Grow with the flow: a latitudinal cline in physiology is associated with more variable precipitation in *Erythranthe cardinalis*

Abstract

Local adaptation is commonly observed in nature: organisms perform well in their natal environment, but poorly outside it. Correlations between traits and latitude, or latitudinal clines, are among the most common pieces of evidence for local adaptation, but identifying the traits under selection and the selective agents is challenging. Here, we investigated a latitudinal cline in growth and photosynthesis across 16 populations of the perennial herb *Erythranthe cardinalis* (Phrymaceae). Using machine learning methods, we identify interannual variation in precipitation as a likely selective agent: Southern populations from more variable environments had higher photosynthetic rates and grew faster. We hypothesize that selection may favor a more annualized life history – grow now rather than save for next year – in environments where severe droughts occur more often. Thus our study provides insight into how species may adapt if Mediterranean climates become more variable due to climate change.

Introduction

1 Local adaptation has been documented within numerous species; populations generally
2 have higher fitness in their native environment, but perform poorly outside it (Schluter,
3 2000; Leimu and Fischer, 2008; Hereford, 2009). However, the prevalance of local adapta-
4 tion remains difficult to assess because researchers rarely test for local adaptation unless
5 there are obvious phenotypic or environmental differences (but see Hereford and Winn
6 2008). When local adaptation occurs, it frequently leads to clines in both phenotypes and

2

allele frequencies when selection varies over environmental gradients (Huxley, 1938; Endler, 1977; Barton, 1999). Phenotypic differences between populations along a cline often have a genetic basis and can be studied in a common garden (Turesson, 1922; Clausen et al., 1940; Hiesey et al., 1942). Despite a long history of studying local adaptation and clines, it remains challenging to identify exactly which traits are under selection and which differ for nonadaptive reasons. In particular, the role that physiological differences play in local adaptation is poorly understood, despite the fact that physiology is frequently assumed to explain adaptation to the abiotic environment. A related problem is identifying which of the myriad and often covarying aspects of the environment cause spatially varying selective pressures.

When populations are locally adapted, reaction norms for fitness will cross, such that local genotypes have higher fitness than foreign genotypes and rank orders change across environments (Kawecki and Ebert, 2004). The traits that underlie local adaptation, however, need not mirror this pattern. Populations can have fixed genetic differences conferring trait values that are adaptive at home but neutral or maladaptive away. Alternatively, the ability to plastically respond to a particular environment or the magnitude of response to an environment could be adaptive. We distinguish between these patterns of adaptive trait differences by referring to 'genetic variation' in trait means and 'genetic variation in plasticity', respectively. Genetic variation in plasticity is synonymous with genotype-by-environment interactions, or simply (G×E). Genetic variation in trait means and plasticity are both involved in adaptation. For example, genetic variation in photoperiod responses (Blackman et al., 2011) and developmental rate (Stinchcombe et al., 2004) allow organisms to properly time their life history with the local environment. Conversely, sun and shade plants do not have intrinsically higher or lower rates of carbon assimilation, but rather, genetic variation in plasticity cause sun plants to assimilate more under high light and shade plants under low light (Givnish, 1988). In plants especially, we know little about the prevalence and adaptive significance of variation in fundamental physiological traits like

34 photosynthesis and their impact on plant performance (Flood et al., 2011).

35 A basic approach to identify candidate traits underlying local adaptation is to find asso-
36 ciations between traits and environments. Either genetic variation in trait means and/or
37 plasticity should vary clinally along environmental gradients. Indeed, clines in ecologically
38 important traits are widespread in nature (Endler, 1977) and often adaptive, but in most
39 cases the selective agent is unknown. For example, in *Drosophila* numerous latitudinal
40 clines exist for traits like thermal tolerance (Hoffmann et al., 2002), body size (Coyne and
41 Beecham (1987) and references therein), and life history (Schmidt et al., 2005). Some
42 *Drosophila* clines have evolved multiple times (Oakeshott et al. (1982); Huey et al. (2000),
43 see also Bradshaw and Holzapfel (2001)) or shifted in response to climate change (Umina
44 et al., 2005), evincing climatic adaptation. Similarly, plant species exhibit latitudinal clines
45 in traits like flowering time (Stinchcombe et al., 2004), cyanogenesis (Kooyers and Olsen,
46 2012), leaf morphology (Hopkins et al., 2008; Stock et al., 2014), and drought resistance
47 (Kooyers et al., 2015) that likely relate to climatic variation.

48 Despite the fact that latitudinal clines have been studied for a long time, latitude *per se*
49 cannot be a selective agent. Latitude may be strongly correlated with one or two key
50 climatic variables, such as temperature, precipitation, or growing degree-days. Latitude
51 may also correlate with the strength of biotic interactions (Schemske et al., 2009) or other
52 nonclimatic aspects of the environment, though as we explain below, we do not yet have
53 compelling data that these are important in our study system. Hence, we focus on whether
54 latitude could be an effective proxy for an underlying climatic driver, in which case we
55 would expect a yet stronger relationship between traits and the key climatic variable(s)
56 driving selection. Alternatively, latitude may be more strongly related to traits than any
57 single climatic variable for at least two reasons. First, latitude may be correlated with
58 several climatic agents of selection that are individually weak, but add up to a strong
59 latitudinal cline. Alternatively, gene flow among neighbouring populations could smooth
60 out local climatic effects, since alleles will experience selection across populations linked

4

by migration (Slatkin, 1978; Paul et al., 2011; Hadfield, 2016). We refer to this as the 'climatic neighborhood'. For example, in mountainous regions average temperature at a given latitude varies widely, but in aggregate, a lower latitude set of populations will experience warmer climate than a higher latitude one. Thus, any particular low latitude population would be warm-adapted, even if it was located in a cooler (e.g. high elevation) site. Because many climatic factors vary latitudinally, and which climatic factors vary latitudinally changes over the earth's surface (e.g. coastal vs. continental), dissecting the evolution of latitudinal clines across many species will help identify generalities, such as whether thermal tolerance maxima or seasonal timing is more important (Bradshaw and Holzapfel, 2008), and whether local or regional climate shapes selective pressures.

In this study, we investigated two major questions: 1) whether genetic variation in physiological trait means or plasticity corresponds with latitude; and 2) what climatic factor(s) could plausibly be responsible for latitudinal clines. Within question 2, we tested three hypotheses outlined in the previous paragraph: latitudinal clines are explained by a single dominant climatic factor, multiple climatic factors, or the climatic neighborhood experienced by nearby population connected through gene flow. These hypotheses are not mutually exclusive since, for example, single or multiple factors in a climatic neighborhood may lead to latitudinal clines. We focused on climate because climate often determines where species are found and also can exert strong selection on populations within species. We acknowledge that other abiotic and biotic factors could contribute to selection and the overall pattern of local adaptation. Furthermore, there is a compelling need to know how populations are (or are not) locally adapted to climate so as to predict how they will respond to climate change (Aitken and Whitlock, 2013).

We examined these questions in *Erythranthe cardinalis* (formerly *Mimulus cardinalis* [Nesom 2014]) because linking physiological traits to potentially complex patterns of local adaptation requires integrating multiple lines of evidence from comparative, experimental, and genomic studies under both lab and field conditions. Many classic and contemporary

5

studies of local adaptation use *Mimulus sensu lato* species because of their natural history, easy propagation, and genetic/genomic resources (Clausen et al., 1940; Hiesey et al., 1971; Bradshaw and Schemske, 2003; Wu et al., 2008; Lowry and Willis, 2010; Wright et al., 2013). Yet, there is a deficiency of links between local adaptation and physiological mechanisms (Angert, 2006; Angert et al., 2008; Wu et al., 2010; Wright et al., 2013). We measured genetic variation in trait means and plasticity in response to temperature and drought among 16 populations distributed over 10.7°of latitude. We found a latitudinal cline of trait means in photosynthesis and growth, but little evidence for variation in plasticity. Interannual variation in precipitation and temperature are associated with this axis of variation, suggesting that climatic variance rather than mean may be an important driver of local adaptation in *E. cardinalis*. The climatic neighborhoods around populations explained trait variation better than local climate, indicating that latitudinal clines may be common because latitude integrates effects of selection on populations connected through gene flow. We place these findings in the context of life history theory and consider future directions in the Discussion.

Material and Methods

Data and annotated source code to reproduce these analyses and manuscript are available on GitHub (https://github.com/cdmuir/card-cline).

Population Selection

E. cardinalis is a perennial forb native to the Western US (California and Oregon). It is predominantly outcrossing, self-compatible, and pollinated primarily by hummingbirds. We used 16 populations from throughout the range of *E. cardinalis* (Table 1). These populations were intentionally chosen to span much of the climatic range of the species

6

based on all known occurrences (see below). Although the elevation range of *E. cardinalis* compresses slightly from south to north, latitude is not strongly confounded by elevation among our focal populations. The correlation between Latitude and Elevation among the 16 focal populations was weak ($r = -0.25$, $P = 0.34$). Seeds were collected in the field from mature, undehisced fruits left open for 2-4 weeks to dry, then stored at room temperature. To control for maternal effects, we grew a large number of field-derived seeds in the greenhouse and generated seed families for this experiment by haphazardly crossing individuals from the same population. We selected seed families to maximize the number of field-derived individuals represented. Thus, we used seeds from 154 greenhouse-derived seed families, 4–12 (mean = 9.6, median = 12) families per population.

Table 1: Latitude, longitude, and elevation (mas = meters above seal level) of 16 focal populations used in this study.

Name	Latitude	Longtiude	Elevation (mas)
Hauser Creek	32.657	-116.532	799
Cottonwood Creek	32.609	-116.7	267
Sweetwater River	32.9	-116.585	1180
Grade Road Palomar	33.314	-116.871	1577
Whitewater Canyon	33.994	-116.665	705
Mill Creek	34.077	-116.873	2050
West Fork Mojave River	34.284	-117.378	1120
North Fork Middle Tule River	36.201	-118.651	1314
Paradise Creek	36.518	-118.759	926
Redwood Creek	36.691	-118.91	1727
Wawona	37.541	-119.649	1224
Rainbow Creek	37.819	-120.007	876
Middle Yuba River	39.397	-121.082	455
Little Jamison Creek	39.743	-120.704	1603
Deep Creek	41.668	-123.11	707
Rock Creek	43.374	-122.957	326

Plant propagation

On 14 April, 2014, 3-5 seeds per family were sown directly on sand (Quikrete Play Sand, Georgia, USA) watered to field capacity in RLC4 Ray Leach cone-tainers placed in RL98 98-well trays (Stuewe & Sons, Inc., Oregon, USA). We used pure sand because *E. cardinalis* typically grows in sandy, riparian soils (A. Angert, pers. obs.). Two jumbo-sized cotton balls at the bottom of cone-tainers prevented sand from washing out. Cone-tainers sat in medium-sized flow trays (FLOWTMD, Stuewe & Sons, Inc., Oregon, USA) to continuously bottom-water plants during germination in greenhouses at the University British Columbia campus in Vancouver, Canada (49°15' N, 123°15' W). Misters thoroughly wetted the top of the sand every two hours during the day. Most seeds germinated between 1 and 2 weeks, but we allowed 3 weeks before transferring seedlings to growth chambers. We recorded germination daily between one to two weeks after sowing, and every 2-3 days thereafter. On 5 May (21 days after sowing), we transferred seedlings to one of two growth chambers (Model E-15 Conviron, Manitoba, Canada). We thinned seedlings to one plant per cone-tainer, leaving the center-most plant. 702 of 768 (91.4%) cone-tainers had plants that could be used in the experiment. We allowed one week at constant, non stressful conditions (day: 20℃, night: 16℃) for plants to acclimate to growth chambers before starting treatments. The initial size of seedlings, measured as the length of the first true leaves, did not differ between populations, families, or treatments (Table S1).

Temperature and drought treatments

We imposed four treatments, a fully-factorial cross of two temperature levels and two watering levels. The temperature levels closely simulated an average growing season at the thermal extremes of the species range, which we designate as Hot and Cool treatments. Watering levels contrasted a perennial and seasonal stream, which we refer to as Well-watered and Drought treatments. A detailed description of treatments is provided in the

8

Supplemental Materials and Methods and summarized in Fig 1. Because growth chambers cannot be subdivided, one chamber was assigned to the Hot treatment level and another to the Cool treatment level. Within each chamber, there were two Well-watered blocks and two Drought blocks. The photosynthetically active radiation in both chambers was approximately 400 μmol quanta m^{-2} s^{-1} and set to a 16:8 light:dark cycle to simulate summer growing conditions. The growth chambers did not control humidity, but because of watering and high plant transpiration rates, the relative humidity was quite high in both temperature levels (data not shown). Lower humidity would have made the drought more severe, but low soil moisture is stressful in and of itself. The total number of plants in each treatment was: $n_{\text{cool,dry}} = 169$; $n_{\text{cool,ww}} = 174$; $n_{\text{hot,dry}} = 176$; $n_{\text{hot,ww}} = 183$. Each population had 8–12 individuals per treatment level (mean = 11, median = 11).

Trait measurements

We measured five traits in response to temperature and watering treatments (Table 2).

Table 2: Key traits measured in this study.

Trait	Units
Days to germination	day
Leaf expansion rate	mm day^{-1}
Stem elongation rate	cm day^{-1}
Photosynthetic rate	μmol CO$_2$ m^{-2} s^{-1}
Mortality	probability of death

Days to germination   We tested for population variation in germination rate, measured as Days to Germination, using a lognormal survival model fit using the survreg function in the R package **survival** version 2.38 (Therneau, 2015). We treated Population as a fixed effect and Family as random effect using a Γ frailty function. Statistical significance of the Population effect was determined using analysis of deviance. Note that, unlike other traits
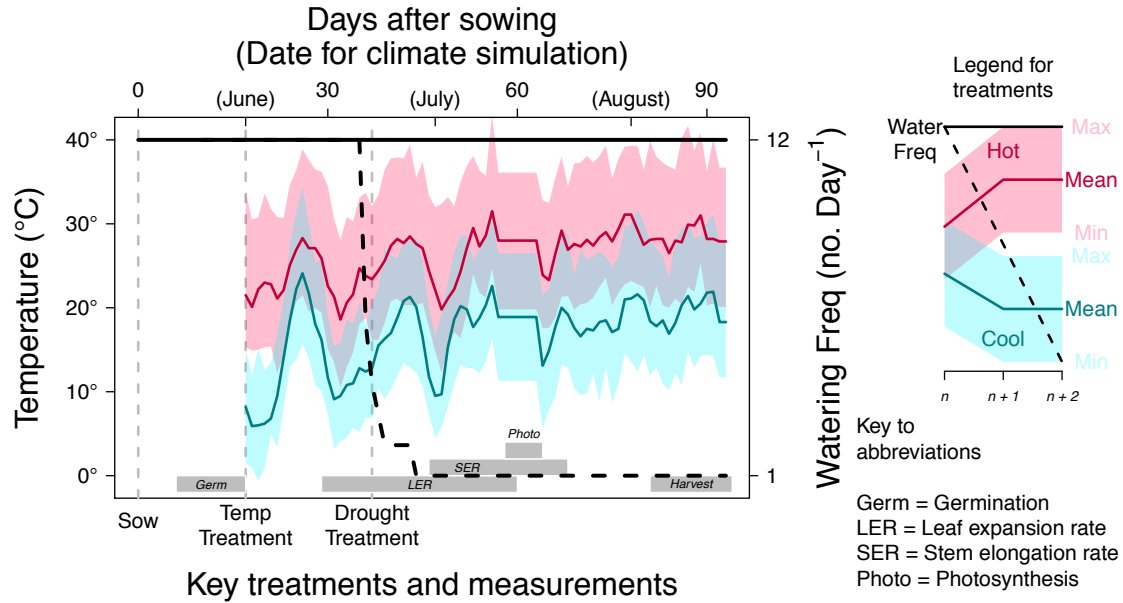
9

Figure 1: Overview of experimental treatments and timing of key trait measurements. All plants germinated within 21 days of sowing. At that time, we began temperature treatments (left axis), simulating a typical June-August weather pattern at Hot (red) and Cool (blue) sites. The bold lines track the average daily temperatures. Within each day, there was a maximum daytime temperature (top of translucent polygons) and minimum nighttime temperature (bottom of translucent polygons). The drought treatment commenced later by ramping down the frequency of bottom-watering episodes (dashed black line; right axis), while watering frequency was maintained in the control treatment (solid black line). Grey boxes on the bottom of the plot outline the period of key measurements described in the Material and Methods.

discussed below, we did not include Block, Treatment, or Population × Treatment interactions because during germination plants had not been placed into blocks and treatments had not yet been applied.

Growth rate: leaf expansion and stem elongation We measured growth rate during two phases: leaf expansion and stem elongation. Growth measurements were taken during the early vegetative stage. We censused leaf length twice per week shortly after the emergence of true leaves from 12 May – 12 June (28–59 days after sowing), resulting

in 10 measurements. We ceased measuring leaf length once it appeared to asymptote and growth shifted to stem elongation. We also censused plant height on 7 occasions (twice per week) between 29 May and 20 June (45 to 67 days after sowing) until plants began to initiate floral buds. Thus all growth measurements occured during the vegetative, pre-reproductive phase. Both leaf expansion and stem elongation were modelled separately as second-order polynomials. We used empirical Bayes' estimates of growth for each individual plant from linear mixed-effects models fit with the R package **lme4** version 1.1-12 (Bates et al., 2015).

Photosynthesis During the week of 10 to 16 June (57 to 63 days after sowing), we measured daytime photosynthetic rate on a subset of 329 plants evenly spread between treatments and families within populations. The youngest, fully-expanded leaf acclimated for 3 minutes to reach steady state in a 6-cm^2 chamber of a LI-COR 6400XT Portable Photosynthesis System (LI-COR Biosciences, Lincoln, Nebraska). We made all measurements at ambient light (400 μmol m^{-2} s^{-1} of photosynthetically active radiation), atmospheric CO_2 (400 ppm), temperature, and moderate relative humidity. All measurements were taken between 9:00 AM and 5:00 PM (3 hours after lights turned on and 5 hours before lights turned off). During this period, we suspended normal day-to-day temperature fluctuations and set daytime temperatures to the average for that period (Cool: 26.5°; Hot: 36.1°) so that all plants within a temperature level could be measured under the same conditions. We measured photosynthesis after dry down had progressed to assess differences in photosynthetic responses to drought.

Mortality We assayed mortality during twice-weekly growth measurements. We analyzed the probability of surviving until the end of the experiment as a function of population, treatment, and their interactions using a Generalized Linear Mixed Model (GLMM) assuming binomially distributed errors. We included Family and Block as random effects.

196 We assessed significance of fixed effects using Type-II Analysis of Deviance with Wald χ^2
197 tests in the R package **car** (Fox and Weisberg, 2011).

Genetic variation in trait means and plasticity

199 For all traits (Table 2) except germination (see above), we tested for Population, Treatment
200 (Temperature, Water, and Temperature $\times$ Water), and Population $\times$ Treatment interac-
201 tions (Population $\times$ Temperature, Population $\times$ Water, and Population $\times$ Temperature $\times$
202 Water). We interpreted significant Population effects to indicate genetic variation in trait
203 means and Population $\times$ Treatment interactions to indicate genetic variation in plastic-
204 ity. As mentioned above, we used survival and GLMM models for germination rate and
205 mortality, respectively. For all other traits, we used mixed model ANOVAs with Family
206 and Block included as random factors. We fit models using restricted maximum likelihood
207 in lmer, a function in the R package **lme4** (Bates et al., 2015). We determined significant
208 fixed effect terms using a step-wise backward elimination procedure implemented with the
209 step function in the R package **lmerTest** version 2.0-32 (Kuznetsova et al., 2016). We used
210 Satterthwaite's approximation to calculate denominator degrees of freedom for F-tests.
211 We also included days to germination as a covariate in growth analyses. To ensure that
212 Population and Treatment effects were specific to a particular growth phase, we included
213 germination day as a covariate in leaf expansion and stem elongation analyses.

214 Failure to detect a significant effect could be the result of Type-2 error, so we complemented
215 step-wise ANOVA (see above) by comparing effect sizes calculated in the full model. The
216 full model contains all main effects, two-way interactions (Population $\times$ Temperature,
217 Population $\times$ Water), a three-way interaction (Population $\times$ Temperature $\times$ Water), and
218 random effects. For linear mixed-effects models (leaf expansion, stem elongation, and pho-
219 tosynthesis) we used mean-squared error as a measure of effect size; for GLMM (mortality)
220 we used χ^2 as a measure of effect size. We did not include germination rate because no

Population × Treatment effects were estimated. The difference in effect size of Population versus Population × Treatment is:

$$\Delta \text{Effect Size}_{\text{Pop}-(\text{Pop}\times\text{Trt})} = \text{Effect Size}_{\text{Pop}} - \text{Effect Size}_{\text{Pop}\times\text{Trt}}$$

We calculated $\Delta \text{Effect Size}_{\text{Pop}-(\text{Pop}\times\text{Trt})}$ for all two- and three-way Population × Treatment interactions for each trait. To determine whether $\Delta \text{Effect Size}_{\text{Pop}-(\text{Pop}\times\text{Trt})}$ was significantly different than 0, we calculated 95% confidence intervals using 1000 parametric bootstrap samples simulated from fitted models. If the 95% confidence interval for a given $\Delta \text{Effect Size}_{\text{Pop}-(\text{Pop}\times\text{Trt})}$, was greater than zero then we concluded that the Population effect size was significantly larger than the Population × Treatment effect size, and vice versa if the confidence internval was less than zero. If the confidence spanned zero, then the effect sizes are not significantly different.

Principal components of germination, growth, and photosynthesis

For each single-trait model above, we extracted the Population coefficient (factoring out Treatment and other effects). The multivariate distribution of these coefficients was then summarized using principal components analysis. The first principal component of these traits (TraitPC1) loaded positively with germination, growth, and photostynthetic rate, therefore we define this as a phenotypic axis delineating fast to slow growth.

Identifying putative selective agents

Latitudinal clines are common, but it is often difficult to ascribe this variation to a particular selective agent. To reiterate, we tested three non-mutually exclusive hypotheses about how such latitudinal clines emerge: 1) one or two climatic variables explain latitudinal trait variation; 2) latitude is a proxy for multiple climatic factors that together shape

13

trait variation; and 3) latitude integrates selection in a broader climatic neighborhood. We found that a population's position along TraitPC1 correlated strongly with the latitude of origin (see Results) and next used Random Forest regression (Liaw and Wiener, 2002) to identify putative climatic factors underlying trait-latitude associations in *E. cardinalis*. We reasoned that if we identified a single climatic factor that explained more trait variation than latitude, then this would suggest that factor is a key selective agent underlying the latitudinal cline (Hypothesis 1). On the other hand, if multiple climatic factors together are necessary to explain trait variation, then this would suggest that many climatic factors together have imposed selection for the latitudinal cline (Hypothesis 2). We hereafter refer to factors identified in this analysis as 'Climate-TraitPC1' variables.

To test Hypothesis 3 about climatic neighborhoods driving selection, we directly competed local with neighborhood climate. The logic is that if the climatic analysis can identify candidate climatic factors important for local adaptation, then stronger correlations with neighbourhood climate would suggest a role for gene flow. We used the immediate collection location for local climate. For climate neighborhoods, we sampled climate at 1000 random points (at 90-m resolution) within a 62-km radius buffer around the collection and took the average. We chose this buffer radius based on population genetic structure, as inferred from $\approx$25,000 restriction-site associated SNPs among 49 populations from across the range (Paul et al., In review). Spatial autocorrelation in allele frequencies persists for 62 km. However radii of 10 km^2 and 100 km^2 resulted in similar outcomes (data not shown). Since *E. cardinalis* is found exclusively in riparian areas, we only selected points along streams using the National Hydrogeoraphy Dataset (United States Geological Survey, 2015). Climatic means and variances (see below) were weighted by their climatic suitability as determined using a multimodel ensemble average of ecological niche models (Angert et al., 2016). In addition to competing local and neighborhood climate, we compared the univariate correlation between local and neighborhood climate with TraitPC1 and Latitude using paired *t*-tests. We adjusted degrees of freedom to account for the fact that many climatic

14

factors are highly correlated and not independent. Specifically, we calculated the effective number of independent climatic factors (M_{eff}) using the formula $M_{\text{eff}} = 1 + (M - 1)(1 - \text{Var}(\lambda)/M)$ (Chevrud, 2001), where M is the original number of climatic factors and λ are the eigenvalues of the correlation matrix of all climatic factors.

To help eliminate potentially spurious correlations between TraitPC1 and climate, we tested for overlap between climatic variables that best predict latitude of all *E. cardinalis* occurrence records (see detail below), not just the 16 focal populations. We refer to these climatic factors as 'Climate-Latitude' variables. The logic is that climatic factors associated with both TraitPC1 and latitude for all populations are more likely to be important selective agents than climatic factors that happen to correlate with TraitPC1 but do not covary with latitude throughout the *E. cardinalis* range. If a climatic factor is driving the latitudinal cline in TraitPC1, then we expect that climatic factor will correlate strongly with latitude of occurrence localities. Therefore, we did not consider Climate-TraitPC1 variables to be candidate selective agents unless the same or very similar variable was found in the Climate-Latitude analysis. However, we do not interpret potential selective agents identified in Climate-Latitude analyses alone, because the goal was to explain the latitudinal clines in traits, not all aspects of climate that vary with latitude.

We selected Climate-Latitude and Climate-TraitPC1 variables independently using Variable Selection Using Random Forest (VSURF) algorithm in the R package **VSURF** version 1.0.3 (Genuer et al., 2016). Random Forest regression is useful for cases like ours when the number of potential predictors is similar to or greater than the number of observations ('high p, low n' problem). VSURF is a multistep algorithm that progressively retains or eliminates variables based on their importance over regression trees in the forest. Variable importance is defined as the average amount a climate variable reduces mean-squared error in the predicted response (TraitPC1 or Latitude), compared to a randomly permuted dataset, across all trees in the random forest (see Genuer et al. [2015] for further detail). Hence, VSURF automatically eliminates unimportant and redundant variables based on

the data without having to arbitrarily choose among colinear climate variables before the analysis. We kept only variables selected for prediction, the most stringent criterion. We visually depict how we selected climatic variables in Fig 2.

For Climate-Latitude analyses, we compiled a representative set of 356 recent (since 2000) known *E. cardinalis* occurrences from a comprehensive set of herbarium records and an exhaustive field survey in 2010-11 (Angert et al., 2016). These occurrences were thinned by 50% to correct for uneven sampling. For both Climate-TraitPC1 analyses (16 focal populations) and Climate-Latitude (many populations), we used a 90-m digital elevation model from HydroSHEDS (Lehner et al., 2006) to extract elevation. Monthly interpolated climate layers were calculated using ClimateWNA version 5.30 (Wang et al., 2012), which accurately downscales climate data specifically for the rugged topography of western North America. For each occurence, we calculated bioclimatic variables using the biovars function in the R package **dismo** version 1.1-1 (Hijmans et al., 2016). We included 24 climatic factors, 9 from ClimateWNA and 15 bioclimatic variables (Table S2). The bioclimatic variables included all permutations of two climatic factors, temperature and precipitation, and six temporal scales (annual average, coldest quarter, warmest quarter, wettest quarter, driest quarter, or seasonality) as well as mean diurnal range, isothermality, and annual temperature range. For each variable, we calculated both a 30-year normal by averaging annual values between 1981 and 2010 and 30-year coefficient of variation, a standardized metric of interannual climatic variation. Temperatures were converted to Kelvin to be on a ratio scale appropriate for calculating the coefficient of variation (CV). In total, the VSURF algorithm selected among 96 climate variables: 24 climatic factors × 2 types (30-year average and CV) × 2 spatial scales (local and neighborhood).

16

Results

A coordinated latitudinal cline in germination, growth, and photosynthesis

There are strong genetically-based trait differences in time to germination, growth, and photosynthetic rate among populations of *E. cardinalis*, as evidenced by large and significant population effects for these traits (Table 3). A single principal component captured 71.6 % of the trait variation among populations, defining an axis of variation from fast to slow growth. A population's position along this axis strongly covaried with its latitude of origin; southern populations grew faster than northern populations (Fig 3). There were similar latitudinal clines for individual traits underlying PC1 (Figures S1 to S4).

Table 3: Summary of Population, Treatment, and Population × Treatment effects. We used different statistical modeling for the diverse traits assayed – glmer: generalized linear mixed model using the R package **lme4** (Bates et al., 2015); lmer: linear mixed model using the R package **lme4** (Bates et al., 2015); survreg: survival regression using the R package **survival** (Therneau, 2015). Note that temperature and water treatments were imposed after germination, hence are not applicable to this trait. Complete analysis of variance/deviance tables for each trait are available in the Supporting Information. Key to statistical significance: *$P < 0.05$; ** $P < 0.01$; *** $P < 0.001$

Trait Statistical model	Germination survreg	Leaf expansion lmer	Stem elongation lmer	Photosynthesis lmer	Mortality glmer
Population	***	***	***	***	
Temperature	NA	***	***	**	***
Water	NA	*			***
Pop × Temp	NA			*	
Pop × Water	NA	*			
Temp × Water	NA				***
Pop × Temp × Water	NA				

17

Little evidence for variation in plasticity

In contrast to the genetic variation in trait means described above, we found little evidence of G×E in *E. cardinalis*. There were only two statistically significant Population × Treatment interactions (Table 3, Fig. S5), but these were not strong compared to Population and Temperature effects. Otherwise, populations responded similarly to treatments: faster growth in the hot treatment, slower growth in the dry treatment, and high mortality in the hot, dry treatment (Table 3). Complete ANOVA tables are available in the Supporting Information (Tables S3 to S6).

The effect size of Population was significantly larger than that for Population × Treatment interactions (Fig. S6) in most cases. For leaf expansion, Population had a significantly larger effect size than Population × Treatment interactions in 2 of 3 comparisons (Fig. S6A). For stem elongation (Fig. S6B) and mortality (Fig. S6D), Population effect sizes were significantly larger than all Population × Treatment interactions. For Photosynthesis, Population and Population × Treatment effect sizes were not significantly different (Fig. S6C), presumably because we had a smaller sample size.

Neighborhood climatic variability best explains latitudinal cline

Interannual variation in climate averaged over each populations's climatic neighborhood correlated most strongly with trait variation and latitude of *E. cardinalis* occurrences (Fig. 4, Table S7). All 16 Climate-Latitude and 7 Climate-TraitPC1 variables were neighborhood rather than local variables (Fig. 4). In fact, neighborhood climate almost always correlated better with TraitPC1 and Latitude than local climate (Fig. 5). On average, neighborhood Climate-TraitPC1 correlation coefficients were 0.16 higher than correlations with local-scale climate variables (paired *t*-test, $t = 7.87$, d.f. $= 33.6$, $P = 3.94 \times 10^{-9}$). Likewise, neighborhood Climate-Latitude correlation coefficients were 0.13 higher than

those for local-scale climate (paired t-test, $t = 6.71$, d.f. $= 36.8$, $P = 7.22 \times 10^{-8}$). Among Climate-Latitude and Climate-TraitPC1 variables, neighborhood climatic variability over 30 years (1981–2010) in either winter precipitation (bio16$_\sigma$) and/or temperature (bio11$_\sigma$) are the strongest candidates to explain the latitudinal cline in *E. cardinalis* (see Table S2 for a key to climate variable abbreviations). Note that the coefficient of variation of a climatic factor is subscripted with σ whereas the mean is subscripted with μ. More specifically, greater winter precipitation variability and lower winter temperature variability are associated with Southern latitudes and higher TraitPC1 values (Fig. 6A,B). Neighborhood interannual variation in winter precipitation (bio16$_\sigma$) was the most important Climate-Latitude variable (Fig. 4A). However, neighborhood bio16$_\sigma$ did not overlap with Climate-TraitPC1 variables (Fig. 4B). We nevertheless consider it a plausible candidate for two reasons. First, neighborhood bio16$_\sigma$ correlated strongly with TraitPC1 (Fig. 6A). Second, one of the most important Climate-TraitPC1 variables (neighborhood bio15$_\sigma$; Fig. 6B,C) is very similar to bio16$_\sigma$. In Mediterranean climates like California, most precipitation occurs in the wettest quarter (winter), so years with low winter precipitation also have low precipitation seasonality. Hence, highly variable year-to-year winter precipitation at lower latitude (Fig. 6D) is closely associated with large swings in precipitation seasonality (Fig. 6C).

Interannual variation in temperature of the coldest quarter (neighborhood bio11$_\sigma$) is another plausible candidate because it was the only variable in both Climate-Latitude and Climate-TraitPC1 analyses (Fig. 4). Neighborhood bio11$_\sigma$ explained more variation in TraitPC1 than latitude (latitude $r^2 = 0.55$ vs. bio11$_\sigma$ $r^2 = 0.6$; Fig. S7), whereas neighborhood bio16$_\sigma$ did slightly worse (bio16$_\sigma$ $r^2 = 0.49$). Models using bio15$_\sigma$ or bio11$_\sigma$ to predict TraitPC1 also had significantly lower Akaike Information Criteria (AIC) than the latitude model (AIC of different models – bio15$_\sigma$: 48.5; bio11$_\sigma$: 52.4; latitude: 54.5). The best two-factor model including both neighborhood bio15$_\sigma$ and bio11$_\sigma$ did not significantly improve explanatory power ($r^2 = 0.71$, AIC= 49.2). In summary, either variation in precip-

itation or temperature seasonality may be important selective agents, but there is no strong evidence that they are both important. The most important Climate-TraitPC1 variable, neighborhood variation in mean diurnal range (bio2$_\sigma$; Fig. 4B) did not have any obvious similarity to Climate-Latitude variables. Given the large number of potential associations, we therefore think this may be a spuriously strong relationship.

Discussion

We found evidence for one of two common signatures of local adaptation in the perennial herb *Erythranthe cardinalis*. Latitudinal clines in germination rate, photosynthesis, and growth suggest adaptive differentiation in important physiological traits of the species. However, we caution that these are candidate adaptive traits and that we cannot yet rule out nonadaptive demographic processes such as a recent range expansion toward higher latitude (Paul et al., In review; Sheth and Angert, 2017). In contrast, we found little evidence for variation in plasticity to temperature or drought. Due to low replication within families, we did not have power to assess within-population genotype-by-environment interactions, which may be present. As we discuss below, low variation in plasticity among populations may indicate that some dimensions of the fundamental abiotic niche are relatively conserved. Note that statistical power to detect significant plasticity is lower than that for differences in trait means, but the effect size of variation in plasticity was significantly less than that for trait means in most cases (Fig. S6). Finally, our results suggest that neighborhood-scale climate and interannual variation are more important selective agents than local averages. In the paragraphs that follow, we tie these results into the broader threads of evolutionary theory that might help explain why variation in physiological trait means changes clinally, whereas plastic responses to temperature and drought are relatively static. One caveat to bear in mind is that we are limited by the size of the climate grid ($\approx 90 \text{ m}^2$) and therefore unable to detect very fine-scale local adaptation.

20

Evolutionary theory indicates that the shape of fitness tradeoffs, demography, and gene flow can constrain adaptation (Levins, 1968; Ronce and Kirkpatrick, 2001; Lenormand, 2002) and hence the type of variation maintained within species. Specifically, adaptive variation can be maintained by spatially varying selection if tradeoffs are not too strong, demography is symmetric, and/or maladaptive gene flow is low. Strong tradeoffs can prevent local adaptation in spatially variable environments because selection favors habitat specialists that track a specific habitat regardless of its frequency in the environment (Levins, 1968). For example, a riparian specialist may experience similar selection in rivers of high rainfall regions and deserts, even though the habitat is much rarer in the latter. In *E. cardinalis* we found substantial genetically based variation among populations along a phenotypic axis from fast to slow growth that varied over a large spatial scale (Fig. 3). If this variation is adaptive, it suggests one of several possibilities to investigate in the future: the fitness tradeoff between low versus high latitude environments is not too strong nor swamped by demographic asymmetry or maladaptive gene flow. That is, alleles favoured at one latitude are not strongly selected against when they flow to another population, allowing locally adaptive genetic variation to be maintained by spatially heterogeneous selection. We also know from previous work that population size does not vary strongly with latitude (Angert, unpub. data). Gene flow appears to be high, but attenuates at broad spatial scales, especially between southern (< 35°N) and northern portions of the range (Paul et al., In review).

Nevertheless, local gene flow from similar environments may shape how selection varies with latitude. Theory predicts that populations will not be perfectly adapted to their immediate habitat when there is gene flow from surrounding populations with different optima (Lenormand, 2002). With spatial heterogeneity and gene flow, traits will not covary perfectly with the local optimum (Slatkin, 1978; Paul et al., 2011; Hadfield, 2016), but should instead better match the average environment experienced by nearby populations connected through gene flow, which we refer to as the climatic neighborhood. Gene flow

21

and spatial heterogeneity may therefore be important in maintaining genetic variation (Yeaman and Jarvis, 2006). As this hypothesis predicts, climatic neighborhoods (62-km buffer around populations) correlated with traits and latitude of occurrences better than local climate (Fig. 4). We interpret this as suggestive evidence that gene flow between neighboring *E. cardinalis* populations shapes selection – populations are locally adapted to prevailing climate in their neighborhood, but perhaps not perfectly adapted to their local climate. This may not greatly constrain local adaptation because local and neighborhood climate values were generally similar in *E. cardinalis* populations (Fig. 5), at least at the resolution of ClimateWNA (90 m^2). Therefore, we would predict in reciprocal transplants that populations whose local climate is farther from their neighborhood average would be less well adapted than those close to their neighborhood average.

An alternative, nonbiological, explanation for why neighborhood climate correlates better than local climate is that averaging climate over a wider area integrates out error in ClimateWNA values. ClimateWNA interpolates between weather stations using a fine-scale digital elevation model to accurately downscale climate data (Wang et al., 2012), but there is obviously some error. If the error were randomly distributed about the true climatic values, then averaging climate over a larger sample of points could give a more precise estimate, which might explain why traits and latitude correlate better with climatic neighbourhood. For this technical artefact to be important, the measurement error in local climate would have to be large relative to true variation in climate among locations within the climatic neighbourhoods. We do not know the relative magnitude of these factors, but future work should compare ClimateWNA-predicted local climate with actual plant-level measurements to estimate the magnitude of this error.

It is reasonable to predict that southern populations, which appear to experience more frequent drought years (see below), might have physiological adaptations to respond to drought stress to survive and grow in drier soil. We found little evidence for this type of drought tolerance; all populations responded to drought and temperature similarly (Ta-

22

ble 3). Plants grew faster in the Hot treatment, but there was little effect of drought on growth. Rather, the effects of drought took longer to materialize but resulted in high mortality, especially in the Hot treatment. However, there was no differential mortality among populations in this treatment. Although our results indicate that this axis of the species niche may be constrained, plants have multiple ways to resist drought through both tolerance and escape (Ludlow, 1989; Kooyers, 2015). Next, we consider why drought tolerance may less important in local adaptation than a form of escape for this species.

We hypothesize that tolerance to dry soil may be constrained by a combination of strong fitness tradeoffs, demographic asymmetry, and gene flow. Soil moisture in riparian habitats where *E. cardinalis* lives is highly heterogeneous at very small spatial scales (several meters). Plants in the stream never have to tolerate drought whereas plants only a few meters away may experience extreme drought since there is little direct precipitation during the growing season in Mediterranean climates of western North America. We hypothesize that alleles confering greater drought tolerance may be quite costly in well-watered soils, and *vice versa*, leading to strong fitness tradeoffs. Such tradeoffs would promote specialization to one soil moisture or another, thereby inhibiting the evolution of broad environmental tolerance within a population. Demography and gene flow may reinforce niche conservatism. A new mutant with increased drought tolerance that could survive at the resource-poor margin of a population would likely be demographically overwhelmed by the larger census populations that can be maintained in higher-resource environments. Infrequent wet years may also produce most seeds, so selection is weighted towards alleles that have high fitness in the wet environment, even if dry years are more frequent (Templeton and Levin, 1979; Brown and Venable, 1986). However, demographic asymmetry should equally hinder the evolution of both drought tolerance and escape, so it should not explain why one mechanism evolves but not the other. Finally, gene flow, which is generally high among *E. cardinalis* populations within the same ecoregion (Paul et al., In review), will thwart local adaptation and reinforce specialization. Thus, the spatial grain of the environment,

23

demographic asymmetry, and gene flow may conspire to constrain local adaptation along this environmental axis. Consistent with this hypothesis, recent record-setting droughts have caused the decline or even local extinction of some natural populations of *E. cardinalis* (Sheth and Angert, 2017).

In sum, these results indicate that genetic differences in physiology and growth are better candidates than plastic responses to temperature and drought as mediators of local adaptation to climate in *E. cardinalis*. Next, we would like to understand why variation in these particular traits may be adaptive. We argue that temporally more variable environments, as experienced by southern populations, select for a more 'annualized' life-history strategy, a form of drought escape. Demographic observations in natural populations of *E. cardinalis* reveal that southern populations tend to flower earlier at a smaller size, while northern populations invest more in vegetative growth (Sheth and Angert, 2017). In this experiment, the fastest growing plants began producing flowers in ~ 60 days (data not shown), suggesting that rapid vegetative develop may likewise affect flowering time. The association between position along the 'fast-slow' continuum and associated traits in *E. cardinalis* is similar to interspecific relationships between growth, functional traits, and life history (Adler et al., 2014; Salguero-Gómez et al., 2016). However, we cannot exclude unexplored factors (e.g. edaphic conditions, competitors, pollinators, etc.) which may also contribute to the latitudinal cline.

Greater investment in aboveground growth, as opposed to belowground storage for future seasons, may be favoured in climates with more frequent drought years, but maladaptive in climates with more consistent precipitation. In a stable environment where winter survivorship is assured in most years, failure to store resources may reduce lifetime fitness. But for perennial herbs in Mediterranean climates, a dry winter (rainy season) can kill the rhizomes (underground stems that store nutrients for future growth) before emergence or aboveground stems before flowering. If drought years occur frequently enough, selection may favour the fast-growing strategy because there is no advantage to storage if drought

24

kills plants before flowering. Considering life-history strategy as a continuum from no storage (annual) to lots of storage (perennial), we hypothesize that the optimal allocation to aboveground growth is more 'annualized' in southern climates that have greater interannual variation in precipitation.This is a form of drought escape in that plants are investing more reproduction in the present to avoid possible drought in subsequent years, but is distinct from classic drought escape syndromes in which plants speed up development early in the season before the onset of drought.

The hypothesis that greater precipitation variability selects for an annualized life history is tentative, but consistent with theory and data from other species. Life history theory shows that less variable environments are one factor that favours the evolution of perenniality (Stearns, 1976; Iwasa and Cohen, 1989; Friedman and Rubin, 2015). Populations of the perennial *Plantago asiatica* show a similar latitudinal cline in growth and allocation to storage (Sawada et al., 1994) but attribute the cline to variation in growing season length. There are also life history clines in the closely related species *E. guttata*, but the underlying traits and climatic drivers are quite different. Annual *E. guttata* flower sooner and produce fewer stolons in response to climates with shorter seasons and more intense summer drought (Lowry and Willis, 2010; Friedman et al., 2015; Kooyers et al., 2015). In contrast, there are no truly annual (monocarpic and semelparous) populations of *E. cardinalis*. Rather, our hypothesis states that climatic variability selects on quantitative variation in allocation to growth versus storage.

In summary, we found evidence for a coordinated latitudinal cline in germination rate, photosynthesis, and growth, suggesting local adaptation. We therefore predict to find different optima for these traits in different climates. We did not find evidence that the relative performance of populations shifts with temperature or watering regime, suggesting relatively little variation in plasticity. Exploratory analysis implicate that more variable precipitation regimes at lower latitude could drive much of the latitudinal cline, though other climatic factors could also contribute. Interestingly, the climatic neighborhood may

25

shape selective pressures more than local climate. In the future, we will use field experiments to test whether greater variation in precipitation selects for faster growth and if selection on temperature/drought responses does not vary among populations. By doing so, we aim to understand why certain physiological and developmental mechanisms, but not others, contribute to local adaptation.

References

Adler, P. B., R. Salguero-Gómez, A. Compagnoni, J. S. Hsu, J. Ray-Mukherjee, C. Mbeau-Ache, and M. Franco, 2014. Functional traits explain variation in plant life history strategies. Proceedings of the National Academy of Sciences of the United States of America 111:740–745.

Aitken, S. N. and M. C. Whitlock, 2013. Assisted gene flow to facilitate local adaptation to climate change. Annual Review of Ecology, Evolution, and Systematics 44:367–388.

Angert, A. L., 2006. Demography of central and marginal populations of monkeyflowers (*Mimulus cardinalis* and *M. lewisii*). Ecology 87:2014–2025.

Angert, A. L., M. J. Bayly, S. N. Sheth, and J. R. Paul, 2016. Testing range-limit hypotheses using range-wide habitat suitability and occupancy for the scarlet monkeyflower (*Mimulus cardinalis*) .

Angert, A. L., H. Bradshaw Jr, and D. W. Schemske, 2008. Using experimental evolution to investigate geographic range limits in monkeyflowers. Evolution 62:2660–2675.

Barton, N. H., 1999. Clines in polygenic traits. Genetics Research 74:223–236.

Bates, D., M. Mächler, B. Bolker, and S. Walker, 2015. Fitting linear mixed-effects models using lme4. Journal of Statistical Software 67:1–48. URL `http://CRAN.R-project.org/package=lme4`. R package version 1.1-7.

563 Blackman, B. K., S. D. Michaels, and L. H. Rieseberg, 2011. Connecting the sun to
564   flowering in sunflower adaptation. Molecular Ecology 20:3503–3512.

565 Bradshaw, H. and D. W. Schemske, 2003. Allele substitution at a flower colour locus
566   produces a pollinator shift in monkeyflowers. Nature 426:176–178.

567 Bradshaw, W. and C. Holzapfel, 2008. Genetic response to rapid climate change: it's
568   seasonal timing that matters. Molecular ecology 17:157–166.

569 Bradshaw, W. E. and C. M. Holzapfel, 2001. Genetic shift in photoperiodic response corre-
570   lated with global warming. Proceedings of the National Academy of Sciences 98:14509–
571   14511.

572 Brown, J. S. and D. L. Venable, 1986. Evolutionary ecology of seed-bank annuals in
573   temporally varying environments. The American Naturalist 127:31–47.

574 Chevrud, J., 2001. A simple correction for multiple comparisons in interval mapping genome
575   scans. Heredity 87:52–58.

576 Clausen, J., D. Keck, and W. Hiesey, 1940. Experimental studies on the nature of species.
577   I. The effects of varied environments on western American plants, vol. 520. Carnegie
578   Institution of Washington, Washington, D.C.

579 Cleveland, W. S., E. Grosse, and W. M. Shyu, 1992. Local regression models. Statistical
580   models in S Pp. 309–376.

581 Coyne, J. A. and E. Beecham, 1987. Heritability of two morphological characters within
582   and among natural populations of *Drosophila melanogaster*. Genetics 117:727–737. URL
583   http://www.genetics.org/content/117/4/727.

584 Endler, J., 1977. Geographic variation, clines, and speciation. Princeton University Press,
585   Princeton, NJ.

586 Flood, P. J., J. Harbinson, and M. G. M. Aarts, 2011. Natural genetic variation in plant

photosynthesis. Trends in Plant Science 16:327–335.

Fox, J. and S. Weisberg, 2011. An R Companion to Applied Regression. Second ed. Sage, Thousand Oaks CA.

Friedman, J. and M. J. Rubin, 2015. All in good time: Understanding annual and perennial strategies in plants. American Journal of Botany 102:497–499.

Friedman, J., A. D. Twyford, J. H. Willis, and B. K. Blackman, 2015. The extent and genetic basis of phenotypic divergence in life history traits in *Mimulus guttatus*. Molecular Ecology 24:111–122.

Genuer, R., J.-M. Poggi, and C. Tuleau-Malot, 2015. VSURF: An R package for variable selection using random forests. The R Journal 7:19–33.

———, 2016. VSURF: Variable Selection Using Random Forests. URL `http://CRAN.R-project.org/package=VSURF`. R package version 1.0.3.

Gilbert, P., 2014. Brief User's Guide: Dynamic Systems Estimation. URL `http://cran.r-project.org/web/packages/dse/vignettes/dse-guide.pdf`.

Givnish, T. J., 1988. Adaptation to sun and shade: a whole-plant perspective. Functional Plant Biology 15:63–92.

Hadfield, J. D., 2010. Mcmc methods for multi-response generalized linear mixed models: The MCMCglmm R package. Journal of Statistical Software 33:1–22. URL `http://www.jstatsoft.org/v33/i02/`.

———, 2016. The spatial scale of local adaptation in a stochastic environment. Ecology Letters 19:780–788.

Hereford, J., 2009. A quantitative survey of local adaptation and fitness trade-offs. The American Naturalist 173:579–588.

Hereford, J. and A. A. Winn, 2008. Limits to local adaptation in six populations of the

annual plant *Diodia teres*. New Phytologist 178:888–896.

Hiesey, W., M. Nobs, and O. Björkman, 1971. Experimental studies on the nature of species. V. Biosystematics, genetics, and physiological ecology of the Erythranthe section of *Mimulus*, vol. 628. Carnegie Institution of Washington, Washington, D.C.

Hiesey, W. M., J. Clausen, and D. D. Keck, 1942. Relations between climate and intraspecific variation in plants. American Naturalist Pp. 5–22.

Hijmans, R. J., S. Phillips, J. Leathwick, and J. Elith, 2016. dismo: Species distribution modeling. URL `http://CRAN.R-project.org/package=dismo`. R package version 1.1-1.

Hoffmann, A. A., A. Anderson, and R. Hallas, 2002. Opposing clines for high and low temperature resistance in *Drosophila melanogaster*. Ecology Letters 5:614–618.

Hopkins, R., J. Schmitt, and J. R. Stinchcombe, 2008. A latitudinal cline and response to vernalization in leaf angle and morphology in *Arabidopsis thaliana* (Brassicaceae). New Phytologist 179:155–164.

Huey, R. B., G. W. Gilchrist, M. L. Carlson, D. Berrigan, and L. Serra, 2000. Rapid evolution of a geographic cline in size in an introduced fly. Science 287:308–309.

Huxley, J. S., 1938. Clines: an auxiliary taxonomic principle. Nature 142:219–220.

Iwasa, Y. and D. Cohen, 1989. Optimal growth schedule of a perennial plant. American Naturalist 133:480–505.

Kawecki, T. J. and D. Ebert, 2004. Conceptual issues in local adaptation. Ecology Letters 7:1225–1241.

Kooyers, N. J., 2015. The evolution of drought escape and avoidance in natural herbaceous populations. Plant Science 234:155–162.

Kooyers, N. J., A. B. Greenlee, J. M. Colicchio, M. Oh, and B. K. Blackman, 2015. Replicate altitudinal clines reveal that evolutionary flexibility underlies adaptation to drought

635   stress in annual *Mimulus guttatus*. New Phytologist 206:152–165.

636 Kooyers, N. J. and K. M. Olsen, 2012. Rapid evolution of an adaptive cyanogenesis cline
637   in introduced north american white clover (*Trifolium repens* L.). Molecular Ecology
638   21:2455–2468.

639 Kuznetsova, A., P. Bruun Brockhoff, and R. Haubo Bojesen Christensen, 2016.
640   lmerTest: Tests in Linear Mixed Effects Models. URL `http://CRAN.R-`
641   `project.org/package=lmerTest`. R package version 2.0-32.

642 Lehner, B., K. Verdin, and A. Jarvis, 2006. HydroSHEDS technical documentation. World
643   Wildlife Fund, Washington, D.C. URL `www.worldwildlife.org/hydrosheds`.

644 Leimu, R. and M. Fischer, 2008. A meta-analysis of local adaptation in plants. PLoS One
645   3:e4010–e4010.

646 Lenormand, T., 2002. Gene flow and the limits to natural selection. Trends in Ecology &
647   Evolution 17:183–189.

648 Levins, R., 1968. Evolution in changing environments: some theoretical explorations.
649   Princeton University Press, Princeton, New Jersey.

650 Liaw, A. and M. Wiener, 2002. Classification and regression by randomforest. R News
651   2:18–22. URL `http://CRAN.R-project.org/doc/Rnews/`.

652 Lowry, D. B. and J. H. Willis, 2010. A widespread chromosomal inversion polymorphism
653   contributes to a major life-history transition, local adaptation, and reproductive isola-
654   tion. PLoS biology 8:2227.

655 Ludlow, M. M., 1989. Strategies of response to water stress. Pp. 269–281, *in* K. H. Kreeb,
656   H. Richter, and T. M. Hinckley, eds. Structural and functional responses to environmental
657   stresses. SPB Academic, The Hague, The Netherlands.

658 Nesom, G. L., 2014. Taxonomy of *Erythranthe* sect. *Erythranthe* (phrymaceae). Phytoneu-

ron 31:1–41.

Oakeshott, J., J. Gibson, P. Anderson, W. Knibb, D. Anderson, and G. Chambers, 1982. Alcohol dehydrogenase and glycerol-3-phosphate dehydrogenase clines in *Drosophila melanogaster* on different continents. Evolution Pp. 86–96.

Paul, J. R., T. L. Parchman, B. Econopouly, C. A. Buerkle, and A. L. Angert, In review. Population genomics and range limits: diversity and differentiation across the geographic range of *Mimulus cardinalis* (Phyrmaceae) .

Paul, J. R., S. N. Sheth, and A. L. Angert, 2011. Quantifying the impact of gene flow on phenotype-environment mismatch: A demonstration with the scarlet monkeyflower *Mimulus cardinalis*. The American Naturalist 178:S62–S79.

Pfaff, B., 2008. VAR, SVAR and SVEC models: Implementation within R package vars. Journal of Statistical Software 27:1–32.

Ronce, O. and M. Kirkpatrick, 2001. When sources become sinks: migrational meltdown in heterogeneous habitats. Evolution 55:1520–1531.

Salguero-Gómez, R., O. R. Jones, E. Jongejans, S. P. Blomberg, D. J. Hodgson, C. Mbeau-Ache, P. A. Zuidema, H. de Kroon, and Y. M. Buckley, 2016. Fast–slow continuum and reproductive strategies structure plant life-history variation worldwide. Proceedings of the National Academy of Sciences of the United States of America 113:230–235.

Sawada, S., Y. Nakajima, M. Tsukuda, K. Sasaki, Y. Hazama, M. Futatsuya, and A. Watanabe, 1994. Ecotypic differentiation of dry matter production processes in relation to survivorship and reproductive potential in *Plantago asiatica* populations along climatic gradients. Functional Ecology 8:400–409.

Schemske, D. W., G. G. Mittelbach, H. V. Cornell, J. M. Sobel, and K. Roy, 2009. Is there a latitudinal gradient in the importance of biotic interactions? Annual Review of Ecology, Evolution, and Systematics 40:245–269.

Schluter, D., 2000. The Ecology of Adaptive Radiation. Oxford University Press.

Schmidt, P. S., L. Matzkin, M. Ippolito, and W. F. Eanes, 2005. Geographic variation in diapause incidence, life-history traits, and climatic adaptation in *Drosophila melanogaster*. Evolution 59:1721–1732.

Sheth, S. N. and A. L. Angert, 2017. Demographic compensation does not rescue *Mimulus cardinalis* populations at the southern edge of the species range. bioRxiv .

Slatkin, M., 1978. Spatial patterns in the distributions of polygenic characters. Journal of Theoretical Biology 70:213–28.

Stearns, S., 1976. Life history tactics: a review of the ideas. Quarterly Review of Biology 51.

Stinchcombe, J. R., C. Weinig, M. Ungerer, K. M. Olsen, C. Mays, S. S. Halldorsdottir, M. D. Purugganan, and J. Schmitt, 2004. A latitudinal cline in flowering time in *Arabidopsis thaliana* modulated by the flowering time gene FRIGIDA. Proceedings of the National Academy of Sciences of the United States of America 101:4712–4717.

Stock, A. J., B. E. Campitelli, and J. R. Stinchcombe, 2014. Quantitative genetic variance and multivariate clines in the ivyleaf morning glory, *Ipomoea hederacea*. Philosophical Transactions of the Royal Society B: Biological Sciences 369:20130259.

Templeton, A. R. and D. A. Levin, 1979. Evolutionary consequences of seed pools. The American Naturalist 114:232–249.

Therneau, T. M., 2015. survival: A Package for Survival Analysis in S. URL http://CRAN.R-project.org/package=survival. R package version 2.38.

Turesson, G., 1922. The genotypic response of the plant species to habitat. Hereditas 3:211–350.

Umina, P., A. Weeks, M. Kearney, S. McKechnie, and A. Hoffmann, 2005. A rapid shift

⁷⁰⁸ in a classic clinal pattern in drosophila reflecting climate change. Science 308:691–693.

⁷⁰⁹ United States Geological Survey, 2015. National Hydrogeoraphy Dataset. United States
⁷¹⁰ Geological Survey, Washington, D.C. URL `http://nhd.usgs.gov/index.html`.

⁷¹¹ Wang, T., A. Hamann, D. L. Spittlehouse, and T. Q. Murdock, 2012. ClimateWNA –
⁷¹² high-resolution spatial climate data for western North America. Journal of Applied
⁷¹³ Meteorology and Climatology 51:16–29.

⁷¹⁴ Wright, K. M., D. Lloyd, D. B. Lowry, M. R. Macnair, and J. H. Willis, 2013. Indirect
⁷¹⁵ evolution of hybrid lethality due to linkage with selected locus in *Mimulus guttatus*. PLoS
⁷¹⁶ Biol 11:e1001497.

⁷¹⁷ Wu, C., D. Lowry, A. Cooley, K. Wright, Y. Lee, and J. Willis, 2008. *Mimulus* is an
⁷¹⁸ emerging model system for the integration of ecological and genomic studies. Heredity
⁷¹⁹ 100:220–230.

⁷²⁰ Wu, C. A., D. B. Lowry, L. I. Nutter, and J. H. Willis, 2010. Natural variation for drought-
⁷²¹ response traits in the *Mimulus guttatus* species complex. Oecologia 162:23–33.

⁷²² Yeaman, S. and A. Jarvis, 2006. Regional heterogeneity and gene flow maintain variance
⁷²³ in a quantitative trait within populations of lodgepole pine. Proceedings of hte Royal
⁷²⁴ Society of London B 273:1587–1593.

Climate–Latitude variables

local   neighborhood

Climate–Trait variables

local   neighborhood

$\text{Climate}_i = x_{i,0}$

$\text{Climate}_i = \sum_{j=1}^{n} \frac{x_{i,j}}{n}$

$\text{Climate}_i = x_{i,0}$

$\text{Climate}_i = \sum_{j=1}^{n} \frac{x_{i,j}}{n}$

62–km buffer

$x_{i,0}$ $x_{i,4}$ $x_{i,1}$ $x_{i,2}$ $x_{i,3}$ $x_{i,5}$

62–km buffer

$x_{i,0}$ $x_{i,4}$ $x_{i,1}$ $x_{i,2}$ $x_{i,3}$ $x_{i,5}$

Latitude of origin: 44° 42° 40° 38° 36° 34° 32°

Climate variable (local or neighborhood)

Trait

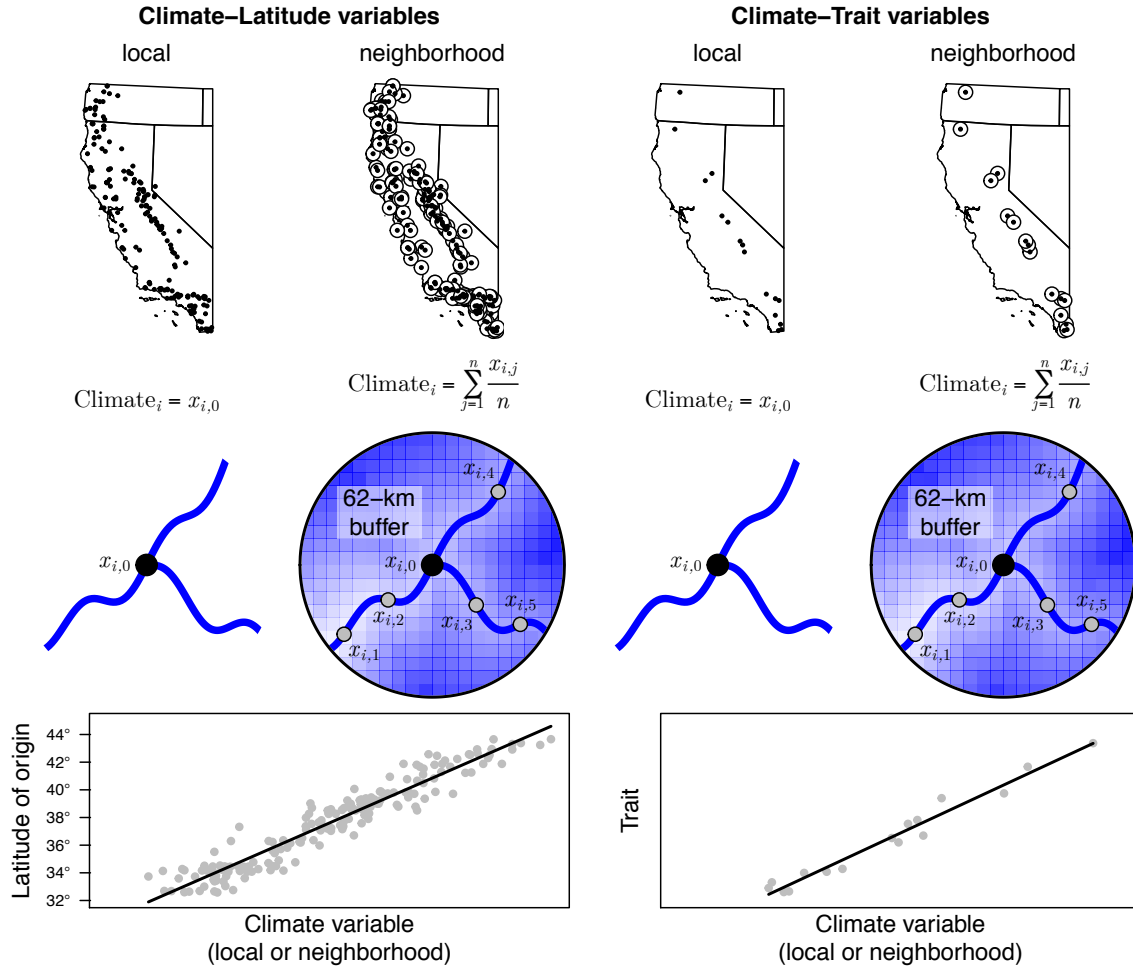Climate variable (local or neighborhood)

Figure 2: Overview of method for identifying putative climatic selective agents underlying latitudinal cline. We looked for climate variables that explained both the latitude of 356 *E. cardinalis* occurrences ('Climate-Latitude variables') and traits ('Climate-Trait variables'). For Climate-Latitude variables we extracted climate data from recent occurrences located throughout California and Oregon, USA (shown in map). For Climate-Trait variables, we extracted climatic data for the 16 focal populations. For both analyses, we extracted local and neighborhood climate. Local climate refers to climate only from where a population was collected ($x_{i,0}$). Neighborhood climate was calculated as the average over 1000 points in a 62-km radius climatic neighborhood ($x_{i,1}, x_{i,2}, \dots$), but only along stream habitats as *E. cardinalis* is riparian. We identified climatic factors that most strongly predicted latitude of occurrences (Climate-Latitude variables) and traits (Climate-Trait variables), as shown for hypothetical data in plots at the bottom of the figure.
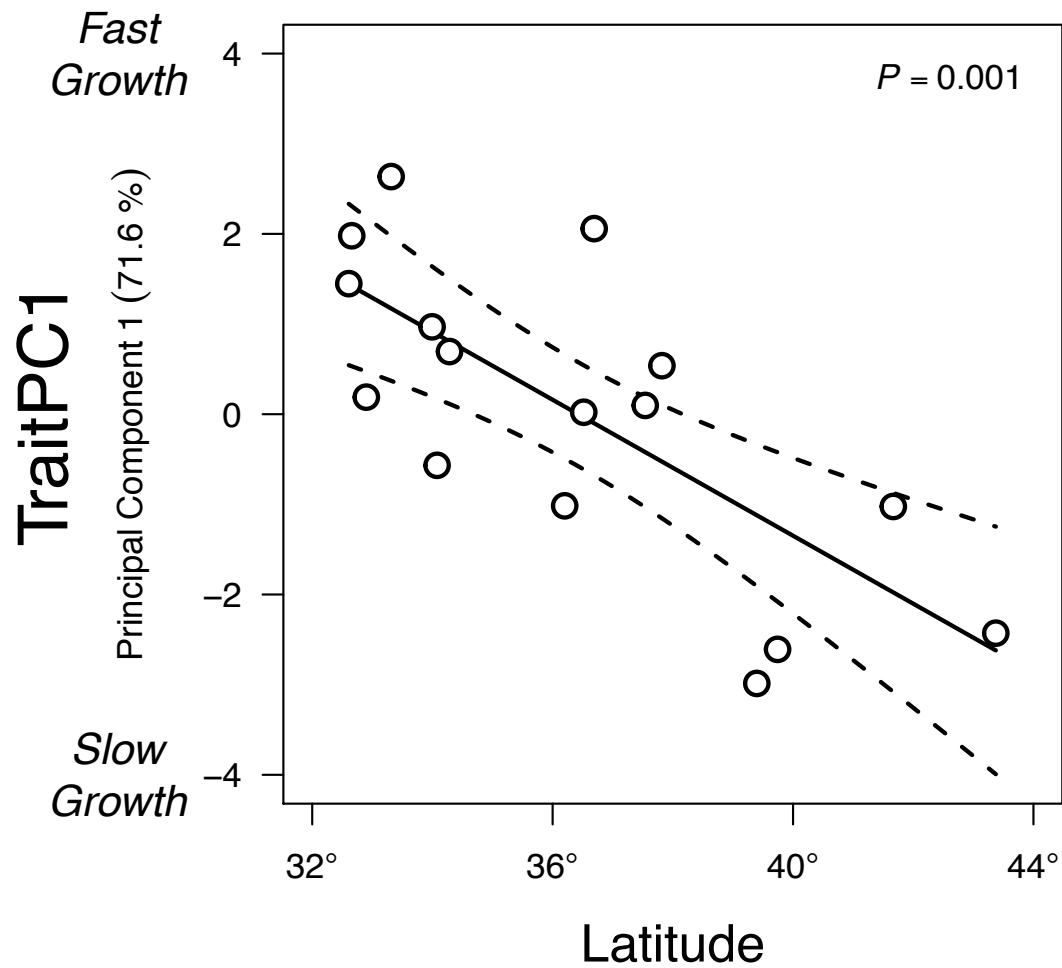
Figure 3: Trait variation, from fast to slow growth, is closely associated with latitude. Each point is a population's latitude of origin (x-axis) and position along the slow to fast growth axis (y-axis), defined as Principal Component 1 of four traits (see Material and Methods). The line and 95% confidence intervals were estimated using linear regression.
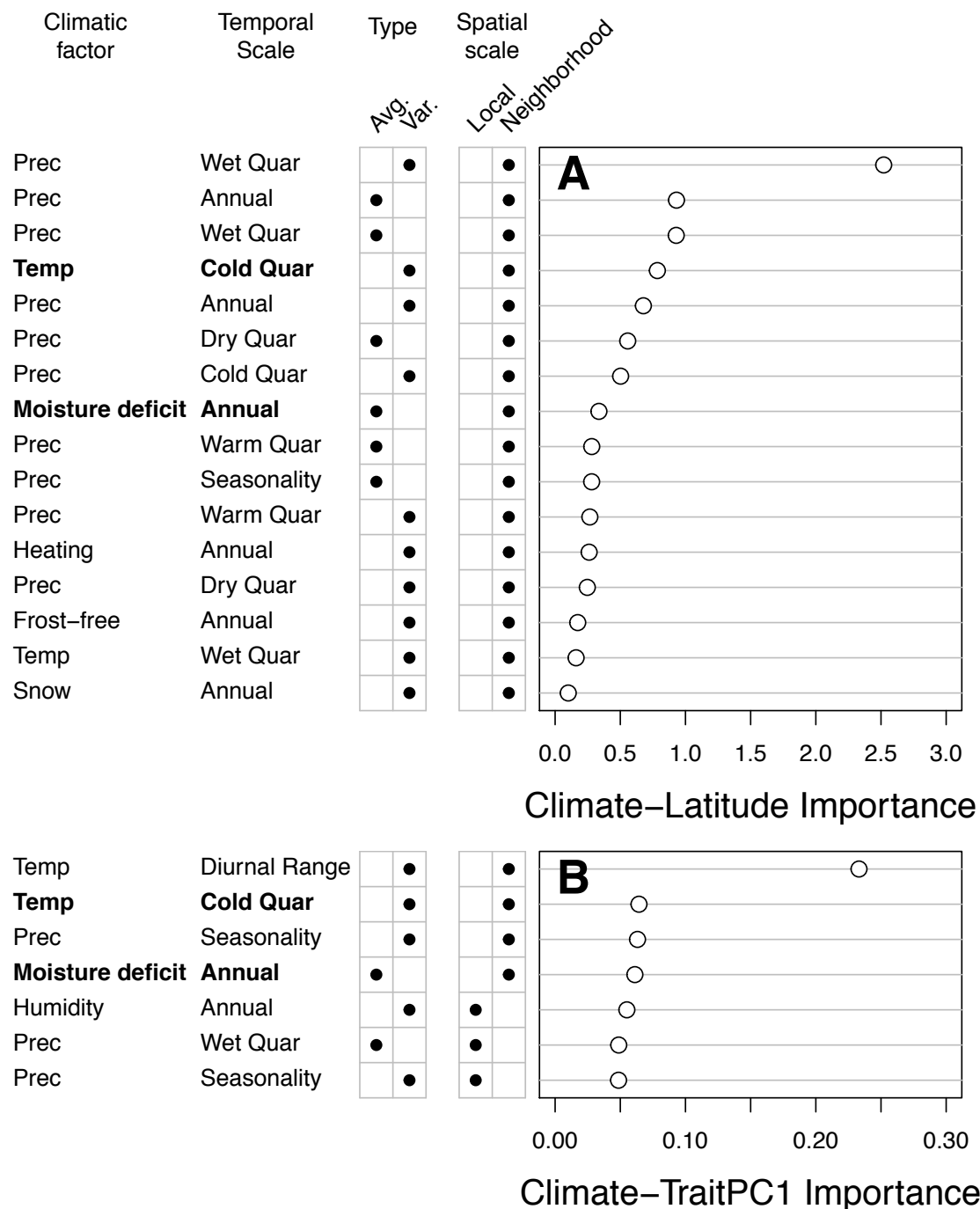
Panel A — Climate–Latitude Importance

Climatic factor	Temporal Scale	Avg.	Var.	Local	Neighborhood
Prec	Wet Quar		•		•
Prec	Annual	•			•
Prec	Wet Quar	•			•
Temp	**Cold Quar**		•		•
Prec	Annual		•		•
Prec	Dry Quar	•			•
Prec	Cold Quar		•		•
Moisture deficit	**Annual**	•			•
Prec	Warm Quar	•			•
Prec	Seasonality	•			•
Prec	Warm Quar		•		•
Heating	Annual		•		•
Prec	Dry Quar		•		•
Frost–free	Annual		•		•
Temp	Wet Quar		•		•
Snow	Annual		•		•

Climate–Latitude Importance (axis: 0.0, 0.5, 1.0, 1.5, 2.0, 2.5, 3.0)

Panel B — Climate–TraitPC1 Importance

Climatic factor	Temporal Scale	Avg.	Var.	Local	Neighborhood
Temp	Diurnal Range		•		•
Temp	**Cold Quar**		•		•
Prec	Seasonality		•		•
Moisture deficit	**Annual**	•			•
Humidity	Annual		•	•	
Prec	Wet Quar	•		•	
Prec	Seasonality		•	•	

Climate–TraitPC1 Importance (axis: 0.00, 0.10, 0.20, 0.30)

Figure 4: Climatic variation integrated over climatic neighborhood is closely correlated with latitude of *E. cardinalis* and trait variation. A. Using Random Forest regression, we identified 16 climatic variables significantly (high importance) associated with latitude of *E. cardinalis* occurrences. B. Only one of of the most important Climate-Latitude variables (in bold) was among the most important Climate-TraitPC1 variables. Variable importance is defined as the average amount a climate variable reduces mean-squared error in the predicted response (TraitPC1 or Latitude), compared to a randomly permuted dataset, across all trees in the random forest (see Genuer et al. [2015] for further detail). Note that the Importance values in A and B are not comparable because the dependent variables (Latitude and Trait PC1, respectively) are on different scales. Climatic variables (left of A; right of B) are defined by four qualities: Climatic factor – Temperature (Temp), Precipitation (Prec), Heating degree-days (Heating), Snow (precipitation as snow); Temporal scale – Annual, Coldest quarter (Cold
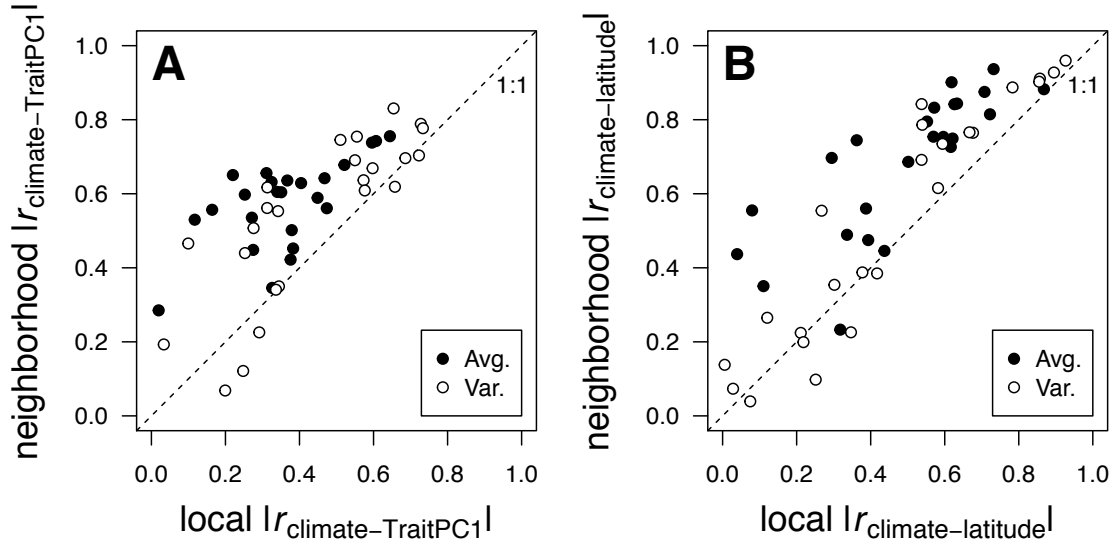
Figure 5: Neighborhood climate predicts TraitPC1 ('Climate-trait', panel A) and Latitude of occurences ('Climate-latitude', panel B) better than local climate. Each point is the absolute value of the Pearson correlation coefficient ($|r|$) between TraitPC1 (A) or latitude (B) for 24 climatic factors, for which we used both the 30-year mean (closed circles) and coefficient of variation (open circles). Most points lie above the 1:1 line, indicating stronger correlations with neighborhood compared to local climate. Neighborhood climate was integrated over a 62-km radius around focal populations (see text for further detail).
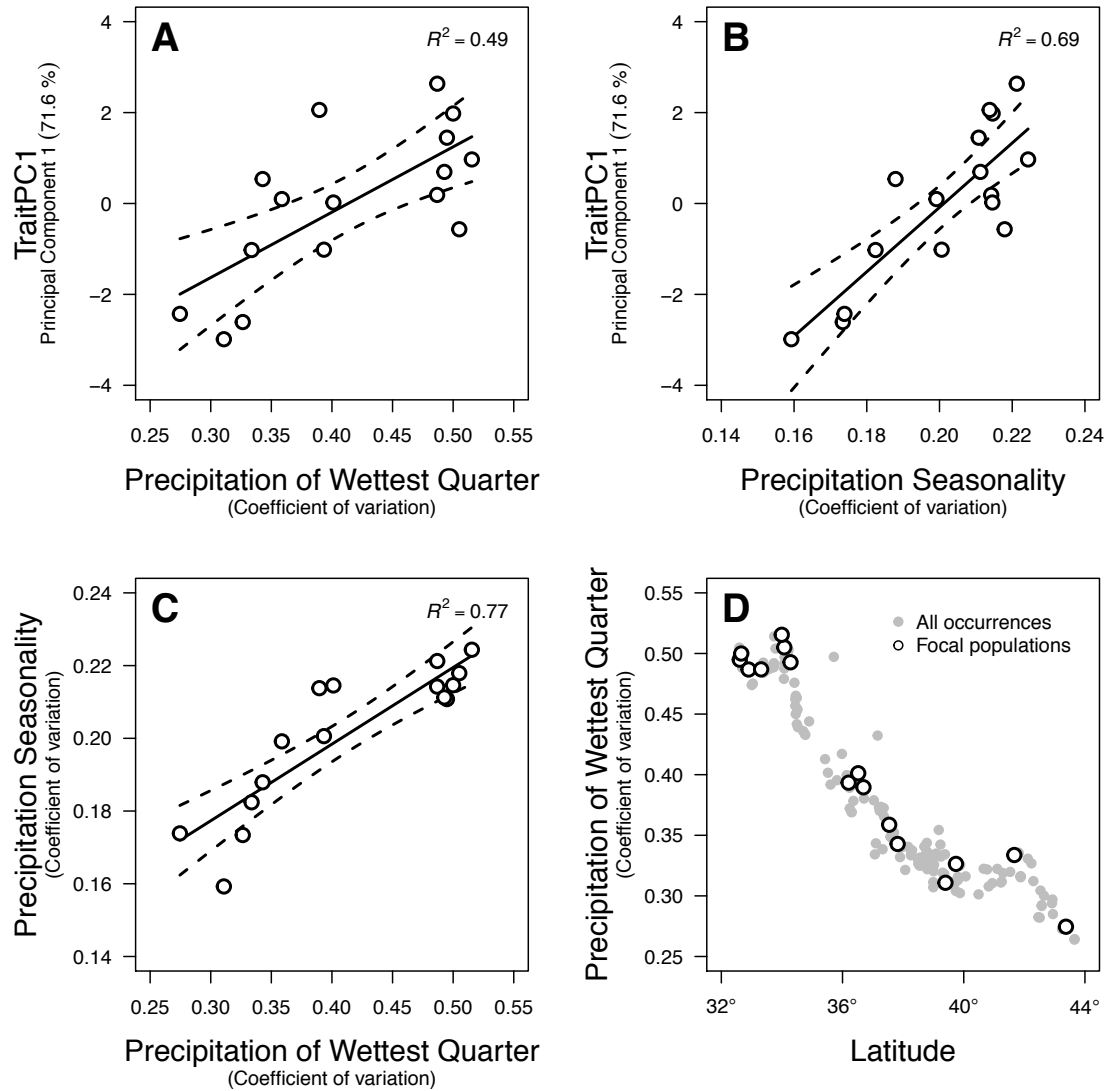
Figure 6: Variation in precipitation is correlated with TraitPC1 and latitude. A. Greater values of TraitPC1 are associated with greater interannual variation in precipitation of the wettest quarter. This was the most important Climate-Latitude variable, but not among the most important Climate-TraitPC1 variables. B. However, a closely related parameter, interannaul variation in precipitation seasonality, was among the most important Climate-TraitPC1 variables. C. Across focal populations, variation in precipitation of the wettest quarter and seasonality are closely correlated. D. Southern populations of *E. cardinalis* experience much greater interannual variationi in precipitation. In all panels, we report climatic neighborhood values (see Material and Methods). Regression lines, 95% confidence intervals, and coefficients of determination (R^2) were calculated using linear regression.

Supporting Information

Supporting Tables

Table S1: Initial size of seedlings did not vary among Populations, Families, or Treatments. We used a censored Gaussian model of initial size at the outset of the experiment (longest leaf length of the first true leaves). The model was censored because we could not accurately measure leaves less than 0.25 mm with digital callipers (217 of 702, 30.9%, were too small). We fit models using a Bayesian MCMC method implemented using the MCMCglmm function with default priors in the R package **MCMCglmm** version 2.17 (Hadfield, 2010). We estimated the posterior distribution from 1000 samples of an MCMC chain run for 10^5 steps after a 10^4 step burn-in. We used step-wise backward elimination procedure to find the best-supported model according to Deviance Information Criterion (DIC).

Model	Random	DIC
Population + Water + Temperature + Population:Water + Population:Temperature + Water:Temperature + Population:Water:Temperature	Family	1638
Population + Water + Temperature + Population:Water + Population:Temperature + Water:Temperature	Family	1605.2
Population + Water + Temperature + Population:Water + Population:Temperature	Family	1603.4
Population + Water + Temperature + Population:Water + Water:Temperature	Family	1577.5
Population + Water + Temperature + Population:Temperature + Water:Temperature	Family	1579.9
Population + Water + Temperature + Population:Water	Family	1577.3
Population + Water + Temperature + Water:Temperature	Family	1550.5
Population + Water + Temperature	Family	1549.3
Population + Water	Family	1541.7
Population + Temperature	Family	1546.8
Water + Temperature	Family	1551.1
Population	Family	1541.9
Water	Family	1543.9
-	Family	1541.7
-	-	1538.3

Table S2: Climatic variables used

Abbreviation	Climate variable
DD_0	degree-days below 0℃(chilling degree-days)
DD5	degree-days above 5℃(growing degree-days)
DD_18	degree-days below 18℃(heating degree-days)
DD18	degree-days above 18℃(cooling degree-days)
NFFD	number of frost-free days
PAS	precipitation as snow (mm) between August in previous year and July in current
Eref	Hargreaves reference evaporation (mm)
CMD	Hargreaves climatic moisture deficit (mm)
RH	mean annual relative humidity
bio1	annual mean temperature
bio2	mean diurnal range (mean of monthly (max temp - min temp))
bio3	isothermality (bio2/bio7) (* 100)
bio4	temperature seasonality (standard deviation *100)
bio5	max temperature of warmest month
bio6	min temperature of coldest month
bio7	temperature annual range (bio5-bio6)
bio8	mean temperature of wettest quarter
bio9	mean temperature of driest quarter
bio10	mean temperature of warmest quarter
bio11	mean temperature of coldest quarter
bio12	annual precipitation
bio15	precipitation seasonality (coefficient of variation)
bio16	precipitation of wettest quarter
bio17	precipitation of driest quarter
bio18	precipitation of warmest quarter
bio19	precipitation of coldest quarter

Table S3: Analysis of varianace (ANOVA) table on leaf expansion rate (LER) using **lmerTest** (Kuznetsova et al., 2016). Family and Block were included as random effects. Abbreviations: SS = sum of squares; MS = mean sum of squares (SS / df1); df1 = numerator degrees of freedom; df2 = denominator degrees of freedom.

	SS	MS	df1	df2	*F-value*	*P*-value
Day to Germination	12.12	12.12	1	637	35.21	4.9×10^{-9}
Population	22.22	1.48	15	118	4.3	2.5×10^{-6}
Temperature	80.42	80.42	1	5	233.61	2.6×10^{-5}
Water	4.1	4.1	1	5	11.92	0.019
Temperature × Water	0.03	0.03	1	4	0.07	0.801
Population × Temperature	2.76	0.18	15	547	0.53	0.925
Population × Water	9.66	0.64	15	562	1.87	0.024
Population × Temperature × Water	4.11	0.27	15	530	0.78	0.700

Table S4: Analysis of varianace (ANOVA) table on stem elongation rate (SER) using **lmerTest** (Kuznetsova et al., 2016). Family and Block were included as random effects. Abbreviations: SS = sum of squares; MS = mean sum of squares (SS / df1); df1 = numerator degrees of freedom; df2 = denominator degrees of freedom.

	SS	MS	df1	df2	*F-value*	*P*-value
Day to Germination	3.6	3.6	1	662	21.1	5.1×10^{-6}
Population	12	0.8	15	113	4.7	5.8×10^{-7}
Temperature	12.4	12.4	1	6	72.8	1.5×10^{-4}
Water	0.6	0.6	1	5	3.7	0.113
Temperature × Water	0.9	0.9	1	4	5.2	0.093
Population × Temperature	3.6	0.2	15	549	1.4	0.126
Population × Water	2.8	0.2	15	536	1.1	0.330
Population × Temperature × Water	1.5	0.1	15	518	0.6	0.874

Table S5: Analysis of varianace (ANOVA) table on photosynthetic rate using **lmerTest** (Kuznetsova et al., 2016). Family and Block were included as random effects. Abbreviations: SS = sum of squares; MS = mean sum of squares (SS / df1); df1 = numerator degrees of freedom; df2 = denominator degrees of freedom.
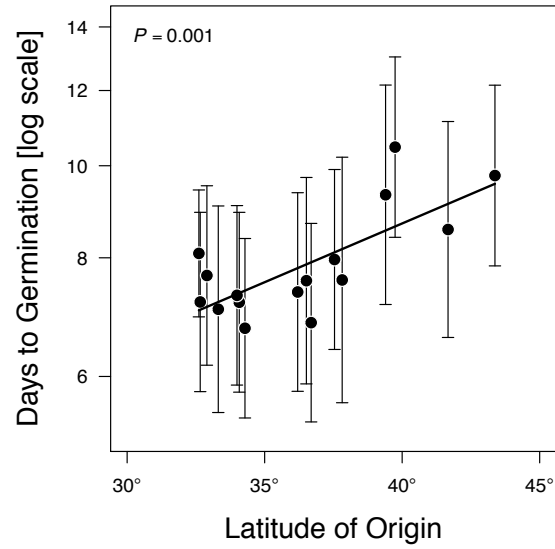
	SS	MS	df1	df2	*F-value*	*P*-value
Population	347.7	23.2	15	78	3.02	7.5×10^{-4}
Temperature	134.1	134.1	1	6	17.46	6.4×10^{-3}
Water	51	51	1	4	6.64	0.066
Temperature × Water	0.7	0.7	1	3	0.09	0.781
Population × Temperature	218.6	14.6	15	263	1.9	0.024
Population × Water	87.7	5.8	15	233	0.76	0.724
Population × Temperature × Water	91.4	6.1	15	208	0.79	0.686

Table S6: Analysis of deviance table on the probability of mortality by the end of the experiment using Type-II Wald χ^2 tests in the R package **car** (Fox and Weisberg, 2011). Family and Block were included as random effects. Abbreviations: df = degrees of freedom

	χ^2	df	*P*-value
Population	32	31	0.419
Temperature	31.8	6	1.8×10^{-5}
Water	69.2	12	4.6×10^{-10}
Temperature × Water	20.7	1	5.3×10^{-6}
Population × Temperature	5.6	15	0.985
Population × Water	8.6	15	0.897
Population × Temperature × Water	0.2	15	1.000

Table S7: Important climatic variables predicting latitude of *E. cardinalis* populations ('Climate-Latitude') and the first principal component of traits measured in a common garden ('Climate-TraitPC1'). Local climatic variables were measured from the exact location of collection; neighborhood climatic variables were averaged from a 62-km neighborhood around population (see Material and Methods). Importance and significance were determined using the variable selection using random forests (VSURF) algorithm (see Material and Methods). Climatic variables are described in Table S2. μ signifies the mean of the climate variables from 1981–2010; σ indicates coeffiecient of variation among years.

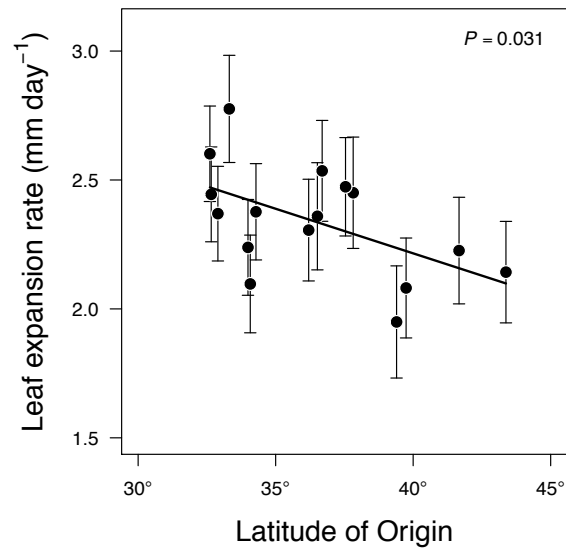| Climate-Latitude variables	Climate-TraitPC1 variables
Precipitation of wettest quarter (σ, neighborhood)	Mean diurnal range (σ, neighborhood)
Annual precipitation (μ, neighborhood)	Mean temperature of coldest quarter (σ, neighborhood)
Precipitation of wettest quarter (μ, neighborhood)	Precipitation seasonality (σ, neighborhood)
Mean temperature of coldest quarter (σ, neighborhood)	Hargreaves climatic moisture deficit (μ, neighborhood)
Annual precipitation (σ, neighborhood)	Annual mean relative humidity (σ, local)
Precipitation of driest quarter (μ, neighborhood)	Precipitation of wettest quarter (μ, local)
Precipitation of coldest quarter (σ, neighborhood)	Precipitation seasonality (σ, local)
Hargreaves climatic moisture deficit (μ, neighborhood)	
Precipitation of warmest quarter (μ, neighborhood)	
Precipitation seasonality (μ, neighborhood)	
Precipitation of warmest quarter (σ, neighborhood)	
Heating degree-days (σ, neighborhood)	
Precipitation of driest quarter (σ, neighborhood)	
Number of frost-free days (σ, neighborhood)	
Mean temperature of wettest quarter (σ, neighborhood)	
Precipitation as snow (σ, neighborhood)	

Supporting Figures

Figure S1: Southern populations germinate faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted days to germination in days under growth chamber conditions (see Material and Methods). Bars around each point are 95% confidence intervals. Predicted time to germination and confidence intervals are based on survival regression (see Materials and Materials). The line is the linear regression of log(model-predicated days to germination) $\sim$ latitude. The P-value of the regression is given in the upper left corner.
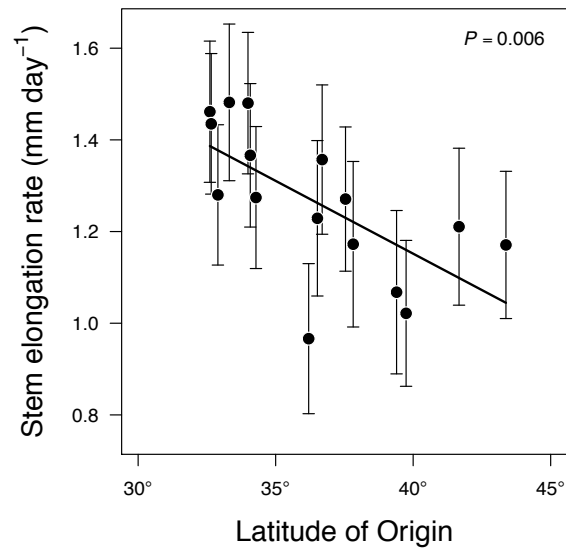
Figure S2: Southern populations grow faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted leaf expansion rate during the rosette phase. Bars around each point are 95% confidence intervals. Predicted leaf expansion rate based least-square mean estimates and confidence intervals were calculated from linear mixed-effects models (see Materials and Materials). The line is the linear regression of model-predicated leaf expansion rate $\sim$ latitude. The P-value of the regression is given in the upper right corner.

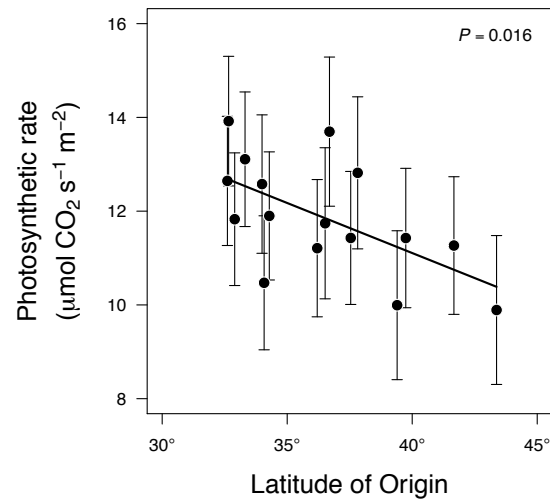**Figure S3:** Southern populations grow faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted stem elongation rate. Bars around each point are 95% confidence intervals. Predicted stem elongation rate based least-square mean estimates and confidence intervals were calculated from linear mixed-effects models (see Materials and Materials). The line is the linear regression of model-predicated stem elongation rate $\sim$ latitude. The P-value of the regression is given in the upper right corner.

Figure S4: Southern populations photosynthesize faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted instantaneous photosynthetic rate. Bars around each point are 95% confidence intervals. Predicted photosynthetic rates based least-square mean estimates and confidence intervals were calculated from linear mixed-effects models (see Materials and Materials). The line is the linear regression of model-predicated photosynthetic rate $\sim$ latitude. The P-value of the regression is given in the upper right corner.
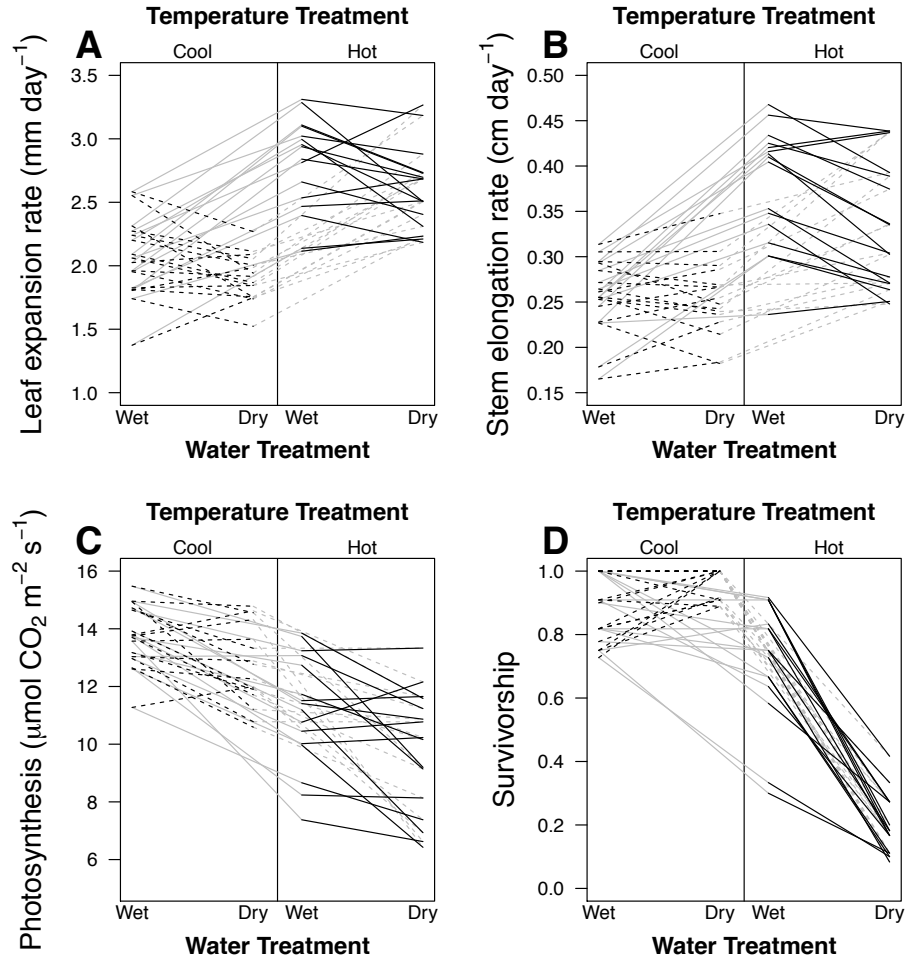
Figure S5: Reaction norms signify little Population × Treatment interactions. For all panels, black lines represent population-level reaction norms from Wet to Dry in the Cool temperature treatment (dashed black lines) and Hot temperature treatment (solid black lines); gray lines represent reaction norms from Cool to Hot in the Wet treatment (solid gray lines) and Dry treatment (dashed gray lines). The responses shown are (A) leaf expansion rate, (B) stem elongation rate, (C) photosynthesis, and (D) survivorship (= 1 - mortality).
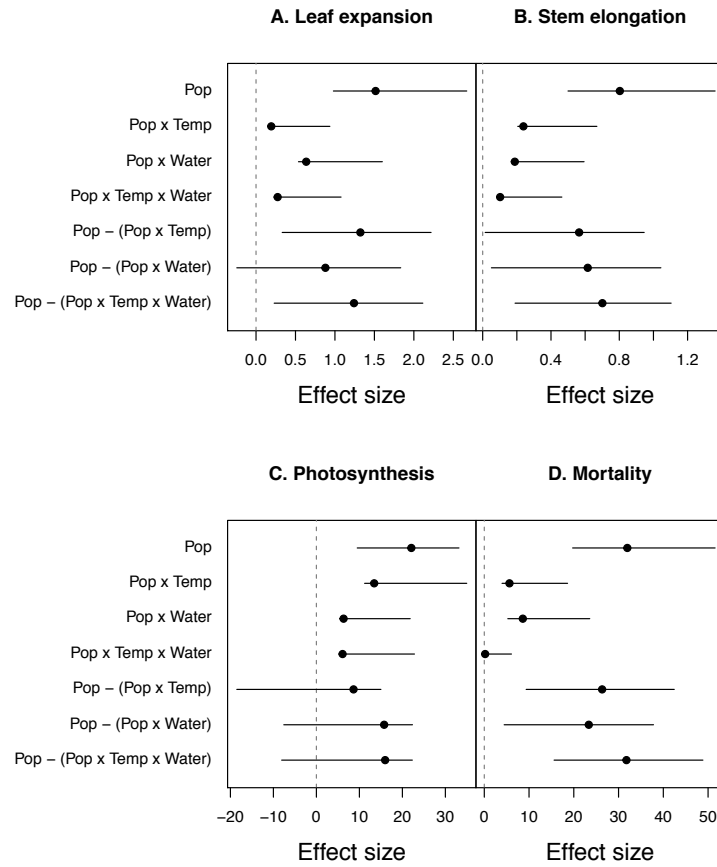
Figure S6: Population effect sizes are usually larger than Population × Treatment effect sizes. In each panel, we plot estimated effect size (points) and 95% confidence intervals (lines) inferred using parametric bootstrap (see Materials and Methods). At the top, we plot the effect sizes of Population ('Pop'), two-way interactions between Population and Temperature ('Pop × Temp') or Water ('Pop × Water'), and the three-way interaction between Population, Temperature, and Water ('Pop × Temp × Water'). Below that we plot the difference in the Population minus the Population × Treatment effect size (e.g. 'Pop - (Pop × Temp)'). When confidence intervals do no overlap zero (dashed line), this means that Population has a significantly greater effect size than the interaction. Effect sizes were measured using unstandarized mean square error for linear mixed-effects models (leaf expansion, stem elongation, and photosynthesis) and χ^2 for GLMM (mortality). Hence, the effect size values are not comparable between different traits.
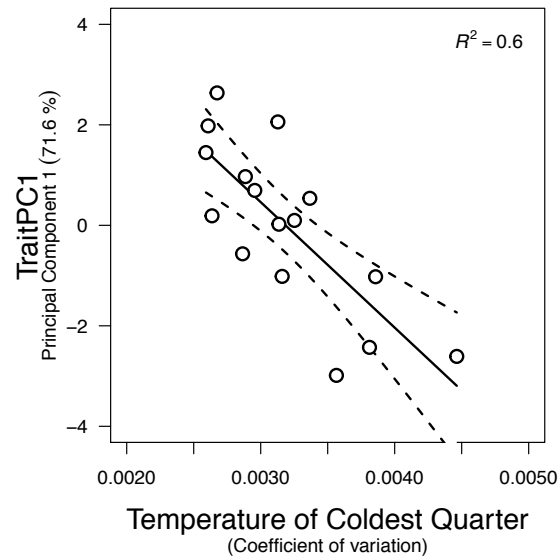
Figure S7: Trait variation, from fast to slow growth, is closely associated with neighborhood variation in temperature of the coldest quarter ($bio11_\sigma$) Each point is a population coefficient of variation in bio11 averaged over a 62-km climatic neighborhood (x-axis) and position along the slow to fast growth axis (y-axis), defined as Principal Component 1 of four traits (see Material and Methods). The line and 95% confidence intervals were estimated using linear regression.

Supporting Material and Methods

Temperature treatments

We simulated typical growing season (June 1 - August 15) air temperatures at the two most thermally divergent focal sites in our study, Whitewater Canyon (WWC, Hot) and Little Jameson (LIJ, Cool). We downloaded daily interpolated mean, minimum, and maximum air temperature from 13 years (2000-2012) at both sites from ClimateWNA (Wang et al., 2012). This range was chosen because seeds used in the experiment were collected around 2012, thus their presence in that location at that time suggests that populations were able to persist there for at least some years before collection. Monthly temperatures from ClimateWNA are highly correlated with the air temperature recorded from data loggers in the field at these sites (A. Angert, unpub. data). Hence, the ClimateWNA temperature profiles are similar to actual thermal regimes experienced by *E. cardinalis* in nature. We simulated realistic temperature regimes by calculating the mean temperature trend from June to August using LOESS (Cleveland et al., 1992). The residuals were highly autocorrelated at both sites (warmer than average days are typically followed by more warm days) and there was strong correlation ($r = 0.65$) between sites (warm days in WWC were also warm in LIJ). The 'VARselect' function in the **vars** package for R (Pfaff, 2008) indicated that a lag two Vector Autoregression (VAR(2)) model best captured the within-site autocorrelation as well as between-site correlation in residuals. We fit and simulated from the VAR(2) model using the package **dse** (Gilbert, 2014) in R. Simulated data closely resembled the autocorrelation and between-site correlation of the actual data. From simulated mean temperature, we next selected minimum and maximum daily temperatures. Mean, min, and max temperature were highly correlated at both sites. We chose min and max temperatures using site-specific fitted linear models between mean, max, and min temperature, with additional variation given by normally distributed random deviates with variance equal to the residual variance of the linear models. For each day, the nighttime

52

(22:00 - 6:00) chamber temperature was set to the simulated minimum temperature. During the middle of the day, temperature was set to the simulated maximum temperature, with a variable period of transition between min and max so that the average temperature was equal the simulated mean temperature.

Watering treatments

For watering treatments, we simulated two extreme types of streams where *E. cardinalis* grows. In the well-watered treatment, we simulated a large stream that never goes dry during the summer growing season. In the drought treatment, we simulated a small stream that has ample flow at the beginning of the season due to rain and snow melt, but gradually dries down through the summer. In both treatments, plants were bottom-watered using water chilled to 7.5℃. Plants in the well-watered treatment were fully saturated every two hours during the day. Watering in the drought treatment gradually declined from every two hours to every day between May 20 (36 days after sowing) and 10 June (57 days after sowing). Simultaneously, the amount of bottom-watering per flood decreased, such that only the bottom of the cone-tainers were wetted by the end of the experiment.