



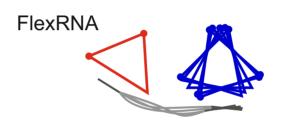
# **MMMx: RigiFlex modelling**

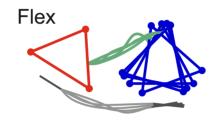
G. Jeschke

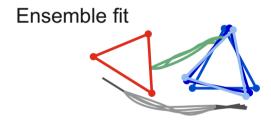
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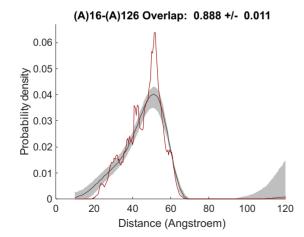


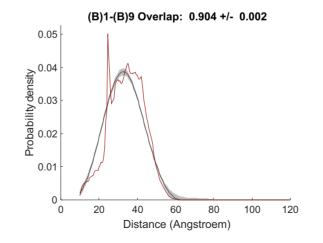


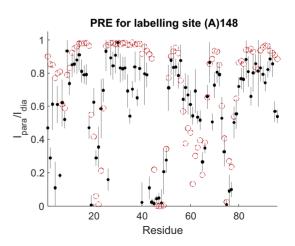












## Modelling SRSF1 in complex with UCAUUGGAU

#### **Steps**

- run Rigi module with hierarchical clustering to generate an ensemble of rigid-body arrangements
- run FlexRNA to generate a two-nucleotide linker UU between binding motifs CA and GGA
- run FlexRNA to generate single 5'-terminal nucleotide U
- run FlexRNA to generate single 3'-terminal nucleotide U
- run Flex to generate peptide linker from residue 90 to 120
- refine all conformers by Yasara

#### At this point, we have generated a raw ensemble by RigiFlex

• run EnsembleFit with the raw ensemble, DEER distance distribution restraints, and PRE restraints

#### **Dependences on third-party software**

- SCWRL4 for generating amino acid side groups in Flex
- Yasara for refining conformers (proprietary software)
- refinement by GROMACS (free) is possible, but requires manual force-field choice

## Making rigid-body arrangements

```
!rigi % call the Rigi module
 rbtemplate BSRS.pdb % load the rigid-body template
 separate on % separate the rigid bodies from each other for spin labelling
 maxtrials 10000 % make 10000 trials within the distance distributions
 models 200 % generate up to 200 models
 save SRSF1 rba % save in MMMx internal rigid-body arrangement format with file name SRSF1 rba.mat
 rigid (A) (B) % define the first rigid body by chains A and B of the template
    (A) 16 mtsl % first reference point of first rigid body, labelled by MTSL
   (A) 37 mtsl % second reference point of first rigid body, labelled by MTSL
   (A) 72 mtsl % third reference point of first rigid body, labelled by MTSL
  .rigid % close block kev
 rigid (C) (D) % define the first rigid body by chains C and D of the template
    (D) 126 mtsl % first reference point of second rigid body, labelled by MTSL
    (D) 148 mtsl % second reference point of second rigid body, labelled by MTSL
    (D) 169 mtsl % third reference point of second rigid body, labelled by MTSL
  .rigid % close block key
 ddr mtsl % define 7 core restraints between reference points in different rigid bodies
   (A) 16 (D) 148
                     32.5 06.0 % Gaussian restraint
   ... % there are six more lines as the previous one
  .ddr % close block key
```

## **Further specifications for rigid-body arrangements**

```
plink % specify the length of a peptide linker
    (A)89 (D)121 32 % 32 residues anchored at sites (A)89 and (D)121, <=3.8 Å/residue
    .plink

nlink % specify the length of a nucleic acid linker
    (B)3 (C)6 3 16 % 3 nucleotides anchored at sites (B)3 and (C)6, maximum length 16 Å
    .nlink

superimpose 2 % superposition is onto rigid body 2
.rigi % close module
```

- Laura Esteban Hofer was running 50000 trials on Euler and she requested 6000 models
- when running Flex on Euler with a fresh MMMx installation, specify the path to SCWRL4 in the Flex block by scwrl4 pathname

(this is generally necessary on Linux or Mac, Matlab fails to find the correct path)

#### **Completing the RNA**

```
!flexrna 0.75 1 0.016667 % 75% coverage of distributions, 1 model, maximum of 1 min per conformer
  expand SRSF1 rba % the input conformers are generated by expanding rigid-body arrangements
  sequence 4 5 UU % add nucleotides number 4 and 5 with sequence UU
  save SRSF1 short UU all % save the models with basis file name 'SRSF1 short UU all'
  anchor 5p (B) 3 % the 5'-terminal anchor nucleotide is nucleotide 3 in chain B
  anchor 3p (C)6 % the 3'-terminal anchor nucleotide is nucleotide 6 in chain C
.flexrna % close module
!flexrna 0.75 1 0.025 % 75% coverage of distributions, 1 model, maximum of 1.5 min per conformer
  addpdb SRSF1 short UU all*.pdb % load conformers generated by the previous module call
  sequence 1 1 U % segment is a single nucleotide U with residue number 1
  save SRSF1 short U1 % save the models with basis file name 'SRSF1 short U1'
  anchor 3p (B) 2 % the 3'-terminal anchor nucleotide is nucleotide 2 in chain C
  ddr dota-gd r5p % specify distance distribution restraints, dota-gd label on protein, r5p on RNA
    (A) 16 (B) 1 34.9 04.1 % Gaussian restraint to RRM1
    (D)148 (B)1 26.5 08.5 % Gaussian restraint to RRM2
  .ddr % close block key
.flexrna % close module
% there is one more FlexRNA block for adding the 3'-terminal nucleotide
```

## Adding the peptide linker

```
!flex 0.75 1 0.25 % 75% coverage of distributions, 1 model, maximum of 15 min per conformer
 addpdb SRSF1 short U9*.pdb % load conformers from previous section
 sequence 90 120 RSGRGTGRGGGGGGGGAPRGRYGPPSRRSE % specify residue numbers and sequence
 n anchor (A)89 % the N-terminal anchor is residue 89 in chain A
 c anchor (D)121 % the C-terminal anchor is residue 121 in chain D
 save SRSF1 short RNA SRSF % save conformers with basis file name 'SRSF1 short RNA'
 ddr mtsl % specify peptide-peptide distance distribution restraints (MTSL label pairs)
   (A) 16 107 29.3 08.7 @deer\C16 A107C short med distr.dat % site 107 is newly generated
   107 (D) 148 25.0 07.3 @deer\A107C C148 short med distr.dat % full distribution is specified
   (A) 37 107 39.7 08.0 @deer\Y37C A107C short med distr.dat
  .ddr % close block key
 ddr dota-gd r5p % specify peptide-RNA distance distribution restraints between dota-gd and r5p
   107 (B) 1 27.2 10.8 @deer\A107C U1 short med distr.dat
  .ddr
 ddr dota-gd r3p % specify peptide-RNA distance distribution restraints between dota-gd and r3p
   107 (B) 9 31.7 11.8 @deer\A107C U9 short med distr.dat
 .ddr
.flex % close module
```

### **Refining conformer models**

```
!yasara 1 % allow for up to 1 hour for refinement
   addpdb SRSF1_short_RNA_i*_m1.pdb % process all output conformers from the previous section
   save SRSF1_short_refined % save output with basis file name 'SRSF1_short_refined'
.yasara % close module

# report % open log file in editor
```

- Yasara cannot be stopped by MMMx
- if Yasara runs take longer than the specified maximum time, more and more Yasara instances are generated
- this can severely slow down a desktop computer
- avoid the # report statement if you run on a server (Euler), use console mode

# Fitting populations and contracting the ensemble

```
!ensemblefit
  addpdb SRSF1 short refined m*.pdb % use all refined conformers in the raw ensemble
  interactive % display figure that visualizes fit progress
  plot % plot figures on fit quality
  csv % save fits in comma-separated value files
  save SRSF1 UCAUUGGAU ensemble fit.ens % output name for MMMx ensemble list
  ddr mtsl % specify distance distribution restraints protein to protein sites
       (A) 16 (A) 148 32.5 06.0 @deer\C16 C148 short med distr.dat
       ... % more lines as the previous one
  .ddr
  ddr dota-gd r5p % specify distance distribution restraints protein site to 5'-terminus of RNA
      (A) 107 (B) 1 27.2 10.8 @deer\A107C U1 short med distr.dat
      ... % more such lines
   .ddr
  ddr dota-gd r3p % specify distance distribution restraints protein site to 3'-terminus of RNA
      (A) 107 (B) 9
                         31.7 11.8 @deer\A107C U9 short med distr.dat
      ... % more such lines
 .ddr
 ddr r5p r3p % specify distance distribution restraint between RNA termini
      (B) 1
           (B) 9 32.6 10.5 \frac{10.5}{10.5} \frac{10.5}{10.5}
 .ddr
```

# **Specifying PRE restraints**

```
% PRE ratio data C16
               site larmor td R2dia taui taur maxrate
  % pre label
        mtsl (A) 16 600.13 12.812 49.66 0.50 11.15 170
  pre
          (A) 18 0.941 0.280 % proton site, PRE ratio, standard deviation
          (A) 19 0.529 0.134 % proton site, PRE ratio, standard deviation
          . . .
  .pre % close PRE block
  % PRE ratio data C148
  % pre label site Larmor td R2dia taui taur maxrate
        mtsl (A) 148 600.13 12.812 49.35 0.50 11.15 170
  pre
          (A) 17 0.470 0.057 % proton site, PRE ratio, standard deviation
          (A) 19 0.289 0.064 % proton site, PRE ratio, standard deviation
  .pre % close PRE block
.ensemblefit % close module
```

• the PRE block specifies the label type, spin label site, proton Larmor frequency in MHz, total INEPT delay td in ms, relaxation rate R2dia for the diamagnetic sample in s<sup>-1</sup>, correlation time taui of internal label motion in ns, rotational correlation time taur of the protein in ns, and a maximum relaxation rate enhancement maxrate in s<sup>-1</sup>