediated predominantly by  $\gamma$ -aminobutyric acid (GABA)-releasing parvalbumin (PV) interneurons (21, 22). PV interneurons in LA and basal amygdala (BA) form an inhibitory network receiving inputs from, and regulating, LA principal neuron activity by forming perisomatic synapses (23, 24). Fear conditioning induced a transient increase in periso-

matic PV immunolabeling (24) surrounding LA principal neurons (1.5 to 6 hours but not 18 to 24 hours, Fig. 4A).

To test whether PV interneurons suppress non-allocated principal neurons, we released LA and BA PV inhibition. Transgenic mice expressing Cre recombinase in PV interneurons (PV-Cre mice) were microinjected with adeno-associated virus (AAV) expressing Cre-dependent hM4Di. Control PV-Cre mice with intact PV function (vehicle-treated) similarly showed impaired event 2 memory (Fig. 3E) if HSV-infected neurons allocated to event 1 were optogenetically silenced during event 2 (6-hour interval, Fig. 4B). However, inhibiting PV neurons before event 2 [clozapine-N-oxide (CNO) to activate hM4Di receptors] permitted event 2 memory formation. Therefore, if principal neurons not allocated to the engram supporting event 1 (losers) are





**Fig. 4. Competition-mediating engram interaction depends on excitation-inhibition.** (A) Fear conditioning transiently increases perisomatic PV immunolabeling (green) surrounding LA principal neurons relative to home-cage (HC), CS alone (6h), or immediate shock (6h, Imm shock);  $*P < 0.001$ . (Right) High or low perisomatic PV, with or without DAPI-labeled nuclei (blue, perisomatic region outlined). Scale bar, 10  $\mu\text{m}$ .  $n = 3$  mice per group. Arrows indicate perisomatic PV immunolabel in example cells designated “high PV” or “low PV.” (B) Event 2 memory is impaired (Fig. 3E) if events are separated by 6 hours and neurons allocated to

event 1 are silenced (RL+) during event 2. Inhibiting PV interneurons (hM4Di+CNO) permits event 2 memory formation, group difference only,  $F(3,23) = 22.61$ ,  $*P < 0.001$ . (Right) HSV-NpACY (YFP, green), AAV-hM4Di (mCherry, red). Scale bar, 200  $\mu\text{m}$ .  $n = 4$  to 8 mice for all groups. (C) HSV-infected neurons inhibited (RL+) during event 1 are not allocated to event 1 engram;  $*P > 0.05$ . Exciting these neurons before event 2 (BL+) allocates them to event 2 engram,  $F(1,7) = 51.02$ ,  $*P < 0.001$ .  $n = 8$  mice. (D) Event 2 memory is enhanced if CS1 is recalled 6 hours, but not 24 hours, before event 2;  $F(2,29) = 5.28$ ,  $*P < 0.05$ .  $n = 8$  to 12 mice for all groups.

released from PV inhibition, they may be allocated to an engram for an event occurring shortly after (become winners). Similarly, relieving PV inhibition before event 2 had no effect when events were separated by 24 hours, showing the time dependency of this competition (Fig. 4B).

We examined whether directly manipulating LA principal neuron excitability, rather than

indirectly relieving PV inhibition, overcomes these competition effects. HSV-infected principal neurons were optogenetically inhibited during event 1 (these neurons were not allocated to event 1 engram; Fig. 4C). Event 2 occurred 6 hours later when neurons allocated to event 1 engram would also be coallocated to event 2 engram. However, immediately before event 2, we optogenetically

excited HSV-infected neurons. These “loser” HSV-infected neurons became critical components of the engram supporting event 2, outcompeting previous winning neurons.

These results reveal that, in the LA, a transient competitive process governs the interaction between engrams to integrate memories for events occurring closely in time and distinguish memories

for events occurring farther apart in time. Coallocation is not limited to linking memories at encoding. Memory recall may engage a similar process to link new with old memories. We trained mice on event 1, 2 days before event 2. Event 2 memory was enhanced if event 1 was recalled 6 hours, not 24 hours, before event 2 (Fig. 4D). Here, we find that excitatory-inhibitory balance determines whether memories are bound or, alternately, segregated in the LA. More broadly, these principles provide a foundation for understanding how memories are organized within associative networks.

**Note added in proof:** During final preparation of this manuscript, a notable study showing time-limited coallocation of hippocampal memory traces was published (25).

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## SUPPLEMENTARY MATERIALS

[www.sciencemag.org/content/353/6297/383/suppl/DC1](http://www.sciencemag.org/content/353/6297/383/suppl/DC1)  
Materials and Methods  
Figs. S1 to S9  
References (26–43)

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## BEHAVIORAL ECOLOGY

# Reciprocal signaling in honeyguide-human mutualism

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Greater honeyguides (*Indicator indicator*) lead human honey-hunters to wild bees' nests, in a rare example of a mutualistic foraging partnership between humans and free-living wild animals. We show experimentally that a specialized vocal sound made by Mozambican honey-hunters seeking bees' nests elicits elevated cooperative behavior from honeyguides. The production of this sound increased the probability of being guided by a honeyguide from about 33 to 66% and the overall probability of thus finding a bees' nest from 17 to 54%, as compared with other animal or human sounds of similar amplitude. These results provide experimental evidence that a wild animal in a natural setting responds adaptively to a human signal of cooperation.

In 1588, João dos Santos, a Portuguese missionary in Sofala (in present-day Mozambique), often noticed a small bird flying through cracks in the walls of his mission church and nibbling wax from the candlesticks within. This kind of bird, he wrote, had another peculiar habit of leading men to bees' nests by calling and flying from tree to tree. After the men harvested the honey, the birds would eat the wax combs left behind (1). We now know this species to have been the greater honeyguide *Indicator indicator* and dos Santos's description to have been accurate. Honeyguides eat beeswax and know where bees' nests are located; humans can subdue the bees and open the nest using fire and tools, thus exposing beeswax for the honeyguides and honey for the humans (2). This interaction remains an extremely rare example of mutualism between free-living wild animals and our own species. Here we show that it is also a specific example of reciprocal communication between birds and humans.

Greater honeyguides (Fig. 1A) seeking a human collaborator approach people and give a loud chattering call (audio S1). This call is distinct from their territorial song and is accompanied by referential gestures (3): the bird flies from tree to tree in the direction of the bees' nest until its human follower finds the nest (2, 4). The honeyguide thus directs a signal of the bees' nest location toward humans, and the honey-hunters use this signal to their mutual advantage. In this study, we ask whether honeyguides in turn exploit specialized signals directed at them by humans. We studied greater honeyguides (hereafter "honeyguides") in the Niassa National Reserve in northern Mozambique. This region has been noted for its honey and beeswax production at least since Arab trading times centuries ago (5, 6). The local Yao people still harvest wild honey using traditional methods, and this practice remains economically important.

First, we confirmed that in northern Mozambique, honeyguides give reliable information to human honey-hunters. To test whether guiding behavior accurately indicates the direction of bees' nests and leads to their successful discovery by humans, we trailed honey-hunters following honeyguides and tracked our movements via GPS. A guiding event was defined as a bout of guiding by an individual bird, sometimes involving consecutive journeys to different bees' nests. Each guiding event probably involved a different individual honeyguide, as the study area was 230 km<sup>2</sup>, and the home ranges of individual honeyguides that we measured using radio telemetry did not exceed 1 km<sup>2</sup> and overlapped with one another (7) (fig. S1). 75.3% of guiding events led to the successful discovery by humans of at least one bees' nest [mean ± SE = 1.00 ± 0.08 nests; range = 0 to 3 nests; *n* = 97 events, excluding controls in the experiment discussed below (7)]. 94.6% of nests shown belonged to the honeybee *Apis mellifera*, and the rest to stingless bee species (7). Nests were located 0 to 832 m (median = 152 m, *n* = 84 nests, only considering the first nest per guiding event) from the point where guiding began. Figure 1B shows that the birds' initial flight direction accurately signaled the ultimate location of the bees' nest, corroborating a classic study from Kenya (2). Overall, 74.5% of bees' nests found by humans (*n* = 149) involved the help of a honeyguide.

Second, we asked whether the signals used by human honey-hunters provide reliable information to honeyguides. Honey-hunters seeking honeyguides often announce their presence with unspecialized sounds such as shouting and chopping wood (4, 8). In some parts of Africa, however, humans also make specialized vocalizations used only when hunting honey. In the Niassa National Reserve (and, more widely, in northern Mozambique and adjacent southern Tanzania), Yao honey-hunters seeking and following honeyguides produce a loud trill followed by a grunt: "brrrr-hm" [audio S1; see (9) for a melodious whistle used in the same context by the Hadza people of northern Tanzania]. To confirm that "brrrr-hm" is a specialized honey-hunting sound, we interviewed 20 Yao honey-hunters, all of whom reported that they used this specific sound when hunting honey but in no other context. When

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## Competition between engrams influences fear memory formation and recall

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Editor's Summary

### How to link and separate memories

Engrams are the changes in brain tissue that store single memories. Neuroscientists can localize and manipulate them, but until now, little was known about how multiple engrams interact to influence memories. Rashid *et al.* examined how neural assemblies in an area called the lateral amygdala interact. If two frightening events occurred within 6 hours, the same set of neurons was used to express fear memories for both events. However, if the events were separated by 24 hours, distinct memory traces were formed.

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