Total Selenium and Selenium (IV) in the James River Estuary and Southern Chesapeake Bay

Kazufumi Takayanagia and George T. F. Wong

Department of Oceanography, Old Dominion University, Norfolk, Virginia 23508, U.S.A.

Received 12 October 1982 and in revised form 4 May 1983

Keywords: selenium; speciation; estuaries; mixing; Chesapeake Bay

The concentrations of total selenium (Se) and Se (IV) were determined in the surface waters of 30 stations located in the James River and southern Chesapeake Bay. The concentrations of total Se and Se (IV) ranged from 0.28 to 1.91 nM and from 0.07 to 1.36 nM, respectively, between salinities of 31.78 and 0.06%. The concentration of Se (VI), calculated as the difference between the concentrations of total Se and Se (IV), ranged from 0.08 to 0.67 nM. While total Se seemed to be conservative in this study area at salinities above 0.36%, Se (IV) might have been removed during estuarine mixing. The removal of Se (IV) occurred primarily at salinities below 4% possibly via the oxidation of Se (IV) to Se (VI).

Introduction

Riverine input has long been recognized as one of the most important terms in the global geochemical cycle (Boyle et al., 1974; Edmond et al., 1981). Chemical and biological processes which occur during the mixing of river water with seawater may lead to the addition or removal, as well as the alteration, of the speciation of dissolved constituents in river waters (Liss, 1976; Aston, 1978), and thus may determine to a significant extent the riverine supply of elements to the oceans. Elements which can exist in different oxidation states are of particular interest because the true redox potential of natural waters is not precisely known (Breck, 1974) and species in the different oxidation states may behave differently during estuarine mixing (Aston, 1978). Selenium (Se) is one such element. The common oxidation states of selenium are -2, 0, +4 and +6. Sillen (1961) suggested that Se (VI) should be the only detectable species of selenium in oxygenated seawater, since the Se (IV) to Se (VI) ratio at equilibrium should be $10^{-11.5}$, and both Se (-II) and Se (O) should also be undetectable. However, the existence of both Se (IV) and Se (VI) have been reported. The concentration of Se (IV) in seawater ranges from undetectable to 0.7 nM, while the concentration of total Se in seawater ranges from 0.3 to 2.5 nM (Sugimura et al., 1976, Cutter, 1978, Measures & Burton, 1980, Measures et al., 1980; Uchida et al.,

^aPresent address: Fisheries and Oceans Canada, Champlain Centre for Marine Science and Surveys, CP15500, Québec, Canada G1K 7Y7.

1980). The concentrations of total Se and Se (IV) in estuarine waters range from 0.2 to 4.5 nM and from undetectable to 0.3 nM, respectively (Kharkar et al., 1968, Measures & Burton, 1978; Uchida et al., 1980).

Kharkar et al. (1968) studied the interactions between dissolved selenium and particulate phases in the laboratory. They reported that at a selenium concentration similar to that in river water, more than 80% of dissolved selenium could be removed by adsorption onto ferric oxide and manganese dioxide in distilled water. The selenium that was adsorbed onto montmorillonite, kaolinite, manganese dioxide and peat could be desorbed when the solid phases were exposed to seawater. If these processes occur in an estuary, a production of selenium may be observed during estuarine mixing. However, Measures and Burton (1978) reported that both total Se and Se (IV) in the River Test were conservative at salinities between 7 and 34‰. In this paper, we shall discuss the speciation of selenium in the James River and southern Chesapeake Bay.

Materials and methods

Study area

Chesapeake Bay is one of the largest estuaries in the United States. It extends in a north-south direction for about 290 km (Newcombe et al., 1939) and its drainage basin is about 166 200 km² (Pelczar, 1972). It has a number of tributaries. The James River is the third largest river which drains into Chesapeake Bay and it contributes 16% of the annual freshwater input (Pritchard, 1952). It is classified as a coastal plain estuary. The tidal portion of the James River extends for 168 km from its mouth at Hampton Roads to Richmond, Virginia. The volume of water in the tidal portion of the James is approximately $2 \cdot 36 \times 10^9$ m³ (Cronin & Pritchard, 1975), and an average discharge measured at Richmond is approximately 240 m³ s⁻¹ (U.S. Geological Survey, 1973). A rough estimate of the residence time of water excluding tidal flushing would be 114 days. The James River has a number of tributaries. The Chickahominy, Warwick, Nansemond and Elizabeth Rivers are the major ones within the study area. However, they have comparatively low discharge rates (Feuillet & Fleischer, 1980). Thirty sampling stations were occupied in the James River, southern Chesapeake Bay and Atlantic Ocean off Cape Henry (Figure 1). Station 1 is approximately 50 km downstream from Richmond.

Sampling and analytical methods

Samples were collected during two cruises: on 8 December 1981 for stns 1 through 23, and on 4 February 1982 for stns 24 through 30. At each station, surface water was collected with a polyethylene bucket and immediately filtered through Gelman A/E glass-fibre filters. The samples were stored in polyethylene bottles, kept in ice, and returned to the laboratory on the same day. In the laboratory, they were stored at about 4 °C and analysed within a week after the collection (Cheam & Agemian, 1980). The samples were analysed for total Se and Se (IV) as described elsewhere (Takayanagi & Wong, 1983). Briefly, total Se was preconcentrated by coprecipitation with tellurium. Se (IV) was preconcentrated by complexing it with ammonium 1-pyrrolidinedithiocarbamate, extracting the complex into chloroform and back-extracting the selenium into nitric acid. The preconcentrated selenium was determined fluorometrically. In this analytical scheme, any chloroform-extractable organic selenium will also be included as Se (IV), while Se (-II) and Se (O) may be included as total Se although the contributions from these species are expected

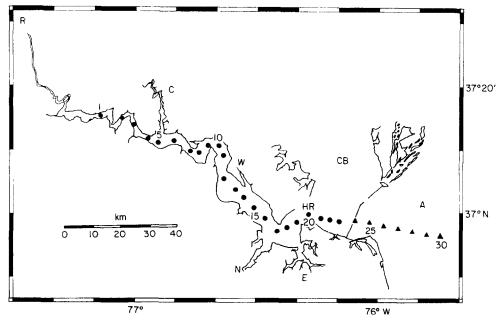


Figure 1. Sampling stations in December (•) and February (•). A: Atlantic Ocean; C: Chickahominy River; CB: Chesapeake Bay; E: Elizabeth River; HR: Hampton Roads; N: Nansemond River; R: Richmond; W: Warwick River.

to be small (Cutter, 1982; Takayanagi, 1982). The salinities were determined with a Guildline Instruments Model 8400 salinometer.

Results and discussion

The concentration of total Se decreased with increasing salinity, ranging from 1.91 nM at 0.06% to 0.28 nM at 31.78% (Figure 2). The concentrations at high salinities are similar to those observed in the surface waters of the Atlantic Ocean (Measures & Burton, 1980). Total Se correlates linearly with salinity with a correlation coefficient of 0.98 between 31 · 78 and 0 · 36‰, implying that total Se is conservative within this range of salinities. The concentration of total Se at 0.06% (1.91 nM), lies significantly above this theoretical dilution line. Since only one data point is involved, the interpretation is somewhat tentative. This sample has been analysed repeatedly and the same result was obtained. We have no reason to suspect that the composition had been modified during sampling or storage. There are two possible reasons for this anomaly. Total Se may have been removed in the waters at salinities below 0.36%. Iron and humic substances have been reported to be removed by flocculation at low salinities (<1\%) (Sholkovitz et al., 1978; Sholkovitz & Copland, 1981). Since some of the dissolved selenium may exist in the colloidal form in natural waters (Sigleo & Helz, 1981; Takayanagi & Wong, 1982), flocculation induced by salinity changes may remove selenium as well. Alternatively, the elevated concentration at 0.06% may be caused by the natural variability of the concentration of total Se in the riverine endmember. Anomalously high concentrations of dissolved organic carbon and ammonia have also been observed at this station (unpublished results).

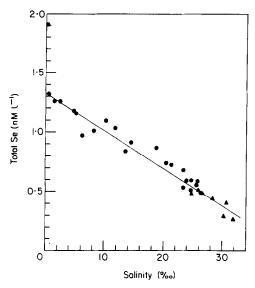


Figure 2. Total Se against salinity in the waters collected in December (\bullet) and February (\triangle). Uncertainty of an individual point is ± 0.02 nM. The linear regression line for the data points with salinities above 0.06% is also shown.

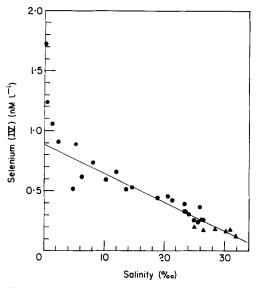


Figure 3. Se (IV) against salinity in the waters collected in December (\bullet) and February (\triangle). Uncertainty of an individual point is ± 0.02 nM. The linear regression line for the data points with salinities above 4% is also shown.

The concentration of Se (IV) also decreased with increasing salinity with a range of $1 \cdot 72$ - $0 \cdot 12$ nM (Figure 3). The concentration of Se (IV) correlates linearly with salinity at salinities above 4‰ with a correlation coefficient of $0 \cdot 93$. Below this salinity, all the data points lie above this dilution line. This suggests that Se (IV) is non-conservative during estuarine mixing and may be removed primarily in the waters at salinities below 4‰. Since only one sample at the riverine endmember was analysed and temporal variations in the concentration of Se (IV) might have occurred, the removal of Se (IV) cannot be

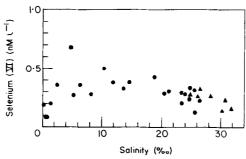


Figure 4. Se (VI) against salinity in the waters collected in December (\bullet) and February (\triangle). Uncertainty of an individual point is ± 0.04 nM.

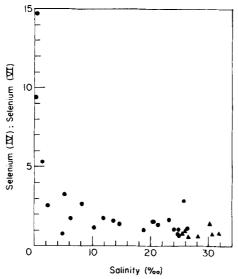


Figure 5. The concentration ratio of Se (IV) to Se (VI) against salinity in the waters collected in December (\bullet) and February (\triangle) .

quantified accurately. However, if the concentration of Se (IV) of 1.72 nM at a salinity of 0.06% does represent the composition of the riverine endmember, the amount of Se (IV) removed in this estuary may be estimated by the method of Boyle et al. (1974). Since the intercept of the dilution line is 0.88 nM, 0.84 nM of Se (IV), or about 50% of the riverine input, has been removed in the waters at salinities less than 4‰. There are several possible mechanisms for the removal of Se (IV) during estuarine mixing: conversion to a particulate form by processes such as biological uptake and adsorption onto suspended particles; or conversion to another dissolved form via processes such as the oxidation to Se (VI). Although the specific biochemical role of selenium is not well understood, it is recognized as an essential trace element for the healthy growth of organisms (Frost & Lish, 1975, Thomson & Robinson, 1980; Mertz, 1981). Accumulation of selenium by zooplankton and marine invertebrates have been reported (Fowler & Benayoun, 1976a, b; Wrench, 1978). Wrench (1978) reported that marine phytoplankton, Tetroselmis tetrathele and Dunaliella minuta, were capable of taking up Se (IV) in culture media. He also suggested that selenium was assimilated as seleno-analogues of the sulphur amino acids. Sandholm et al. (1973) however, reported that the uptake of Se (IV) by green algae, Scenedesmus

dimorphus, was negligible in 60 min. If Se (IV) is actively consumed by organisms, biological uptake may be one of the possible removal mechanisms during estuarine mixing. Since Se (IV) is known to be easily adsorbed onto ferric hydroxide (Geering et al., 1968), a major carrier phase in estuaries, adsorption onto suspended particles is also a plausible removal mechanism. However, since total dissolved Se is approximately conservative between 31.78 and 0.36%, the conversion of Se (IV) to a particulate form is unlikely, at least within this range of salinities.

The concentration of Se (VI) may be estimated as the difference between the concentrations of total Se and Se (IV). Since total Se is conservative while Se (IV) is removed in the waters at salinities above 0.36%, this model will indicate that Se (VI) is produced during estuarine mixing by the oxidation of Se (IV) to Se (VI). Indeed, most data points lie above the theoretical dilution line connecting the riverine and oceanic endmembers (Figure 4). Rosenfeld & Beath (1964) reported that the rate of transformation between Se (IV) and Se (VI) is slow in pure inorganic systems. However, in the natural water system, organisms may be able to increase the rates of oxidative reactions significantly. Morris et al. (1978) suggested that in the low salinity regions of the Tamar Estuary, oxygen utilizing bacteria could be responsible for the oxidation of organic matter and other reduced species such as Fe (II) and Mn (II). Emerson et al. (1982) also observed the bacterial-catalysed oxidation of Mn (II) in the waters of Saanich Inlet.

Se (IV) is the predominant species of selenium in river waters. With increasing salinity, the concentration ratio of Se (IV) to Se (VI) decreases from 15 at 0.36% to around 0.9 at 31.78% (Figure 5). The decrease was most dramatic at salinities below 10%. At salinities above 10%, the ratio was rather constant. However, even at the oceanic endmember, the ratio is still much higher than the value predicted by thermodynamic equilibrium, and it is only slightly higher than the values reported for open ocean seawaters, which range from 0 to 0.7 (Measures & Burton, 1980; Measures et al.; 1980).

Conclusions

Se (IV), a thermodynamically unstable species, was the dominant dissolved inorganic species of selenium in the James River. Total Se was conservative at salinities above 0.36%. However, a significant amount of Se (IV) was removed during estuarine mixing at salinities below 4% possibly as a result of the oxidation of Se (IV) to Se (VI).

Acknowledgements

We thank the crew members of the Old Dominion University research vessel *Linwood Holton* for assisting us sampling the waters and one of us (K.T.) also thanks Old Dominion University for a student research grant.

References

Aston, S. R. 1978 Estuarine chemistry. In Chemical Oceanography, Vol. 7, 2nd Edn (Riley, J. P. & Chester, R., Eds). Academic Press, London, pp. 361-440.

Boyle, E., Collier, R., Dengler, A. T., Edmond, J. M., Ng, A. C. & Stallard, R. F. 1974 On the chemical mass balance in estuaries. Geochimica et Cosmochimica Acta 38, 1719-1728.

Breck, W. G. 1974 Redox levels in the sea. In The Sea, Vol. 5 (Goldberg, E. D., Ed.). John Wiley, New York, pp. 153-179.

Cheam, V. & Agemian, H. 1980 Preservation and stability of inorganic selenium compounds at ppb levels in water samples. Analytica Chimica Acta 113, 237-245.

Cronin, W. B. & Pritchard, D. W. 1975 Additional Statistics on the Dimensions of the Chesapeake Bay and its

Tributaries: Cross-Section Width and Segment Volumes per Meter Depth. Chesapeake Bay Institute Special Report 42, 475 pp. Baltimore, Maryland.

Cutter, G. A. 1978 Species determination of selenium in natural waters. Analytica Chimica Acta 98, 59-66.

Cutter, G. A. 1982 Selenium in reducing waters. Science 217, 829-831.

Edmond, J. M., Boyle, E. A., Grant, B. & Stallard, R. F. 1981 The chemical mass balance in the Amazon plume I: The nutrients. Deep-Sea Research 28, 1339-1374

Emerson, S., Kalhorn, S., Jacobs, L., Tebo, B. M., Nealson, K. H. & Rosson, R. A. 1982 Environmental oxidation rate of manganese (II): bacterial catalysis. *Geochimica et Cosmochimica Acta* 46, 1073-1079.

Feuillet, J. & Fleischer, P. 1980 Estuarine circulation: controlling factor of clay mineral distribution in the James River Estuary, Virginia. *Journal of Sedimentary Petrology* 50, 267-279.

Fowler, S. W. & Benayoun, G. 1976a Accumulation and distribution of selenium in mussel and shrimp tissues.

Bulletin of Environmental Contamination and Toxicology 16, 339-346.

Fowler, S. W. & Benayoun, G. 1976b Selenium kinetics in marine zooplankton. Marine Science Communications 2, 43-67.

Frost, D. V. & Lish, P. M. 1975 Selenium in Biology. Annual Review of Pharmacology 15, 259-284.

Geering, H. R., Cary, E. E., Jones, L. H. P. & Allaway, W. H. 1968 Solubility and redox criteria for the possible forms of selenium in soils. Soil Science Society of America Proceedings 32, 35-40.

Kharkar, D. P., Turekian, K. K. & Bertine, K. K. 1968 Stream supply of dissolved silver, molybdenum, antimony, selenium, chromium, cobalt, rubidium and cesium to the oceans. *Geochimica et Cosmochimica Acta* 32, 285-298.

Liss, P. S. 1976 Conservative and nonconservative behaviour of dissolved constituents during estuarine mixing. In *Estuarine Chemistry* (Burton, J. D. & Liss, P. S., Eds). Academic Press, London, pp. 93-130.

Measures, C. I. & Burton, J. D. 1978 Behaviour and speciation of dissolved selenium in estuarine waters. *Nature* 273, 293-295.

Measures, C. I. & Burton, J. D. 1980 The vertical distribution and oxidation states of dissolved selenium in the northeast Atlantic Ocean and their relationship to biological processes. *Earth and Planetary Science Letters* 46, 385-396.

Measures, C. I., McDuff, R. E. & Edmond, J. M. 1980 Selenium redox chemistry at GEOSECS I re-occupation. Earth and Planetary Science Letters 49, 102-108.

Mertz, W. 1981 The essential trace elements. Science 213, 1332-1338.

Morris, A. W., Mantoura, R. F. C., Bale, A. J. & Howland, R. J. M. 1978 Very low salinity regions of estuaries: important sites for chemical and biological reactions. *Nature* 274, 678-680.

Newcombe, C. L., Horne, W. A. & Shepherd, B. B. 1939 Studies on the physics and chemistry of estuarine waters in Chesapeake Bay. *Journal of Marine Research* 2, 87-116.

Pelczar Jr, J. M. 1972 The current status of the Chesapeake Bay. Opening remarks. Journal of the Washington Academy of Sciences 62, 54-55.

Pritchard, D. W. 1952 Salinity distribution and circulation in the Chesapeake Bay estuarine system. Journal of Marine Research 11, 106-123.

Rosenfeld, I. & Beath, D. 1964 Selemum. Academic Press, New York, 411 pp.

Sandholm, M., Oksanen, H. E. & Pesonen, L. 1973 Uptake of selenium by aquatic organisms. Limnology and Oceanography 18, 496-499.

Sholkovitz, E. K., Boyle, E. R. & Price, N. B. 1978 Removal of dissolved humic acid and iron during estuarine mixing. Earth and Planetary Science Letters 40, 130-136.

Sholkovitz, E. R. & Copiand, D. 1981 The coagulation, solubility and adsorption properties of Fe, Mn, Cu, Ni, Cd, Co and humic acids in a river water. Geochimica et Cosmochimica Acta 45, 181-189.

Sigleo, A. C. & Helz, G. R. 1981 Composition of estuarine colloidal material: major and trace metals. *Geochimica et Cosmochimica Acta* 45, 2501-2509.

Sillen, L. G. 1961 The physical chemistry of seawater. In Oceanography (Sears, M., Ed.). American Association of the Advancement of Science, Washington, D.C., pp. 549-581.

Sugimura, Y., Suzuki, Y. & Miyake, Y. 1976 The content of selenium and its chemical form in seawater. Journal of the Oceanographical Society of Japan 32, 235-241.

Takayanagi, K. 1982 The marine geochemistry of selenium. Ph.D. dissertation, Old Dominion University, 121 pp.

Takayanagi, K. & Wong, G. T. F. 1982 Colloidal and organic selenium in estuarine and river waters. Transaction, American Geophysical Union (EOS) 63, 977.

Takayanagi, K. & Wong, G. T. F. 1983 Fluorimetric determination of selenium (IV) and total selenium in natural waters. *Analytica Chimica Acta* 148, 263–269.

Thomson, C. D. & Robinson, M. F. 1980 Selenium in human health and disease with emphasis on those aspects peculiar to New Zealand. *American Journal of Clinical Nutrition* 33, 303-323.

Uchida, H., Shimoishi, Y. & Toei, K. 1980 Gas chromatographic determination of selenium (-II, 0), -(IV) and -(VI) in natural waters. *Environmental Science and Technology* 14, 541-544.

U.S. Geological Survey 1973 Surface Water Supply of Virginia. James River Bulletin No. 79.

Wrench, J. J. 1978 Selenium metabolism in the marine phytoplankters Tetraselmis tetrathele and Dunaliella minua. Marine Biology 49, 231-236.