

CEST exchange rate fitting agreement in non-steady state multi-B₁ multi-concentration buffered solutions

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INTRODUCTION:

Accurate determination of analyte exchange rate is necessary for quantitative CEST (qCEST) and optimizing saturation schemes for sensitivity and specificity, as determined experimentally in VDMP^[1]. Existing fitting methods largely depend on steady state (SS) solutions of the Bloch equations for multiple B₁, which do not disentangle CEST proton pool size from transverse dephasing through exchange or relaxation^[1]. Non-SS (NSS) measurements address some of these challenges by constraining the CEST effect by exchange rate, but NSS solutions rely on assumptions that do not translate well to fast-exchanging analytes such as in APT or hydroxylCEST^[2]. Here, we present experimental data on the agreement between exchange rate fitting of ammonium chloride (NH₄Cl) for SS and NSS experiments with and without potassium hydrogen phthalate (KHP) buffer, using single- and multi-concentration fitting at multiple B₁.

METHODS:

NH₄Cl solutions (5mM to 1M) were prepared through serial dilution in either 275mM KHP or deionized water, both doped with 200μM Gadavist to reduce T₁ for shorter experimental protocols. Samples were loaded into 750μL culture tubes and sealed with Teflon tape. Imaging was performed on a 300MHz Bruker Ascend vertical bore at 0.47 mm in-plane resolution, 1 mm slice thickness. Temperature was maintained within ±0.1°C using a water bath and fibre optic temperature probe. An inversion recovery T₁ map, WASSR B₀ map, and a multi-flip angle (FA) high FA B₁ map were acquired for Z-spectrum correction. Saturation durations were 200, 585, 1710, or 5000 ms, with saturation B₁ at 0.3, 0.6, or 1.2μT, covering NSS to SS saturation conditions for TR = 5100 ms. The Z-spectrum was sampled at 0.1 ppm intervals across -4 to 4 ppm. ROIs were drawn conservatively to reduce partial volume effects in Aedes, and ROI-mean spectra were fit in MATLAB 2023a using either a NSS discrete time step solver or a SS fit with single- or multi-concentration fits. Initial fit parameters were closest order of magnitude from literature values to mitigate stalling. Markov chain Monte Carlo (MCMC) sampling of the fitting space was used to ensure a minimum was found.

RESULTS:

Fitting Method\Parameter	R _{ex} (Hz)	T ₂ (ms)
Buffered NSS MC ST Fitting	175 ± 9	13 ± 1
Buffered NSS ST Fitting	190 ± 60	16 ± 4
Buffered SS MC ST Fitting	178 ± 7	15 ± 2
Buffered SS ST Fitting	192 ± 33	15 ± 5
Unbuffered MC NSS ST Fitting	129 ± 11	11 ± 1
Unbuffered NSS ST Fitting	53 ± 4	43 ± 4
Unbuffered SS ST Fitting	52 ± 3	32 ± 2
Lit. Unbuffered SS ST Fitting [3]	53 ± 5	13 ± 1
Lit. Buffered SS ST Fitting [4]	160 ± 7	40 ± 2

Table 1. Comparison of Z-spectrum fitting parameters for exchange rate, R_{ex}, and T₂ of the solute pool C1 (NH₄Cl) by experiment type and fitting method. MC indicates a multi-concentration fit, SS indicates a steady state fit on SS data, and NSS indicates a NSS fit across all saturation durations. Literature values for both citrate-buffered and unbuffered NH₄Cl solutions are provided for comparison

DISCUSSION:

The presence and identity of buffer affected R_{ex} most. To quantify solute R_{ex} for clinical translation of qCEST, buffers that accurately emulate physiological conditions will be necessary. Multi-concentration fitting substantially reduced fitting uncertainty, providing greater precision in R_{ex} determination. SS and NSS experiments and corresponding fitting show agreement, with SS fitting of SS experimental data providing higher precision. Buffered MC SS and unbuffered MC SS fitted R_{ex} agree with their respective literature values.

CONCLUSION:

Accurate determination of CEST R_{ex} was most precise when using an MCMC MC SS fitting. Future characterization of R_{ex} for CEST solute quantification and clinical translation of qCEST will require determination of buffers that accurately emulate physiological conditions in the compartments in which clinical CEST measurements will be made.

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