

Grow with the flow: a latitudinal cline in physiology is associated with more variable precipitation in *Erythranthe cardinalis*

Christopher D. Muir^{1,*} and Amy L. Angert¹

¹ Biodiversity Research Centre, University of British Columbia, Vancouver, BC, Canada

*corresponding author: Chris Muir, cdmuir@biodiversity.ubc.ca

Running Head: Latitudinal cline and climate in *Erythranthe*

Key words: local adaptation, cline, photosynthesis, growth rate, *Erythranthe*

Data will be archived on Dryad upon publication.

Acknowledgements

Erin Warkman and Lisa Lin helped collect data. CDM was supported by a Biodiversity Postdoctoral Fellowship funded by the NSERC CREATE program. ALA was supported by an NSERC Discovery Grant and a grant from the National Science Foundation (DEB 0950171).

Grow with the flow: a latitudinal cline in physiology is associated with more variable precipitation in *Erythranthe cardinalis*

Abstract

Local adaptation is one of the most ubiquitous observations in nature: organisms perform well in their natal environment, but poorly outside it. Correlation between traits and latitude, or latitudinal clines, are among the most common pieces of evidence for local adaptation, but identifying the traits under selection and the selective agents are challenging. Here, we investigated a latitudinal cline in growth and photosynthesis across 16 populations of the perennial herb *Erythranthe cardinalis* (Phrymaceae). Using machine learning methods, we identify interannual variation in precipitation as a likely selective agent: Southern populations from more variable environments had higher photosynthetic rates and grew faster. We hypothesize that selection may favor a more annualized life history – grow now rather than save for next year – in environments where severe droughts occur more often. Thus our study provides insight into how species may adapt if Mediterranean climates become more variable due to climate change.

Introduction

1 Local adaptation within species is ubiquitous; populations generally have higher fitness in
2 their native environment, but perform poorly outside it (Schluter, 2000; Hereford, 2009).
3 Local adaptation also frequently leads to clines in both phenotypes and allele frequencies
4 when selection varies over environmental gradients (Huxley, 1938; Endler, 1977; Barton,
5 1999). Phenotypic differences between populations along a cline most often have a ge-
6 netic basis and can be studied in a common garden (Turesson, 1922; Clausen et al., 1940;

7 Hiesey et al., 1942). Despite a long history of studying local adaptation and clines, it
8 remains challenging to identify exactly which traits are under selection and which differ
9 for nonadaptive reasons. In particular, the role that physiological differences play in local
10 adaptation is poorly understood, despite the fact that physiology is frequently assumed to
11 explain adaptation to the abiotic environment. A related problem is identifying which of
12 the myriad and often covarying aspects of the environment causes spatially varying selective
13 pressures.

14 When populations are locally adapted, reaction norms for fitness will cross, such that local
15 genotypes have higher fitness than foreign genotypes and rank orders change across envi-
16 ronments (Kawecki and Ebert, 2004). The traits that underlie local adaptation, however,
17 need not mirror this pattern. Populations can have fixed genetic differences conferring trait
18 values that are adaptive at home but neutral or maladaptive away. Alternatively, genotype-
19 by-environment interactions could indicate that variation in plasticity mediates local adap-
20 tation. We distinguish between these patterns of adaptive trait differences by referring to
21 ‘intrinsic’ and ‘plastic’ trait variation, respectively. Adaptation involves both intrinsic and
22 plastic trait variation. For example, intrinsic differences in photoperiod (Blackman et al.,
23 2011) and developmental rate (Stinchcombe et al., 2004) allow organisms to properly time
24 their life history with the local environment. Conversely, sun and shade plants do not
25 have intrinsically higher or lower rates of carbon assimilation, but rather, genotype-by-
26 environment interactions cause sun plants to assimilate more under high light and shade
27 plants under low light (Givnish, 1988). In plants especially, we know little about the
28 prevalence and adaptive significance of variation in fundamental physiological traits like
29 photosynthesis and their impact on plant performance.

30 A basic approach to identify candidate traits underlying local adaptation is to find asso-
31 ciations between traits and environments. Either intrinsic and/or plastic variation should
32 vary clinally along environmental gradients. Indeed, clines in ecologically important traits
33 are widespread in nature (Endler, 1977) and often adaptive, but in most cases the selective

34 agent is unknown. For example, in *Drosophila* numerous latitudinal clines exist for traits
35 like thermal tolerance (Hoffmann et al., 2002), body size (Coyne and Beecham (1987) and
36 references therein), and life history (Schmidt et al., 2005). Some *Drosophila* clines have
37 evolved multiple times (Oakeshott et al. (1982); Huey et al. (2000), see also Bradshaw and
38 Holzapfel (2001)) or shifted in response to climate change (Umina et al., 2005), evincing
39 climatic adaptation. Similarly, plant species exhibit latitudinal clines in traits like flowering
40 time (Stinchcombe et al., 2004), cyanogenesis (Kooyers and Olsen, 2012), leaf morphology
41 (Hopkins et al., 2008; Stock et al., 2014), and drought response (Kooyers et al., 2015) that
42 likely relate to climatic variation.

43 Despite the fact that latitudinal clines have been studied for a long time, latitude *per se*
44 cannot be a selective agent. Latitude may be strongly correlated with one or two key
45 climatic variables, such as temperature, precipitation, or growing degree-days. Hence, lat-
46 itude is an effective proxy for the underlying climatic driver, but we would expect a yet
47 stronger relationship between traits and the key climatic variable(s) driving selection. Al-
48 ternatively, latitude may be more strongly related to traits than any single climatic variable
49 for at least two reasons. First, latitude may be correlated with several climatic agents of
50 selection that are individually weak, but add up to a strong latitudinal cline. Alterna-
51 tively, gene flow among neighbouring populations could smooth out local climatic effects,
52 since alleles will experience selection across populations linked by migration (Slatkin, 1978;
53 Paul et al., 2011; Hadfield, 2016). We refer to this as the ‘climatic neighborhood’. For
54 example, in mountainous regions average temperature at a given latitude varies widely,
55 but in aggregate, a lower latitude set of populations will experience warmer climate than a
56 higher latitude one. Thus, any particular low latitude population would be warm-adapted,
57 even if it was located in a cooler (e.g. high elevation) site. Because many climatic factors
58 vary latitudinally, and which climatic factors vary latitudinally changes over the earth’s
59 surface (e.g. coastal vs. continental), dissecting the evolution of latitudinal clines across
60 many species will help identify generalities, such as whether thermal tolerance maxima or

61 seasonal timing is more important (Bradshaw and Holzapfel, 2008), or whether local versus
62 regional climate shape selective pressures.

63 In this study, we investigated two major questions: 1) whether intrinsic or plastic physiolog-
64 ical trait variation corresponds with latitude; and 2) what climatic factor(s) could plausibly
65 be responsible for latitudinal clines. Within question 2, we tested three hypotheses outlined
66 in the previous paragraph: latitudinal clines are explained by a single dominant climatic
67 factor, multiple climatic factors, or the climatic neighborhood experienced by nearby pop-
68 ulation connected through gene flow. These hypotheses are not mutually exclusive since,
69 for example, single or multiple factors in a climatic neighborhood may lead to latitudinal
70 clines.

71 We examined these questions in *Erythranthe cardinalis* (formerly *Mimulus cardinalis* [Ne-
72 som 2014]) because linking physiological traits to potentially complex patterns of local
73 adaptation requires integrating multiple lines of evidence from comparative, experimental,
74 and genomic studies under both lab and field conditions. Many classic and contemporary
75 studies of local adaptation use *Mimulus sensu lato* species because of its natural history,
76 easy propagation, and genetic/genomic resources (Clausen et al., 1940; Hiesey et al., 1971;
77 Bradshaw and Schemske, 2003; Wu et al., 2008; Lowry and Willis, 2010; Wright et al.,
78 2013). Yet, there is a conspicuous deficiency of links between local adaptation and physio-
79 logical mechanisms (Angert (2006); Angert et al. (2008); Wu et al. (2010), but see Wright
80 et al. (2013) on mechanisms of copper tolerance). We measured genetic and genotype-
81 by-environment variation in response to temperature and drought among 16 populations
82 distributed over 10.7° of latitude. We found a latitudinal cline of intrinsic variation in pho-
83 tosynthesis and growth, but little evidence for variation in plasticity. Interannual variation
84 in precipitation and temperature are associated with this axis of variation, suggesting that
85 climatic variance rather than mean may be an important driver of local adaptation in
86 *E. cardinalis*. The climatic neighborhoods around populations explained trait variation
87 better than local climate, indicating that latitudinal clines may be common because lati-

88 tude integrates effects of selection on populations connected through gene flow. We place
89 these findings in the context of life history theory and consider future directions in the
90 Discussion.

91 Material and Methods

92 Population Selection

93 We used 16 populations from throughout the range of *E. cardinalis* (Table 1). Seeds were
94 collected in the field from mature, undehisced fruit left open for 2-4 weeks to dry, then
95 stored at room temperature.

Table 1: Geographic region, latitude, longitude, and elevation (mas = meters above seal level) of 16 focal populations used in this study.

Name	Region	Latitude	Longtiude	Elevation (mas)
Hauser Creek	South Margin	32.657	-116.532	799
Cottonwood Creek	South Margin	32.609	-116.7	267
Sweetwater River	South Margin	32.9	-116.585	1180
Grade Road Palomar	South Margin	33.314	-116.871	1577
Whitewater Canyon	Transverse	33.994	-116.665	705
Mill Creek	Transverse	34.077	-116.873	2050
West Fork Mojave River	Transverse	34.284	-117.378	1120
North Fork Middle Tule River	South Sierras	36.201	-118.651	1314
Paradise Creek	South Sierras	36.518	-118.759	926
Redwood Creek	South Sierras	36.691	-118.91	1727
Wawona	Central Sierras	37.541	-119.649	1224
Rainbow Creek	Central Sierras	37.819	-120.007	876
Middle Yuba River	North Sierras	39.397	-121.082	455
Little Jamison Creek	North Sierras	39.743	-120.704	1603
Deep Creek	North Coast	41.668	-123.11	707
Rock Creek	North Margin	43.374	-122.957	326

96 **Plant propagation**

97 On 14 April, 2014, 3-5 seeds per family were sown directly on sand (Quikrete Play Sand,
98 Georgia, USA) watered to field capacity in RLC4 Ray Leach cone-tainers placed in RL98
99 98-well trays (Stuewe & Sons, Inc., Oregon, USA). We used pure sand because *E. cardinalis*
100 typically grows in sandy, riparian soils (A. Angert, pers. obs.). Two jumbo-sized cotton
101 balls at the bottom of cone-tainers prevented sand from washing out. Cone-tainers sat in
102 medium-sized flow trays (FLOWTMD, Stuewe & Sons, Inc., Oregon, USA) to continuously
103 bottom-water plants during germination in greenhouses at the University British Columbia
104 campus in Vancouver, Canada (49°15' N, 123°15' W). Mistlers thoroughly wetted the top of
105 the sand every two hours during the day. Most seeds germinated between 1 and 2 weeks,
106 but we allowed 3 weeks before transferring seedlings to growth chambers. We recorded
107 germination daily between one to two weeks after sowing, and every 2-3 days thereafter.
108 On 5 May (21 days after sowing), we transferred seedlings to one of two growth chambers
109 (Convion, Manitoba, Canada). We thinned seedlings to one plant per cone-tainer, leaving
110 the center-most plant. 702 of 768 (91.4%) had plants that could be used in the experiment.
111 We allowed one week at constant, non stressful conditions (day: 20°C, night: 16°C) for plants
112 to acclimate to growth chambers before starting treatments. The initial size of seedlings,
113 measured as the length of the first true leaves, did not differ between populations, families,
114 or treatments (Table S1).

115 **Temperature and drought treatments**

116 We imposed four treatments, a fully-factorial cross of two temperature levels and two
117 watering levels. The temperature levels closely simulated an average growing season at the
118 thermal extremes of the species range, which we designate as Hot and Cool treatments.
119 Watering levels contrasted a perennial and seasonal stream, which we refer to as Well-
120 watered and Drought treatments. A detailed description of treatments is provided in the

Supplemental Materials and Methods and summarized in Fig 1. Because growth chambers cannot be subdivided, one chamber was assigned to the Hot treatment level and another to the Cool treatment level. Within each chamber, there were two Well-watered blocks and two Drought blocks. The photosynthetically active radiation in both chambers was approximately $400 \mu\text{mol quanta m}^{-2} \text{s}^{-1}$. The growth chambers did not control humidity, but because of watering and high plant transpiration rates, the relative humidity was quite high in both temperature levels (data not shown).

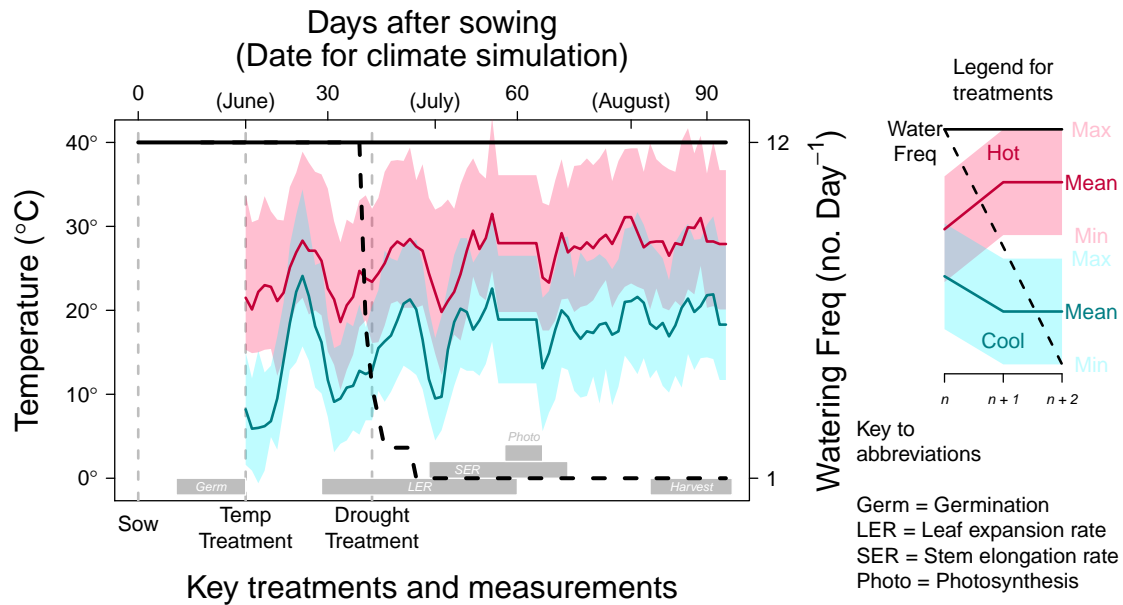


Figure 1: Overview of experimental treatments and timing of key trait measurements. All plants germinated within 21 days of sowing. At that time, we began temperature treatments (left axis), simulating a typical June-August weather pattern at Hot (red) and Cool (blue) sites. The bold lines track the average daily temperatures. Within each day, there was a maximum daytime temperature (top of translucent polygons) and minimum nighttime temperature (bottom of translucent polygons). The drought treatment commenced later by ramping down the frequency of bottom-watering episodes (dashed black line; right axis), while watering frequency was maintained in the control treatment (solid black line). Grey boxes on the bottom of the plot outline the period of key measurements described in the Material and Methods.

128 Trait measurements

129 We measured five traits in response to temperature and watering treatments (Table 2).

Table 2: Key traits measured in this study.

Trait	Units
Days to germination	day
Leaf expansion rate	mm day ⁻¹
Stem elongation rate	cm day ⁻¹
Photosynthetic rate	$\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$
Mortality	probability of death

130 **Days to germination** We tested for population variation in germination rate, measured
 131 as Days to Germination, using a lognormal survival model fit using the survreg function
 132 in the R package **survival** version 2.38 (Therneau, 2015). We treated Population as a fixed
 133 effect and Family as random effect using a Γ frailty function. Statistical significance of the
 134 Population effect was determined using analysis of deviance. Note that, unlike other traits
 135 discussed below, we did not include Block, Treatment, or Population \times Treatment inter-
 136 actions because during germination plants had not been placed into blocks and treatments
 137 had not yet been applied.

138 **Growth rate: leaf expansion and stem elongation** We measured growth rate during
 139 two phases: leaf expansion and stem elongation. We censused leaf length twice per week
 140 from 12 May – 12 June (28–59 days after sowing), resulting in 10 measurements. We
 141 ceased measuring leaf length once it appeared to asymptote and growth shifted to stem
 142 elongation. We also censused plant height on 7 occasions (twice per week) between 29 May
 143 and 20 June (45 to 67 days after sowing). Both leaf expansion and stem elongation were
 144 modelled separately as second-order polynomials. We used empirical Bayes’ estimates of
 145 growth for each individual plant from linear mixed-effects models fit with the R package

146 **lme4** version 1.1-12 (Bates et al., 2015).

147 **Photosynthesis** During the week of 10 to 16 June (57 to 63 days after sowing), we
148 measured daytime photosynthetic rate on a subset of 329 plants evenly spread between
149 treatments and families within populations. The youngest, fully-expanded leaf acclimated
150 for 3 minutes to reach steady state in a 6-cm² chamber of a LI-COR 6400XT Portable Pho-
151 tosynthesis System (LI-COR Biosciences, Lincoln, Nebraska). We made all measurements
152 at ambient light (400 $\mu\text{mol m}^{-2} \text{s}^{-1}$ of photosynthetically active radiation), atmospheric
153 CO₂ (400 ppm), temperature, and moderate relative humidity. During this period, we sus-
154 pended normal day-to-day temperature fluctuations and set daytime temperatures to the
155 average for that period (Cool: 26.5°; Hot: 36.1°) so that all plants within a temperature
156 level could be measured under the same conditions.

157 **Mortality** We assayed mortality during twice-weekly growth measurements. We ana-
158 lyzed the probability of surviving until the end of the experiment as a function of popula-
159 tion, treatment, and their interactions using a Generalized Linear Mixed Model (GLMM)
160 assuming binomially distributed errors. We included Family and Block as random effects.
161 We assessed significance of fixed effects using Type-II Analysis of Deviance with Wald χ^2
162 tests in the R package **car** (Fox and Weisberg, 2011).

163 **Intrinsic variation and plasticity**

164 For all traits (Table 2) except germination (see above), we tested for Population, Treat-
165 ment (Temperature, Water, and Temperature \times Water), and Population \times Treatment
166 interactions (Population \times Temperature, Population \times Water, and Population \times Temper-
167 ature \times Water). We interpreted significant Population effects to indicate intrinsic variation
168 and Population \times Treatment interactions to indicate variation in plasticity. As mentioned

above, we used survival and GLMM models for germination rate and mortality, respectively. For all other traits, we used mixed model ANOVAs with Family and Block included as random factors. We fit models using restricted maximum likelihood in `lmer`, a function in the R package **lme4** (Bates et al., 2015). We determined significant fixed effect terms using a step-wise backward elimination procedure implemented with the `step` function in the R package **lmerTest** version 2.0-32 (Kuznetsova et al., 2016). This package uses Satterthwaite’s approximation to calculate denominator degrees of freedom for F -tests. We also included days to germination as a covariate in growth analyses. To ensure that Population and Treatment effects were specific to a particular growth phase, we included germination day as a covariate in leaf expansion and stem elongation analyses.

Principal components of germination, growth, and photosynthesis

For each single-trait model above, we extracted the Population coefficient (factoring out Treatment and other effects). The multivariate distribution of these coefficients was then summarized using principal components analysis. The first principal component of these traits (TraitPC1) loaded positively with germination, growth, and photosynthetic rate, therefore we define this as a phenotypic axis delineating fast to slow growth.

Identifying putative selective agents

Latitudinal clines are common, but it is often difficult to ascribe this variation to a particular selective agent. To reiterate, we tested three non-mutually exclusive hypotheses about how such latitudinal clines emerge: 1) one or two climatic variable explains latitudinal trait variation; 2) latitude is a proxy for multiple climatic factors that together shape trait variation; and 3) latitude integrates selection in a broader climatic neighborhood. We found that a population’s position along TraitPC1 correlated strongly with the latitude of origin (see Results) and next used Random Forest regression (Liaw and Wiener, 2002) to

193 identify putative climatic factors underlying trait-latitude associations in *E. cardinalis*. We
194 reasoned that if we identified a single climatic factor that explained more trait variation
195 than latitude, then this would suggest that factor is a key selective agent underlying the
196 latitudinal cline (Hypothesis 1). On the other hand, if multiple climatic factors together
197 are necessary to explain trait variation, then this would suggest that many climatic factors
198 together have imposed selection for the latitudinal cline (Hypothesis 2). We hereafter refer
199 to factors identified in this analysis as ‘Climate-TraitPC1’ variables. To test Hypothesis
200 3 about climatic neighborhoods driving selection, we directly compared local with neigh-
201 borhood climate. We used the immediate collection location for local climate. For climate
202 neighborhoods, we sampled climate at 1000 random points (at 90-m resolution) within a
203 62-km radius buffer around the collection and took the average. We chose this buffer size
204 because neutral genetic differentiation increases slowly with geographic distance, indicating
205 significant gene flow between nearby populations (Paul et al., In review). Significant spatial
206 autocorrelation persisted for approximately 62 km. Since *E. cardinalis* is found exclusively
207 in riparian areas, we only selected points along streams using the National Hydrography
208 Dataset (United States Geological Survey, 2015). Climatic means and variances (see be-
209 low) were weighted by their climatic suitability as determined using a multimodel ensemble
210 average of ecological niche models (Angert et al., 2016).

211 In addition, to help eliminate potentially spurious correlations between TraitPC1 and cli-
212 mate, we tested for overlap between climatic variables that best predict latitude of all *E.*
213 *cardinalis* occurrence records (see detail below), not just the 16 focal populations. We refer
214 to these climatic factors as ‘Climate-Latitude’ variables. The logic is that climatic factors
215 associated with both TraitPC1 and latitude for all populations are more likely to be impor-
216 tant selective agents than climatic factors that happen to correlate with TraitPC1 but do
217 not covary with latitude throughout the *E. cardinalis* range. Therefore, we did not consider
218 Climate-TraitPC1 variables to be candidate selective agents unless the same or very simi-
219 lar variable was found in the Climate-Latitude analysis. We selected Climate-Latitude and

220 Climate-TraitPC1 variables independently using Variable Selection Using Random Forest
221 (VSURF) algorithm in the R package **VSURF** version 1.0.3 (Genuer et al., 2016). VSURF
222 ranks variables by their importance over regression trees in the forest. We kept only vari-
223 ables selected for prediction, the most stringent criterion. A visual overview of how we
224 selected climatic variables is depicted in Fig 2.

225 For Climate-Latitude analyses, we compiled a representative set of 356 recent (since 2000)
226 known *E. cardinalis* occurrences. These occurrences were thinned by 50% to correct for
227 uneven sampling from a comprehensive set of herbarium records and an exhaustive field
228 survey in 2010-11 (Angert et al., 2016). For both Climate-TraitPC1 analyses (16 focal
229 populations) and Climate-Latitude (many populations), we used a 90-m digital elevation
230 model from HydroSHEDS (Lehner et al., 2006) to extract elevation. Monthly interpolated
231 climate layers were calculated using ClimateWNA version 5.30 (Wang et al., 2012), which
232 accurately downscales climate data specifically for the rugged topography of western North
233 America. For each occurrence, we calculated bioclimatic variables using the biovars func-
234 tion in the R package **dismo** version 1.1-1 (Hijmans et al., 2016). We included 24 climatic
235 factors, 9 from ClimateWNA and 15 bioclimatic variables (Table S2). The bioclimatic
236 variables included all permutations of two climatic factors, temperature and precipitation,
237 and six temporal scales (annual average, coldest quarter, warmest quarter, wettest quarter,
238 driest quarter, or seasonality) as well as mean diurnal range, isothermality, annual temper-
239 ature range. For each variable, we calculated both a 30-year normal by averaging annual
240 values between 1981 and 2010 and 30-year coefficient of variation, a standardized metric
241 of interannual climatic variation. Temperatures were converted to Kelvin to be on a ratio
242 scale appropriate for calculating the coefficient of variation (CV). In total, the VSURF al-
243 gorithm selected among 96 climate variables: 24 climatic factors \times 2 types (30-year average
244 and CV) \times 2 spatial scales (local and neighborhood).

245 Results

246 A coordinated latitudinal cline in germination, growth, and photosynthe- 247 sis

248 There are strong genetically-based trait differences in time to germination, growth, and
249 photosynthetic rate among populations of *E. cardinalis*, as evidenced by large and signif-
250 icant population effects for these traits (Table 3). A single principal component captured
251 71.6 % of the trait variation among populations, defining an axis of variation from fast
252 to slow growth (Fig 3). As we explain below, intrinsic differences between populations in
253 terms of plant function (photosynthesis) and performance (growth) contrasted with the
254 low amount of variation in plasticity. There were similar latitudinal clines for individual
255 traits underlying PC1 (Figures S1 to S4).

Table 3: Summary of Population, Treatment, and Population \times Treatment effects. We used different statistical modeling for the diverse traits assayed – glmer: generalized linear mixed model using the R package **lme4** (Bates et al., 2015); lmer: linear mixed model using the R package **lme4** (Bates et al., 2015); survreg: survival regression using the R package **survival** (Therneau, 2015). Note that temperature and water treatments were imposed after germination, hence are not applicable to this trait. Complete analysis of variance/deviance tables for each trait are available in the Supporting Information. Key to statistical significance: * $P < 0.05$; ** $P < 0.01$; *** $P < 0.001$

Trait	Germination	Leaf expansion	Stem elongation	Photosynthesis	Mortality
Statistical model	survreg	lmer	lmer	lmer	glmer
Population	***	***	***	***	
Temperature	NA	***	***	**	***
Water	NA	*			***
Pop \times Temp	NA			*	
Pop \times Water	NA	*			
Temp \times Water	NA				***
Pop \times Temp \times Water	NA				

256 Little evidence for variation in plasticity

257 Genotype \times environment (G \times E) interactions are also a common signature of local adap-
258 tation. We found little evidence of G \times E in *E. cardinalis*. There were only two statisti-
259 cally significant Population \times Treatment interactions (Table 3, Fig. ??), but these were
260 not strong compared to Population and Temperature effects. Otherwise, populations re-
261 sponded similarly to treatments: faster growth in the hot treatment, slower growth in the
262 dry treatment, and high mortality in the hot, dry treatment (Table 3). Note that inter-
263 actions were calculated after factoring out intrinsic trait differences, necessarily reducing
264 statistical power to detect significant interactions relative to main effects. However, the
265 fact that the Population and Temperature effects were often highly significant ($P \ll 0.001$
266 in most cases) suggests that statistical power alone cannot explain why we failed to de-
267 tect Population \times Treatment interactions. Complete ANOVA tables are available in the
268 Supporting Information (Tables S3 to S6)

269 Neighborhood climatic variability best explains latitudinal cline

270 Interannual variation in climate averaged over 62-km radius climatic neighborhoods around
271 populations correlated most strongly with trait variation and latitude of *E. cardinalis* oc-
272 currences (Fig. 4, Table S7). All 16 Climate-Latitude and 3 Climate-TraitPC1 variables
273 were neighborhood rather than local variables (Fig. 4). This pattern was generally true for
274 most Climate-TraitPC1 and Climate-Latitude variables (Fig. S6). On average neighbor-
275 hood Climate-TraitPC1 correlation coefficients were 0.16 higher (paired t -test, $t = 7.87$,
276 d.f. = 33.6, $P = 3.94 \times 10^{-9}$) Climate-Latitude correlation coefficients were 0.13 higher
277 (paired t -test, $t = 6.71$, d.f. = 36.8, $P = 7.22 \times 10^{-8}$). Note that we adjusted degrees
278 of freedom and paired t -tests to account for the fact that many climatic factors are highly
279 correlated and not independent. Specifically, we calculated the effective number of indepen-
280 dent climatic factors (M_{eff}) using the formula $M_{\text{eff}} = 1 + (M - 1)(1 - \text{Var}(\lambda)/M)$ (Chevrud

281 2001), where M is the original number of climatic factors and λ are the eigenvalues of the
 282 correlation matrix of all climatic factors. The most important climatic variables related to
 283 the interannual variation rather than average climate.

284 Among Climate-Latitude and Climate-TraitPC1 variables, neighborhood climatic variabil-
 285 ity over 30 years (1981–2010) in either winter precipitation (bio16_σ) and/or temperature
 286 (bio11_σ) are the strongest candidates driving the latitudinal cline in *E. cardinalis* (see Ta-
 287 ble S2 for a key to climate variable abbreviations). Note that the coefficient of variation of
 288 a climatic factor is subscripted with σ whereas the mean is subscripted with μ . More specif-
 289 ically, greater winter precipitation variability and lower winter temperature variability are
 290 associated with Southern latitudes and higher TraitPC1 values (Fig. 5A,B). Neighborhood
 291 interannual variation in winter precipitation (bio16_σ) was the most important Climate-
 292 Latitude variable (Fig. 4A). However, neighborhood bio16_σ did not overlap with Climate-
 293 TraitPC1 variables (Fig. 4B). We nevertheless consider it a plausible candidate for two
 294 reasons. First, neighborhood bio16_σ correlated strongly with TraitPC1 (Fig. 5A). Second,
 295 one of the most important Climate-TraitPC1 variables (neighborhood bio15_σ ; Fig. 5B,C)
 296 is very similar to bio16_σ . In Mediterranean climates like California, most precipitation
 297 occurs in the wettest quarter (winter), so years with low winter precipitation also have
 298 low precipitation seasonality. Hence, highly variable year-to-year winter precipitation at
 299 lower latitude (Fig. 5D) is closely associated with large swings in precipitation seasonality
 300 (Fig. 5C).

301 Interannual variation in temperature of the coldest quarter (neighborhood bio11_σ) is an-
 302 other plausible candidate because it was only variable in both Climate-Latitude and Climate-
 303 TraitPC1 analyses (Fig. 4). Neighborhood bio11_σ explained more variation in TraitPC1
 304 than latitude (latitude $r^2 = 0.55$ vs. bio11_σ $r^2 = 0.6$; Fig. S7), whereas neighborhood
 305 bio16_σ did slightly worse (bio16_σ $r^2 = 0.49$). Models using bio15_σ or bio11_σ to predict
 306 TraitPC1 also had significantly lower Akaike Information Criteria (AIC) than the latitude
 307 model (AIC of different models – bio15_σ : 48.5; bio11_σ : 52.4; latitude: 54.5). The best two-

factor model including both neighborhood bio15_σ and bio11_σ did not significantly improve explanatory power ($r^2 = 0.71$, $\text{AIC} = 49.2$). In summary, either variation in precipitation or temperature seasonality may be important selective agents, but there is no strong evidence that they are both important. The most important Climate-TraitPC1 variable, neighborhood variation in mean diurnal range (bio2_σ ; Fig. 4B) did not have any obvious similarity to Climate-Latitude variables. Given the large number of potential associations, we therefore think this may be a spuriously strong relationship and do not consider it further.

Discussion

We found evidence for one of two common signatures of local adaptation in the perennial herb *Erythranthe cardinalis*. Latitudinal clines in germination rate, photosynthesis, and growth suggest adaptive differentiation in important physiological traits of the species. However, we found little evidence that populations respond differently to temperature or drought. As we discuss below, this latter result may indicate that some dimensions of the fundamental abiotic niche **is** relatively conserved. Finally, our results suggest that neighborhood-scale climate and interannual variation are more important selective agents than local averages. In the paragraphs that follow, we tie these results into the broader threads of evolutionary theory that might help explain why intrinsic variation in physiology changes clinally, whereas plastic responses to temperature and drought are relatively similar.

Evolutionary theory indicates that the shape of fitness tradeoffs, demography, and gene flow can constrain adaptation (Levins, 1968; Ronce and Kirkpatrick, 2001; Lenormand, 2002) and hence the type of variation maintained within species. Specifically, adaptive variation cannot be maintained by spatially varying selection if tradeoffs are too strong, demography is strongly asymmetric, and/or maladaptive gene flow is too high. In *E. cardinalis* we

333 found substantial genetically based variation among populations along a phenotypic axis
334 from fast to slow growth that varied over a large spatial scale (Fig. 3). If this variation is
335 adaptive, it suggests that the fitness tradeoff between low versus high latitude environments
336 is not too strong nor swamped by demographic asymmetry or maladaptive gene flow.
337 That is, alleles favoured at one latitude are not strongly selected against when they flow
338 to another population, allowing locally adaptive genetic variation to be maintained by
339 spatially heterogeneous selection. We also know from previous work that population size
340 does not vary strongly with latitude. Gene flow appears to be high, but attenuates at
341 broad spatial scales, especially between Southern ($< 35^{\circ}\text{N}$) and Northern portions of the
342 range (Paul et al., In review).

343 Nevertheless, local gene flow from ~~similar~~ environments may shape how selection varies
344 with latitude. Theory predicts that populations will not be perfectly adapted to their
345 immediate habitat when there is gene flow from surrounding populations with different
346 optima (Lenormand, 2002). With spatial heterogeneity and gene flow, traits will not co-
347 vary perfectly with the local optimum (Slatkin, 1978; Paul et al., 2011; Hadfield, 2016),
348 but should instead better match the average environment experienced by nearby popula-
349 tions connected through gene flow, which we refer to as the climatic neighborhood. Gene
350 flow and spatial heterogeneity may therefore be important in maintaining genetic variation
351 (Yeaman and Jarvis, 2006). As this hypothesis predicts, climatic neighborhoods (62-km
352 buffer around populations) correlated with ~~TraitPC1~~ and latitude of occurrences better
353 than local climate (Fig. 4). We interpret this as suggestive evidence that gene flow be-
354 tween nearby *E. cardinalis* populations **shapes selection** – populations are locally adapted
355 to prevailing climate in their neighborhood, but perhaps not perfectly adapted to their
356 local climate. This may not greatly constrain local adaptation because local and neighbor-
357 hood climate values were generally similar in *E. cardinalis* populations (data not shown).
358 Therefore, we would predict in reciprocal transplants that populations whose local climate
359 is farther from their neighborhood average would be less well adapted than those close to

360 their neighborhood average.

361 Southern populations, which appear to experience more frequent drought years (see be-
362 low), might have adapted by tolerating drought better than northern populations, thereby
363 expanding the fundamental niche of the species as a whole. We found no evidence for this;
364 all populations responded to drought and temperature similarly (Table 3). Plants grew
365 faster in the Hot treatment, but there was little effect of drought on growth. Rather, the
366 effects of drought took longer to materialize but resulted in high mortality, especially in
367 the Hot treatment. However, there was no differential mortality among populations in this
368 treatment. We hypothesize that evolution of this dimension of the fundamental niche may
369 be constrained by a combination of strong fitness tradeoffs, demographic asymmetry, and
370 gene flow. Riparian habitats where *E. cardinalis* live are highly heterogeneous at small
371 spatial scales. Plants in the stream never have to tolerate drought whereas plants only a
372 few meters away may experience extreme drought since there is little direct precipitation
373 during the growing season in Mediterranean climates of western North America. We hy-
374 pothesize alleles that confer greater drought tolerance may be quite costly in well-watered
375 soils, and *vice versa*, leading to strong fitness tradeoffs. Such tradeoffs would promote
376 specialization to one soil ~~type~~ or another, thereby inhibiting the evolution of broad envi-
377 ronmental tolerance within a population. Demography and gene flow may reinforce niche
378 conservatism. A new mutant with increased drought tolerance that could survive at the
379 resource-poor margin of a population would likely be demographically overwhelmed by the
380 larger census populations that can be maintained in higher-resource environments. Infre-
381 quent wet years may also produce most seeds, so selection is weighted towards alleles that
382 have high fitness in the wet environment, even if dry years are more frequent (Templeton
383 and Levin, 1979; Brown and Venable, 1986). Finally, gene flow, which is generally high
384 among *E. cardinalis* populations within the same ecoregion (Paul et al., In review), will
385 thwart local adaptation and reinforce specialization. Thus, the spatial grain of the environ-
386 ment, demographic asymmetry, and gene flow may conspire to constrain local adaptation

387 via altered fundamental niche.

388 In sum, these results indicate that differences in physiology and growth, but not responses
389 to temperature and drought, mediate local adaptation to climate in *E. cardinalis*. Next,
390 we would like to understand why variation in these particular traits may be adaptive. We
391 argue that temporally more variable environments, as experienced by southern populations,
392 select for a more ‘annualized’ life-history strategy. The association between position along
393 the ‘fast-slow’ continuum and associated traits in *E. cardinalis* is similar to interspecific
394 relationships between growth, functional traits, and life history (Adler et al., 2014; Salguero-
395 Gómez et al., 2016).

396 Greater investment in aboveground growth, as opposed to belowground storage for future
397 seasons, may be favoured in climates with more frequent drought years, but maladaptive
398 in climates with more consistent precipitation. Suppose plants that grow quickly and
399 allocate new resources to continued growth rather than storage have higher fitness over a
400 single growing season. However, by not allocating resources to storage, these fast-growing
401 plants begin future seasons at a deficit. Therefore, in a stable environment where winter
402 survivorship is assured in most years, failure to store resources may reduce lifetime fitness.
403 But the environment is not constant. For perennial herbs in Mediterranean climates, a
404 dry winter (rainy season) can kill the rhizomes (underground stems that store nutrients
405 for future growth) before emergence or aboveground stems before flowering. If drought
406 years occur frequently enough, selection may favour the fast-growing strategy because
407 there is no advantage to storage if drought kills plants before flowering. Considering life-
408 history strategy as a continuum from no storage (annual) to lots of storage (perennial),
409 we hypothesize that the optimal allocation to aboveground growth is more ‘annualized’ in
410 southern climates that have greater interannual variation in precipitation.

411 The hypothesis that greater precipitation variability selects for an annualized life history
412 is tentative, but consistent with theory and data from other species. Life history theory

413 shows that less variable environments are one factor that favours the evolution of perenni-
 414 ality (Stearns, 1976; Iwasa and Cohen, 1989; Friedman and Rubin, 2015). Populations of
 415 the perennial *Plantago asiatica* show a similar latitudinal cline in growth and allocation to
 416 storage (Sawada et al., 1994), though these authors attribute the cline to variation in grow-
 417 ing season length. There are also life history clines in the closely related species *E. guttata*,
 418 but the underlying traits and climatic drivers are quite different. Annual *E. guttata* flower
 419 sooner and produce fewer stolons in response to climates with shorter seasons and more
 420 intense summer drought (Lowry and Willis, 2010; Friedman et al., 2015; Kooyers et al.,
 421 2015). In contrast, there are no truly annual (monocarpic and semelparous) populations of
 422 *E. cardinalis*. Rather, our hypothesis states that climatic variability selects on quantitative
 423 variation in allocation to growth versus storage. This hypothesis makes several indepen-
 424 dent, testable predictions. The allocation tradeoff predicts that northern populations will
 425 provision more assimilate to rhizomes compared with southern populations. If southern
 426 populations are indeed more ‘annualized’ because more frequent droughts cause mortality,
 427 then we predict that species distribution models using recent climate would best predict
 428 occurrences in the south, whereas longer term climate would be a better predictor in the
 429 north. Finally, we predict that southern populations would show greater variation in the
 430 size of recruits and higher maximum population growth rates. Exploratory analysis also
 431 pointed to variation in winter temperature as a potential selective agent. Specifically, win-
 432 ter temperature is more variable in the north, which may lead to less predictability in the
 433 start of the growing season or late frost. This could affect selection on phenology, which we
 434 did not study here, but it is not obviously connected to the latitudinal cline in physiological
 435 traits.

436 In summary, we found evidence for a coordinated latitudinal cline in germination rate,
 437 photosynthesis, and growth, suggesting local adaptation. These differences between popu-
 438 lations suggest different trait optima in different climates. We did not find evidence that the
 439 relative performance of populations shifts with temperature or watering regime, suggesting

440 relatively little variation in plasticity. Exploratory analysis implicate that more variable
441 precipitation regimes at lower latitude drive much of the latitudinal cline, though other
442 climatic factors could also contribute. Interestingly, the climatic neighborhood may shape
443 selective pressures more than local climate. In the future, we will use field experiments to
444 test whether greater variation in precipitation selects for faster growth and that selection
445 on temperature/drought responses does not vary among populations. By doing so, we aim
446 to understand why certain physiological and developmental mechanisms, but not others,
447 contribute to local adaptation.

448 References

- 449 Adler, P. B., R. Salguero-Gómez, A. Compagnoni, J. S. Hsu, J. Ray-Mukherjee, C. Mbeau-
450 Ache, and M. Franco, 2014. Functional traits explain variation in plant life history
451 strategies. *Proceedings of the National Academy of Sciences of the United States of*
452 *America* 111:740–745.
- 453 Angert, A. L., 2006. Demography of central and marginal populations of monkeyflowers
454 (*Mimulus cardinalis* and *M. lewisii*). *Ecology* 87:2014–2025.
- 455 Angert, A. L., M. J. Bayly, S. N. Sheth, and J. R. Paul, 2016. Testing range-limit hypothe-
456 ses using range-wide habitat suitability and occupancy for the scarlet monkeyflower
457 (*Mimulus cardinalis*) .
- 458 Angert, A. L., H. Bradshaw Jr, and D. W. Schemske, 2008. Using experimental evolution
459 to investigate geographic range limits in monkeyflowers. *Evolution* 62:2660–2675.
- 460 Barton, N. H., 1999. Clines in polygenic traits. *Genetics Research* 74:223–236.
- 461 Bates, D., M. Mächler, B. Bolker, and S. Walker, 2015. Fitting linear mixed-effects

models using lme4. Journal of Statistical Software 67:1–48. URL <http://CRAN.R-project.org/package=lme4>. R package version 1.1-7.

Blackman, B. K., S. D. Michaels, and L. H. Rieseberg, 2011. Connecting the sun to flowering in sunflower adaptation. Molecular Ecology 20:3503–3512.

Bradshaw, H. and D. W. Schemske, 2003. Allele substitution at a flower colour locus produces a pollinator shift in monkeyflowers. Nature 426:176–178.

Bradshaw, W. and C. Holzapfel, 2008. Genetic response to rapid climate change: it’s seasonal timing that matters. Molecular ecology 17:157–166.

Bradshaw, W. E. and C. M. Holzapfel, 2001. Genetic shift in photoperiodic response correlated with global warming. Proceedings of the National Academy of Sciences 98:14509–14511.

Brown, J. S. and D. L. Venable, 1986. Evolutionary ecology of seed-bank annuals in temporally varying environments. The American Naturalist 127:31–47.

Chevrud, J., 2001. A simple correction for multiple comparisons in interval mapping genome scans. Heredity 87:52–58.

Clausen, J., D. Keck, and W. Hiesey, 1940. Experimental studies on the nature of species. I. The effects of varied environments on western American plants, vol. 520. Carnegie Institution of Washington, Washington, D.C.

Cleveland, W. S., E. Grosse, and W. M. Shyu, 1992. Local regression models. Statistical models in S Pp. 309–376.

Coyne, J. A. and E. Beecham, 1987. Heritability of two morphological characters within and among natural populations of *Drosophila melanogaster*. Genetics 117:727–737. URL <http://www.genetics.org/content/117/4/727>.

Endler, J., 1977. Geographic variation, clines, and speciation. Princeton University Press,

486 Princeton, NJ.

487 Fox, J. and S. Weisberg, 2011. An R Companion to Applied Regression. Second ed. Sage,
488 Thousand Oaks CA.

489 Friedman, J. and M. J. Rubin, 2015. All in good time: Understanding annual and perennial
490 strategies in plants. *American Journal of Botany* 102:497–499.

491 Friedman, J., A. D. Twyford, J. H. Willis, and B. K. Blackman, 2015. The extent and
492 genetic basis of phenotypic divergence in life history traits in *Mimulus guttatus*. *Molecular*
493 *Ecology* 24:111–122.

494 Genuer, R., J.-M. Poggi, and C. Tuleau-Malot, 2016. VSURF: Variable Selection Using
495 Random Forests. URL <http://CRAN.R-project.org/package=VSURF>. R package ver-
496 sion 1.0.3.

497 Gilbert, P., 2014. Brief User’s Guide: Dynamic Systems Estimation. URL [http://cran.r-](http://cran.r-project.org/web/packages/dse/vignettes/dse-guide.pdf)
498 [project.org/web/packages/dse/vignettes/dse-guide.pdf](http://cran.r-project.org/web/packages/dse/vignettes/dse-guide.pdf).

499 Givnish, T. J., 1988. Adaptation to sun and shade: a whole-plant perspective. *Functional*
500 *Plant Biology* 15:63–92.

501 Hadfield, J. D., 2010. Mcmc methods for multi-response generalized linear mixed mod-
502 els: The MCMCglmm R package. *Journal of Statistical Software* 33:1–22. URL
503 <http://www.jstatsoft.org/v33/i02/>.

504 ———, 2016. The spatial scale of local adaptation in a stochastic environment. *Ecology*
505 *Letters* 19:780–788.

506 Hereford, J., 2009. A quantitative survey of local adaptation and fitness trade-offs. *The*
507 *American Naturalist* 173:579–588.

508 Hiesey, W., M. Nobs, and O. Björkman, 1971. Experimental studies on the nature of
509 species. V. Biosystematics, genetics, and physiological ecology of the *Erythranthe* section

510 of *Mimulus*, vol. 628. Carnegie Institution of Washington, Washington, D.C.

511 Hiesey, W. M., J. Clausen, and D. D. Keck, 1942. Relations between climate and intraspe-
512 cific variation in plants. *American Naturalist* Pp. 5–22.

513 Hijmans, R. J., S. Phillips, J. Leathwick, and J. Elith, 2016. dismo: Species distribution
514 modeling. URL <http://CRAN.R-project.org/package=dismo>. R package version 1.1-1.

515 Hoffmann, A. A., A. Anderson, and R. Hallas, 2002. Opposing clines for high and low
516 temperature resistance in *Drosophila melanogaster*. *Ecology Letters* 5:614–618.

517 Hopkins, R., J. Schmitt, and J. R. Stinchcombe, 2008. A latitudinal cline and response to
518 vernalization in leaf angle and morphology in *Arabidopsis thaliana* (Brassicaceae). *New*
519 *Phytologist* 179:155–164.

520 Huey, R. B., G. W. Gilchrist, M. L. Carlson, D. Berrigan, and L. Serra, 2000. Rapid
521 evolution of a geographic cline in size in an introduced fly. *Science* 287:308–309.

522 Huxley, J. S., 1938. Clines: an auxiliary taxonomic principle. *Nature* 142:219–220.

523 Iwasa, Y. and D. Cohen, 1989. Optimal growth schedule of a perennial plant. *American*
524 *Naturalist* 133:480–505.

525 Kawecki, T. J. and D. Ebert, 2004. Conceptual issues in local adaptation. *Ecology Letters*
526 7:1225–1241.

527 Kooyers, N. J., A. B. Greenlee, J. M. Colicchio, M. Oh, and B. K. Blackman, 2015. Repli-
528 cate altitudinal clines reveal that evolutionary flexibility underlies adaptation to drought
529 stress in annual *Mimulus guttatus*. *New Phytologist* 206:152–165.

530 Kooyers, N. J. and K. M. Olsen, 2012. Rapid evolution of an adaptive cyanogenesis cline
531 in introduced north american white clover (*Trifolium repens* L.). *Molecular Ecology*
532 21:2455–2468.

533 Kuznetsova, A., P. Bruun Brockhoff, and R. Haubo Bojesen Christensen, 2016.

534 lmerTest: Tests in Linear Mixed Effects Models. URL [http://CRAN.R-](http://CRAN.R-project.org/package=lmerTest)
535 [project.org/package=lmerTest](http://CRAN.R-project.org/package=lmerTest). R package version 2.0-32.

536 Lehner, B., K. Verdin, and A. Jarvis, 2006. HydroSHEDS technical documentation. World
537 Wildlife Fund, Washington, D.C. URL www.worldwildlife.org/hydrosheds.

538 Lenormand, T., 2002. Gene flow and the limits to natural selection. Trends in Ecology &
539 Evolution 17:183–189.

540 Levins, R., 1968. Evolution in changing environments: some theoretical explorations.
541 Princeton University Press, Princeton, New Jersey.

542 Liaw, A. and M. Wiener, 2002. Classification and regression by randomforest. R News
543 2:18–22. URL <http://CRAN.R-project.org/doc/Rnews/>.

544 Lowry, D. B. and J. H. Willis, 2010. A widespread chromosomal inversion polymorphism
545 contributes to a major life-history transition, local adaptation, and reproductive isola-
546 tion. PLoS biology 8:2227.

547 Nesom, G. L., 2014. Taxonomy of *Erythranthe* sect. *Erythranthe* (phrymaceae). Phytoneu-
548 ron 31:1–41.

549 Oakeshott, J., J. Gibson, P. Anderson, W. Knibb, D. Anderson, and G. Chambers, 1982.
550 Alcohol dehydrogenase and glycerol-3-phosphate dehydrogenase clines in *Drosophila*
551 *melanogaster* on different continents. Evolution Pp. 86–96.

552 Paul, J. R., T. L. Parchman, B. Econopouly, C. A. Buerkle, and A. L. Angert, In review.
553 Population genomics and range limits: diversity and differentiation across the geographic
554 range of *Mimulus cardinalis* (Phrymaceae) .

555 Paul, J. R., S. N. Sheth, and A. L. Angert, 2011. Quantifying the impact of gene flow
556 on phenotype-environment mismatch: A demonstration with the scarlet monkeyflower
557 *Mimulus cardinalis*. The American Naturalist 178:S62–S79.

558 Pfaff, B., 2008. VAR, SVAR and SVEC models: Implementation within R package vars.
559 Journal of Statistical Software 27:1–32.

560 Ronce, O. and M. Kirkpatrick, 2001. When sources become sinks: migrational meltdown
561 in heterogeneous habitats. Evolution 55:1520–1531.

562 Salguero-Gómez, R., O. R. Jones, E. Jongejans, S. P. Blomberg, D. J. Hodgson, C. Mbeau-
563 Ache, P. A. Zuidema, H. de Kroon, and Y. M. Buckley, 2016. Fast–slow continuum and
564 reproductive strategies structure plant life-history variation worldwide. Proceedings of
565 the National Academy of Sciences of the United States of America 113:230–235.

566 Sawada, S., Y. Nakajima, M. Tsukuda, K. Sasaki, Y. Hazama, M. Futatsuya, and
567 A. Watanabe, 1994. Ecotypic differentiation of dry matter production processes in re-
568 lation to survivorship and reproductive potential in *Plantago asiatica* populations along
569 climatic gradients. Functional Ecology 8:400–409.

570 Schluter, D., 2000. The Ecology of Adaptive Radiation. Oxford University Press.

571 Schmidt, P. S., L. Matzkin, M. Ippolito, and W. F. Eanes, 2005. Geographic variation in di-
572 apause incidence, life-history traits, and climatic adaptation in *Drosophila melanogaster*.
573 Evolution 59:1721–1732.

574 Slatkin, M., 1978. Spatial patterns in the distributions of polygenic characters. Journal of
575 Theoretical Biology 70:213–28.

576 Stearns, S., 1976. Life history tactics: a review of the ideas. Quarterly Review of Biology
577 51.

578 Stinchcombe, J. R., C. Weinig, M. Ungerer, K. M. Olsen, C. Mays, S. S. Halldorsdottir,
579 M. D. Purugganan, and J. Schmitt, 2004. A latitudinal cline in flowering time in *Ara-
580 bidopsis thaliana* modulated by the flowering time gene FRIGIDA. Proceedings of the
581 National Academy of Sciences of the United States of America 101:4712–4717.

582 Stock, A. J., B. E. Campitelli, and J. R. Stinchcombe, 2014. Quantitative genetic variance

583 and multivariate clines in the ivyleaf morning glory, *Ipomoea hederacea*. Philosophical
584 Transactions of the Royal Society B: Biological Sciences 369:20130259.

585 Templeton, A. R. and D. A. Levin, 1979. Evolutionary consequences of seed pools. The
586 American Naturalist 114:232–249.

587 Therneau, T. M., 2015. survival: A Package for Survival Analysis in S. URL
588 <http://CRAN.R-project.org/package=survival>. R package version 2.38.

589 Turesson, G., 1922. The genotypic response of the plant species to habitat. Hereditas
590 3:211–350.

591 Umina, P., A. Weeks, M. Kearney, S. McKechnie, and A. Hoffmann, 2005. A rapid shift
592 in a classic clinal pattern in drosophila reflecting climate change. Science 308:691–693.

593 United States Geological Survey, 2015. National Hydrogeography Dataset. United States
594 Geological Survey, Washington, D.C. URL <http://nhd.usgs.gov/index.html>.

595 Wang, T., A. Hamann, D. L. Spittlehouse, and T. Q. Murdock, 2012. ClimateWNA –
596 high-resolution spatial climate data for western North America. Journal of Applied
597 Meteorology and Climatology 51:16–29.

598 Wright, K. M., D. Lloyd, D. B. Lowry, M. R. Macnair, and J. H. Willis, 2013. Indirect
599 evolution of hybrid lethality due to linkage with selected locus in *Mimulus guttatus*. PLoS
600 Biol 11:e1001497.

601 Wu, C., D. Lowry, A. Cooley, K. Wright, Y. Lee, and J. Willis, 2008. *Mimulus* is an
602 emerging model system for the integration of ecological and genomic studies. Heredity
603 100:220–230.

604 Wu, C. A., D. B. Lowry, L. I. Nutter, and J. H. Willis, 2010. Natural variation for drought-
605 response traits in the *Mimulus guttatus* species complex. Oecologia 162:23–33.

606 Yeaman, S. and A. Jarvis, 2006. Regional heterogeneity and gene flow maintain variance

607 in a quantitative trait within populations of lodgepole pine. Proceedings of the Royal
608 Society of London B 273:1587–1593.

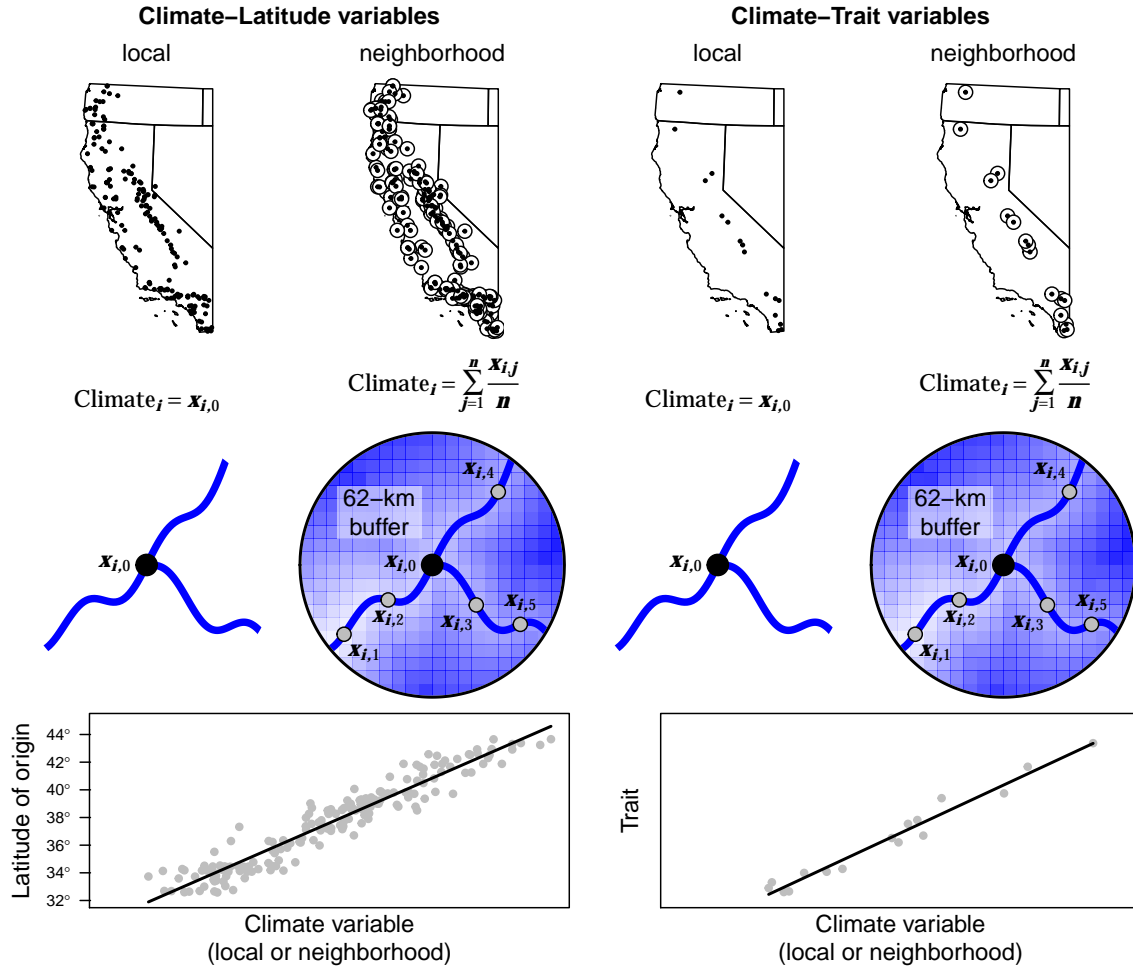


Figure 2: Overview of method for identifying putative climatic selective agents underlying latitudinal cline. We looked for climate variables that explained both the latitude of 356 *E. cardinalis* occurrences ('Climate-Latitude variables') and with traits ('Climate-Trait variables'). For Climate-Latitude variables we extracted climate data from recent occurrences located throughout California and Oregon, USA (shown in map). For Climate-Trait variables, we extracted climatic data for the 16 focal populations. For both analyses, we extracted local and neighborhood climate. Local climate refers to climate only from where a population was collected ($x_{i,0}$). Neighborhood climate was calculated as the average over 1000 points in a 62-km radius climatic neighborhood ($x_{i,1}, x_{i,2}, \dots$), but only along stream habitats as *E. cardinalis* is riparian. We identified climatic factors that most strongly predicted latitude of occurrences (Climate-Latitude variables) and traits (Climate-Trait variables), as shown for hypothetical data in plots at the bottom of the figure.

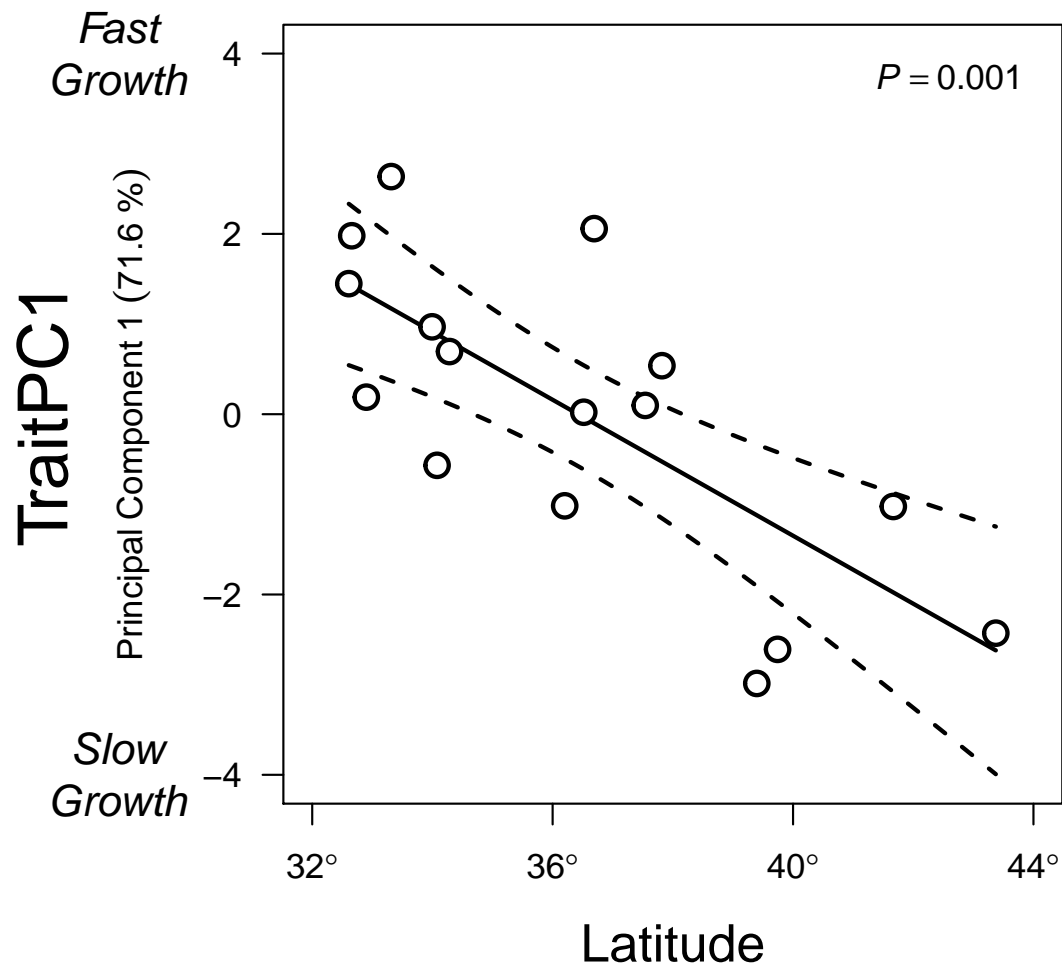


Figure 3: Trait variation, from fast to slow growth, is closely associated with latitude. Each point is a population's latitude of origin (x-axis) and position along the slow to fast growth axis (y-axis), defined as Principal Component 1 of four traits (see Material and Methods). The line and 95% confidence intervals were estimated using linear regression.

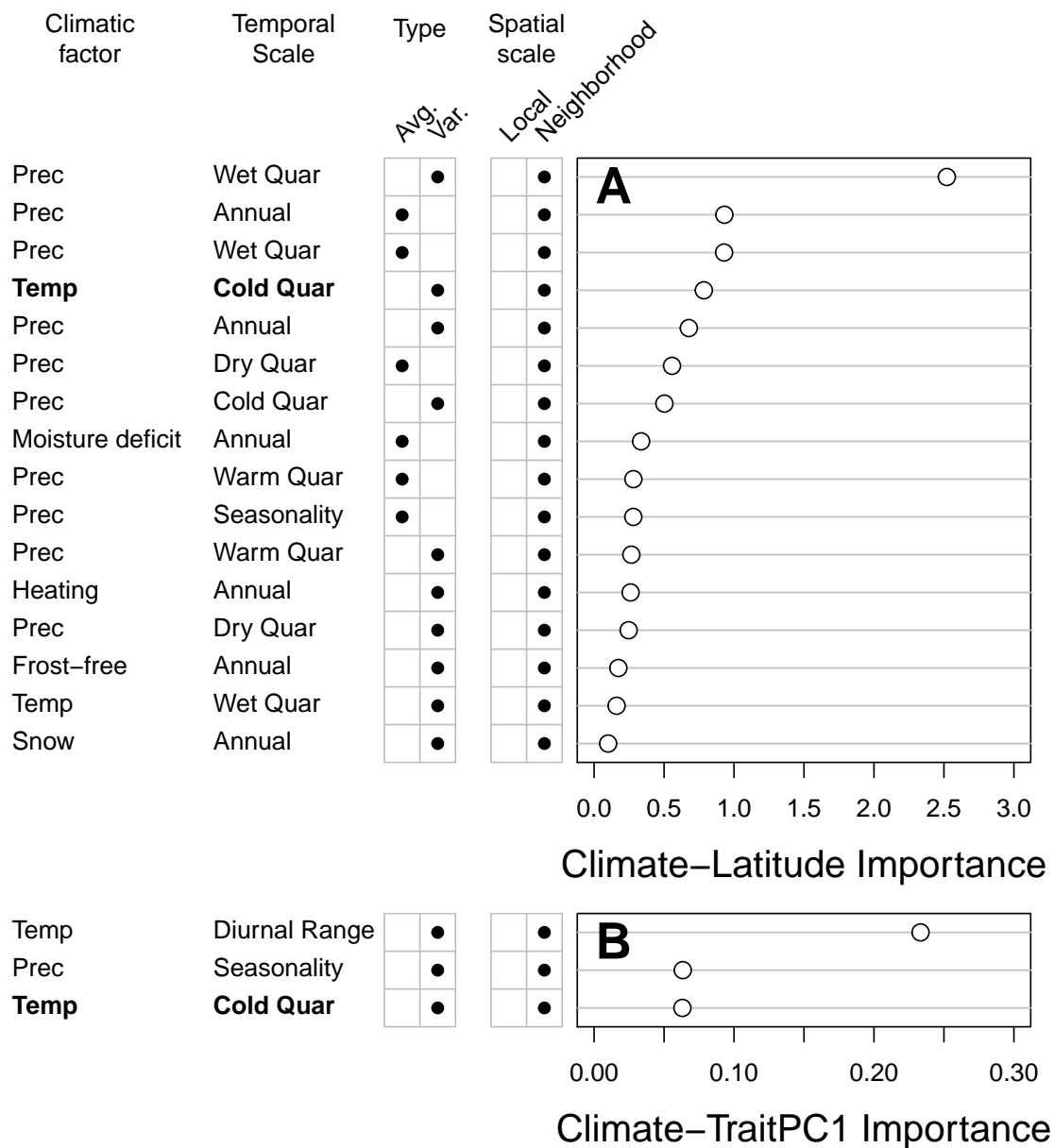


Figure 4: Climatic variation integrated over climatic neighborhood is closely correlated with latitude of *E. cardinalis* and trait variation. A. Using Random Forest regression, we identified 16 climatic variables significantly (high importance) associated with latitude of *E. cardinalis* occurrences. B. Only one of the most important Climate-Latitude variables (in bold) was among the most important Climate-TraitPC1 variables. Note that the Importance values in A and B are not comparable because the dependent variables (Latitude and Trait PC1, respectively) are on different scales. Climatic variables (left of A; right of B) are defined by four qualities: Climatic factor – Temperature (Temp), Precipitation (Prec), Heating degree-days (Heating), Snow (precipitation as snow); Temporal scale – Annual, Coldest quarter (Cold Quar), Warmest Quarter (Warm Quar), Wettest quarter (Wet Quar), Driest Quarter (Dry Quar), or Seasonality; Type – 30-year average (Avg.) or coefficient of variation (Var.); Spatial scale – local or 62-km radius climatic neighborhood.

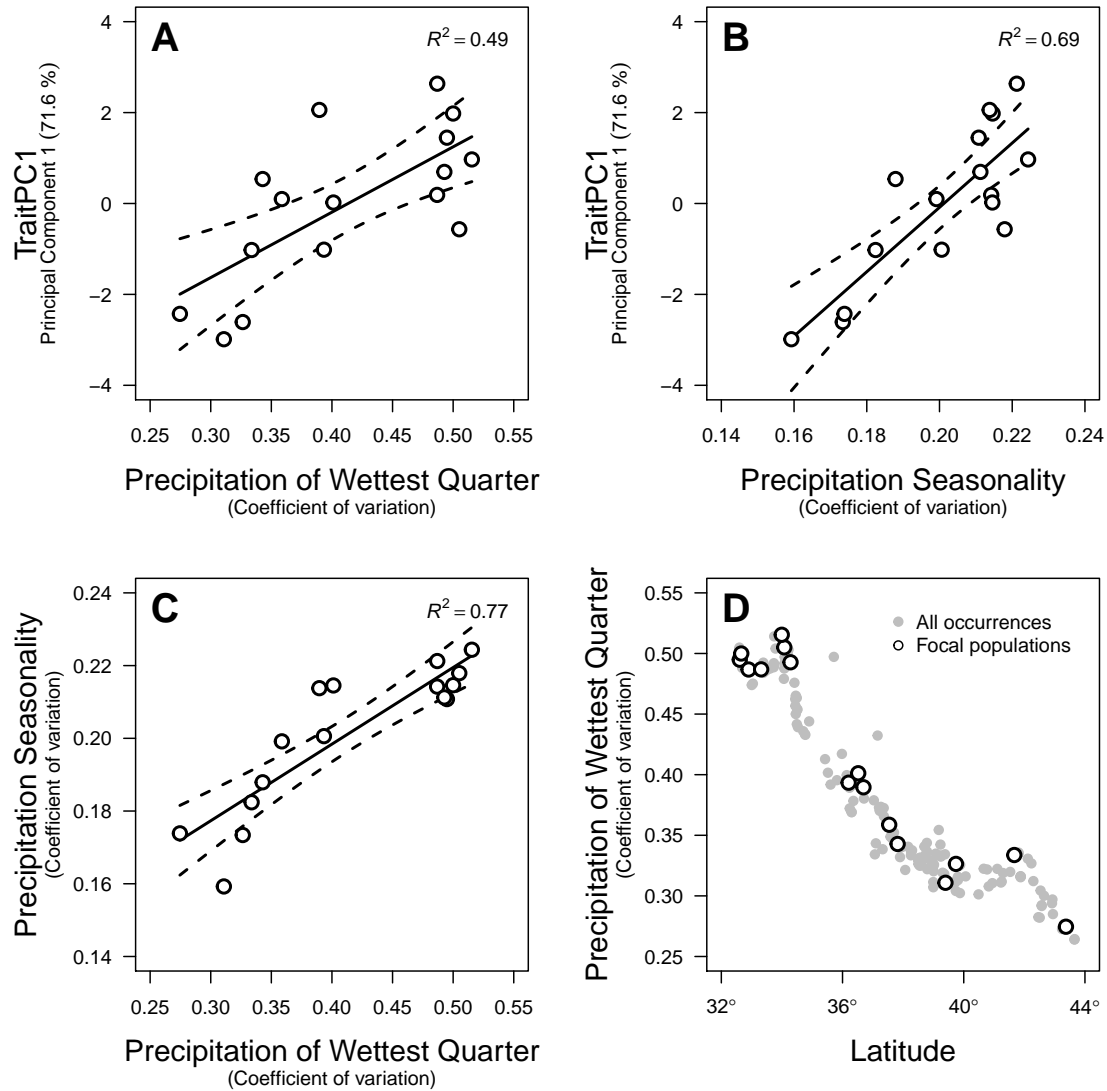


Figure 5: Variation in precipitation is correlated with TraitPC1 and latitude. A. Greater values of TraitPC1 are associated with greater interannual variation in precipitation of the wettest quarter. This was the most important Climate-Latitude variable, but not among the most important Climate-TraitPC1 variables. B. However, a closely related parameter, interannual variation in precipitation seasonality, was among the most important Climate-TraitPC1 variables. C. Across focal populations, variation in precipitation of the wettest quarter and seasonality are closely correlated. D. Southern populations of *E. cardinalis* experience much greater interannual variation in precipitation. In all panels, we report climatic neighborhood values (see Material and Methods). Regression lines, 95% confidence intervals, and coefficients of determination (R^2) were calculated using linear regression.

609 **Supporting Information**

610 **Supporting Tables**

Table S1: Initial size of seedlings did not vary among Populations, Families, or Treatments. We used a censored Gaussian model of initial size at the outset of the experiment (longest leaf length of the first true leaves). The model was censored because we could not accurately measure leaves less than 0.25 mm with digital callipers (217 of 702, 30.9%, were too small). We fit models using a Bayesian MCMC method implemented using the MCMCglmm function with default priors in the R package **MCMCglmm** version 2.17 (Hadfield, 2010). We estimated the posterior distribution from 1000 samples of an MCMC chain run for 10^5 steps after a 10^4 step burn-in. We used step-wise backward elimination procedure to find the best-supported model according to Deviance Information Criterion (DIC).

Model	Random	DIC
Population + Water + Temperature + Population:Water + Population:Temperature + Water:Temperature + Population:Water:Temperature	Family	1638
Population + Water + Temperature + Population:Water + Population:Temperature + Water:Temperature	Family	1605.2
Population + Water + Temperature + Population:Water + Population:Temperature	Family	1603.4
Population + Water + Temperature + Population:Water + Water:Temperature	Family	1577.5
Population + Water + Temperature + Population:Temperature + Water:Temperature	Family	1579.9
Population + Water + Temperature + Population:Water	Family	1577.3
Population + Water + Temperature + Water:Temperature	Family	1550.5
Population + Water + Temperature	Family	1549.3
Population + Water	Family	1541.7
Population + Temperature	Family	1546.8
Water + Temperature	Family	1551.1
Population	Family	1541.9
Water	Family	1543.9
-	Family	1541.7
-	-	1538.3

Table S2: Climatic variables used

Abbreviation	Climate variable
DD_0	degree-days below 0°C(chilling degree-days)
DD5	degree-days above 5°C(growing degree-days)
DD_18	degree-days below 18°C(heating degree-days)
DD18	degree-days above 18°C(cooling degree-days)
NFFD	number of frost-free days
PAS	precipitation as snow (mm) between August in previous year and July in current
Eref	Hargreaves reference evaporation (mm)
CMD	Hargreaves climatic moisture deficit (mm)
RH	mean annual relative humidity
bio1	annual mean temperature
bio2	mean diurnal range (mean of monthly (max temp - min temp))
bio3	isothermality (bio2/bio7) (* 100)
bio4	temperature seasonality (standard deviation *100)
bio5	max temperature of warmest month
bio6	min temperature of coldest month
bio7	temperature annual range (bio5-bio6)
bio8	mean temperature of wettest quarter
bio9	mean temperature of driest quarter
bio10	mean temperature of warmest quarter
bio11	mean temperature of coldest quarter
bio12	annual precipitation
bio15	precipitation seasonality (coefficient of variation)
bio16	precipitation of wettest quarter
bio17	precipitation of driest quarter
bio18	precipitation of warmest quarter
bio19	precipitation of coldest quarter

Table S3: Analysis of variance (ANOVA) table on leaf expansion rate (LER) using **lmerTest** (Kuznetsova et al., 2016). Family and Block were included as random effects. Abbreviations: SS = sum of squares; MS = mean sum of squares (SS / df1); df1 = numerator degrees of freedom; df2 = denominator degrees of freedom.

	SS	MS	df1	df2	<i>F-value</i>	<i>P-value</i>
Day to Germination	12.12	12.12	1	637	35.21	4.9×10^{-9}
Population	22.22	1.48	15	118	4.3	2.5×10^{-6}
Temperature	80.42	80.42	1	5	233.61	2.6×10^{-5}
Water	4.1	4.1	1	5	11.92	0.019
Temperature \times Water	0.03	0.03	1	4	0.07	0.801
Population \times Temperature	2.76	0.18	15	547	0.53	0.925
Population \times Water	9.66	0.64	15	562	1.87	0.024
Population \times Temperature \times Water	4.11	0.27	15	530	0.78	0.700

Table S4: Analysis of variance (ANOVA) table on stem elongation rate (SER) using **lmerTest** (Kuznetsova et al., 2016). Family and Block were included as random effects. Abbreviations: SS = sum of squares; MS = mean sum of squares (SS / df1); df1 = numerator degrees of freedom; df2 = denominator degrees of freedom.

	SS	MS	df1	df2	<i>F-value</i>	<i>P-value</i>
Day to Germination	3.6	3.6	1	662	21.1	5.1×10^{-6}
Population	12	0.8	15	113	4.7	5.8×10^{-7}
Temperature	12.4	12.4	1	6	72.8	1.5×10^{-4}
Water	0.6	0.6	1	5	3.7	0.113
Temperature \times Water	0.9	0.9	1	4	5.2	0.093
Population \times Temperature	3.6	0.2	15	549	1.4	0.126
Population \times Water	2.8	0.2	15	536	1.1	0.330
Population \times Temperature \times Water	1.5	0.1	15	518	0.6	0.874

Table S5: Analysis of variance (ANOVA) table on photosynthetic rate using **lmerTest** (Kuznetsova et al., 2016). Family and Block were included as random effects. Abbreviations: SS = sum of squares; MS = mean sum of squares (SS / df1); df1 = numerator degrees of freedom; df2 = denominator degrees of freedom.

	SS	MS	df1	df2	<i>F-value</i>	<i>P-value</i>
Population	347.7	23.2	15	78	3.02	7.5×10^{-4}
Temperature	134.1	134.1	1	6	17.46	6.4×10^{-3}
Water	51	51	1	4	6.64	0.066
Temperature \times Water	0.7	0.7	1	3	0.09	0.781
Population \times Temperature	218.6	14.6	15	263	1.9	0.024
Population \times Water	87.7	5.8	15	233	0.76	0.724
Population \times Temperature \times Water	91.4	6.1	15	208	0.79	0.686

Table S6: Analysis of deviance table on the probability of mortality by the end of the experiment using Type-II Wald χ^2 tests in the R package **car** (Fox and Weisberg, 2011). Family and Block were included as random effects. Abbreviations: df = degrees of freedom

	χ^2	df	<i>P-value</i>
Population	32	31	0.419
Temperature	31.8	6	1.8×10^{-5}
Water	69.2	12	4.6×10^{-10}
Temperature \times Water	20.7	1	5.3×10^{-6}
Population \times Temperature	5.6	15	0.985
Population \times Water	8.6	15	0.897
Population \times Temperature \times Water	0.2	15	1.000

Table S7: Important climatic variables predicting latitude of *E. cardinalis* populations ('Climate-Latitude') and the first principal component of traits measured in a common garden ('Climate-TraitPC1'). Local climatic variables were measured from the exact location of collection; neighborhood climatic variables were averaged from a 62-km neighborhood around population (see Material and Methods). Importance and significance were determined using the variable selection using random forests (VSURF) algorithm (see Material and Methods). Climatic variables are described in Table S2. μ signifies the mean of the climate variables from 1981–2010; σ indicates coefficient of variation among years.

Climate-Latitude variables	Climate-TraitPC1 variables
Precipitation of wettest quarter (σ , neighborhood)	Mean diurnal range (σ , neighborhood)
Annual precipitation (μ , neighborhood)	Precipitation seasonality (σ , neighborhood)
Precipitation of wettest quarter (μ , neighborhood)	Mean temperature of coldest quarter (σ , neighborhood)
Mean temperature of coldest quarter (σ , neighborhood)	
Annual precipitation (σ , neighborhood)	
Precipitation of driest quarter (μ , neighborhood)	
Precipitation of coldest quarter (σ , neighborhood)	
Hargreaves climatic moisture deficit (μ , neighborhood)	
Precipitation of warmest quarter (μ , neighborhood)	
Precipitation seasonality (μ , neighborhood)	
Precipitation of warmest quarter (σ , neighborhood)	
Heating degree-days (σ , neighborhood)	
Precipitation of driest quarter (σ , neighborhood)	
Number of frost-free days (σ , neighborhood)	
Mean temperature of wettest quarter (σ , neighborhood)	
Precipitation as snow (σ , neighborhood)	

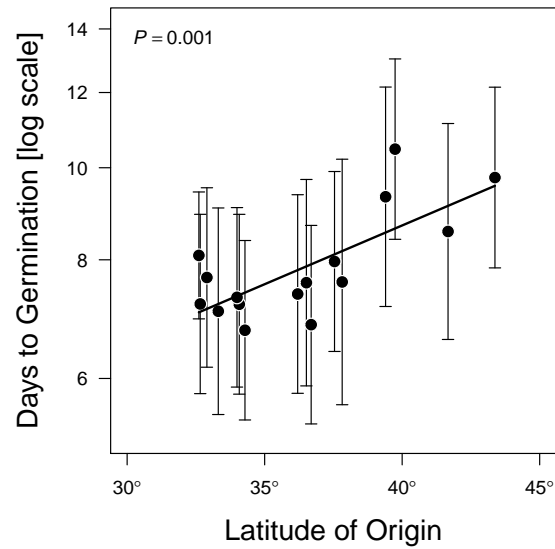


Figure S1: Southern populations germinate faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted days to germination in days under growth chamber conditions (see Material and Methods). Bars around each point are 95% confidence intervals. Predicted time to germination and confidence intervals are based on survival regression (see Materials and Materials). The line is the linear regression of $\log(\text{model-predicted days to germination}) \sim \text{latitude}$. The P -value of the regression is given in the upper left corner.

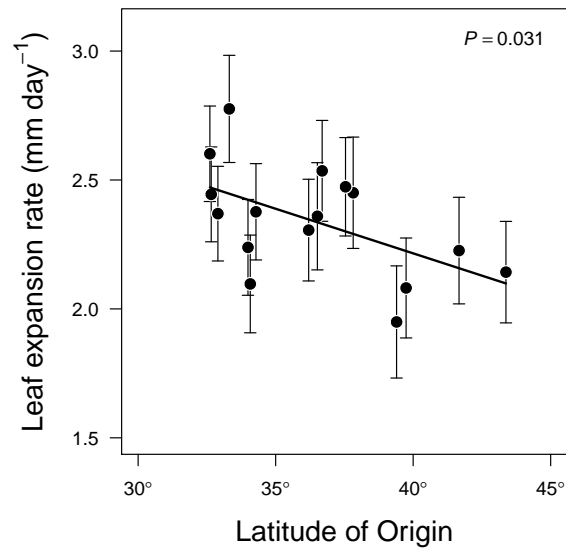


Figure S2: Southern populations grow faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted leaf expansion rate during the rosette phase. Bars around each point are 95% confidence intervals. Predicted leaf expansion rate based least-square mean estimates and confidence intervals were calculated from linear mixed-effects models (see Materials and Materials). The line is the linear regression of model-predicated leaf expansion rate \sim latitude. The P -value of the regression is given in the upper right corner.

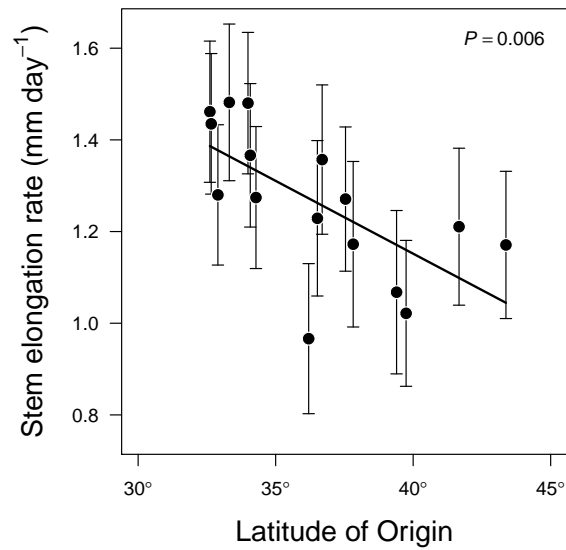


Figure S3: Southern populations grow faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted stem elongation rate. Bars around each point are 95% confidence intervals. Predicted stem elongation rate based least-square mean estimates and confidence intervals were calculated from linear mixed-effects models (see Materials and Materials). The line is the linear regression of model-predicated stem elongation rate \sim latitude. The P -value of the regression is given in the upper right corner.

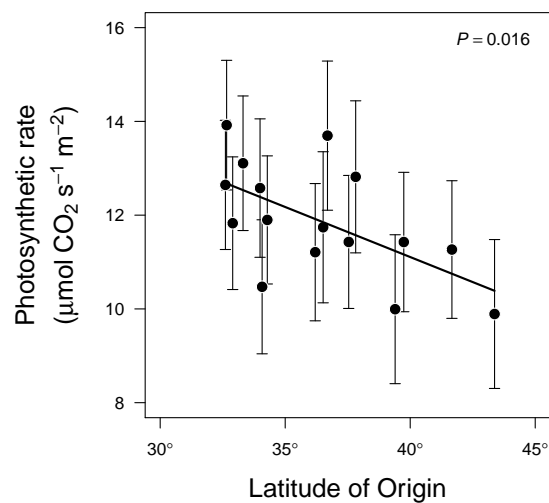


Figure S4: Southern populations photosynthesize faster. Each point is a population of *E. cardinalis* showing its latitude of origin (x-axis) and model-predicted instantaneous photosynthetic rate. Bars around each point are 95% confidence intervals. Predicted photosynthetic rates based least-square mean estimates and confidence intervals were calculated from linear mixed-effects models (see Materials and Materials). The line is the linear regression of model-predicted photosynthetic rate \sim latitude. The P -value of the regression is given in the upper right corner.

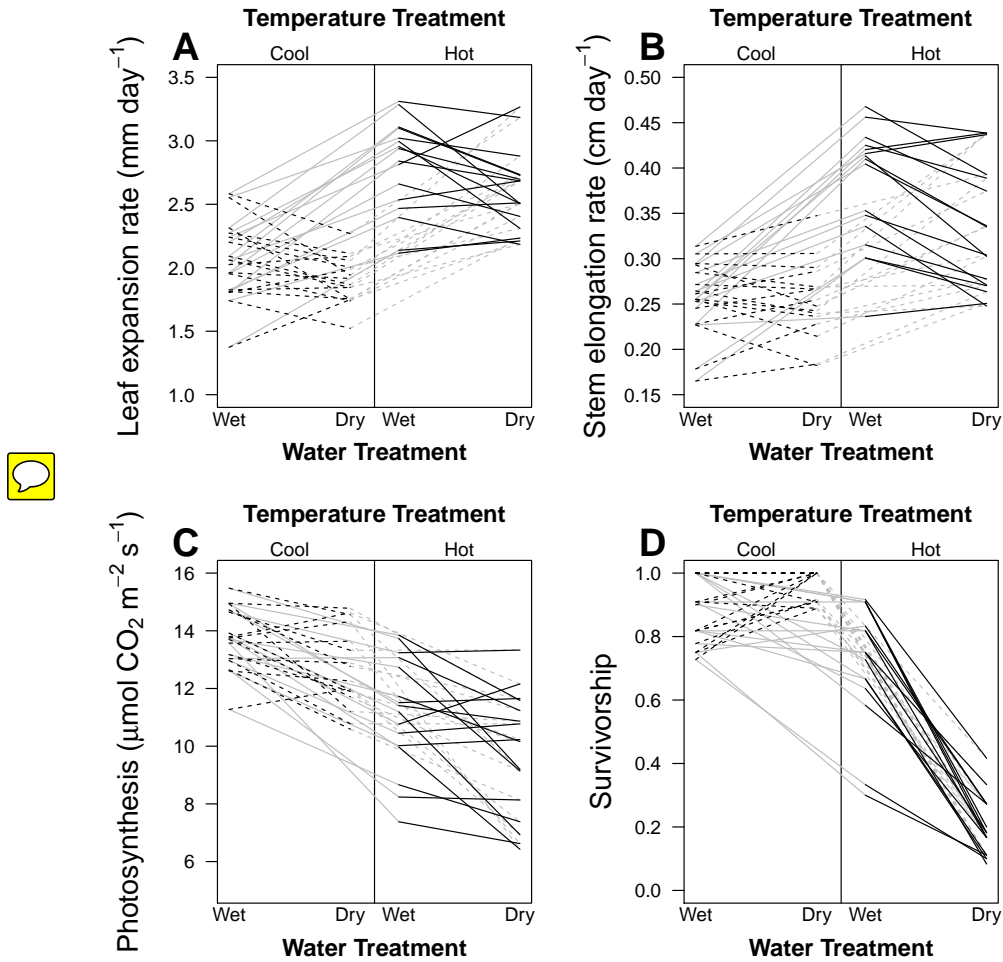


Figure S5: Reaction norms signify little Population \times Treatment interactions. For all panels, black lines represent population-level reaction norms from Wet to Dry in the Cool temperature treatment (dashed black lines) and Hot temperature treatment (solid black lines); gray lines represent reaction norms from Cool to Hot in the Wet treatment (solid gray lines) and Dry treatment (dashed gray lines). The responses shown are (A) leaf expansion rate, (B) stem elongation rate, (C) photosynthesis, and (D) survivorship (= 1 - mortality).

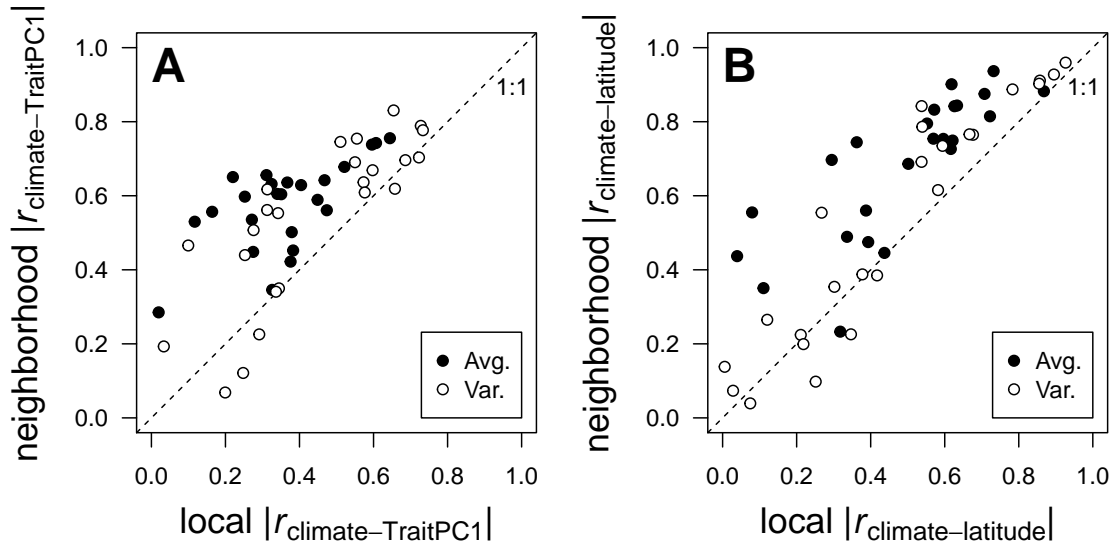


Figure S6: Neighborhood climate predicts TraitPC1 ('Climate-trait', panel A) and Latitude of occurrences ('Climate-latitude', panel B) better than local climate. Each point is the absolute value of the Pearson correlation coefficient ($|r|$) between TraitPC1 (A) or latitude (B) for 24 climatic factors, for which we used both the 30-year mean (closed circles) and coefficient of variation (open circles). Most points lie above the 1:1 line, indicating stronger correlations with neighborhood compared to local climate. Neighborhood climate was integrated over a 62-km radius around focal populations (see text for further detail).

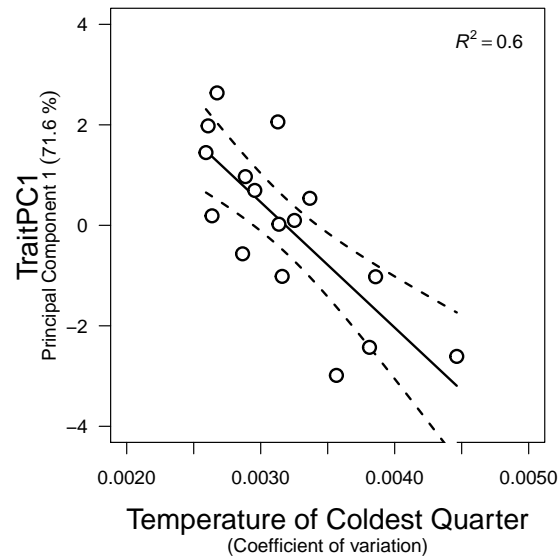


Figure S7: Trait variation, from fast to slow growth, is closely associated with neighborhood variation in temperature of the coldest quarter (bio11_σ). Each point is a population coefficient of variation in bio11 averaged over a 62-km climatic neighborhood (x-axis) and position along the slow to fast growth axis (y-axis), defined as Principal Component 1 of four traits (see Material and Methods). The line and 95% confidence intervals were estimated using linear regression.

612 Supporting Material and Methods

613 Temperature treatments

614 We simulated typical growing season (June 1 - August 15) air temperatures at the two most
615 thermally divergent focal sites in our study, Whitewater Canyon (WWC, Hot) and Little
616 Jameson (LIJ, Cool). We downloaded daily interpolated mean, minimum, and maximum
617 air temperature from 13 years (2000-2012) at both sites from ClimateWNA (Wang et al.,
618 2012). This range was chosen because seeds used in the experiment were collected around
619 2012, thus their presence in that location at that time suggests that populations were able
620 to persist there for at least some years before collection. Monthly temperatures from Cli-
621 mateWNA are highly correlated with the air temperature recorded from data loggers in
622 the field at these sites (A. Angert, unpub. data). Hence, the ClimateWNA temperature
623 profiles are similar to actual thermal regimes experienced by *E. cardinalis* in nature. We
624 simulated realistic temperature regimes by calculating the mean temperature trend from
625 June to August using LOESS (Cleveland et al., 1992). The residuals were highly autocor-
626 related at both sites (warmer than average days are typically followed by more warm days)
627 and there was strong correlation ($r = 0.65$) between sites (warm days in WWC were also
628 warm in LIJ). The ‘VARselect’ function in the **vars** package for R (Pfaff, 2008) indicated
629 that a lag two Vector Autoregression (VAR(2)) model best captured the within-site auto-
630 correlation as well as between-site correlation in residuals. We fit and simulated from the
631 VAR(2) model using the package **dse** (Gilbert, 2014) in R. Simulated data closely resem-
632 bled the autocorrelation and between-site correlation of the actual data. From simulated
633 mean temperature, we next selected minimum and maximum daily temperatures. Mean,
634 min, and max temperature were highly correlated at both sites. We chose min and max
635 temperatures using site-specific fitted linear models between mean, max, and min tem-
636 perature, with additional variation given by normally distributed random deviates with
637 variance equal to the residual variance of the linear models. For each day, the nighttime

638 (22:00 - 6:00) chamber temperature was set to the simulated minimum temperature. Dur-
639 ing the middle of the day, temperature was set to the simulated maximum temperature,
640 with a variable period of transition between min and max so that the average temperature
641 was equal the simulated mean temperature.

642 **Watering treatments**

643 For watering treatments, we simulated two extreme types of streams where *E. cardinalis*
644 grows. In the well-watered treatment, we simulated a large stream that never goes dry
645 during the summer growing season. In the drought treatment, we simulated a small stream
646 that has ample flow at the beginning of the season due to rain and snow melt, but gradually
647 dries down through the summer. In both treatments, plants were bottom-watered using
648 water chilled to 7.5°C. Plants in the well-watered treatment were fully saturated every two
649 hours during the day. Watering in the drought treatment gradually declined from every
650 two hours to every day between May 20 (36 days after sowing) and 10 June (57 days after
651 sowing). Simultaneously, the amount of bottom-watering per flood decreased, such that
652 only the bottom of the cone-tainers were wetted by the end of the experiment.