

Understanding the Clinical Microbiome Biological Engineering Thesis Proposal

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Abstract

In spite of the recent increase in research on the human microbiome, there is not a clear consensus on the relationship between human microbial communities and disease. Microbes colonize our entire bodies, supplementing our body's functions, priming and training our immune systems, providing resistance to colonization by pathogens, and contributing to maintenance of health or progression of disease. However, major knowledge gaps exist in this field. The microbiota of certain body sites have been much less studied than others, and even the most extensively studied body sites lack a synthesized understanding of the clinical relevance of human-microbe associations. Additionally, translating microbial associations into biological hypotheses remains challenging due to the lack of centralized tools and databases for assigning biological meaning to groups of microbes.

This thesis will expand our understanding of the clinical microbiome in three ways. First, I will characterize the relationships between the microbial communities of the aerodigestive tract and their associations with clinical factors, increasing our basic understanding of this under-studied system. Next, I will perform a meta-analysis of published case-control gut microbiome studies across many disease states, synthesizing many existing studies by identifying consistent microbial markers of health and disease. Finally, I will curate groups of biologically related microbes to enable enrichment analyses and generalizable interpretations of results from microbiome studies.

1 Overall objectives and specific aims

1.1 Overall objectives

The research presented in this thesis is united by a common purpose: advancing our understanding of the clinical human microbiome. First, I will enrich our basic understanding of an under-studied microbial system by characterizing the relationships between the microbial communities of the aerodigestive tract and their associations with clinical factors. Second, I will synthesize results across many studies of a well-studied system, moving the field toward a better understanding of and consensus on the associations between gut microbial communities and human disease. To do this, I will perform a meta-analysis of published case-control gut microbiome studies across many disease states to identify consistent microbial markers of health and disease. Finally, I will curate existing knowledge on microbial communities and develop a tool for inferring generalizable biological hypotheses from existing and future microbiome studies. Together, this work will contribute new knowledge to the exciting field of human microbiome research and will empower researchers to draw clinically meaningful insights from their existing and future analyses.

1.2 Specific Aims

Aim 1 Apply standard methods to identify microbial community characteristics associated with gastro-esophageal reflux disease and aspiration.

1. Quantify relationships between lung, gastric, and throat microbial communities.
2. Identify clinical modulators of lung, gastric, and throat microbial communities.

Aim 2 Perform a meta-analysis of gut microbiome studies to identify consistent microbial signatures within and across multiple diseases.

1. Compile and process publicly available case-control gut microbiome studies with a standardized method.
2. Determine whether certain microbes are consistently associated disease in general or with specific diseases.
3. Identify relationships between physiologically-related diseases by comparing their microbial characteristics.

Aim 3 Enable generalizable interpretations of microbiome analyses by assigning bacteria to groups with similar characteristics and known associations with disease.

1. Combine existing databases with targeted literature searches to define *microbe sets* based on known biological relationships.
2. Use machine-learning techniques to extract disease-associated *microbe sets* from datasets collected in Aim 2.
3. Develop these *microbe sets* into a collaborative tool for use in interpreting new microbiome studies.

2 Background and significance

The topics addressed in this thesis are broad, and are all connected by the motivation to better understand clinically-relevant associations between microbes and their human hosts. My work will focus on the microbial communities of two major body systems: the aerodigestive and gastrointestinal tracts. To study these, I will use a combination of traditional analytical techniques, supplemented by novel methods as required. This section will provide background on the aerodigestive tract, the gut microbiome, and current analytical techniques used in microbiome studies.

2.1 Aerodigestive tract

2.1.1 Physiology and disease

- The aerodigestive tract is linked and the microbial exchange across sites is unknown.
- Gastro-esophageal reflux disease (GERD) and aspiration are both complex and prevalent conditions that are thought to be related to respiratory diseases.

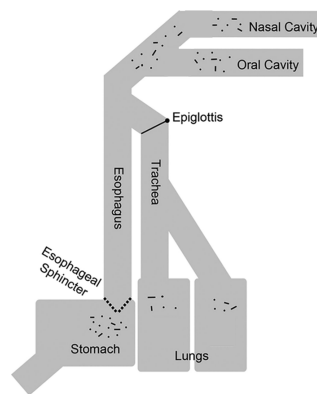


Figure 1: Schematic of the flow relationship between sites of the aerodigestive tract. Adapted from [1].

2.1.2 Microbiome of the aerodigestive tract

- Lung and gastric microbiomes have historically been poorly studied. In the lung, a few diseases have been examined. In the stomach, it's mostly been related to *H. pylori*.
- The relationship between communities across the aerodigestive tract is still up for debate.

2.2 Gut microbiome

2.2.1 Gut microbiome in health and disease

- The gut is important to human health and disease, and gut microbes are important to the gut.

- Lots of diseases have been hypothesized to be associated with the gut microbiome. Here’s one or two sentences about IBD, CRC, and CDI.

2.2.2 Existing understanding of the gut microbiome

- It’s been hard to find consensus on specific disease associations.
- But we do know that the microbiome is stable and modifiable. People generally think that diversity and Bacteroides/Firmicutes ratios matter.
- Existing meta-analyses haven’t found much consensus either and they have focused on one or two diseases only.

2.3 Analytical background and significance

2.3.1 Data generation, analysis and associated challenges

- Turning next generation 16S sequencing data into OTU tables is a process with lots of steps which are not standardized.
- Resulting 16S data is hard to analyze because it is high-dimensional, sparse, and has lots of batch effects.
- People find associations with disease by looking at alpha and beta diversity and doing univariate tests on OTU abundances.

2.3.2 Interpreting taxonomy-based microbiome analyses

- When you do univariate analyses, you get a list of OTUs that are associated with the phenotype. Then it’s up to you, the researcher, to figure out what those OTUs mean.
- Enrichment analysis is a direct way to identify biologically meaningful patterns in data. It’s been used in RNA data a lot (GSEA).
- For enrichment analyses to work, you need to have curated groups of related features (i.e. microbes). This doesn’t really exist for microbes.

3 Research design and methods

The research presented in this thesis is united by a common purpose: advancing our understanding of the clinical human microbiome. First, I will enrich our basic understanding of an under-studied microbial system. Second, I will collect and synthesize results from many studies of a well-studied system, to move the field toward a better understanding of the interactions between microbial communities and human disease. Finally, I will curate existing knowledge on microbial communities to develop a tool for inferring generalizable biological hypotheses from existing and future microbiome studies.

3.1 Aim 1: Aerodigestive microbiota associated with GERD and aspiration

GERD, aspiration, and respiratory infections are three related conditions with complex and unclear interactions. We know that aspirating patients are at a higher risk for respiratory infections, and that many patients who present with idiopathic respiratory problems have a high prevalence of GERD. Furthermore, the microbial communities of the aerodigestive tract are connected and likely exchange bacterial members, which may contribute to respiratory infections. We hypothesize that the microbial communities of the aerodigestive tract are extensively exchanging microbes, and that certain clinical conditions like aspiration or GERD may modulate the amount of exchange happening across various sites.

To address this hypothesis, we will first identify which microbes are exchanged across sites and define a metric to quantify the “extent” of this exchange. To define this metric, we will incorporate both the co-occurrence and the abundance of microbes in the two sites and calculate it for each site-combination. Next, we will identify clinical factors that have an effect on microbial exchange in the aerodigestive tract. Specifically, we will investigate how aspiration, reflux, and PPI use affects the similarity of communities and the exchange of microbes between sites in the aerodigest tract. We hypothesize that aspiration will increase the lung-throat connection and that reflux and PPI use will strengthen the stomach-lung connection. Quantitatively describing the amount of microbial exchange happening in the aerodigestive tract and determining clinical modulators of this exchange will contribute new knowledge to our current understanding of the aerodigestive microbiome, and could inform future aerodigestive investigations and treatments.

3.1.1 Aerodigestive patient cohort

- Our cohort consists of 261 patients recruited by Rachel Rosen at Boston Children’s over the past 6 years.
- We have throat swabs, gastric fluid, and BAL. We also have aspiration and reflux metadata.

Sites	N
gastric, throat, & BAL	87
gastric & throat	45
gastric & BAL	34
BAL & throat	9

Table 1: Samples in study

3.1.2 Quantify exchange of microbes between lung, gastric, and throat communities

- First, we’ll define exchanged microbes and quantify them as the percentage of patients who are exchanging that microbe across their two sites (p_s), (Figure 2).

- It's possible that the gastric-lung exchanged microbes come from the environment, because of the low biomass of these sites. We'll look at them on a tree and in the literature to make sure that's not the case.

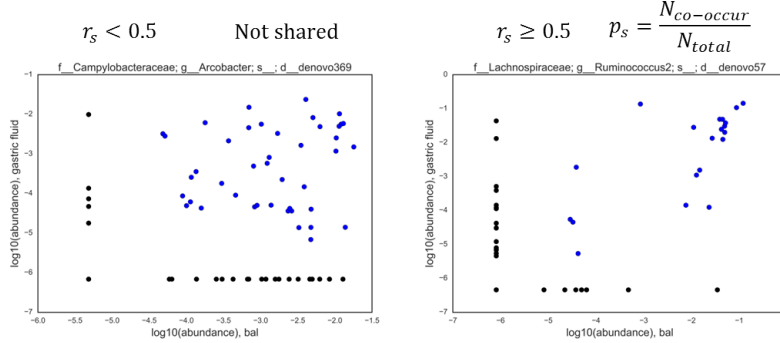


Figure 2: If the abundance of a microbe when it is present in both sites is correlated, then we consider it exchanged across those sites (blue points, right panel). p_s is then calculated as the percentage of patients who have the microbe present in both sites (blue points divided by total points, where each point represents one patient). Example of microbes which are (A) not exchanged and (B) exchanged between stomach and lungs.

3.1.3 Identify clinical modulators of lung, gastric, and throat microbial communities

- We hypothesize that aspiration, reflux, and PPI use will affect exchange across sites. We'll calculate p_s and look at beta diversity for the different groups.
- We hypothesize that aspiration will modulate the amount of throat-lung and gastric-lung exchange, that reflux will modulate the amount of gastric-lung exchange, and that PPI use may also affect the gastric-lung connection.
- Some caveats to this work are that we're not really addressing directionality of the exchange and we don't have metadata on whether patients developed respiratory infections.

3.2 Aim 2: Meta-analysis of gut microbiome studies

By combining results from existing gut microbiome case-control studies, we can move the field toward a consolidated understanding of consistent microbial markers of gut-related diseases. We hypothesize that certain bacteria will often be associated with disease, and that some of these bacteria will be associated with many different types of diseases while others will be unique to one or two conditions. Additionally, we hypothesize that microbial signatures of health and disease will be more similar in similar diseases (i.e. in diabetes and obesity vs. in diabetes and autism).

To address our hypotheses, we will first acquire a comprehensive collection of case-control gut microbiome datasets and process them with standardized methods. We will analyze each dataset individually and synthesize the results from all datasets with basic meta-analysis

techniques to identify microbes consistently associated with health or general disease. We will also perform a similar intra-disease meta-analysis for studies analyzing the same disease to identify microbes that may be specific markers of certain disease and not others. Finally, we will identify relationships between physiologically-related diseases by comparing their microbial characteristics across multiple datasets. This comprehensive pan-disease meta-analysis will consolidate the findings from many existing gut 16S microbiome studies, synthesizing our existing knowledge and generating new hypotheses to inform future mechanistic experiments and case-control analyses.

3.2.1 Compile and process gut microbiome datasets

- We'll collect and process all 16S case-control stool datasets that we can find through a standard pipeline.
- We'll start our analyses by collapsing to genus level, but if that doesn't work we'll also look at higher-order taxonomic levels.

3.2.2 Identify microbial markers of disease

- We'll do univariate analyses on each dataset individually, and combine the results across *all* datasets using a modified Fisher's method for combining p-values.
- Next, we'll do an intra-disease meta-analysis, combining results from datasets looking at the same disease. We hope to find microbes that are associated with a specific disease but not with disease in general, or that have a different directionality of association.
- We might not be able to find anything, probably because of batch effects. Developing robust methods to overcome technical batch effects in 16S studies is not within the scope of this work, but there are many simpler options available to help mitigate severe batch effects.

3.2.3 Compare results between studies for related diseases

- Our next hypothesis is that similar diseases will have similar signatures of dysbiosis. We'll summarize each dataset with a "microbial signature" and see which datasets/diseases cluster together.
- We expect diseases with strong microbiome associations to cluster together. We may also see diseases with similar underlying causes (i.e. inflammation) clustering together (Figure 3).

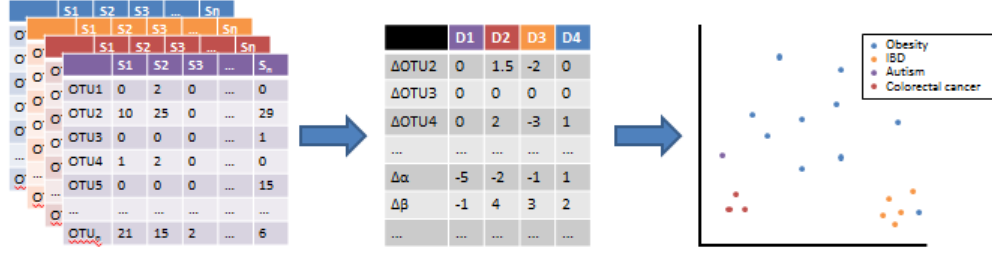


Figure 3: Defining microbial signatures

3.3 Aim 3: Assigning bacteria to groups with similar functions and disease associations

In this aim, we will curate biologically-motivated *microbe sets* to enable easier interpretation of results from 16S microbiome analyses. By defining groups of related microbes, we will enable enrichment analyses like GSEA, but for microbiome data (Section 2.3.2). Enrichment analyses will allow for better biological understanding of results from individual studies as well as more consistent comparisons of results across studies reported in the literature.

To define *microbe sets* that facilitate enrichment analyses, we will begin by searching the literature for existing microbial annotation databases and papers which characterize broad groups of microbes. In parallel, we will also mine the datasets and results from Aim 2 for meaningful microbial associations with human phenotypes such as disease or inflammation (Figure 4). Finally, we will combine and package this information in a format that is easy to use and update as future studies contribute to the field. This tool will enable future microbiome scientists to extract more meaningful information from their microbiome studies, thus contributing significantly to increasing our understanding of the clinical and scientific relevance of the human microbiome.

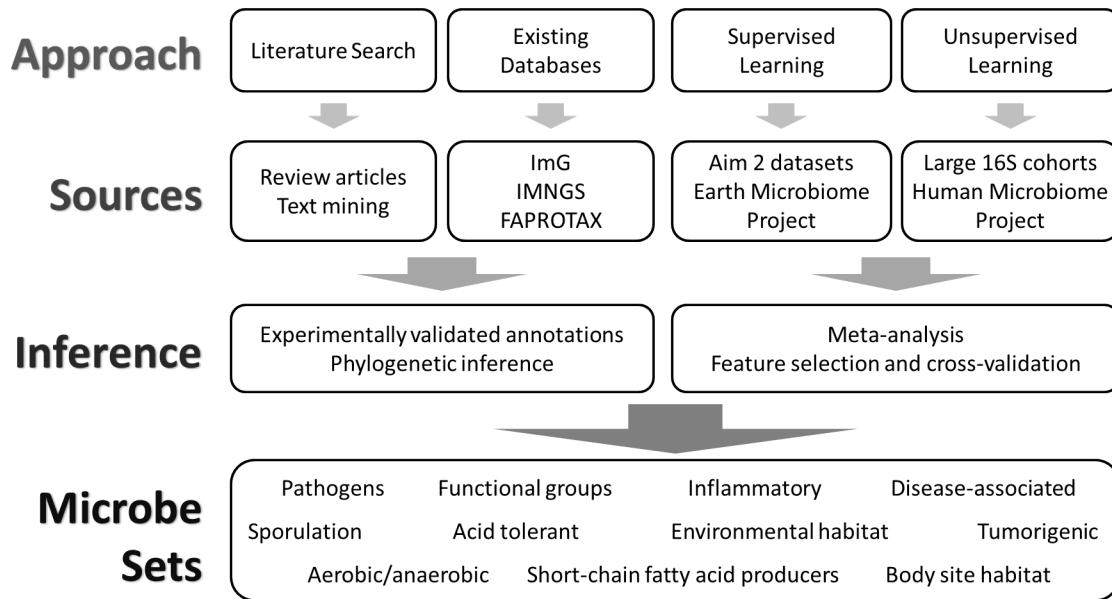


Figure 4: A variety of approaches will be used to define microbe sets, including manual curation from literature and database searches and data-driven methods (“Approach”). Many different kinds of resources will be drawn upon (“Sources”) to infer groups of related microbes (“Inference”) based on different categories (“Microbe Sets”).

3.3.1 Define microbe sets based on known biological relationships

- We’ll start with a literature and database search to see what’s out there in terms of microbial annotations. We’ll start with the ImG database and see what’s in it and what needs to be filled out.
- In collaboration with Ilana, we’ll do a combination of literature mining and bioinformatics inference to fill out our annotations.
- We’ll also build a tree from the 16S sequences of all the bacteria we annotate and maybe do some phylogenetic inference to fill out more annotations.

3.3.2 Extract disease-associated microbe sets from datasets in Aim 2

- We’ll also define some microbe sets based on the results from meta-analyses in Aim 2, and from machine-learning magic on the datasets from Aim 2 (Table 2).
- The machine-learning stuff may not work so we’ll try a few other approaches.

Microbe set association	Classification task
General health/disease	All healthy vs. all disease
Diarrhea	CDI, EDD, IBS-D vs. controls
Neurological	Autism, Parkinson’s vs. controls
Liver	NASH, MHE vs. controls
Metabolic syndrome	T1D, T2D, obesity, metabolic syndrome vs. controls
Autoimmune/inflammatory	T1D, rheumatoid arthritis, psoriatic arthritis, Crohn’s disease vs. controls or non-autoimmune patients

Table 2: Classification tasks to identify groups of phenotype-associated microbes

3.3.3 Develop collaborative tool for interpreting microbiome studies

- We’ll make our annotated microbe sets available to researchers and will build a tool that does the enrichment analyses on their data for them.
- This is going to be hard and it’s okay if it’s not perfect or comprehensive because it’s mainly going to be used as a hypothesis-generation tool.

4 Preliminary studies

4.1 Aim 1

4.1.1 Microbiome community exchange

- We found a lot of exchanged microbes, the majority of which were between the throat and stomach (as expected).
- Interestingly, we also found lots of exchange/similarity between the stomach and lungs. This might have to do with microaspiration of gastric contents.

4.1.2 Modulators of community exchange

- We found that aspirators share more between and have more similar throat and lung communities than non-aspirators.
- Our first pass with reflux showed that there might be more stomach-lung exchange in patients with more frequent full-column reflux. Interestingly, this wasn’t reflected in the “exchanged microbes” but rather in the beta-diversity of stomach and lung communities, indicating that the exchange might be more stochastic here.

4.2 Aim 2

4.2.1 Collecting and reprocessing 16S case-control datasets

- We’ve collected and processed about 30 datasets. These are summarized in a table that I still need to compile (N patients, sequencing depth, sequencing technology).

4.2.2 Identify general patterns of health and diseases

- First we looked at alpha diversity, and saw that it was only consistently different in diarrheal diseases.
- Then we did univariate analyses for each study. Looking at the pattern, we noticed that diarrheal diseases have broad community shifts. The other diseases are less clear.
- We also did the Fisher's method for combining p-values. Results from this are TBD this weekend. I think we'll find a handful of genera that are significantly associated with health or disease when we combine *all* diseases together.

References

- [1] C.M. Bassis, J.R. Erb-Downward, R.P. Dickson, C.M. Freeman, T.M. Schmidt, V.B. Young, J.M. Beck, J.L. Curtis, and G.B. Huffnagle. Analysis of the upper respiratory tract microbiotas as the source of the lung and gastric microbiotas in healthy individuals. *MBio*, 6(2):e00037–15, 2015. doi: 10.1128/mBio.00037-15. URL <http://dx.doi.org/10.1128/mBio.00037-15>.