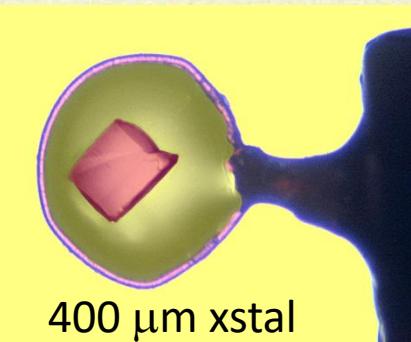
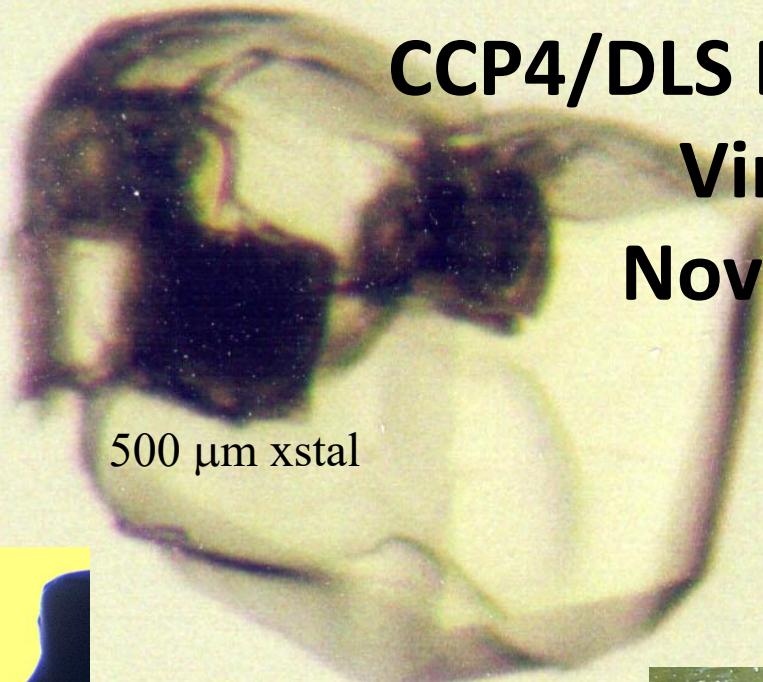


CRYOPROTECTION for CRYO-COOLING: why and how?

CCP4/DLS Data Collection Course

Virtual Intro Day

November 3rd 2025



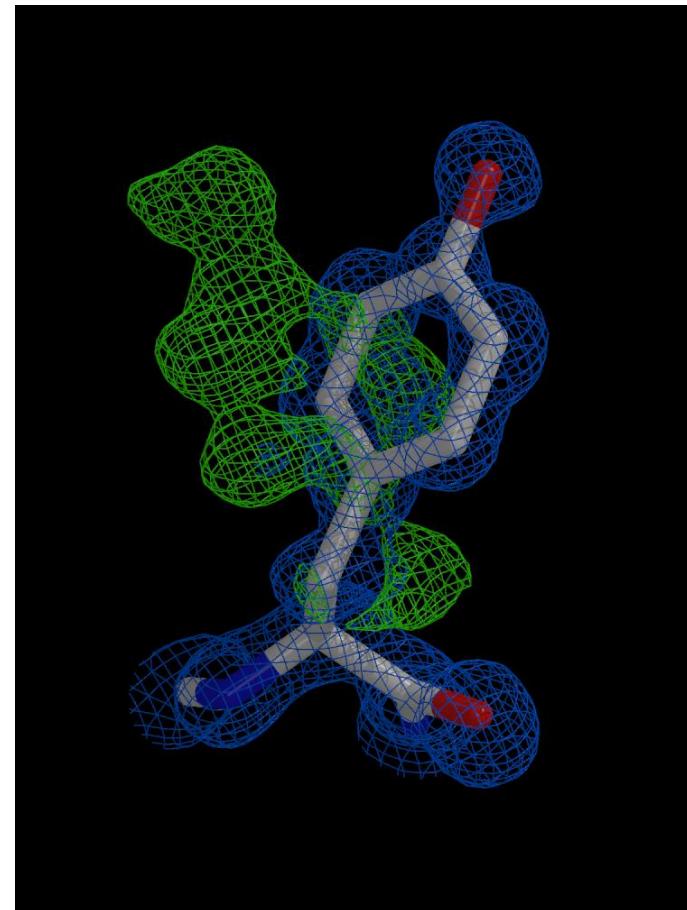
Elspeth Garman,
Biochemistry, Oxford

elspeth.garman@bioch.ox.ac.uk

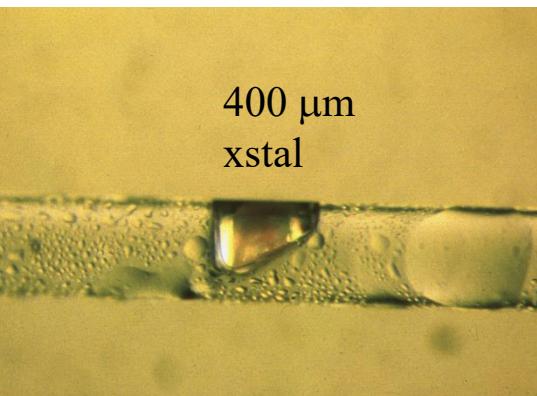


Data quality pivotally affects the amount of biological information we obtain: cryo-cooling is LAST Experimental step which determines ultimate result.

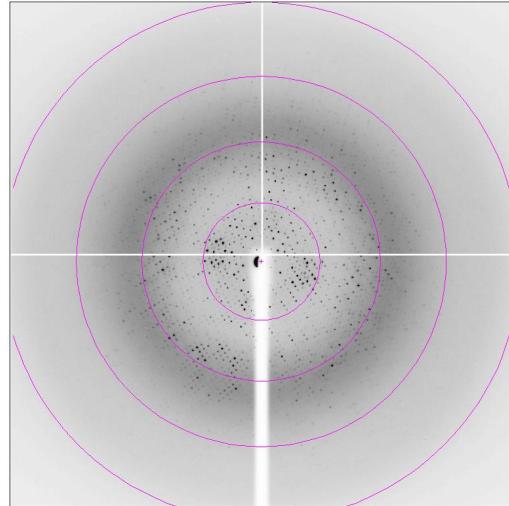
Copyrighted
Gary Larson
Cartoon of cave
man finding
Structure of dirt.



Detailed biological information



The Plan:

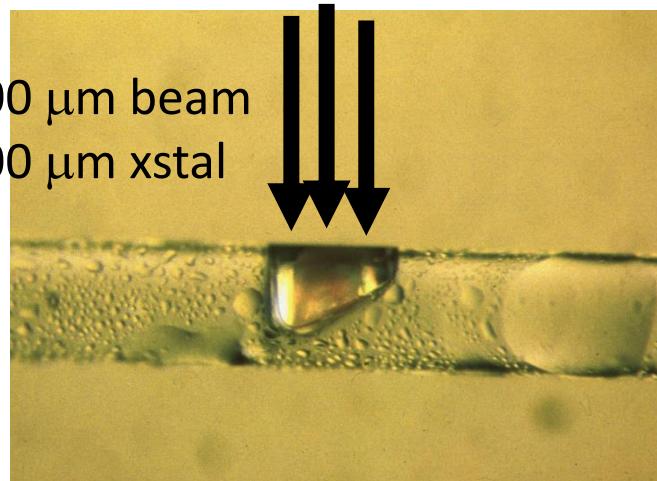


- Cryo techniques
 - **Why cool? Radiation damage.**
 - Optimising cryoprotection.
 - Testing at room temperature.
 - Storage and retrieval.
 - If nothing works...

ROOM TEMPERATURE

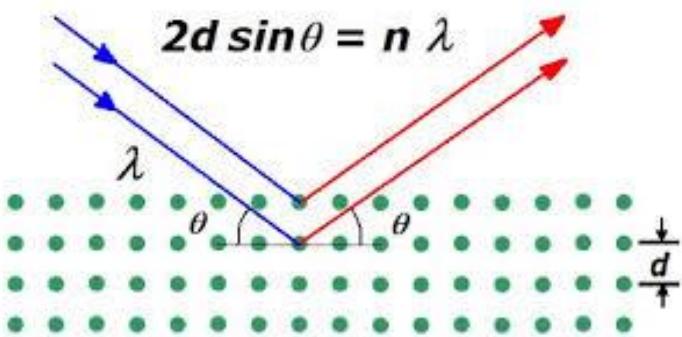
Intensity decrease
with increasing X-ray
exposure

300 μm beam
400 μm xstal

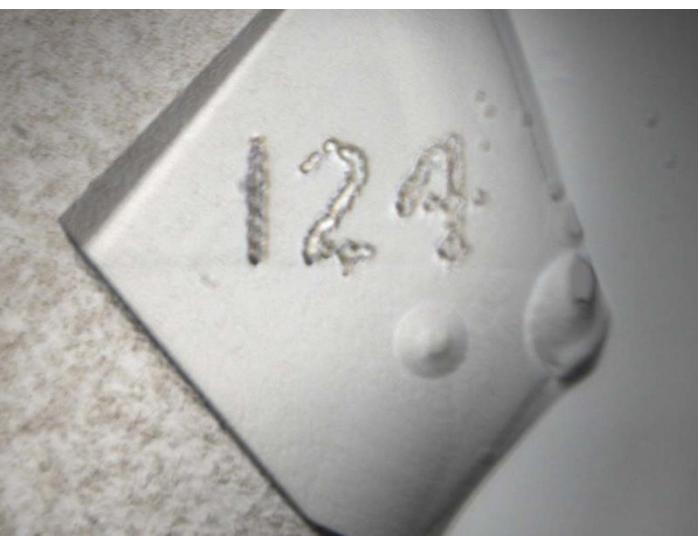
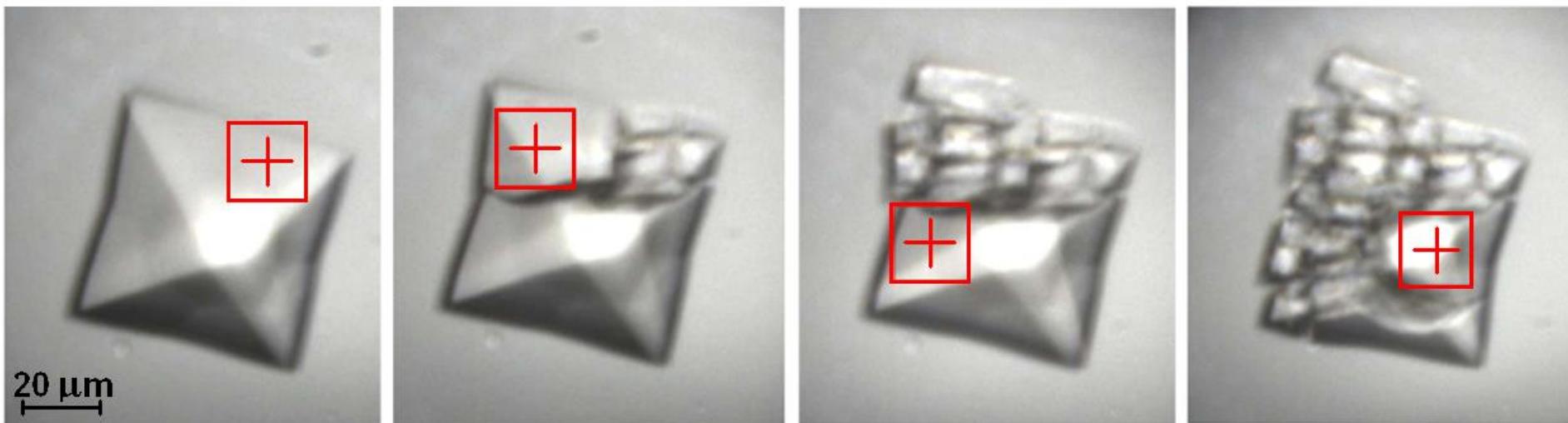


Loss of diffraction
Incomplete data
from crystals at
RT

$$2d \sin \theta = n \lambda$$



I24, Diamond, *in situ* data collection from a
Bovine Enterovirus 2 crystal, room temperature, 0.5 s
20 µm x 20 µm beam

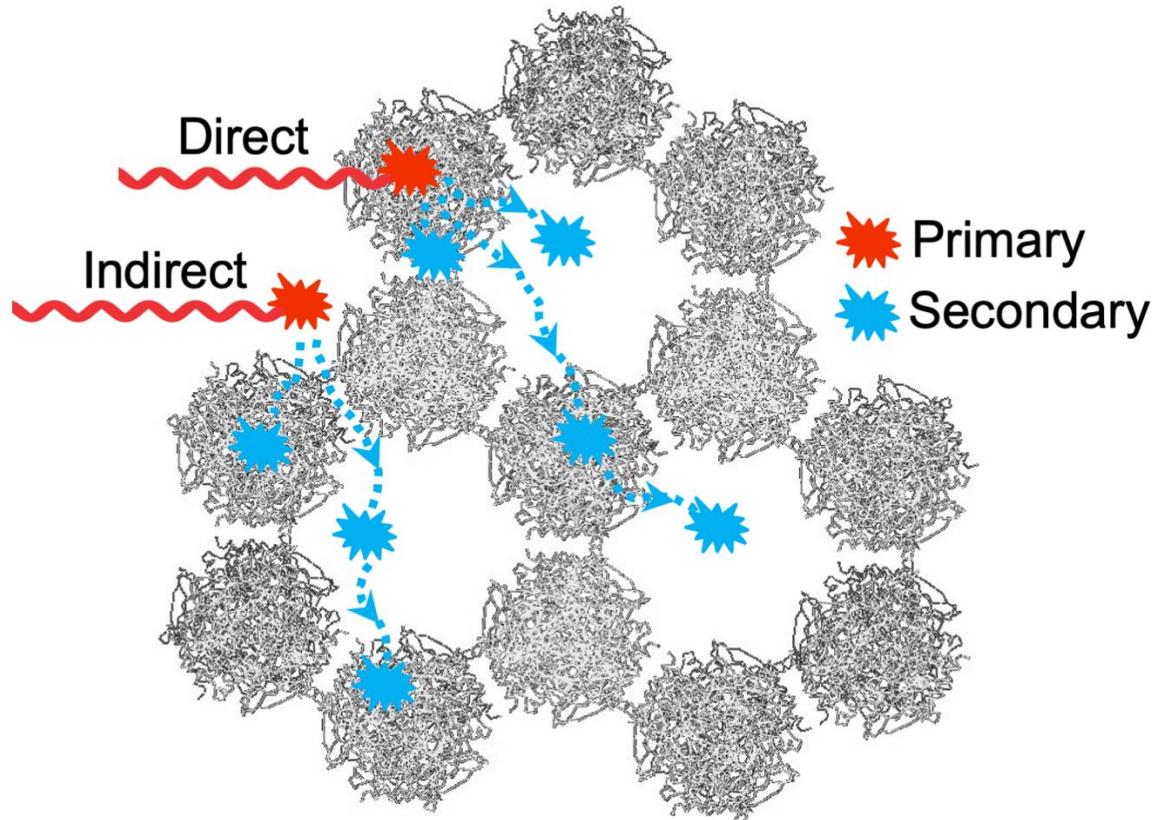
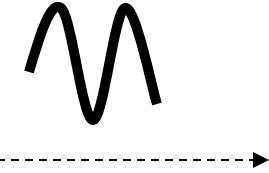


Axford *et al.*,
Acta Cryst D (2012) 592

Beamline logo I24
(Gwyndaf Evans *et al.*)

Radiation damage

Primary
Secondary

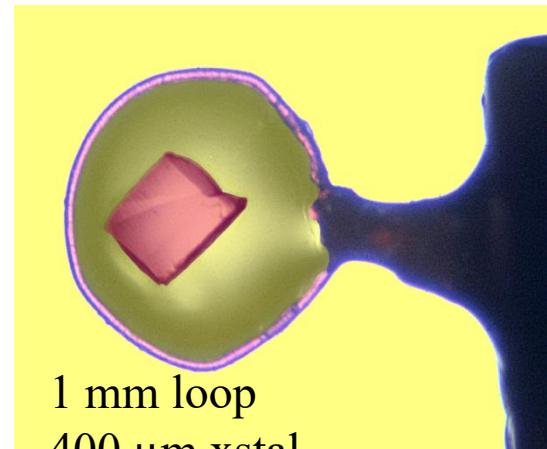
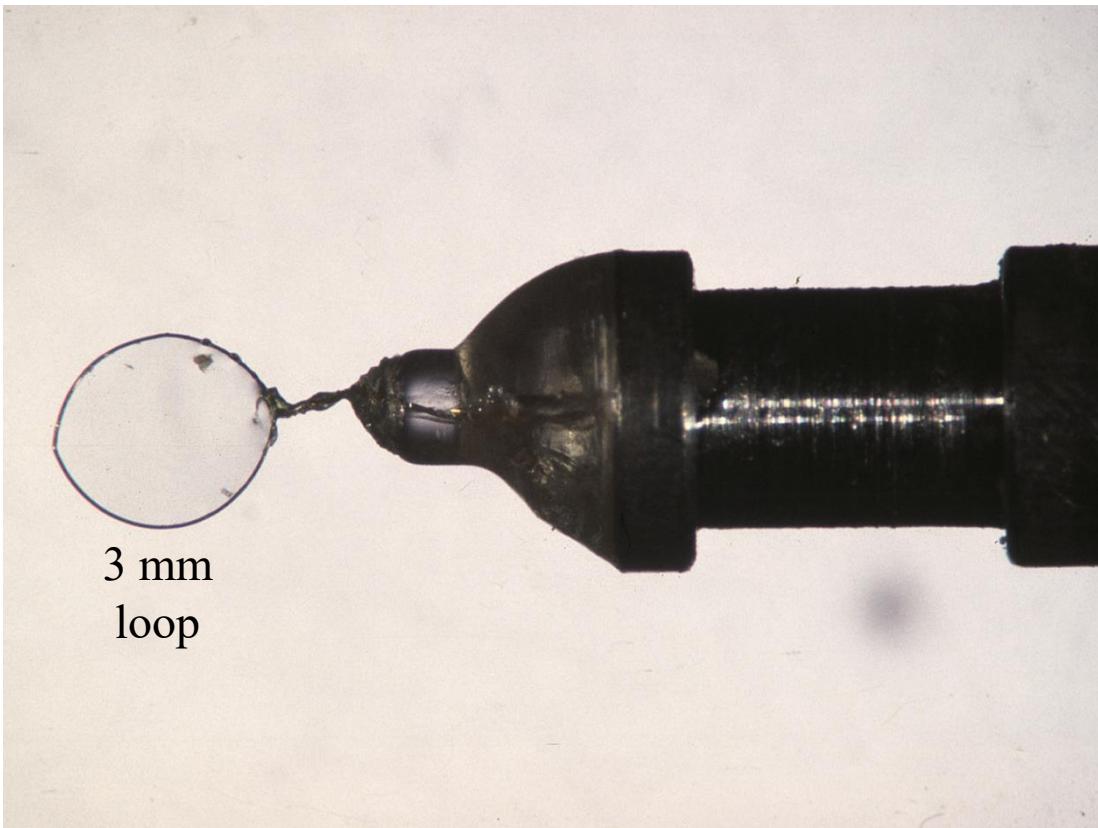


Protein: direct
Solvent: indirect



PRIMARY; inevitable, a fact of physics! Neutralise it?
SECONDARY, can we control it?

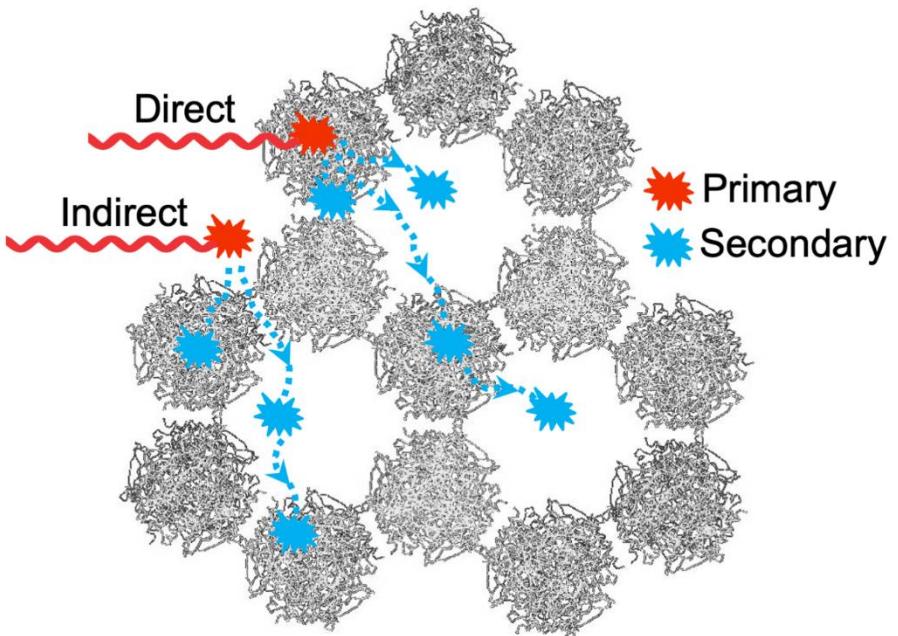
Loop mounting: T-Y.Teng (1990) J.Appl.Cryst, 23, 387-391 [APS!].
Used wire loops



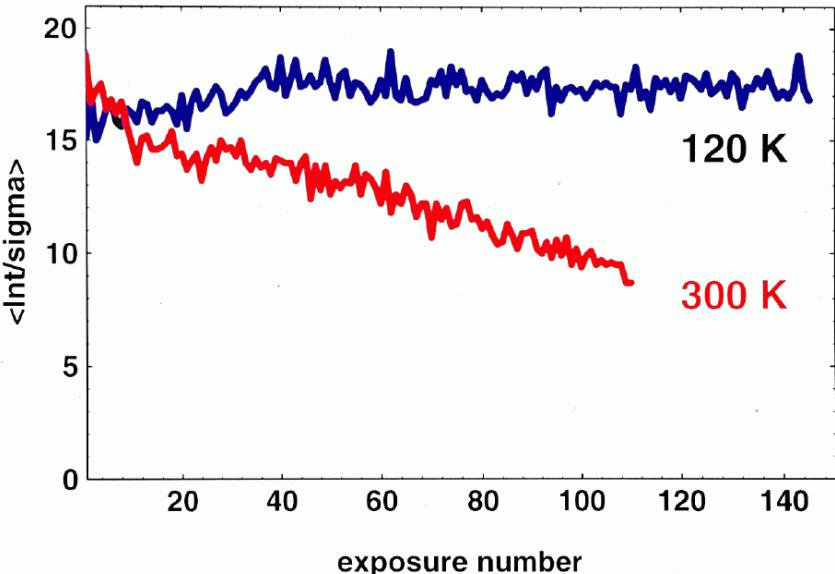
Also, a commercially available and easy to use cryostat (Cosier and Glazer 1986) made the technique accessible to many labs.

[Garman and Schneider, J.Appl.Cryst, (1997) 23]

Radiation damage

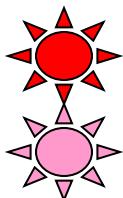
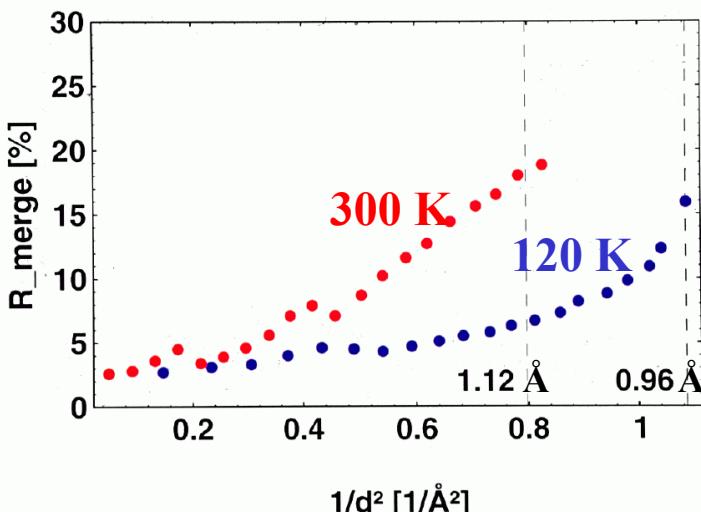


Significantly reduced at
100 K: time factor of ~ 70
[Nave and Garman *JSR* (2005), **12**, 257-260].



SP445: Data Quality

[T.Schneider]



PRIMARY; inevitable, a fact of physics! Proportions?



SECONDARY, can we control it?

Garman and Schneider *J Appl Cryst* 1997

Temperatures of X-ray structures released by the PDB to 26/5/2025



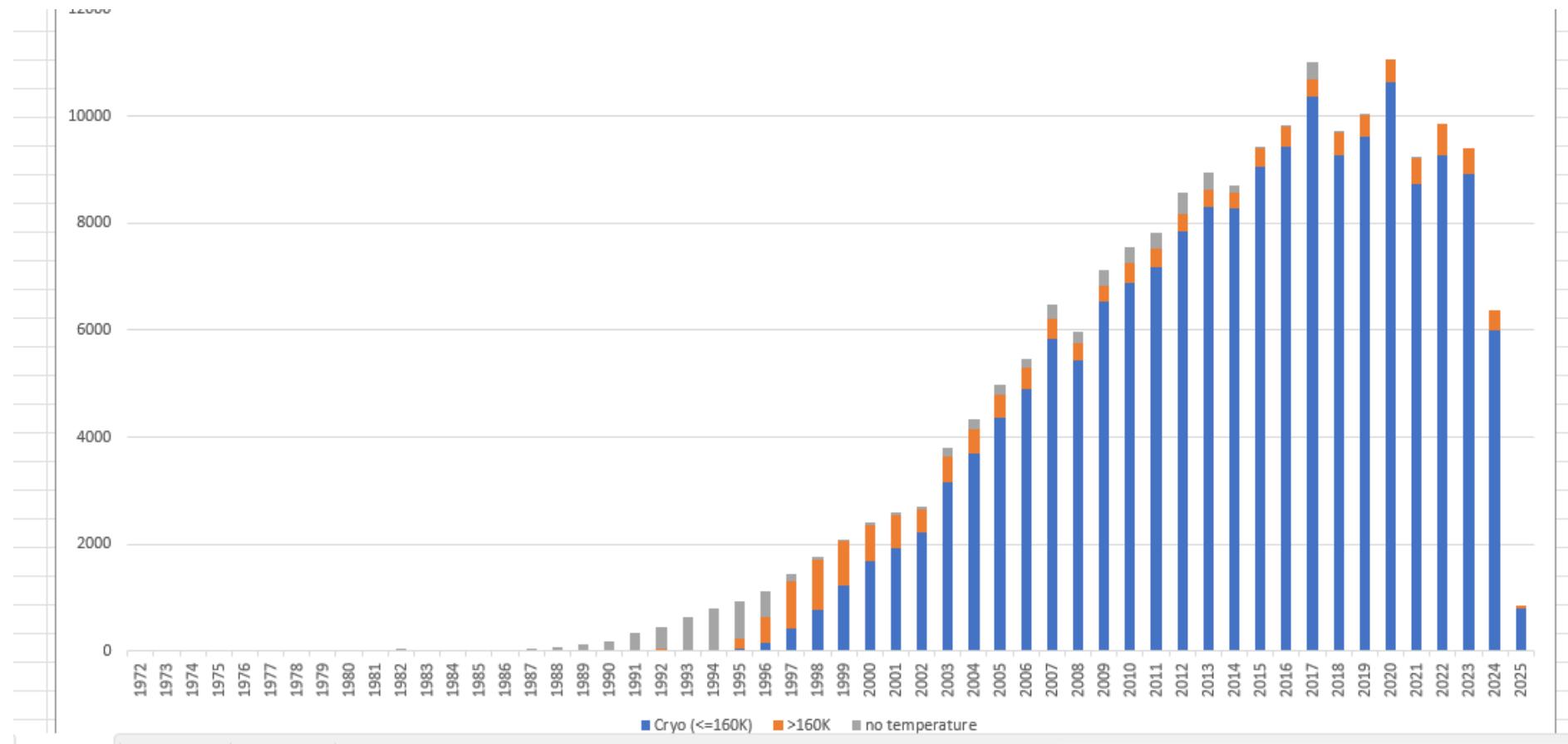
Cryo (<160 K)



>160 K



No T specified



BUT we also see degradation at 100 K
(see RadDam lecture at third virtual day)

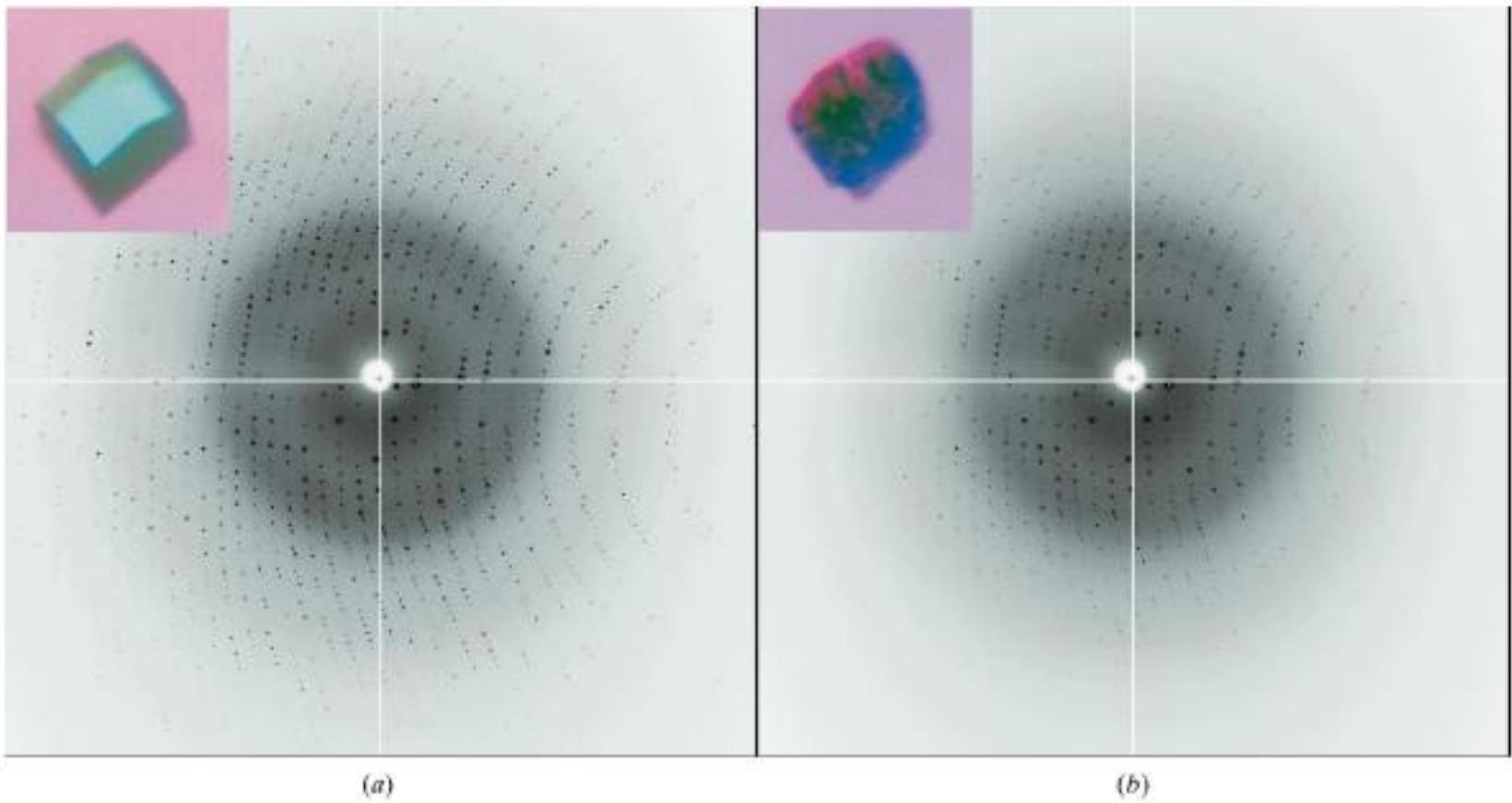
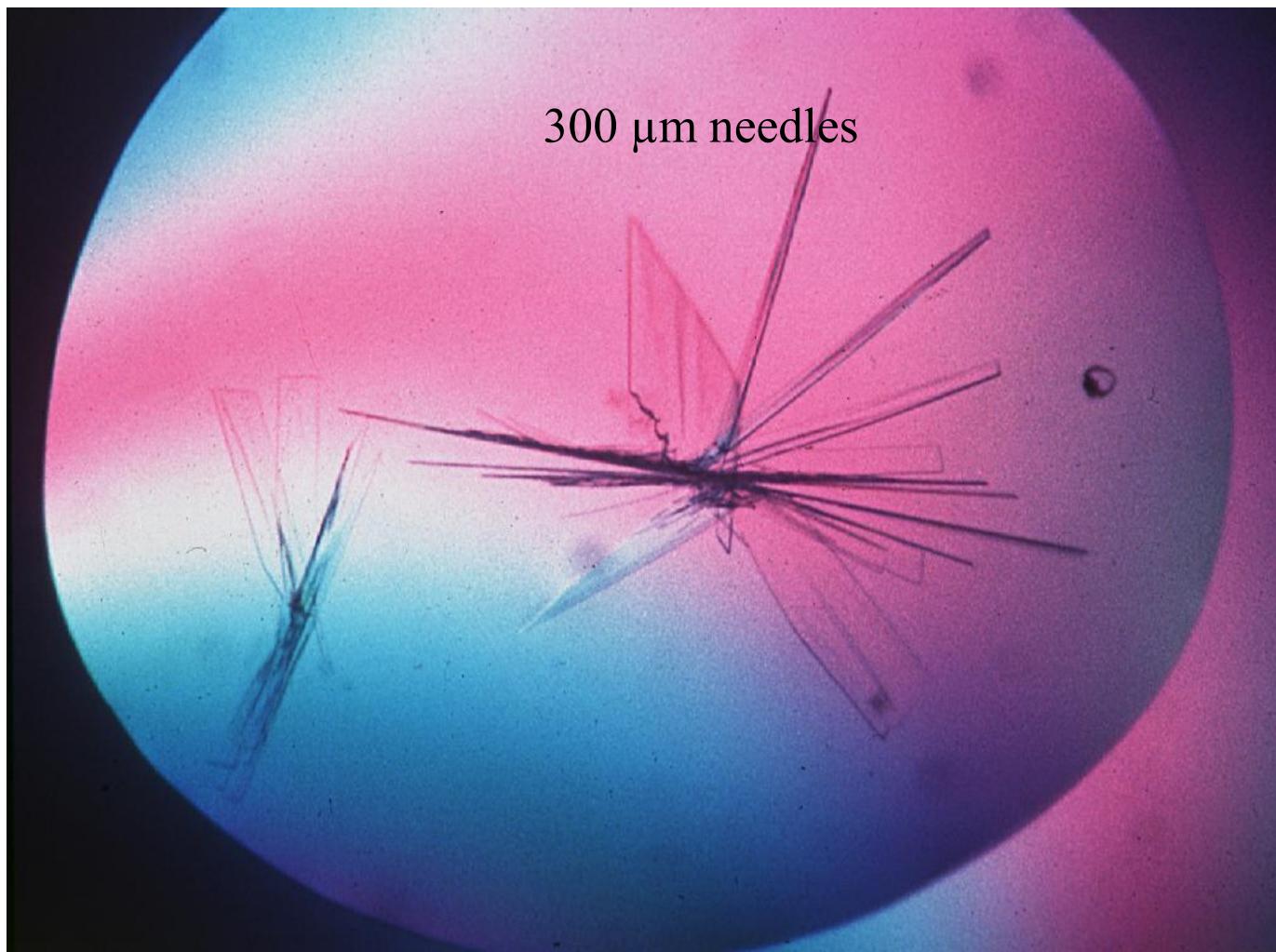


Figure 1

Diffraction images of a lysozyme crystal during a 1.36×10^3 s X-ray exposure at the 14-BM-C beamline. The resolution of diffraction is 1.6 Å at the edge of the image. Two images were taken with identical X-ray dosage. (a) The first image; during its exposure 1.2×10^4 Gy were absorbed. (b) The last image; after accumulating 1.6×10^7 Gy of absorbed energy. The inserts are photomicrographs of the crystal before and after X-ray exposures. The size of this crystal is $\sim 110 \times 110 \times 60$ µm. The crystal was maintained at 100 ± 1 K during the experiment.

Loop mounting is a MUCH gentler technique than capillary mounting.

e.g. cyclin A, 5 μ m x 100 μ m x 300 μ m

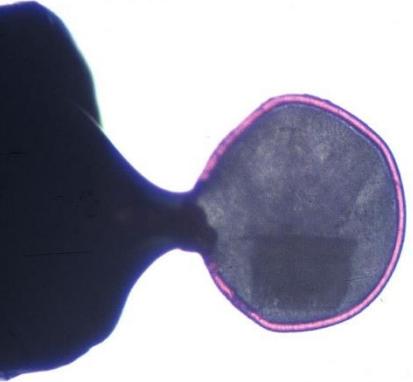


Other advantages:

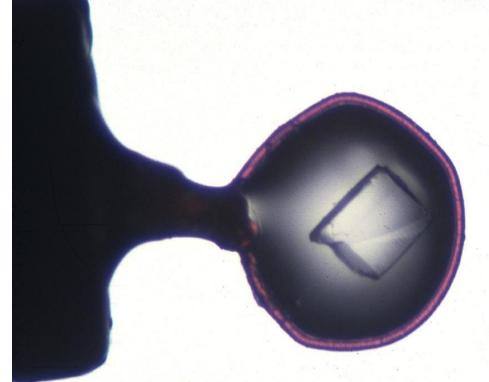
- Usually get a whole data set from a single crystal \Rightarrow higher QUALITY data.
For MAD, the systematic errors are minimised by using only one crystal.
- Can harvest and store crystals while they are in peak condition.
- Small crystals and flat plates can be mounted easily.
- Reduced disorder and thermal movement
- Much lower solvent background.
- No secondary radiation damage during storage.
- New experiments are possible.

NOTE: Crystals cost about the same as diamonds!





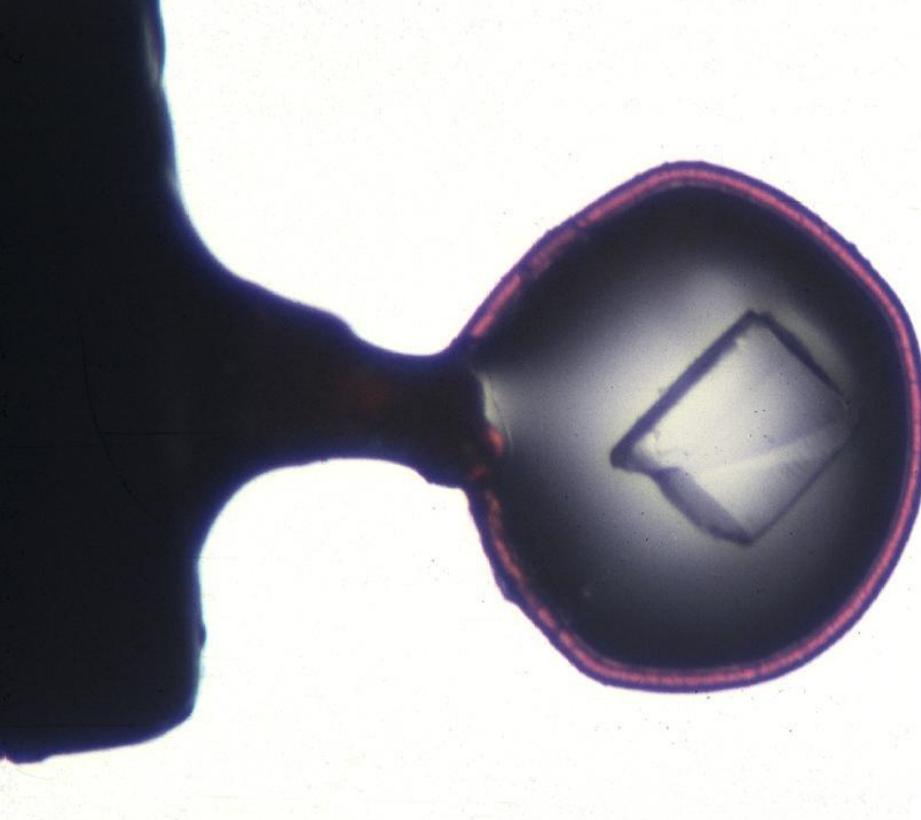
The Plan:



- Cryo techniques
 - Why cool? Radiation damage.
 - **Optimising cryoprotection.**
 - Testing at room temperature.
 - Storage and retrieval.
 - If nothing works...

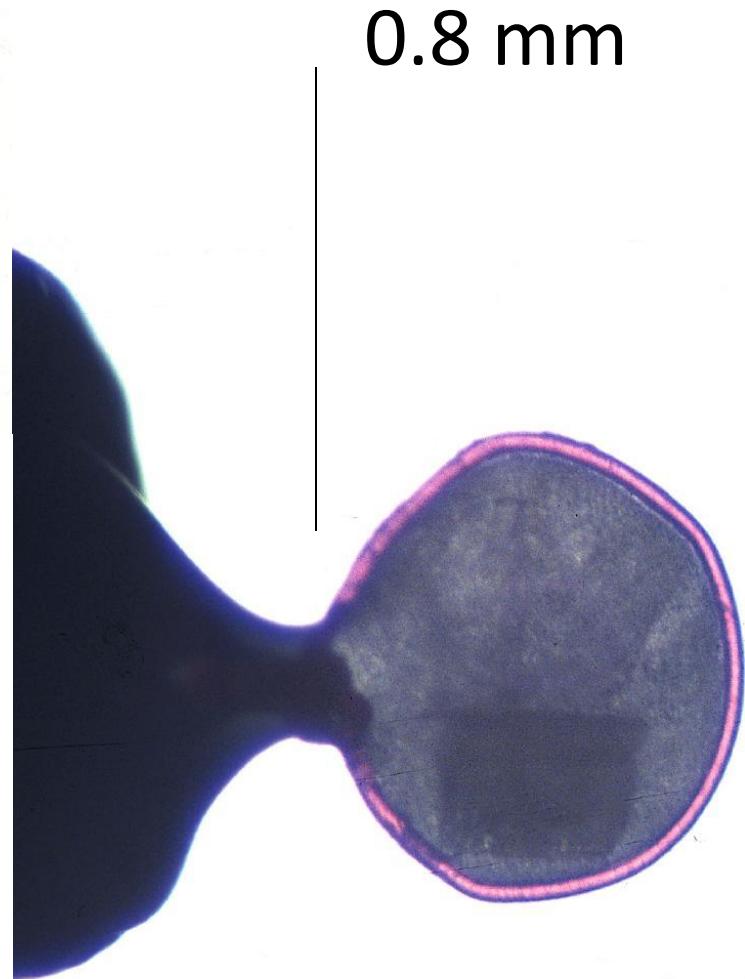
Cryocooling: HOW?

- **COOL** the crystal so fast that the water in the solvent channels is vitrified and does not form crystalline ice. [Pure water: $\sim 10^{-5}$ s for typical protein crystal sized drop]
 - Add ‘antifreeze’ to increase time for cooling process. [1-2 s] i.e. ‘**cryoprotect**’
- N.B. WE DO **NOT** WANT TO ‘FREEZE’ the crystal!
- Collect data at around 100K
[below 130 K and always below 155 K [Weik, 2001]]

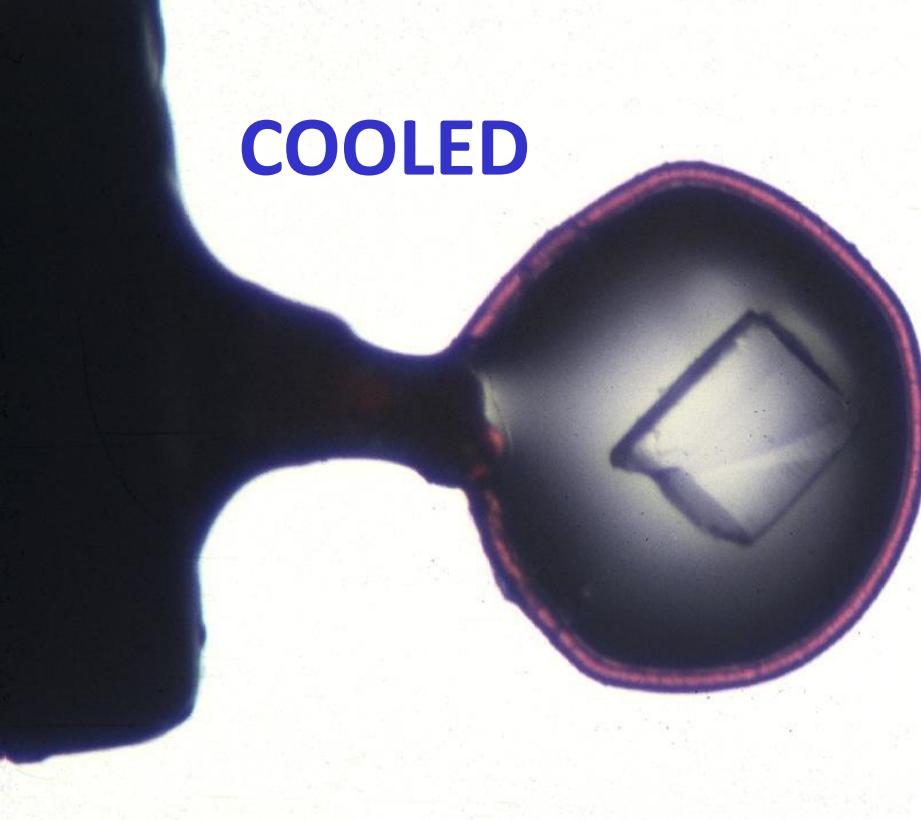


Crystalline ice has 7% more volume than liquid water.

Thus all the solvent channels will expand if no cryoprotectant is added, and crystal order is compromised

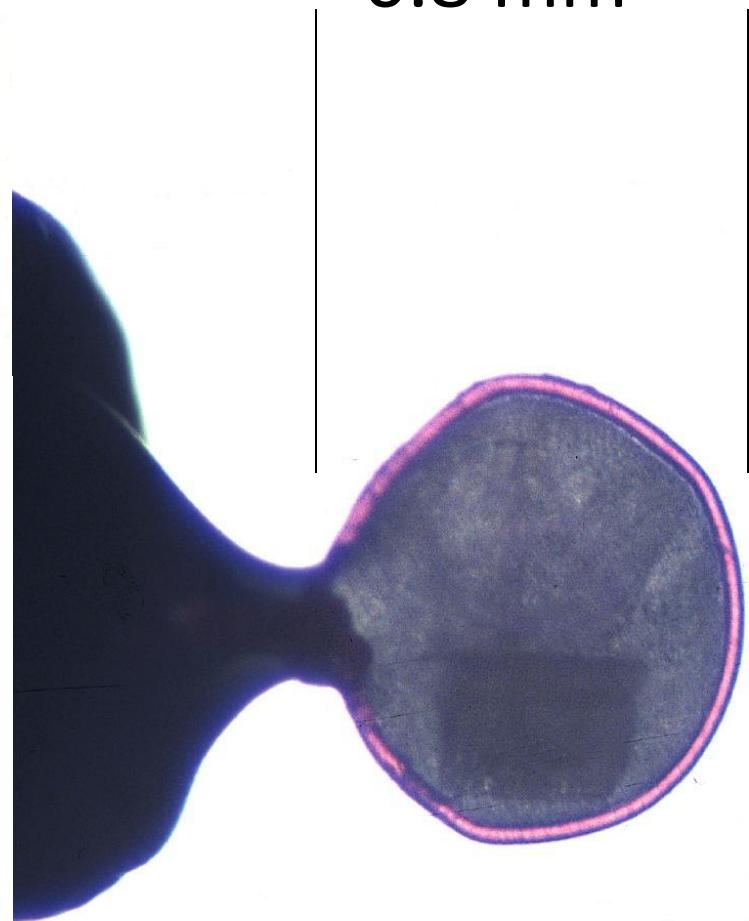


COOLED



FROZEN

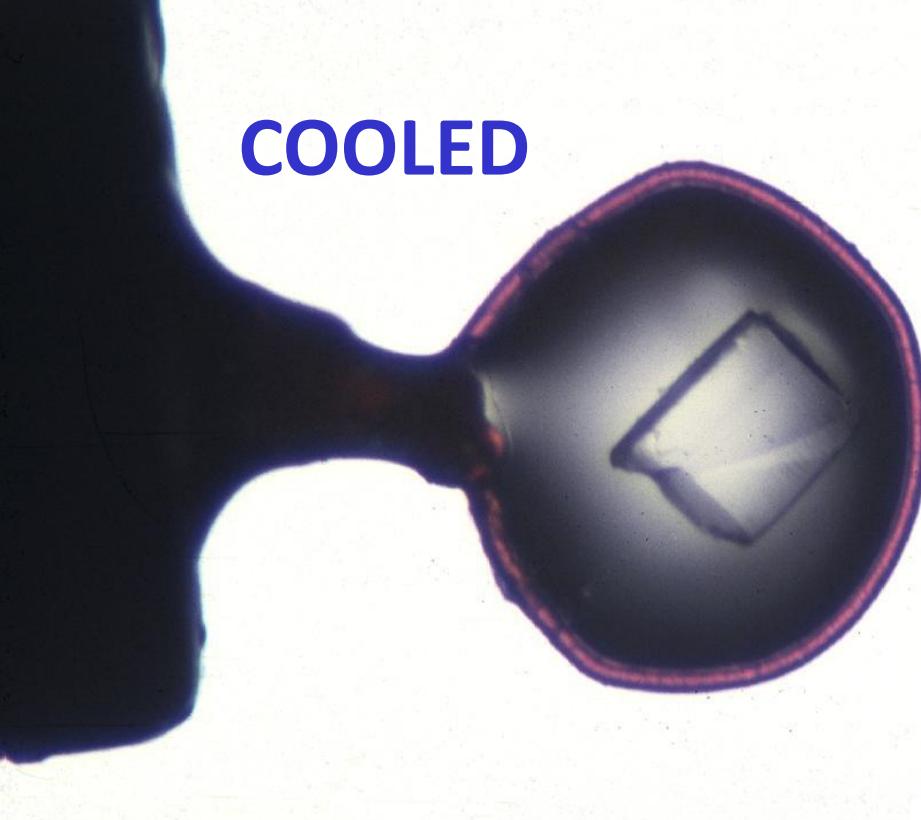
0.8 mm



Crystalline ice has 7% more volume than liquid water.

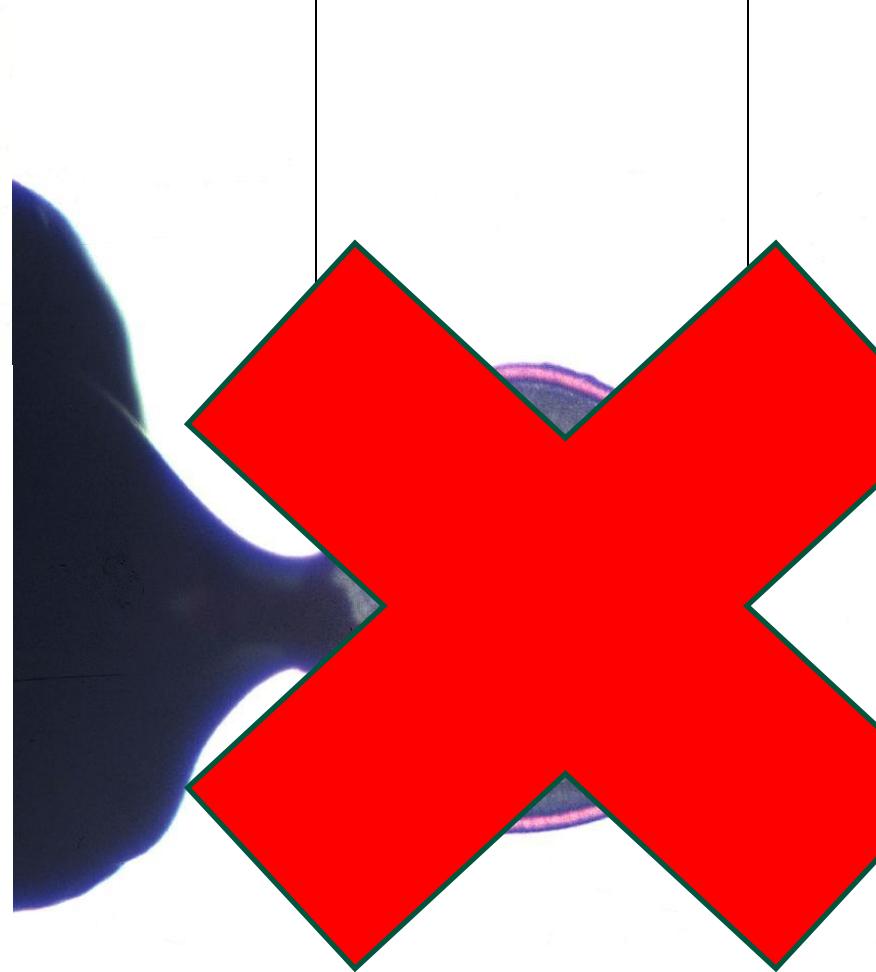
Thus all the solvent channels will expand if no cryoprotectant is added, and crystal order is compromised

COOLED



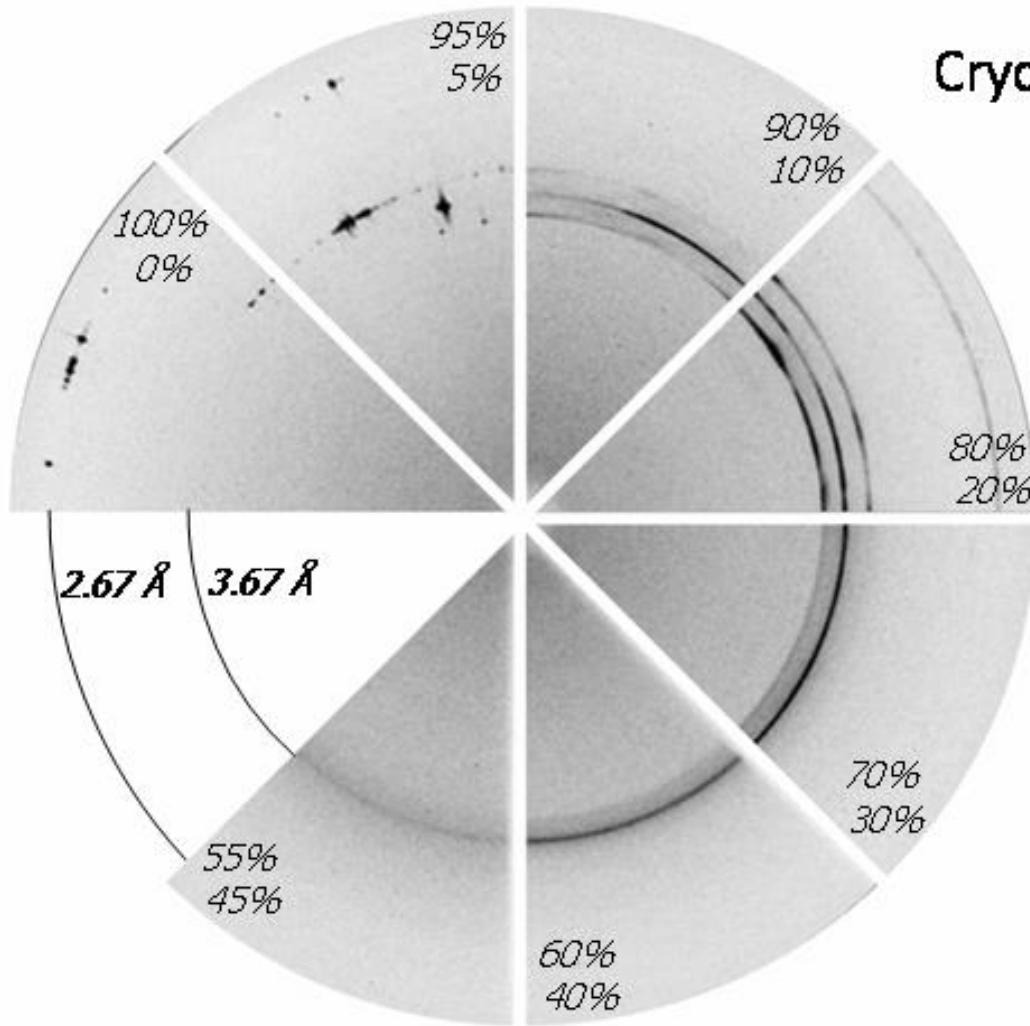
FROZEN

0.8 mm



Crystalline ice has 7% more volume than liquid water.

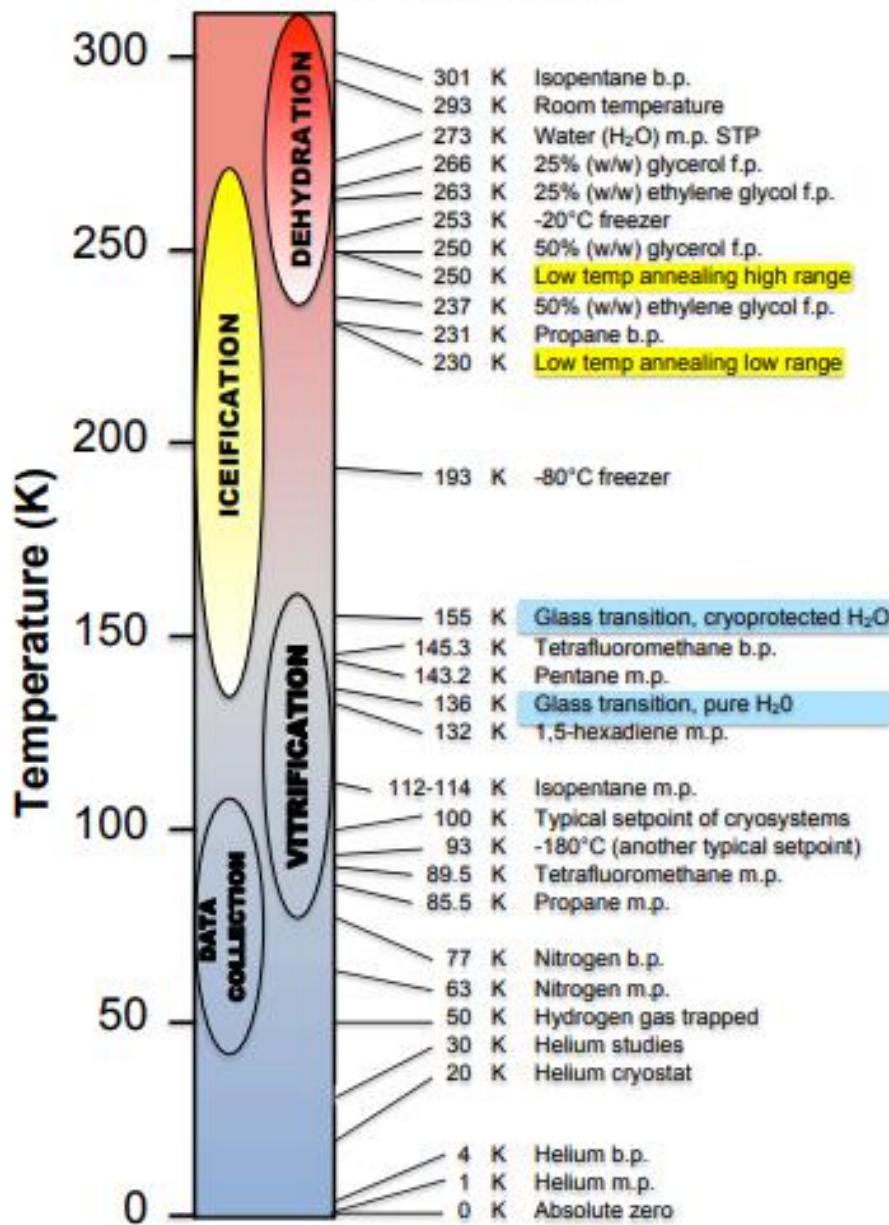
Thus all the solvent channels will expand if no cryoprotectant is added, and crystal order is compromised



Cryo-buffer optimisation

- replace the water in mother liquor with cryo-agent, rather than diluting the mother liquor.
- test cryo-buffer alone in loop.

Notable Temperatures in Cryocrystallography



Cryo-buffers.

- PEG < 4K → increase PEG, add small PEGs
- PEG \geq 4K → add small PEGs
- 2/3rds of cases → add 15 - 25% glycerol
- MPD → harvesting buffer, increase MPD concentration.
- Salt → add MPD and/or ethylene glycol or glycerol
 - increase conc/add salt. Lithium salts good.

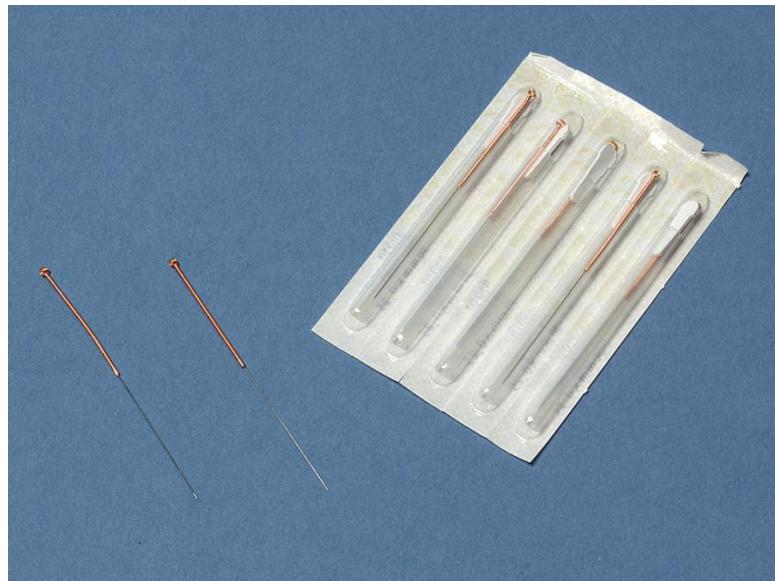
[Rubinson et al, Acta D (2000), D56, 996-1001.]

→ Exchange salt. e.g. 100% 8M Na Formate.

N.B. Low salt needs > concentration of cryoprotectant than high salt.

Sugars, paratone N, combinations, +++

CRYSTAL MANIPULATION:



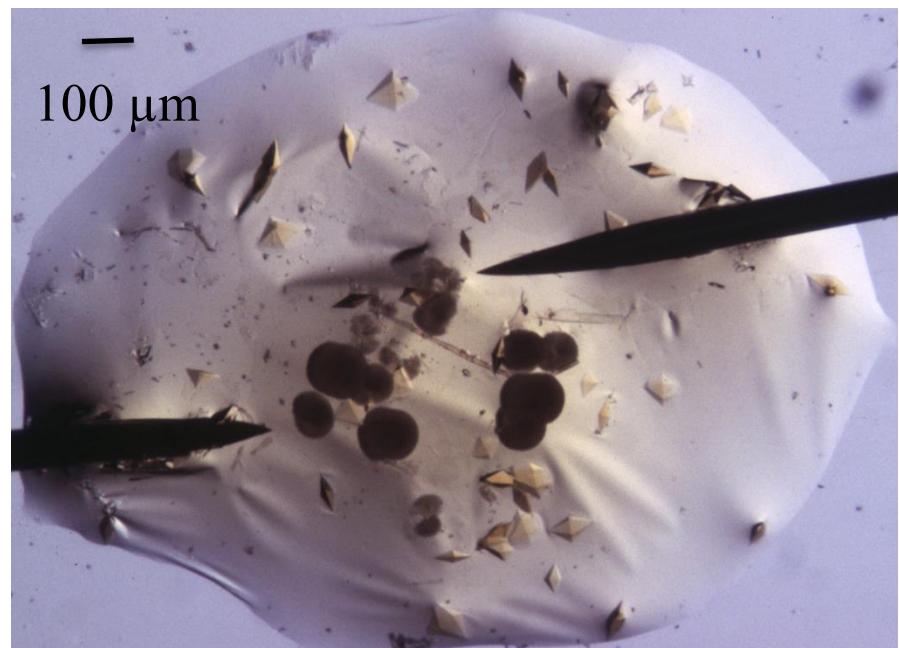
e.g. 'Skin' on protein drop:

Gentle surgery

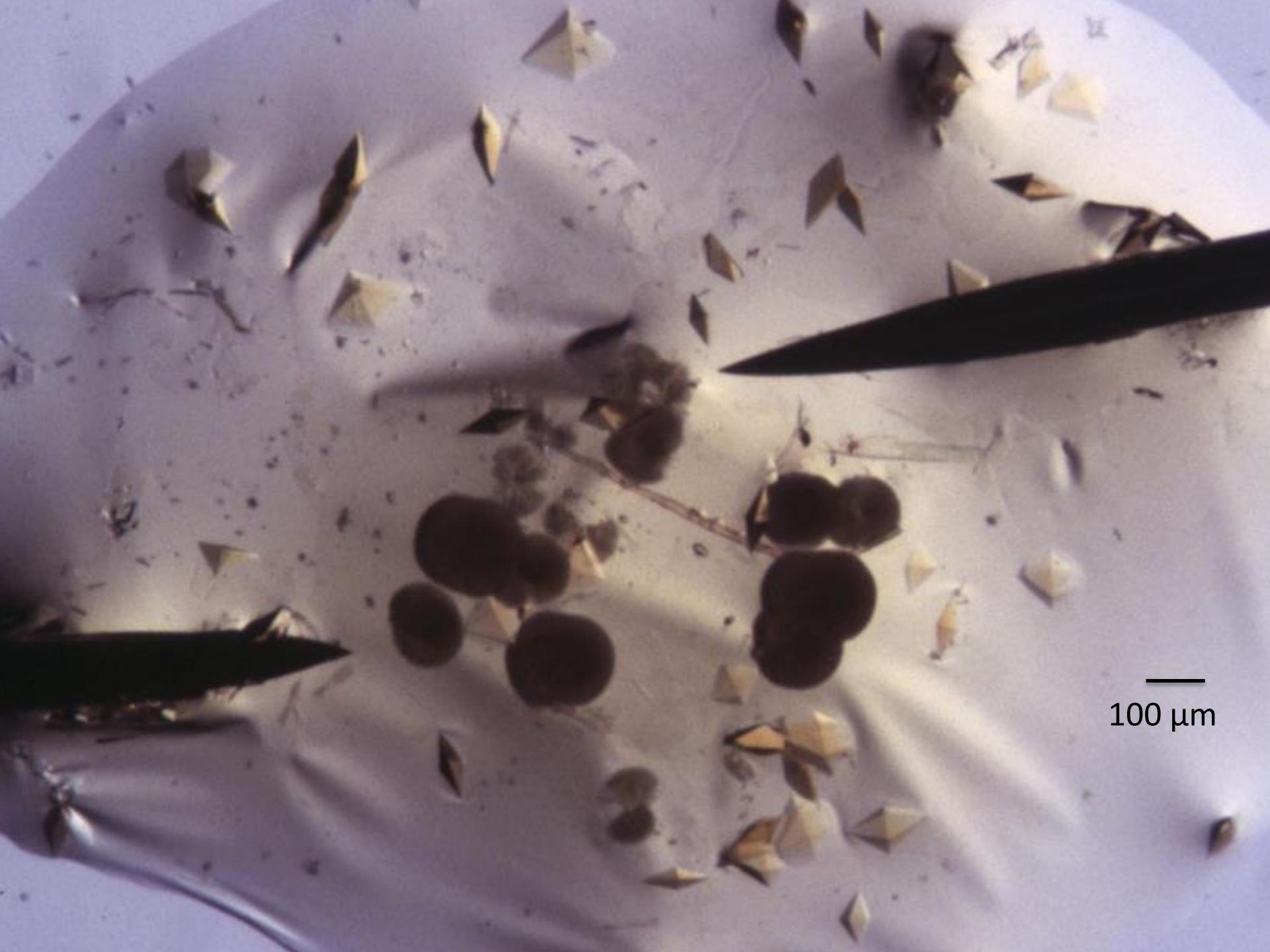
Skin doesn't diffract: just
increases background

Acupuncture needles:
[free samples available from me]

- No loss of liquid
- Slightly flexible
- Fine
- Different sizes available.



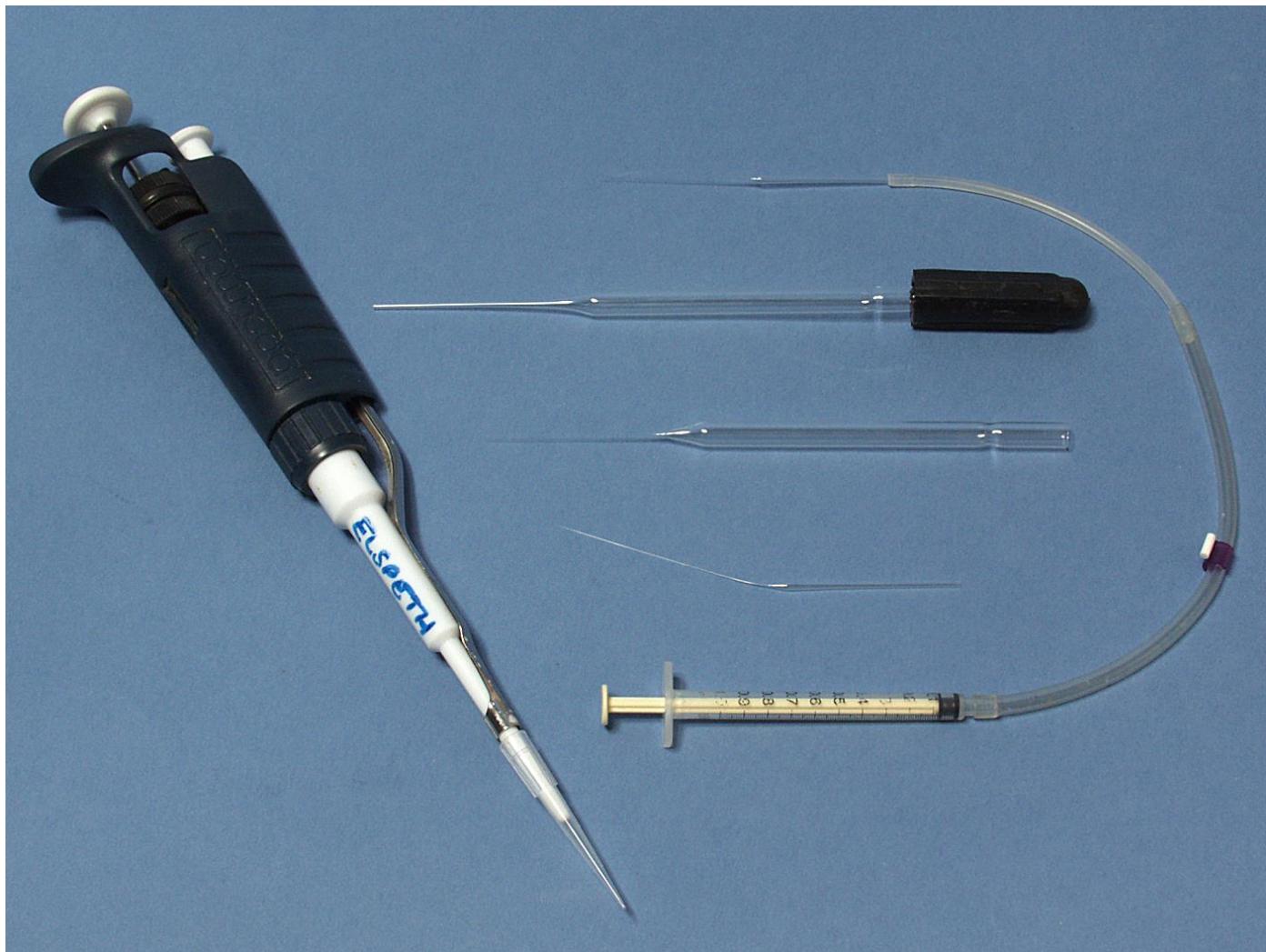
Cats' whiskers, horse eye lashes, horse tail hairs, etc etc



A scanning electron micrograph showing a surface covered in numerous small, yellow, pyramid-shaped particles. Interspersed among these are several larger, dark, irregularly shaped clusters. A scale bar in the bottom right corner indicates 100 μm.

100 μm

Manipulation of crystal from growth drop. A cryo-loop, or a syringe with flexible hose give much better control than a Gilson.



Illegible table of minimum concentrations of glycerol required to cryoprotect Hampton Screen I.

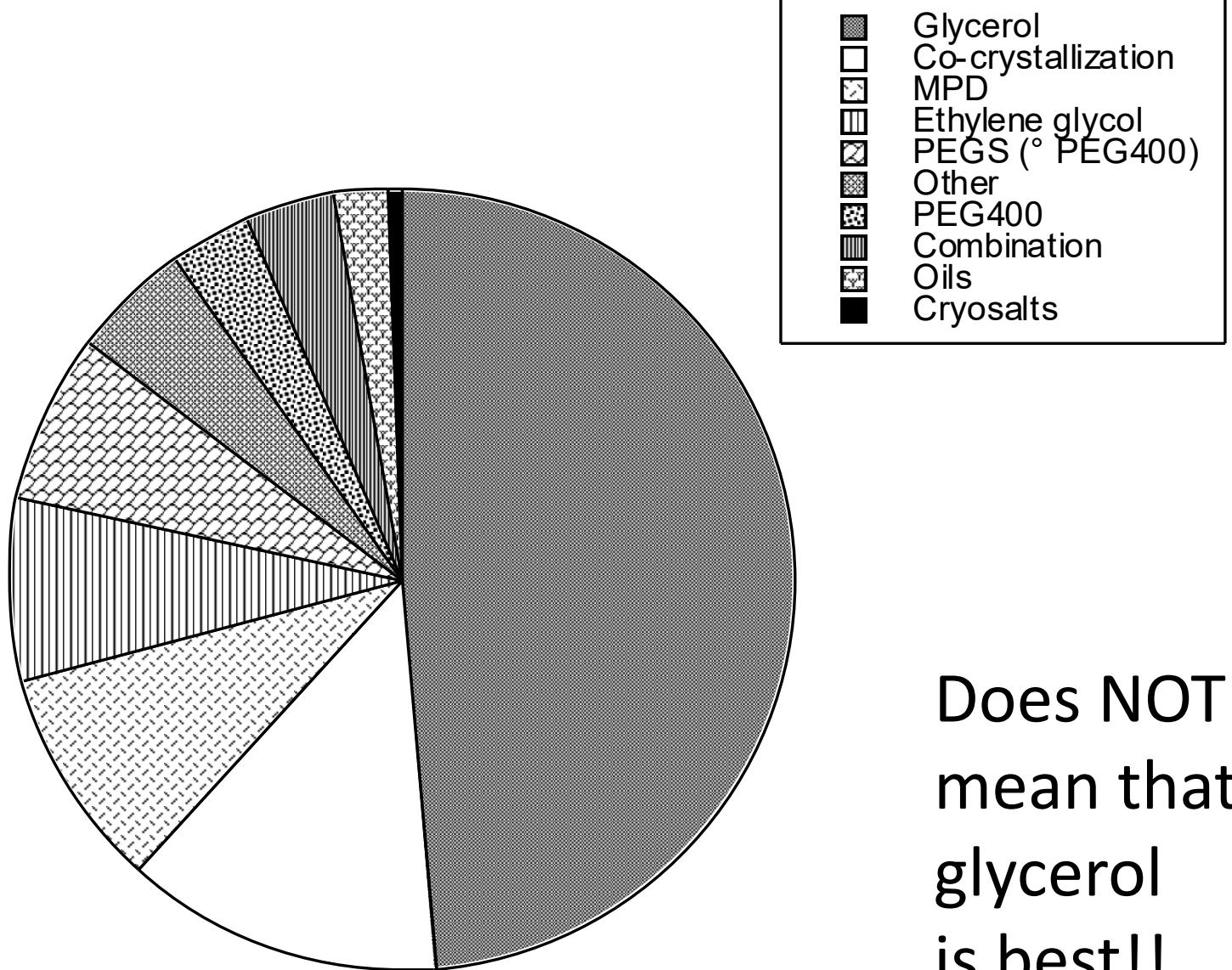
For emergency use only (done by dilution)

[Mitchell and Garman,
J.Appl.Cryst. (1996) 29, 584
McFerrin and Snell
J.Appl.Cryst (2002) 35, 538]

N.B. CDK2: 8M Na Formate excellent cryoprotectant: affects salt bridge ionisation?

TABLE 1. Minimum concentration of glycerol to be added to solutions 1 to 50 of the Hampton Research Crystal Screen^(TM) I Reagent Components to provide cryoprotection when frozen to 100K. It must be noted that glycerol was added to the Crystal Screen solutions resulting in a DILUTION of the original components. From Garman and Mitchell J. Appl. Cryst.(1996) 29, 584-587

Solution number, SALT	BUFFER	PRECIPITANT	GLYCEROL CONC. v/v
1. 0.02 M Ca Chloride	0.1 M Na Acetate pH 4.6	30% v/v 2-methyl-2,4-pentanediol	0
2. None	None	0.4 M K, Na Tartrate	35
3. None	None	0.4 M NH4 Phosphate	35
4. None	0.1 M Tris HCl pH 8.5	2.0 M NH4 Sulphate	25
5. 0.2 M Na Citrate	0.1 M Na Hepes pH 7.5	30% v/v 2-methyl-2,4-pentanediol	0
6. 0.2 M Mg Chloride	0.1 M Tris HCl pH 8.5	30% w/v PEG 4000	20
7. None	0.1 M Na Cacodylate pH 6.5	1.4 M Na Acetate	30
8. 0.2 M Na Citrate	0.1 M Na Cacodylate pH 6.5	30% v/v 2-propanol	30
9. 0.2 M NH4 Acetate	0.1 M Na Citrate pH 5.6	30% w/v PEG 4000	15
10. 0.2 M NH4 Acetate	0.1 M Na Acetate pH 4.6	30% w/v PEG 4000	15
11. None	0.1 M Na Citrate pH 5.6	1.0 M NH4 Phosphate	20
12. 0.2 M Mg Chloride	0.1 M Na Hepes pH 7.5	30% v/v 2-propanol	10
13. 0.2 M Na Citrate	0.1 M Tris HCl pH 8.5	30% v/v PEG 400	0
14. 0.2 M Ca Chloride	0.1 M Na Hepes pH 7.5	28% v/v PEG 400	5
15. 0.2 M NH4 Sulphate	0.1 M Na Cacodylate pH 6.5	30% w/v PEG 8000	15
16. None	0.1 M Na Hepes pH 7.5	1.5 M Li Sulphate	25
17. 0.2 M Li Sulphate	0.1 M Tris HCl pH 8.5	30% PEG 4000	15
18. 0.2 M Mg Acetate	0.1 M Na Cacodylate pH 6.5	20% PEG 8000	20
19. 0.2 M NH4 Acetate	0.1 M Tris HCl pH 8.5	30% v/v 2-propanol	20
20. 0.2 M NH4 Sulphate	0.1 M Na Acetate pH 4.6	25% w/v PEG 4000	20
21. 0.2 M Mg Acetate	0.1 M Na Cacodylate pH 6.5	30% v/v 2-methyl-2,4-pentanediol	0
22. 0.2 M Na Acetate	0.1 M Tris HCl pH 8.5	30% w/v PEG 4000	15
23. 0.2 M Mg Chloride	0.1 M Na Hepes pH 7.5	30% v/v PEG 400	0
24. 0.2 Ca Chloride	0.1 M Na Acetate pH 4.6	20% v/v 2-propanol	30
25. None	0.1 M Imidazole pH 6.5	1.0 M Na Acetate	30
26. 0.2 M NH4 Acetate	0.1 M Na Citrate pH 5.6	30% v/v 2-methyl-2,4-pentanediol	0
27. 0.2 M Na Citrate	0.1 M Na Hepes pH 7.5	20 % v/v 2-propanol	30
28. 0.2 M Na Acetate	0.1 M Na Cacodylate pH 6.5	30% w/v PEG 8000	15
29. None	0.1 M Na Hepes pH 7.5	0.8 M K,Na Tartrate	35
30. 0.2 M NH4 Sulphate	None	30% w/v PEG 8000	15
31. 0.2 M NH4 Sulphate	None	30% w/v PEG 4000	15
32. None	None	2.0 M NH4 Sulphate	25
33. None	None	4.0 M Na Formate	10
34. None	0.1 M Na Acetate pH 4.6	2.0 M Na Formate	30
35. None	0.1 M Na Hepes pH 7.5	1.6 M Na, K Phosphate	25
36. None	0.1 M Tris HCl pH 8.5	8% w/v PEG 8000	35
37. None	0.1 M Na Acetate pH 4.6	8% w/v PEG 4000	30
38. None	0.1 M Na Hepes pH 7.5	1.4 M Na Citrate	10
39. None	0.1 M Na Hepes pH 7.5	2% v/v PEG 400 and 2.0 M NH4 Sulphate	15
40. None	0.1 M Na Citrate pH 5.6	20% v/v 2-propanol and 20% w/v PEG 4000	5
41. None	0.1 M Na Hepes pH 7.5	10% v/v 2-propanol and 20% w/v PEG 4000	15
42. 0.05 M K Phosphate	None	20% w/v PEG 8000	20
43. None	None	30% v/v PEG 1500	20
44. None	None	0.2 M Mg Formate	50
45. 0.2 M Zn Acetate	0.1 M Na Cacodylate pH 6.5	18% w/v PEG 8000	20
46. 0.2 M Ca Acetate	0.1 M Na Cacodylate pH 6.5	18% w/v PEG 8000	20
47. None	0.1 M Na Acetate pH 4.6	2.0 M NH4 Sulphate	20
48. None	0.1 M Tris HCl pH 8.5	2.0 M NH4 Phosphate	20
49. 1.0 M Li Sulphate	None	2% w/v PEG 8000	20
50. 0.5 M Li Sulphate	None	15% w/v PEG 8000	20



Acta D structures 2001. [Garman and Doublie, Methods in Enzymology
2003, 368, 188.]

Transfer of crystal into cryobuffer:

1) Dialysis of cryoprotectant.

[Fernandez *et al*, JAPC (2000) 33, 168-171]

2) Co-crystallisation with cryoprotectant agent: glycerol is already known to help in some cases

[Sousa, Acta Cryst D51 (1995) 271-277.]

3) Rapid transfer

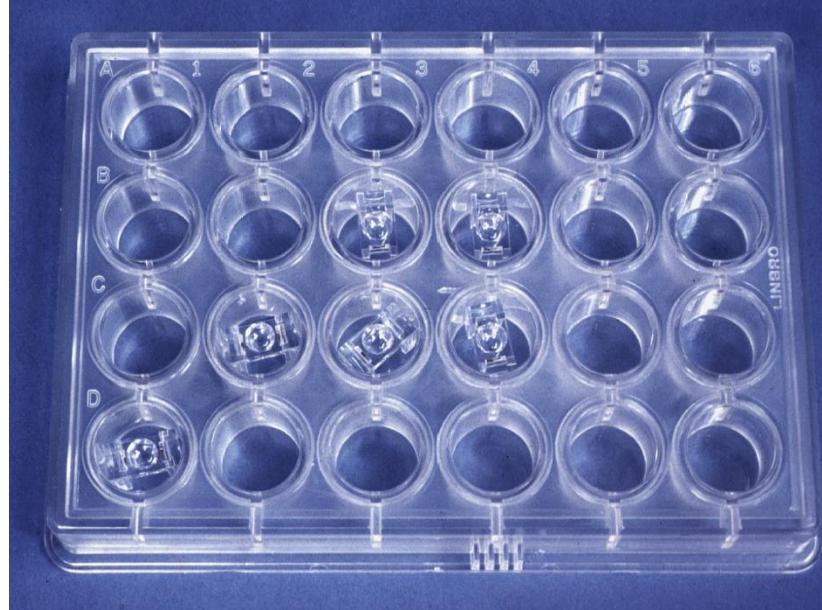
- Straight into final concentration for up to 5 mins
- can just ‘sweep through’- 0.5 sec
- Sequential soaks in increasing concentrations.

WANT TO MINIMISE HANDLING, as handling
can increase the mosaic spread. i.e. 2) is BEST:....

SOLUBILITY versus OSMOTIC shock

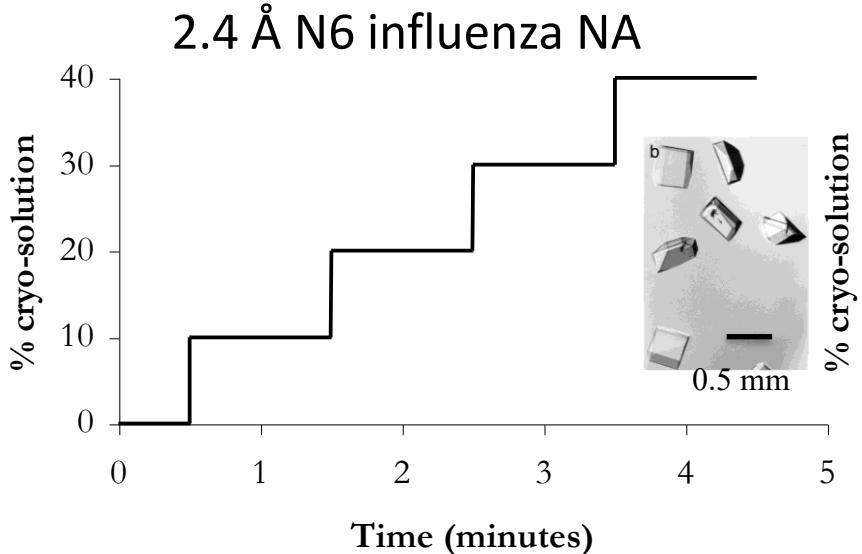
Crystal transfer optimisation:

- balance osmotic shock and attack of crystal surface
- serial transfers: minimise handling and dehydration

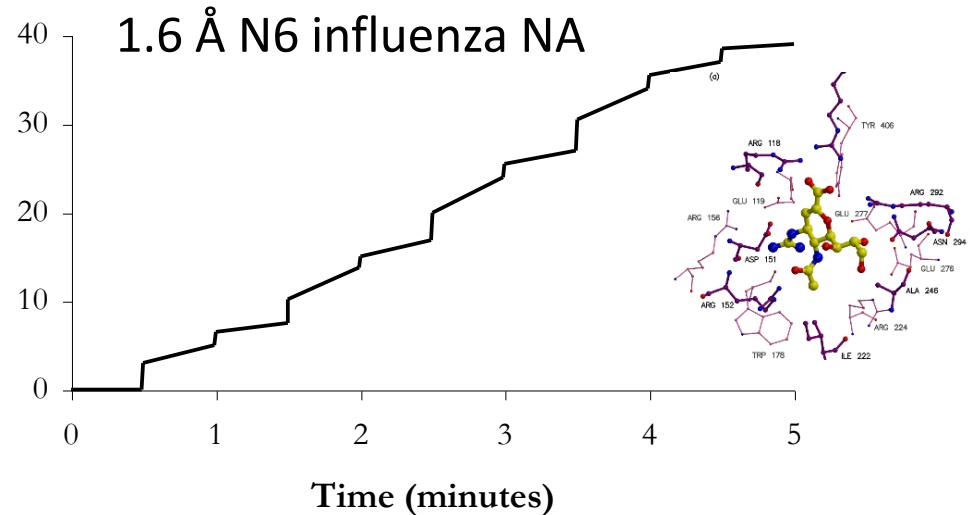


Quick 'dunk': 3.2 Å N6 influenza NA

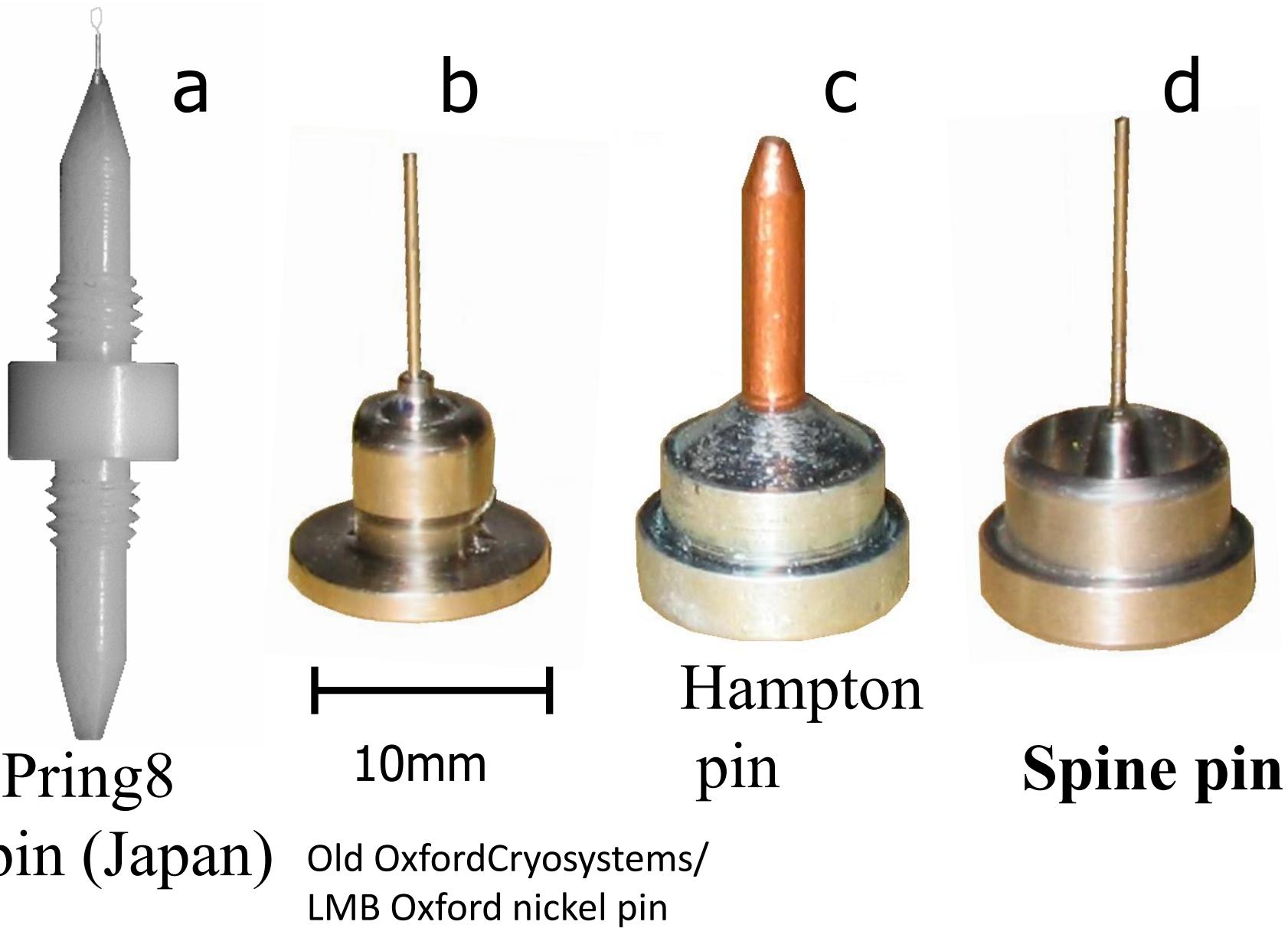
a) Move crystal between solutions



b) Solution pipetted onto crystal



Hardware development: standardisation...





Litho-loops,
etched mylar,
ActiLoop polymer
Molecular Dimensions



Check loop vs
crystal size
BEFORE
opening drop



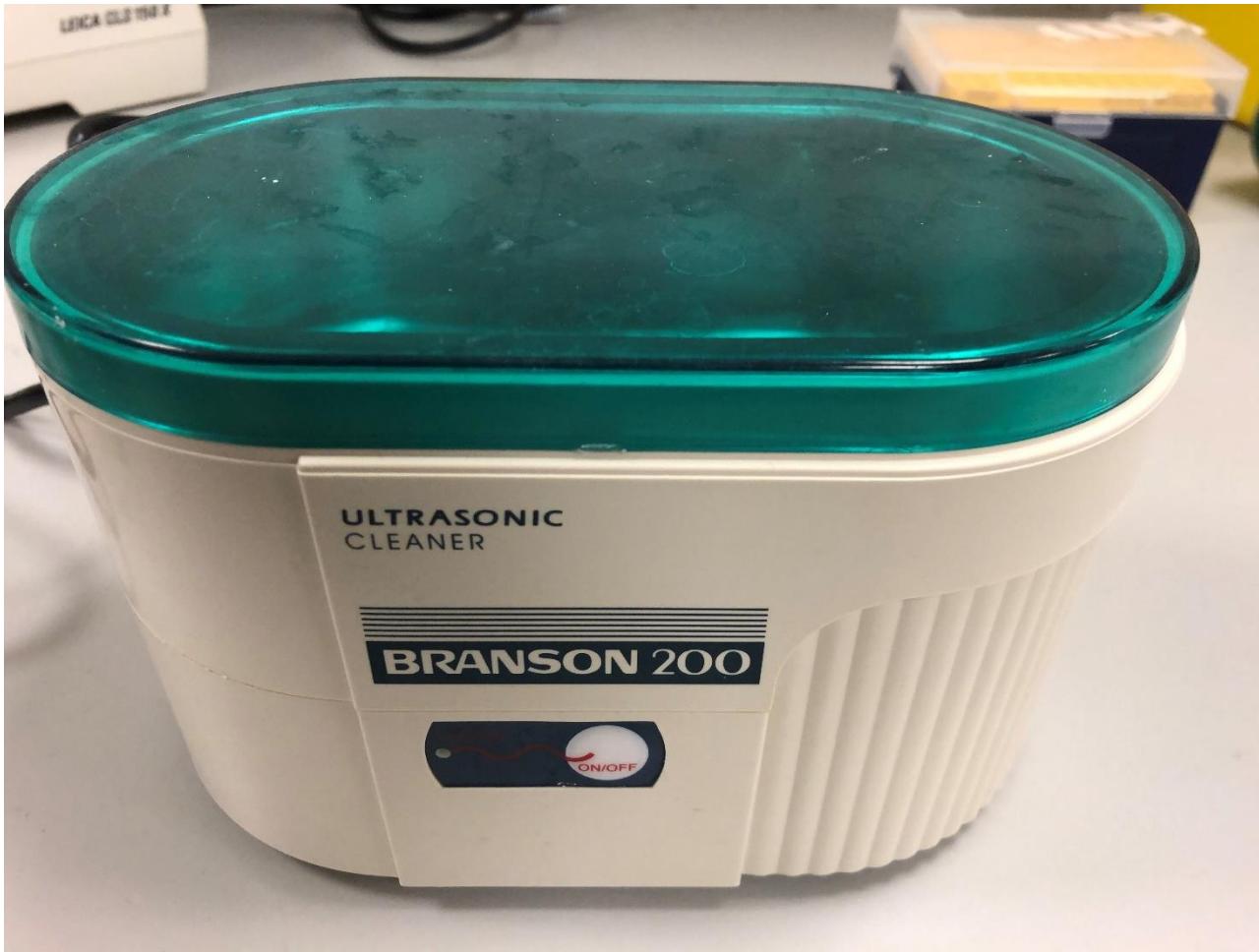
Nylon loops
Hampton
Research
NB: CAN VIBRATE
IN N₂ STREAM!



Expensive and fragile but
stable in N₂ stream.

Micro-mounts,
microfabricated
polyimide film. **MiTegen**
[Thorne *et al* (2003), JAPC 36, 1455.]

Loop Cleaning: important!

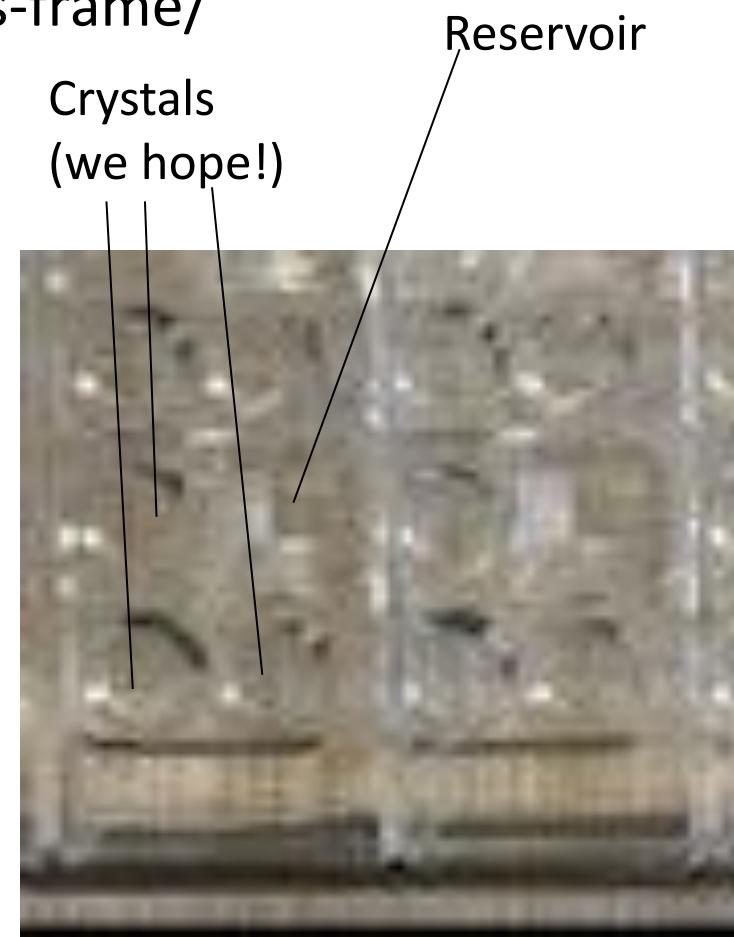
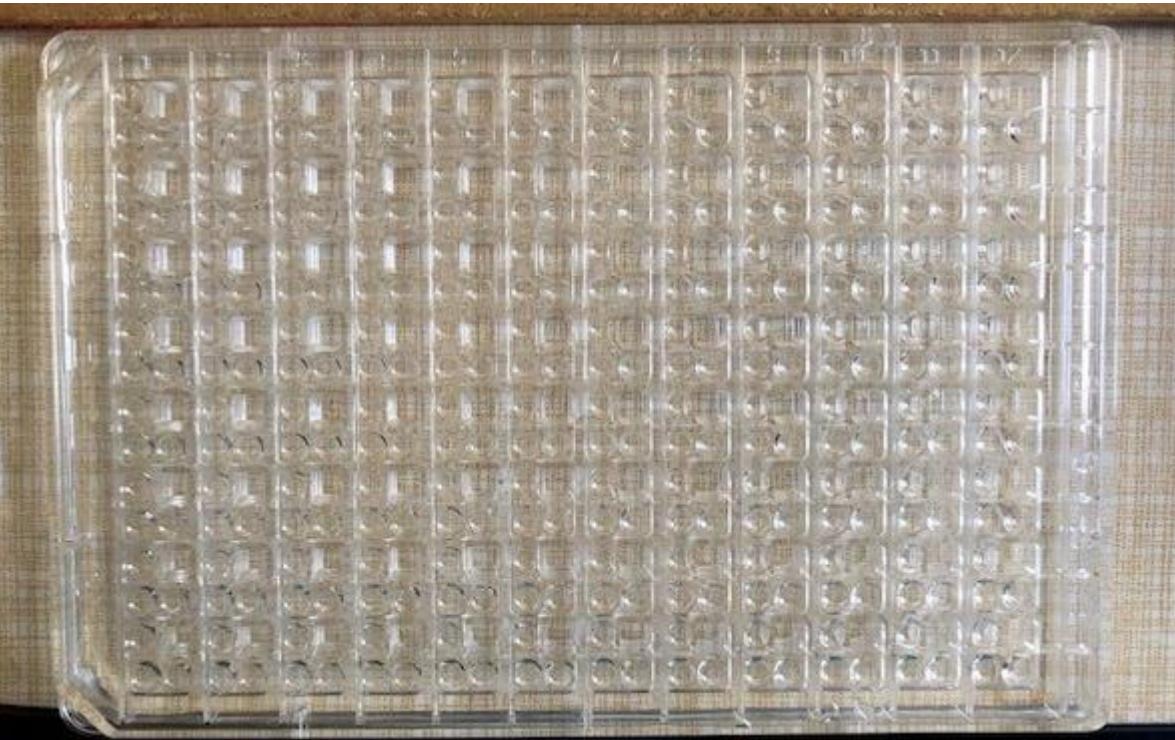


2 mins/cycle, 2 cycles.

Detergent (1 cycle) then water (1 cycle)

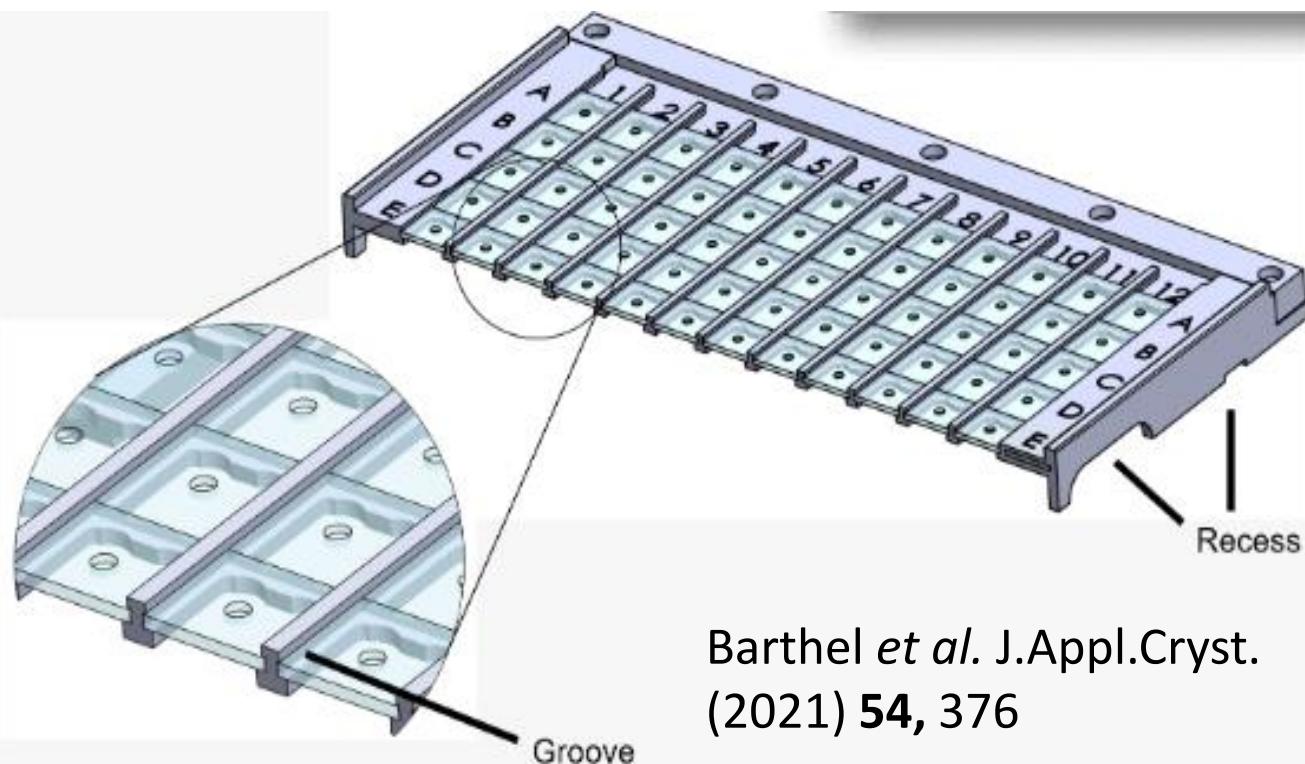
Crystallisation trays:

- 96 well plates –small volumes!
- Round bottom wells MUCH easier than square cross sections
- Dehydrate quickly when you take off the sticky tape lid: easy access frame, <https://cfg-biotech.de/easy-access-frame/>

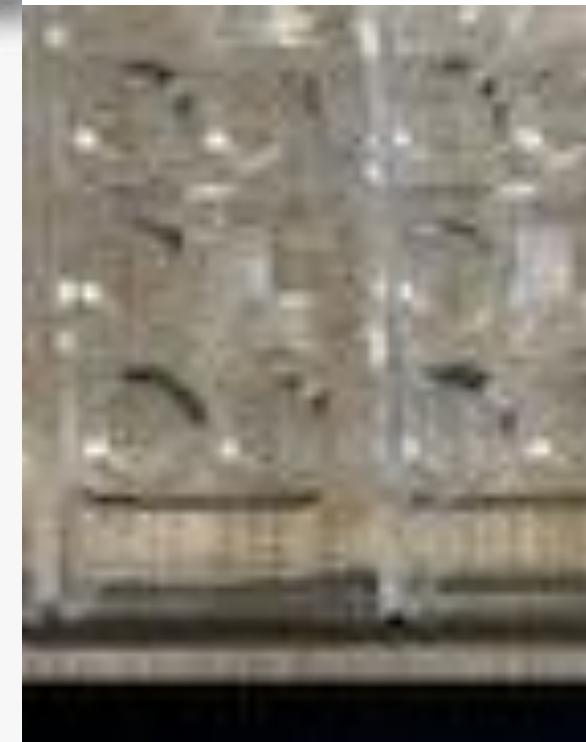


Crystallisation trays:

- 96 well plates –small volumes!
- Round bottom wells MUCH easier than square cross sections
- Dehydrate quickly when you take off the sticky tape lid: easy access frame, <https://cfg-biotech.de/easy-access-frame/>
- Fits MRC plates



Barthel et al. J.Appl.Cryst.
(2021) 54, 376

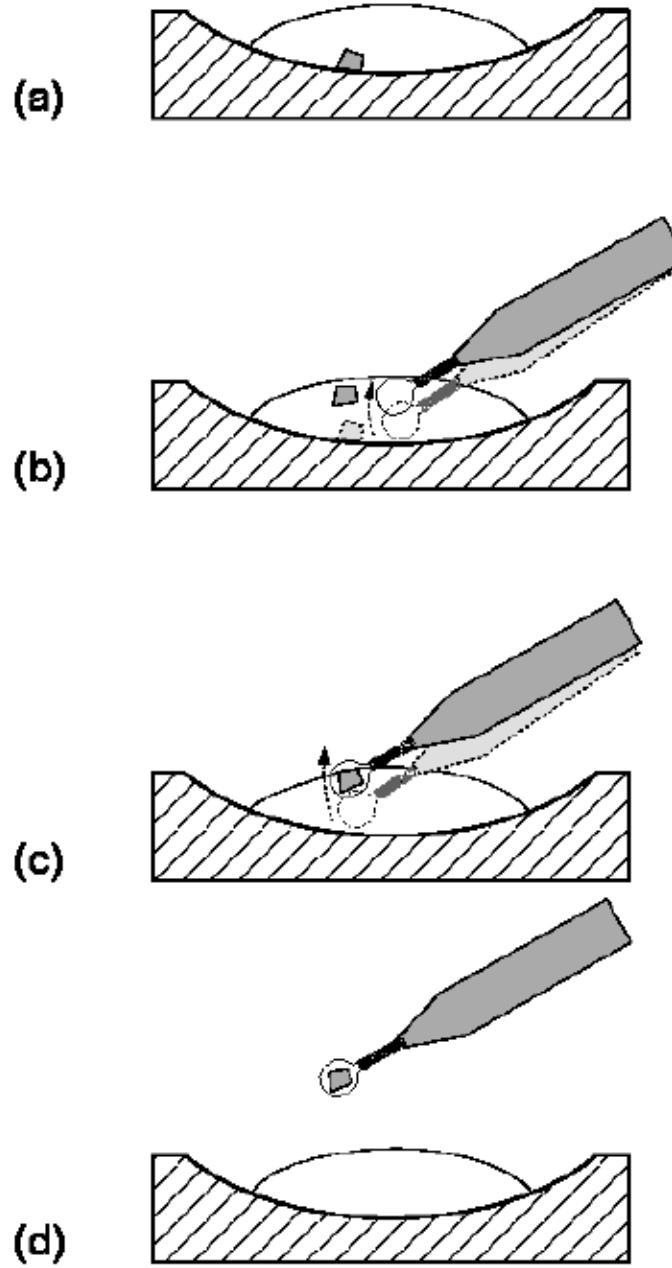


Fishing:

- minimise handling
- minimise liquid round crystal
- fish near cryogen
- loop perpendicular to liquid

NB: if drop too thick, background increases by factor of 10. Touch on plastic surface to get rid of it.

N.B. Acupuncture needles.
Salt crystals in loop.



Fishing: what can go wrong?

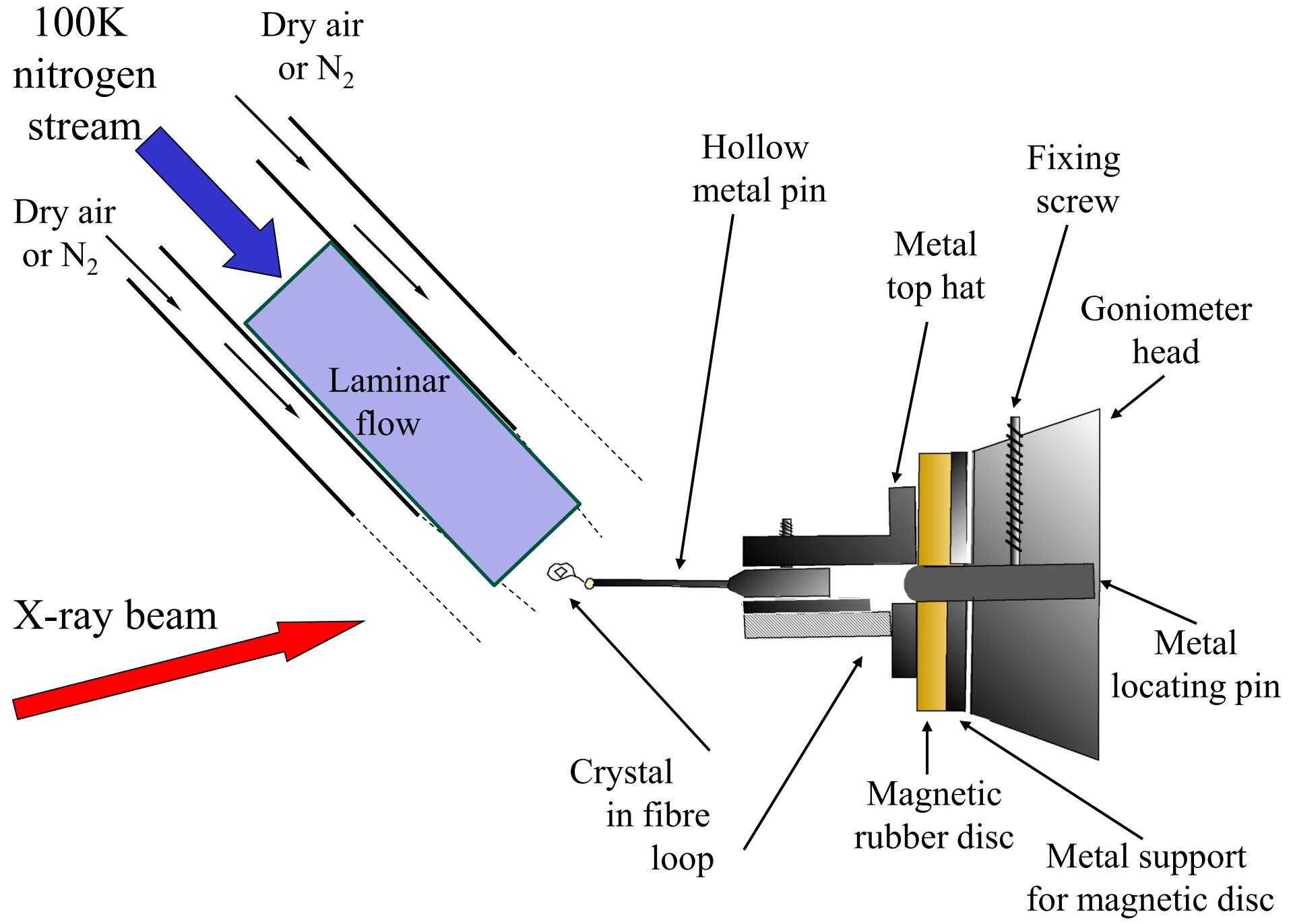
- Skin on drop
- Crystal stuck to plastic
- Loop size is wrong
- Takes too long: drop dries out
- Damage crystals trying to catch them
- Too much solvent on loop



Flash **cool** (NOT ‘freeze’)

into cryogen:

- Straight into liquid nitrogen:
**have a standard pin length AND
blow/flap away surface cold nitrogen from
FULL Dewar.** [Warkentin *et al* 39, (2006) 805-811]
- Stream cool into nitrogen gas stream held
at around 100K: pre-align pin.
**have a standard pin length and block the
stream!** **PRACTICE!!!**
(but not with your most precious crystals)



DIFFRACTION?

- NO ...

Does the crystal diffract at room temperature??

NO



Back to crystallisation
trials

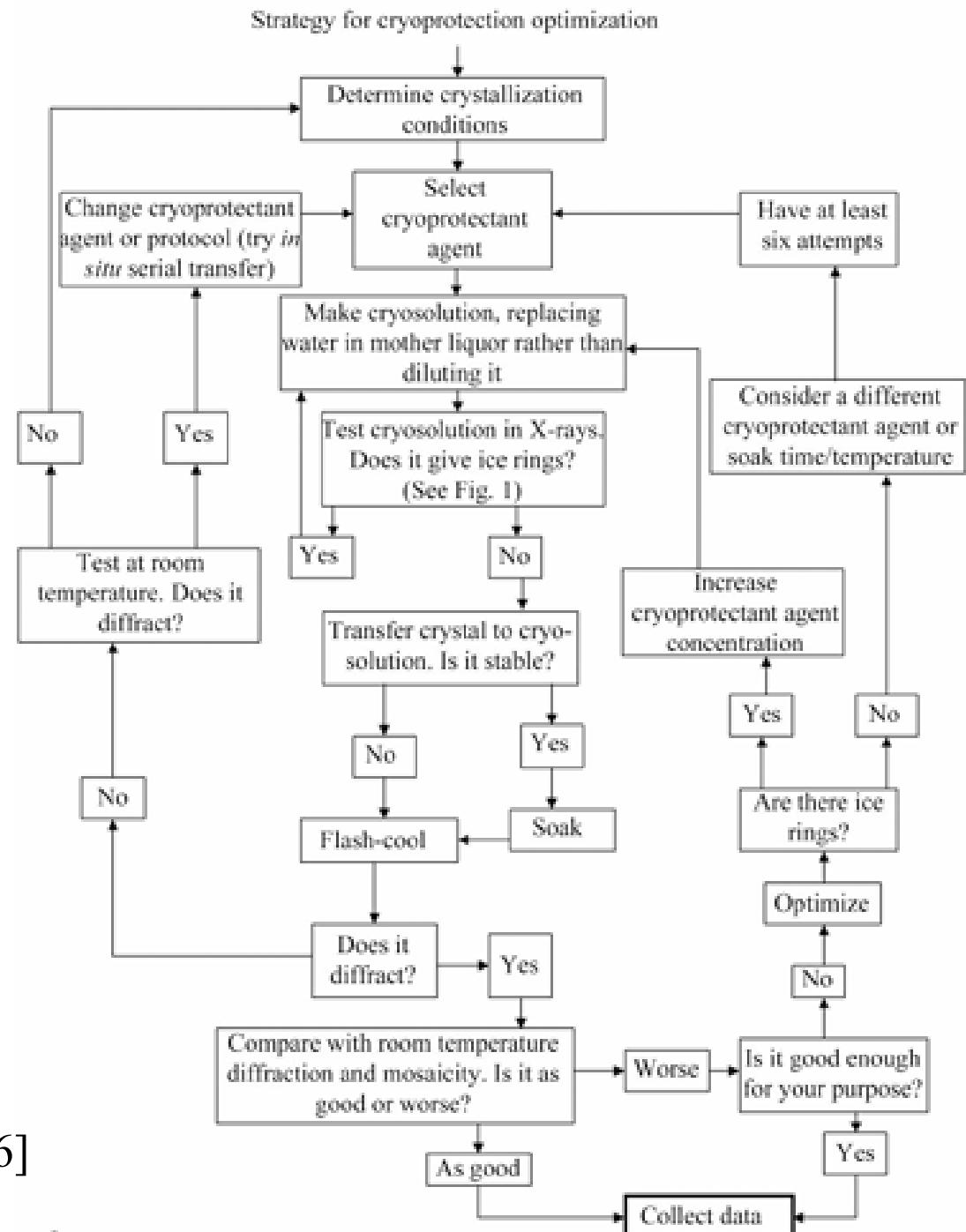
YES



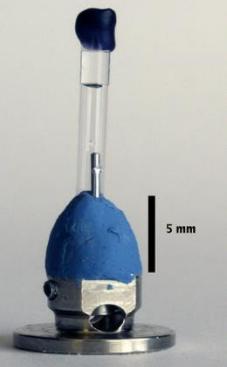
Change cryo-protocol

- YES... then take data and/or store it in a Dewar.

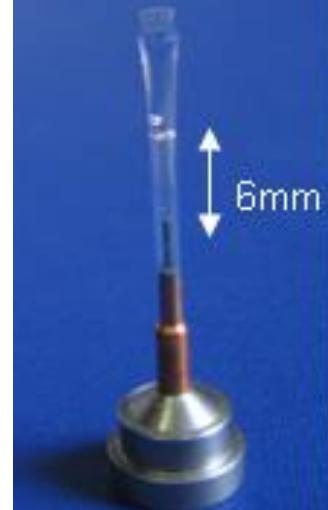
Strategy for cryoprotection Optimisation



[Garman & Owen Acta D 2006]



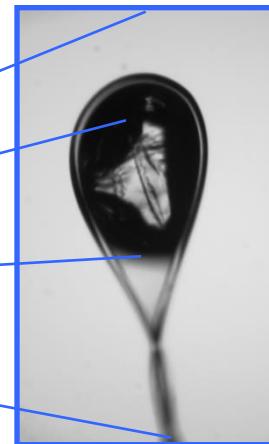
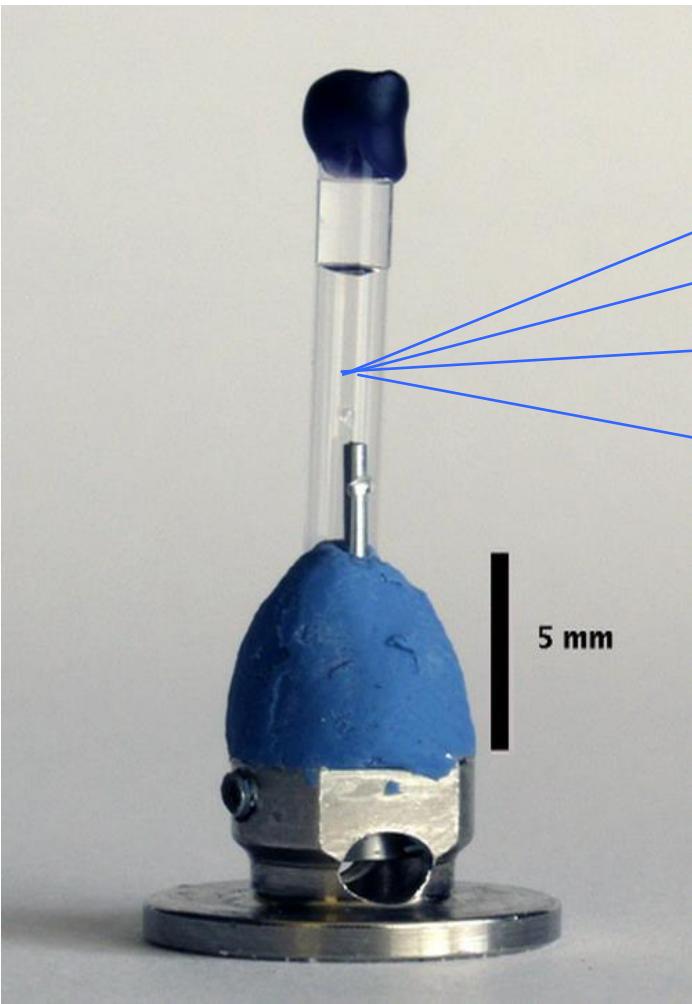
The Plan:



- Cryo techniques
 - Why cool? Radiation damage.
 - Optimising cryoprotection.
 - **Testing at room temperature.**
 - Storage and retrieval.
 - If nothing works...

(Much) easier RT mounting method

Allows protein crystals to be mounted at room temperature in loop.

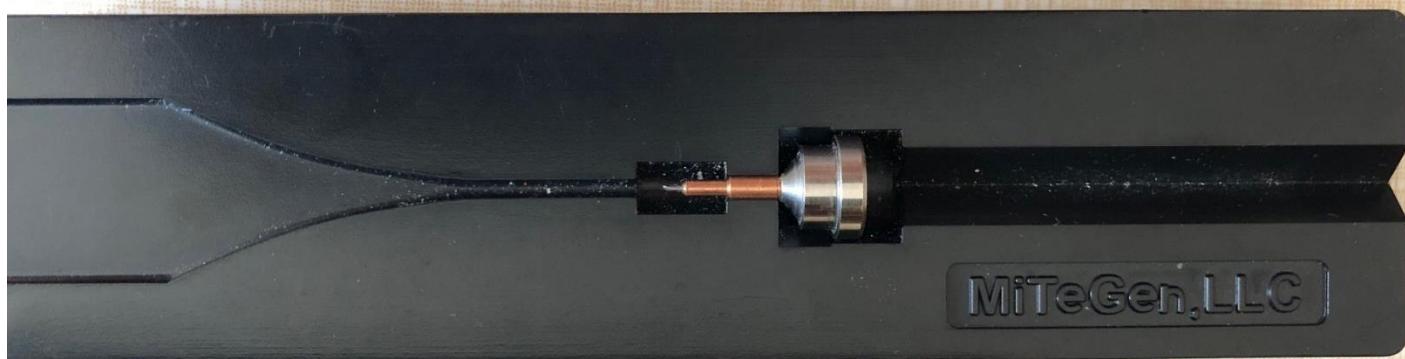
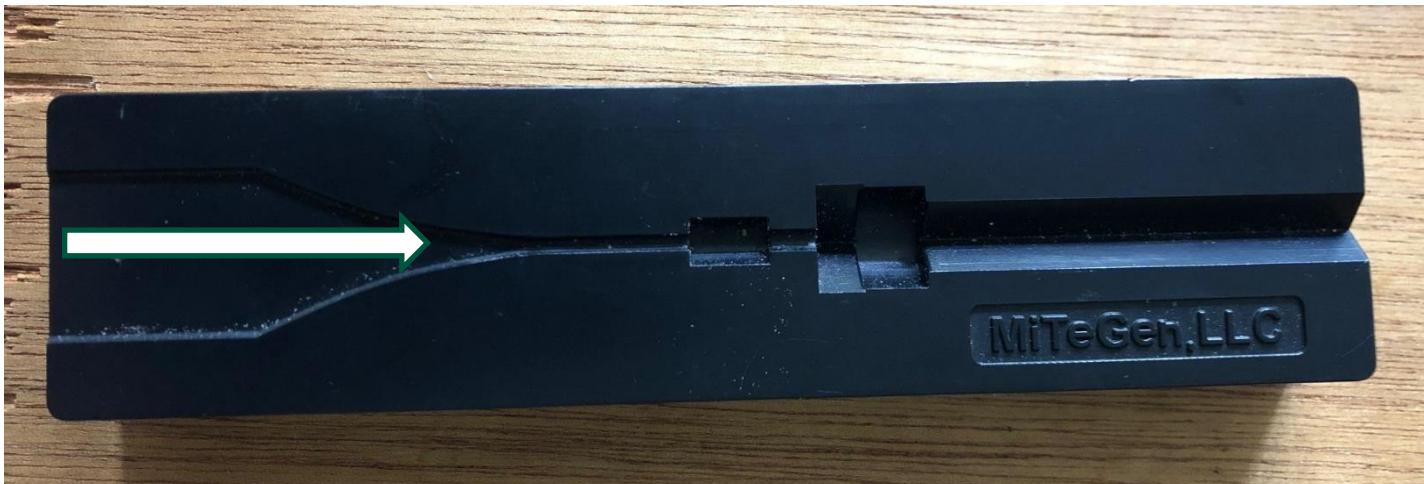


Capillary prevents drop drying out at room temperature.

Also eliminates crystal manipulation between room temperature and cryo-cooled datasets.

Skrzypczak-Jankun *et al* Acta Cryst. (1996).
D52, 959-965

RT mounting

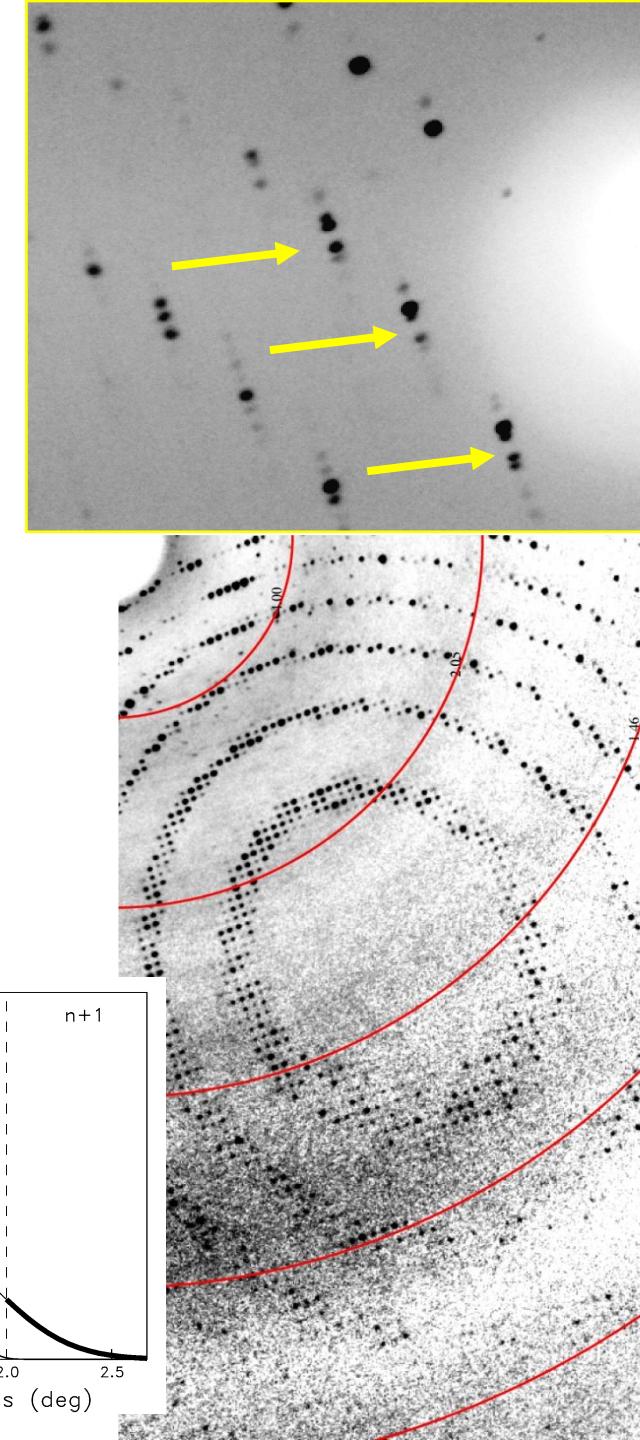
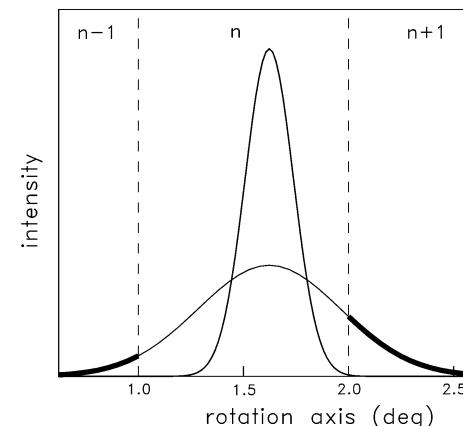


Kalinin *et al.* (2005).
J. App. Cryst. 38, 333-339.

The black MiTeGen plastic former helps you thread the thin plastic sleeve over the copper pin holding the crystal.
Then seal the bottom of the tube against the pin with grease.

MOSAICITY (‘spread’ of a reflection in 3-D space):

- Want to minimise mosaicity but it often increases from RT values.
- Are cryoconditions optimised?
 - solutions
 - transfer
- Speed of transfer to cryogen
- Speed of cooling
- Size of crystal:



Cryo-protocol Optimisation: maximising resolution, [minimise mosaicity]

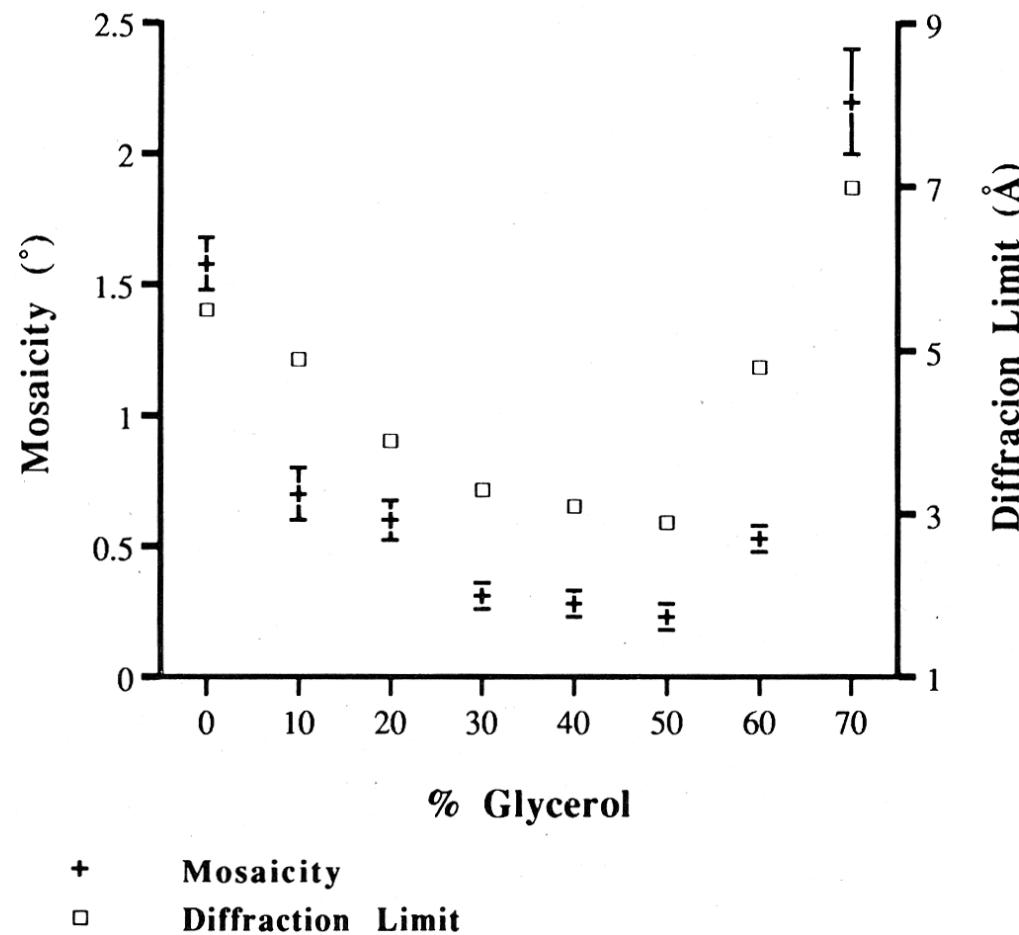
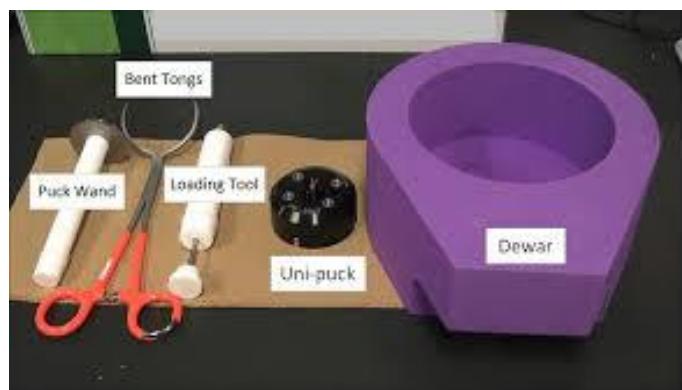


Fig. 2. Variation of mosaicity and diffraction limit of GPb crystals with percentage of glycerol in the buffer. The error bars represent statistical counting errors only.



The Plan:



- Cryo techniques
 - Why cool? Radiation damage.
 - Optimising cryoprotection.
 - Testing at room temperature.
 - **Storage and retrieval**
 - If nothing works...

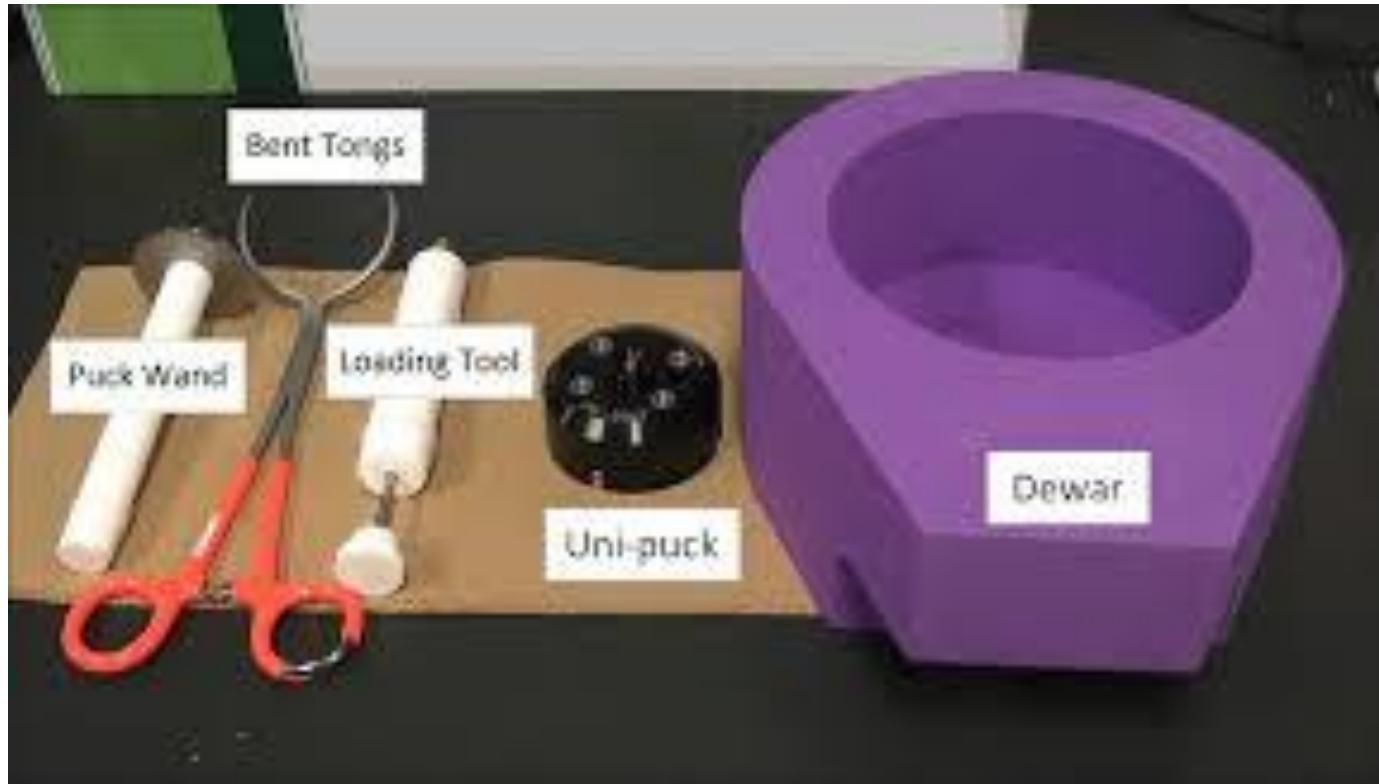
Crystal
storage
and
retrieval

Copyrighted
Gary Larson
Cartoon showing
Man pulling freezer
power out of wall

Hyperquenching: want a FAST cool

Keep Dewar full or remove gas layer from above liquid.

ALSO keep hand moving in liquid nitrogen to escape insulating bubble layer round sample.



[Warkentin et al., Thorne, J. Appl. Cryst. (2006) 39, 805]



STORAGE:

- Label the vials beforehand.
- Label the canes.
- Keep notes on puck positions.
- Allow transport Dewars to dry out after each trip.



Dewar drying rack, Dept Biochemistry, Oxford, UK



Excellent Dewar testing protocol

[http://smb.slac.stanford.edu/facilities/hard
ware/cryotools/shipping-dewar-
testing.html](http://smb.slac.stanford.edu/facilities/hardware/cryotools/shipping-dewar-testing.html)

GOOGLE search (first hit):

ssrl dewar testing

[See also Owen, Pritchard & Garman. *J. Appl. Cryst.* (2004) **37**, 1000-1003]

Change storage LN₂ every 3 months.



Crystal Handling under LN₂

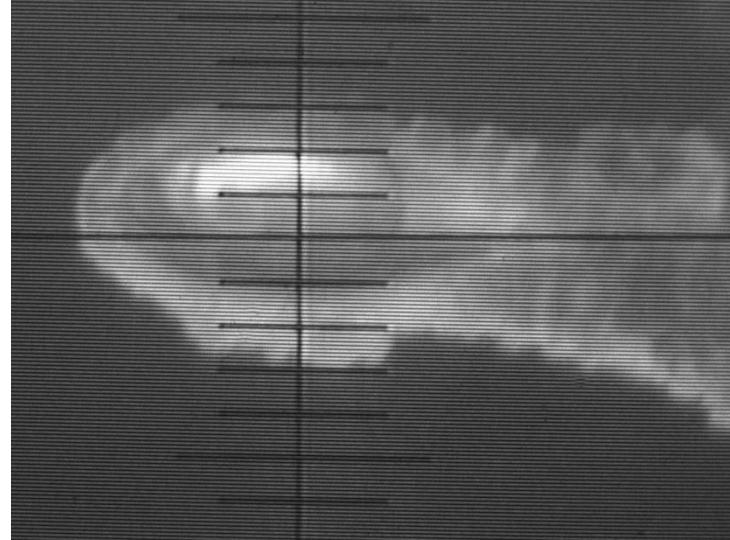
- Use appropriate tools
- Have a small working Dewar: change the LN₂ in it often.
- Wait for LN₂ to stop bubbling before trying any manipulations.
- Only move one hand at a time
- Steady the stationary hand on edge of Dewar

SAFETY: gloves and goggles



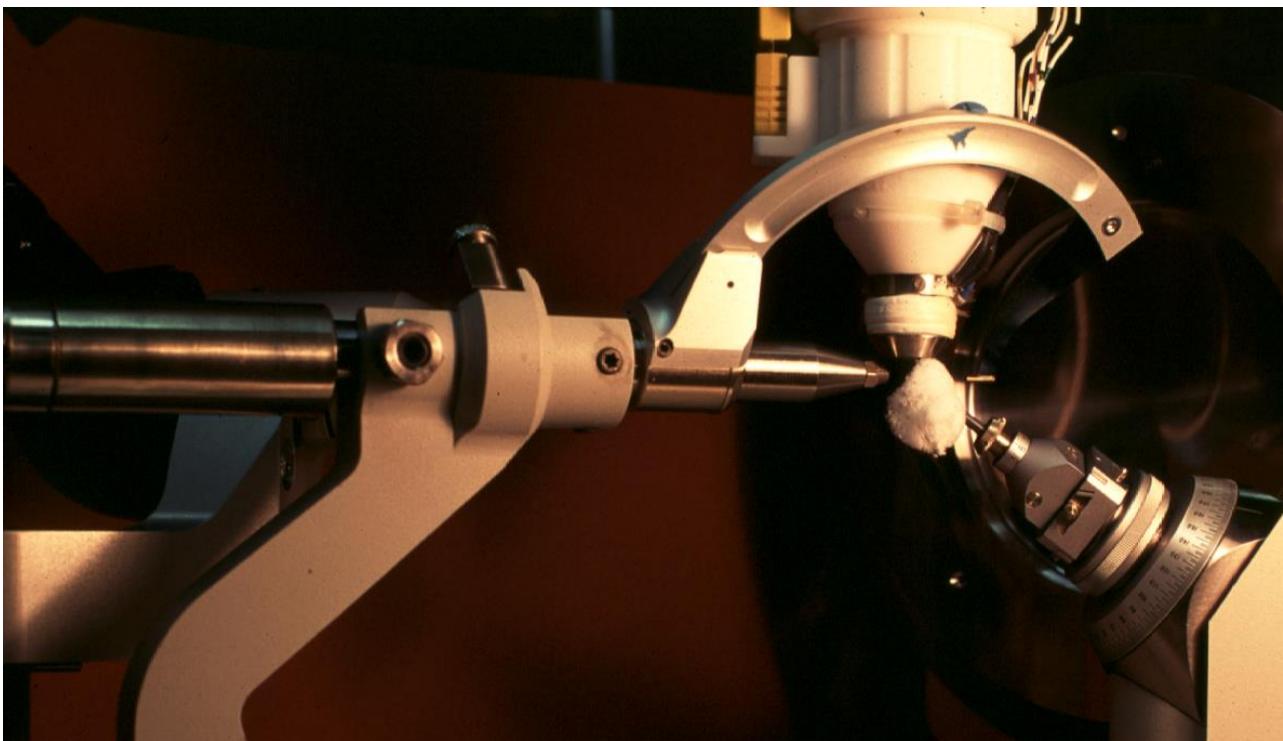
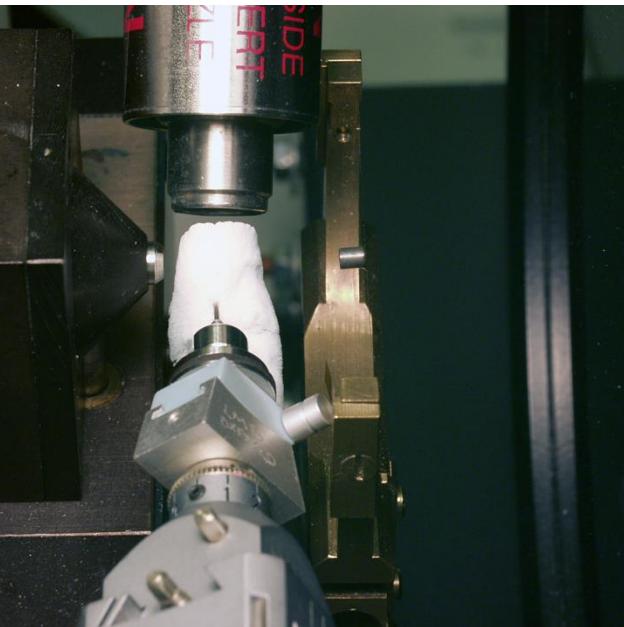
What am I doing wrong?





Absolutely NO ice of ANY sort.

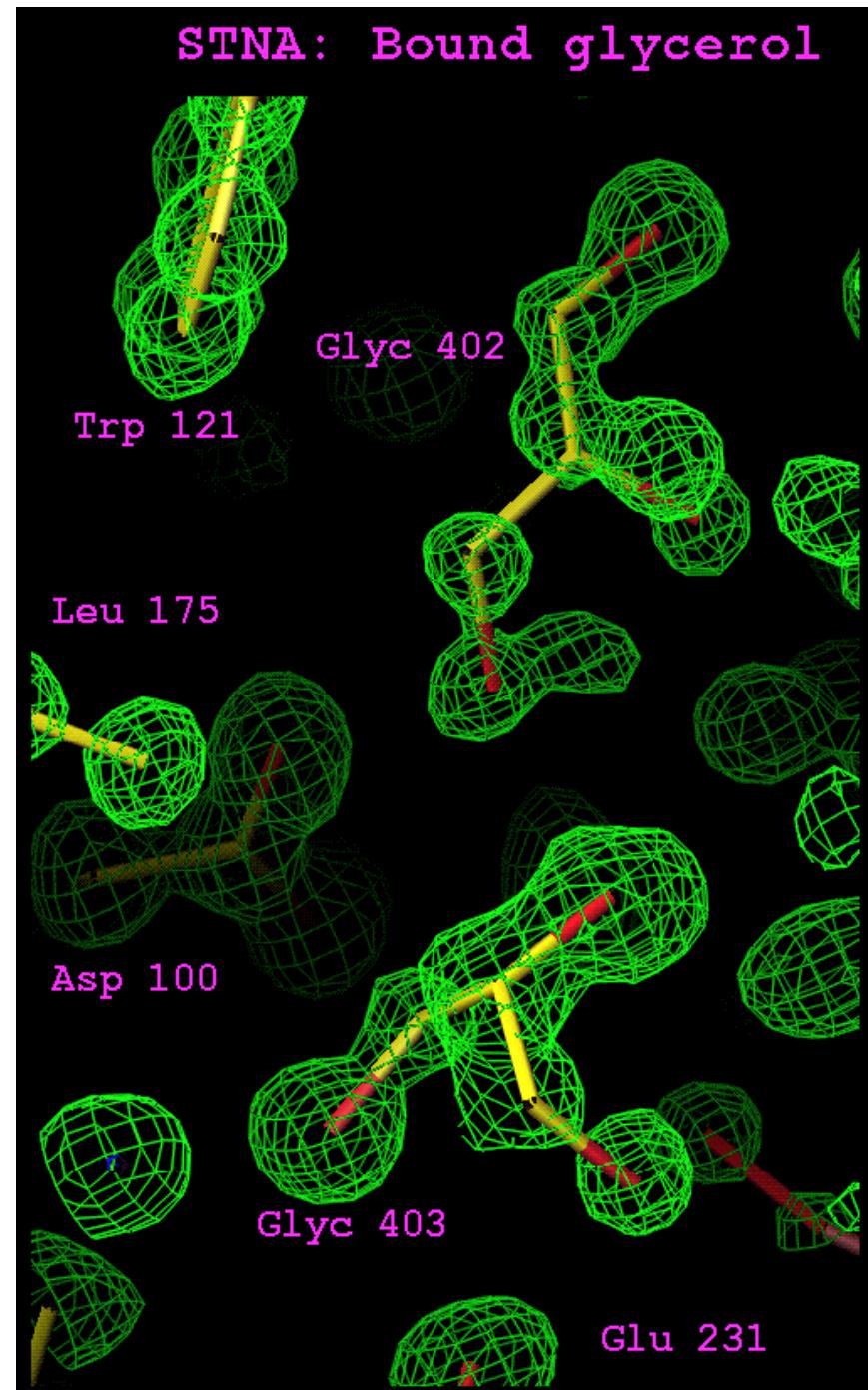
See Garman and Schneider J. Appl. Cryst (1997) for trouble shooters guide.



Bound cryoprotectant agent.

Beware competitive inhibition with a wanted substrate.

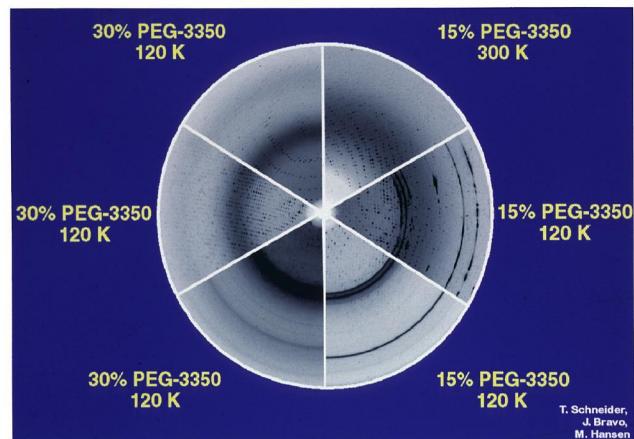
Put substrate in cryo-buffer.





The Plan:

- Cryo techniques
 - Why cool? Radiation damage.
 - Optimising cryoprotection.
 - Testing at room temperature.
 - Storage and retrieval.
 - **If nothing seems to work....**



If nothing seems to work:

- Diffraction at room temperature?
- Cryo-solutions
- Transfer/handling/soaking – vary time and temperature [e.g. try 4° overnight]
- Cryogen choice: nitrogen gas/liquid
- Osmolarity matching
- Crystal annealing
- Swap buffer
- Try more than once.

Theoretical study: starts to rationalise experimental practice.

Heat transfer study by a proper mechanical engineer!

Most to least important factors:

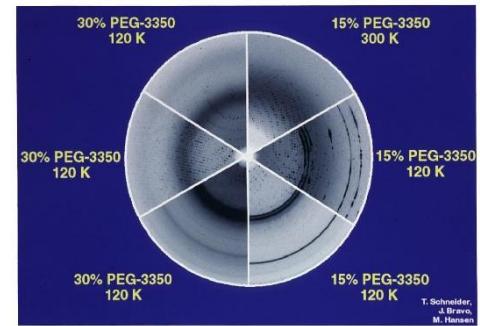
- 1) Crystal solvent content and solvent composition.
- 2) Crystal size and shape.
- 3) Amount of residual liquid around the crystal
- 4) Cooling method (liquid plunge *versus* gas stream).
- 5) Choice of gas/liquid.
- 6) Relative speed between cooling fluid and crystal.

[S. Kriminski, M. Kazmierczak and R.E. Thorne

`Heat transfer from protein crystals: implications for flash-cooling and X-ray beam heating. Acta Cryst. (2003) D59, 697-708.]

If nothing seems to work:

- Diffraction at room temperature?
- Cryo-solutions
- Transfer/handling/soaking – vary time and temperature [try 4° overnight]
- Cryogen choice
- **Osmolarity matching**
- **Crystal annealing**
- Swap buffer
- Try more than once
- Dehydration



Copyrighted
Gary Larson
Cartoon ‘Slug
vacation disasters’.
Slugs on way to
Great Salt Lake.

Osmolarity matching:

Osmotic shock: compresses crystal \Rightarrow cracks

\Rightarrow Mosaic spread increases \Rightarrow resolution lower.

‘Quick dips’ give greatest osmotic shock but minimise time for cryoprotectant attack.

Method:

- 1) Look up osmotic pressure of mother liquor in CRC Handbook of Physics and Chemistry, Section D-232.
11th column is O: Os/kg
- 2) Look up o.p. of cryoprotectant agent.
- 3) Modify conc. of mother liquor to minimise change in osmotic pressure.

See (**Garman Acta D 1999, Section 6.4**)

Crystal annealing: remove ice, reduce mosaicity and increase resolution

- Block stream temporarily (1-10 secs) OR
- Put crystal back in cryo-buffer solution.
- Then flash cool again.
- Worth a try (can repeat several times).
- Works sometimes.

[misnomer: slow heat and fast cool is really
‘tempering’]

Yeh and Hol (1998) *Acta Cryst.* D54, 479- 480.

Harp *et al.* (1998) *Acta Cryst* D54, 622-628, and (1999) D55, 1129-1134.

Hanson *et al.* (2003) *Meth Enzym* 368, 217

Understanding why annealing sometimes works.

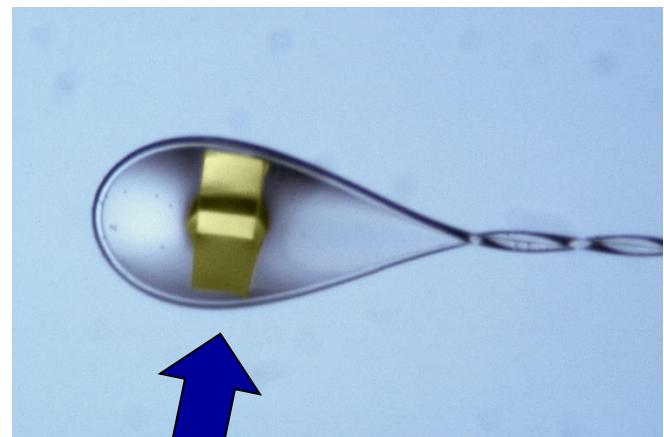
Cryoprotectant agent concentration: match contraction of lattice to contraction of bulk solvent to avoid lattice distortion, higher mosaicity etc.

Not enough cryoprotectant



Water exported

Too much cryoprotectant

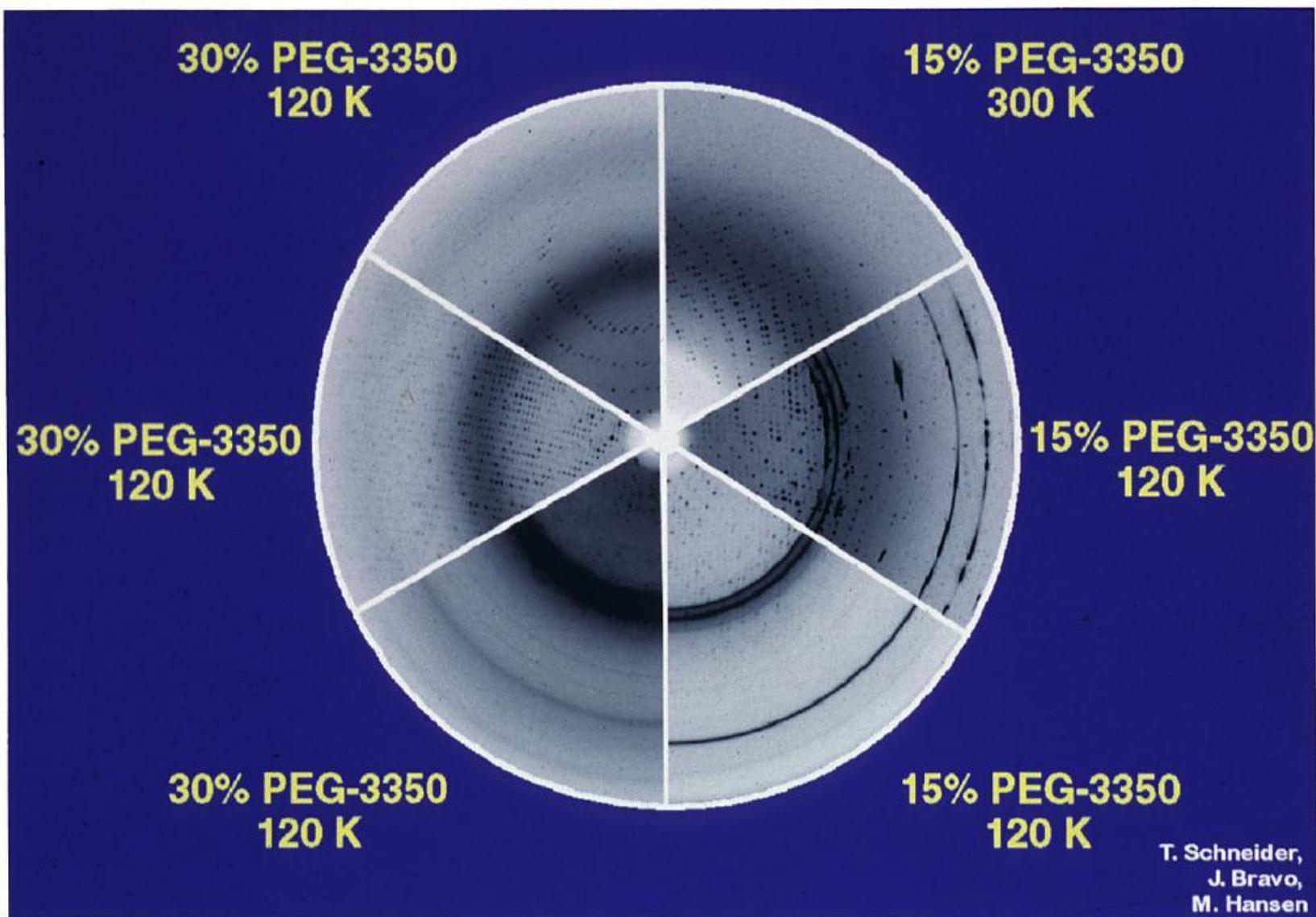


Water imported

If nothing seems to work:

- Diffraction at room temperature?
- Cryo-solutions
- Transfer/handling/soaking – vary time and temperature [4°]
- Cryogen choice
- Osmolarity matching
- Crystal annealing
- **Swap buffer**
- **Try more than once.**
- Dehydration

Have at least 6 goes...



Dehydration (if desperate!)

- Solvent channels effect:

https://www.researchgate.net/publication/51655454_Direct_cryocooling_of_naked_crystals_Are_cryoprotection_agents_always_necessary

- Combined dehydration and dry mounting results:

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4346222/>

CRYO-COOLING:

Advantages

- Reduced radiation damage rate (~ factor 70).
- Gentler mounting
- Lower background
- Higher resolution
- Fewer crystals
- Can ship crystals
- Use crystals when ready.

Disadvantages

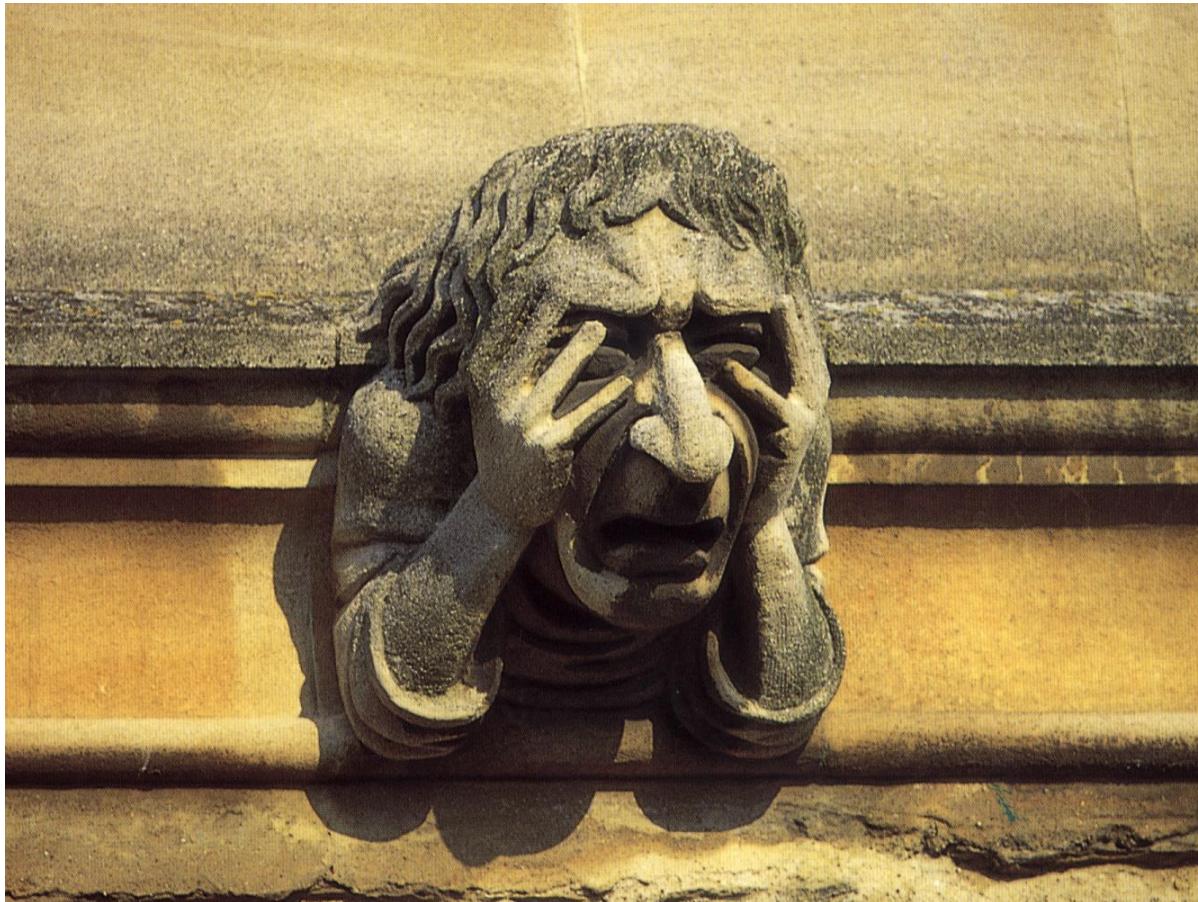
- Expensive equipment
- Increase in mosaic spread.
- Need to invest time and PRACTICE fishing!

**DON'T just do what your colleagues say 'always works'.
THINK about it!**

Cryocrystallography: for more gory details see

- Rodgers, D (1997) Methods in Enzymology 276, 183-203 and in International Tables of Crystallography Volume F (2001) 202-208.
- Hope, H in International Tables of Crystallography Volume F (2001) 197-201.
- Garman and Schneider (1997) J.Appl. Cryst, 30, 211-237.
- Parkin and Hope (1998) J.Appl. Cryst. 31, 945-953.
- Garman (1999) Acta D55, 1641-1653.
- Garman and Doublié (2003) Methods in Enzym. 396, 188-216.
- Garman (2003) Curr. Opin. Struct. Biol. 13, 545-551.
- Garman and Owen, (2006) *Acta Cryst. D* 62, 32-47.
- Pflugrath (2015) *Acta Cryst F* 71, 622-642.

lots of questions are expected and very welcome



e.g. what is this?



Lorna Dougan,
Leeds, LOOminaries

Workshop Advice from the Organisers

There is a practical purpose to the poster session, which is to enable the tutors present at the workshop to best help you with your crystals/data. For that reason, keep your poster and presentation clear and concise. Remember it is perfectly valid for your poster to contain questions for the tutors!

Please note, for your poster to be eligible for the prize we request that you follow these instructions:

- Posters must be printed **max A0 in size**, and **portrait** orientation
- Describe your project in terms of the problems you are facing. What have you tried? What hasn't worked? How do you think you might solve them?
- Don't recycle an old conference poster! We are interested in the work in progress, not necessarily the most complete scientific story.
- Please include a **photograph** of yourself to help the tutors match the projects to attendees during the workshop.
- Be prepared to present your project by your poster during the scheduled poster session.
- By the time of the in-person poster session, please make sure to also upload a PDF of your poster to your individual project channel on the Slack workspace.

Elspeth's Poster Judging Check List. 2025

1) Do I WANT to read it?

i.e. a) Can colour blind people read this poster?

b) are the colours good for my eyes or is it garish

c) Is the design eye-catching

d) is it so full of text that I don't know where to start

e) is it many small grotty panels stuck together, or is it a proper A0 or A1 poster (preferably laminated).

f) are the figures and figure labels so small I don't want to make the effort to work out what they are about. Are there units on ALL the axes labels?

g) are the Figures referred to in the text and then I have to find them on the poster or is the flow clear?

h) If there are crystal pictures are there any scale bars?

i) if there is a structure is the data collection temperature specified?



2) Is there a clear aim to the work stated succinctly at the start?



3) Are there any references at all (a lot of posters fail this one).

4) Are there any acknowledgements (ditto).



5) Balance: is the Intro so long there are no results.

6) Is there enough detail regarding the Methods to judge the experimental/theoretical competence?

7) Is the title appropriate and/or has it got jargon in it?

8) Is the author beside the poster? Is there a photo of the author on the poster so I can find them easily at the coffee break if I am interested in the poster?

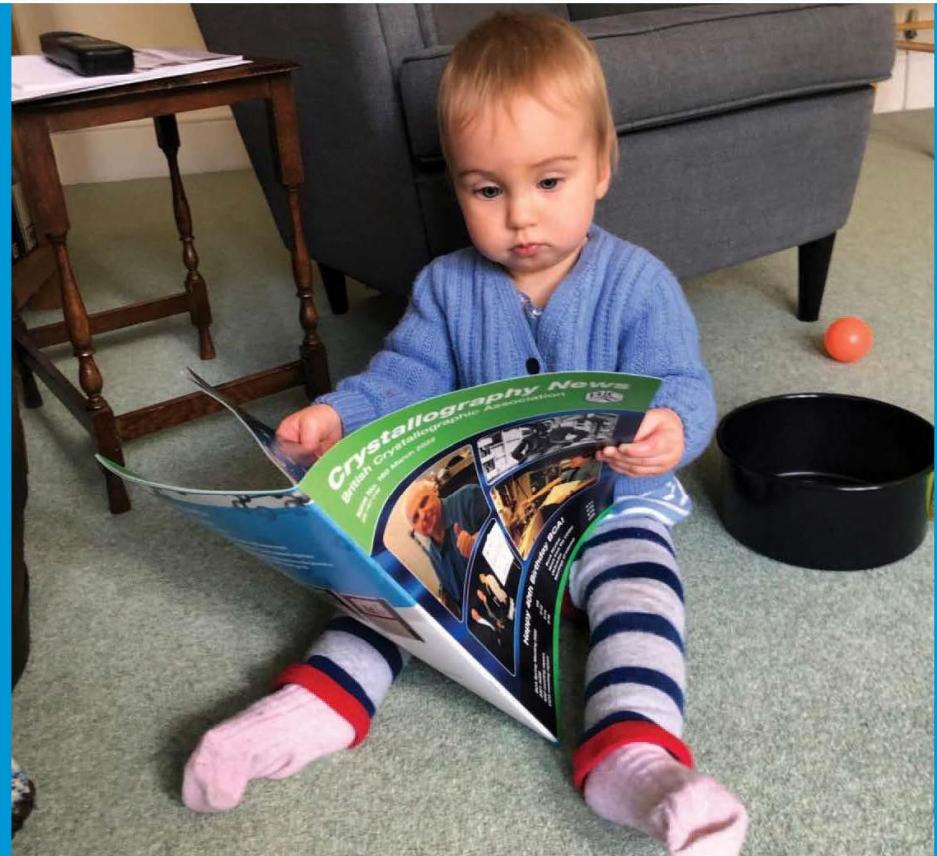
Never too young to start learning crystallography...

Madeleine, aged 9 months, reading BCA Crystallography News!

Crystallography News

British Crystallographic Association

Issue No. 162 September 2022
ISSI 1467-2790



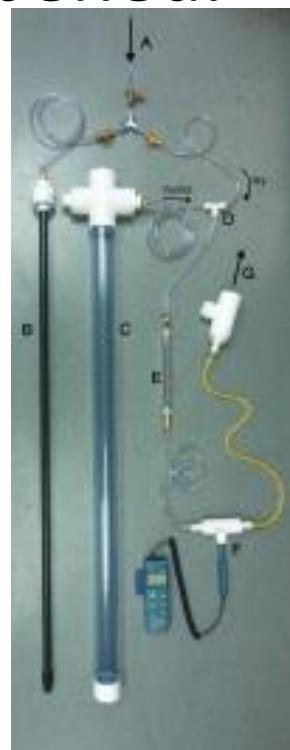
AlphaFold's implications for structure determination

AlphaFold: the Start of Something Bigger?	p6	CryoEM Book Review	p20
Spring Meeting 2023	p12	ECA Max Perutz Prize	p21
National Crystallography Service	p14	UKRI Infrastructure Fund	p22
Meeting Reports	p16	News from the CCDC	p24

- Extra slides for questions/information

Humidity device to avoid dehydration during mounting

Especially effective for those mother liquors which phase separate when crystallisation drop is opened.



Preamble

NATURE BRIEFING

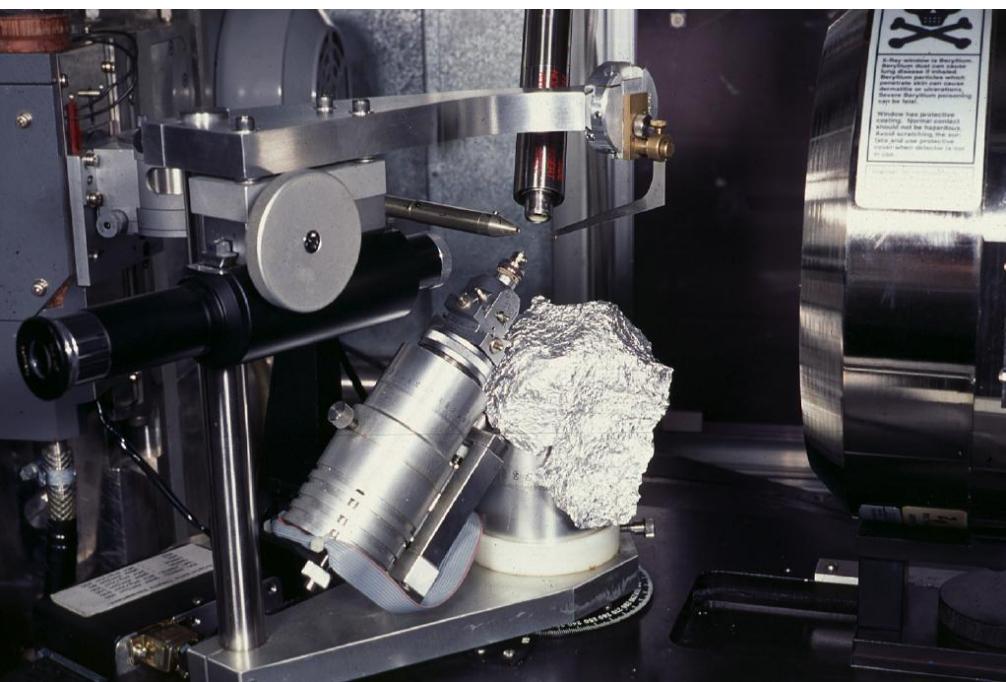
QUOTE OF THE DAY 22/3/23

“Most of the time, if you speak up, at least 30% of the people in the room will have the same question and weren’t brave enough to ask it.”

Nicola Fox, who as NASA’s new science chief will oversee more than 100 space missions, says:

“there are no stupid questions.”

SO PLEASE ASK LOTS OF QUESTIONS!!!



- Pre-centre loop.
- Speed of transfer from drop
- Block stream [avoid dehydration]
- Speed of cooling

Why bother to cover the nitrogen stream?

- Avoid crystal being dehydrated by dry nitrogen/air.
- You may wave crystal in and out of stream while cooling it: slow cooling will give ice.
- Want to cool it FAST. Much easier to do that if you whip the cover away once crystal is in position.

23 GLYCEROL, CH₂OHCHOHCH₂OH

MOLECULAR WEIGHT = 92.09

RELATIVE SPECIFIC REFRACTIVITY = 1.109

0.00 % by wt. dil.

For Values of O

A % by wt.	ρ D ₄ ²⁰	D ₄ ²⁰	C, g/l	M g-mol/l	C _o g/l	(C _o - C) g/l	(n - n _o) $\times 10^4$	n	Δ °C	O Os/kg	S g-mol/l	η n.
0.50	0.9994	1.0011	5.0	0.054	994.4	3.9	6	1.3336	0.072	0.039	0.020	1.009
1.00	1.0005	1.0023	10.0	0.109	990.5	7.7	12	1.3342	0.180	0.097	0.051	1.020
2.00	1.0028	1.0046	20.1	0.218	982.7	15.5	23	1.3353	0.411	0.221	0.119	1.046
3.00	1.0051	1.0069	30.2	0.327	974.9	23.3	35	1.3365	0.627	0.337	0.182	1.072
4.00	1.0074	1.0092	40.3	0.438	967.1	31.2	46	1.3376	0.849	0.456	0.247	1.098
5.00	1.0097	1.0115	50.5	0.548	959.2	39.0	58	1.3388	1.078	0.580	0.315	1.125
6.00	1.0120	1.0138	60.7	0.659	951.3	46.9	70	1.3400	1.316	0.708	0.385	1.155
7.00	1.0144	1.0162	71.0	0.771	943.4	54.9	82	1.3412	1.561	0.839	0.457	1.186
8.00	1.0167	1.0185	81.3	0.883	935.4	62.9	94	1.3424	1.811	0.974	0.530	1.218
9.00	1.0191	1.0209	91.7	0.996	927.4	70.9	106	1.3436	2.064	1.110	0.603	1.253
10.00	1.0215	1.0233	102.1	1.109	919.3	78.9	118	1.3448	2.323	1.249	0.678	1.288
12.00	1.0262	1.0281	123.1	1.337	903.1	95.1	142	1.3472	2.880	1.548	0.837	1.362
14.00	1.0311	1.0329	144.4	1.568	886.7	111.5	167	1.3496	3.469	1.865	1.004	1.442
16.00	1.0360	1.0378	165.8	1.800	870.2	128.0	191	1.3521	4.094	2.201	1.177	1.530
18.00	1.0409	1.0428	187.4	2.035	853.6	144.7	217	1.3547	4.756	2.557	1.359	1.627
20.00	1.0459	1.0478	209.2	2.272	836.8	161.5	242	1.3572	5.46	2.93	1.546	1.734
24.00	1.0561	1.0580	253.5	2.752	802.6	195.6	294	1.3624	7.01	3.77	1.944	1.984
28.00	1.0664	1.0683	298.6	3.243	767.8	230.4	347	1.3676	8.77	4.71	2.370	2.274
32.00	1.0770	1.0789	344.6	3.742	732.3	265.9	400	1.3730	10.74	5.78	2.814	2.632
36.00	1.0876	1.0896	391.5	4.252	696.1	302.2	455	1.3785	12.96	6.97	3.276	3.082
40.00	1.0984	1.1003	439.4	4.771	659.0	339.2	511	1.3841	15.50	8.33	3.757	3.646
44.00	1.1092	1.1112	488.1	5.300	621.2	377.1	567	1.3897				4.434
48.00	1.1200	1.1220	537.6	5.838	582.4	415.8	624	1.3954				5.402
52.00	1.1308	1.1328	588.0	6.385	542.8	455.4	681	1.4011				6.653
56.00	1.1419	1.1439	639.4	6.944	502.4	495.8	739	1.4069				8.332
60.00	1.1530	1.1551	691.8	7.513	461.2	537.0	799	1.4129				10.66
64.00	1.1643	1.1663	745.1	8.091	419.1	579.1	859	1.4189				13.63
68.00	1.1755	1.1775	799.3	8.680	376.1	622.1	919	1.4249				18.42
72.00	1.1866	1.1887	854.3	9.277	332.2	666.0	980	1.4310				27.57
76.00	1.1976	1.1997	910.2	9.883	287.4	710.8	1040	1.4370				40.49
80.00	1.2085	1.2106	966.8	10.498	241.7	756.5	1101	1.4431				59.78
84.00	1.2192	1.2214	1024.2	11.121	195.1	803.2	1162	1.4492				84.17
88.00	1.2299	1.2320	1082.3	11.752	147.6	850.7	1223	1.4553				147.2
92.00	1.2404	1.2426	1141.1	12.392	99.2	899.0	1284	1.4613				383.7
96.00	1.2508	1.2530	1200.7	13.039	50.0	948.2	1344	1.4674				778.9
100.00	1.2611	1.2633	1261.1	13.694	0.0	998.2	1405	1.4735				1487.0

24 HYDROCHLORIC ACID, HCl

Osmotic Pressure Matching: Example

Osmality: Os/kg

• 2.0M NaCl	
50mM pH 7.8 Tris HCl	3.95
• Need 20% glycerol	<u>2.9</u>
	1.05

From Tables, 0.55M NaCl has o.p. 1.05 Os/kg

→ Try 0.55 M NaCl, 20% glycerol

50mM pH 7.8 Tris HCl