

Real-time detection of gene promoter activity: quantitation of toxin gene transcription

Kandiah Jeyaseelan,[a](#) Donghui Ma, and Arunmozhiarasi Armugam

Department of Biochemistry, Faculty of Medicine, National University of Singapore, 10 Medical Drive, 119260, Singapore

Received 2001 Feb 19; Revised 2001 Apr 27; Accepted 2001 Apr 27.

Copyright © 2001 Oxford University Press

This article has been [cited by](#) other articles in PMC.

ABSTRACT

[Go to:](#) ☐ [Go to:](#) ☐

We have developed a new method for quantification of promoter activity in cell lines transfected with recombinant plasmids containing the reporter gene encoding chloramphenicol acetyl transferase (CAT) by real-time PCR. As the efficiency of transfection has a direct influence on the total mRNA produced, we have used the neomycin-resistance gene present within the same vector DNA to normalize the measurement of mRNA levels. Three promoters from genes encoding toxins (pre-synaptic neurotoxin phospholipase A₂, post-synaptic α neurotoxin and cardiotoxin), believed to have evolved from the same ancestor but exhibiting different promoter activities, have been employed in this study to demonstrate the feasibility and accuracy of the method in CAT gene reporter analysis.

INTRODUCTION

[Go to:](#) ☐ [Go to:](#) ☐

Methods available for investigating promoter activity usually involve linking the promoter from a gene of interest to the coding sequence of an unrelated reporter gene. The reporter gene usually codes for a specific enzyme, which is absent in the host and can easily be detected. To date, a number of reporter gene systems have been established for measuring the gene expression. These include the genes encoding chloramphenicol acetyltransferase (CAT; [1](#)), firefly luciferase ([2](#)) and β -galactosidase ([3](#)). In the CAT assay, the CAT enzyme activity is measured from the expressed proteins, using ¹⁴C-labeled chloramphenicol as the substrate. In the luciferase assay, a bioluminescent reaction catalyzed by luciferase is used and in the β -galactosidase assays, the β -galactosidase activity is measured colorimetrically. Although the method of measuring the CAT enzyme activity can be correlated to gene expression, it does suffer from disadvantages such as the use of radioisotopes, reduced sensitivity of the assay and extensive procedures such as thin layer chromatography, autoradiography, etc. These will undoubtedly affect the accuracy of measurements of the reporter construct. Furthermore, when dealing with an inducible system, the duration of protein turnover must also be taken into account ([4](#)). Thus, measurement of mRNA levels will be the most direct method for investigating the gene activity. Techniques such as northern hybridization, S1 nuclease and RNase I protection assays can be used for such purposes. However, all these procedures are again considerably laborious and are not always useful for simultaneous analysis of many different gene constructs. Reverse transcription–polymerase

chain reaction (RT–PCR) has been widely used for the detection of the mRNA molecules and offers a possible alternative method to quantifying gene expression level. The quantification of PCR products mainly relies on the phase of the reaction. Various laboratories have developed competitive PCR assays. However, competitive PCR requires accurate quantification of target and competitor amplicons at the end of the reaction and this usually entails laborious post-PCR processing steps. Recently, the ABI PRISM™ 7700 Sequence Detection System (SDS) coupled with TaqMan® chemistry has been shown to provide a rapid, sensitive method for quantifying nucleic acids (5–7). In this assay, reactions are characterized on a real-time basis at the onset of PCR, using gene-specific fluorogenic probes which are subjected to 5′ nucleotidase activity of the DNA polymerase. The higher the copy number of the mRNAs or cDNAs, the sooner a significant increase in fluorescence is observed which is detected as a C_T (threshold cycle) value. The system can support multiplex PCR and, hence, more than one species of mRNA/DNA per sample can be measured simultaneously. In this communication, we report the use of the real-time PCR in the quantification of promoter activity of three related toxin genes, using CAT mRNAs as the target.

MATERIALS AND METHODS

Go to: ☐ Go to: ☐

Cell culture and DNA transfection

The Chinese hamster ovary (CHO) cell line was maintained in α-MEM (8) medium supplemented with 10% fetal bovine serum, 50 U penicillin and 50 µg/ml streptomycin. The day before transfection, CHO cells were subcultured, and trypsinized cells at a density of 3.0×10^5 cells per well were inoculated into a six-well plate. Transfections were carried out with 6 µg of reporter plasmids by calcium phosphate method (9). After 24 h, transfection medium was replaced with fresh α-MEM and incubated at 37°C for a further 24 h before harvesting cells for CAT assays or RNA isolation. To normalize the transfection efficiencies, 6 µg of pSV-β-Gal, a plasmid expressing β-galactosidase (Promega, WI, USA) was co-transfected with the test plasmids in each experiment.

For CAT assays, cytoplasmic extracts of cells were prepared by the freeze–thaw method and the enzyme activity was determined by using a Beckman liquid scintillation counter as described by Seed and Sheen (10). All results were normalized by using pSV-β-gal as an internal control and are the means of determinations ± SD from six individual experiments.

Plasmid construction

Plasmid pMAMneoCAT (Clontech, CA, USA) was digested with *Nhe*I and *Nde*I to remove the mouse mammary tumor virus (MMTV) promoter. Toxin gene promoters with different promotional strength available in the laboratory were first amplified by PCR, and then subcloned into promoterless CAT reporter gene vector pMAMneoCAT plasmid in normal and reverse direction. The sequences of the constructs have also been verified by DNA sequencing (Automated DNA sequencer, Applied Biosystems, CA, USA).

In vitro transcription and translation

In vitro transcription was carried out by using RiboMAX Kit (Promega). The reactions were carried out according to instructions from the manufacturer.

RNA isolation and real-time PCR

Total RNA was isolated by a single step method using TRIZOL reagent (Life Technologies, CA, USA) from CHO cells 48 h after transfection. Before real-time PCR, the RNA samples were further treated by RNase-free DNase I at 37°C for 30 min.

TaqMan probes and primers

SDS primers and TaqMan probes for the CAT gene and the Neomycin gene have been designed using PrimerExpress 1.0 software (Perkin Elmer, MA, USA). The SDS forward and reverse primers for the Neo gene were 5'-CTCCTGCCGAGAAAGTATCCA-3' and 5'-GCCGGATCAAGCGTATGC-3', respectively, while the SDS forward and reverse primers for the CAT gene were 5'-GCCGCTGGCGATTTCAG-3' and 5'-TTCATTAAGCATTCTGCCGACAT-3'. The TaqMan probe for CAT gene, 5'-TCATCATGCCGTCTGTGATGGCTTC-3' and the probe for Neo gene, 5'-CGCCGCATTGCATCAGCCAT-3', have been labeled with a reporter fluorescent dye, 6-carboxyfluorescein (FAM), and VIC dye, respectively, at their 5' ends. The 3' ends of the probes have been labeled with a fluorescent dye, 6-carboxy-tetramethyl-rhodamine (TAMRA), as the quencher. The specificity of SDS primers was examined under the standard PCR conditions prior to quantitation by real-time PCR. The primers and probes have been synthesized by Applied Biosystems.

Real-time quantitative PCR

Real-time PCR was carried out using the TaqMan[®] EZ RT-PCR reagents. Both RT and DNA polymerization were carried out in the same tube without subsequent addition of enzymes or buffer.

The RT-PCR was performed in a MicroAmp Optical 96-well reaction plate using *rTth* DNA polymerase. This enzyme functions as both a thermostable reverse transcriptase and a thermostable DNA polymerase. Each RT-PCR reaction contained the following: 1× TaqMan buffer, 3 mM manganese acetate, 300 μM each of dATP, dCTP and dGTP, 600 μM of dUTP, 200 nM primers, 100 nM TaqMan probe, 2.0 U *rTth* DNA polymerase, 0.5 U AmpErase UNG and 50 ng total RNA, in 50 μl of reaction volume. RT was carried out for 30 min at 60°C following an initial 2 min incubation at 50°C for UNG activity to prevent carryover reactions. The reaction was terminated by heating at 95°C for 5 min. The PCR amplification was then performed for 40 cycles with each cycle at 94°C for 20 s and 60°C for 1 min. All reactions were carried out in triplicate using the ABI Prism 7700 SDS. The threshold cycle, C_T , values were averaged from the values obtained from each reaction, and the promoter activity was calculated as a relative level of expression (CAT gene using FAM-labeled probe) to the calibrator (Neomycin gene using VIC-labeled probe). The FAM and VIC spectral data were collected from reactions carried out in separate tubes using the same stock of RNA and other reagents to avoid the spectral overlap among VIC/FAM/TAMRA and to avoid limitation of reagents and dNTPs.

RESULTS AND DISCUSSION

Go to: ☒ Go to: ☒

CAT assays on cardiotoxin-2, neurotoxin-2 and phospholipase A₂ promoters

The promoters of cardiotoxin-2 (CTX-2; [11](#)), neurotoxin-2 (NTX-2; [12](#)) and phospholipase A₂ (PLA₂; [13](#)) genes were sub-cloned upstream of CAT gene into pMAMneoCAT promoterless reporter vector in both correct and reverse orientations and transfected separately into CHO cells. We have shown previously ([11](#)) that the epithelial-derived CHO cells yielded comparatively similar levels of expression of toxin genes and hence can be used as a cell line for the investigation of the toxin genes' promoter activity. A deletion construct of CTX promoter containing the highly active TATA box (Del 4; [11](#)) was

also used. Both the CAT assay and real-time PCR analysis were carried out for all the constructs. Figure 1 shows that the promoter activity of the CTX-2 gene, is ~5-fold higher than that of the PLA₂ gene. However, lower promoter activity was observed for NTX promoter (4-fold less than that of PLA₂). The CTX-deletion construct showed the highest promoter activity (~3-fold higher than the intact CTX-2 promoter). The promoters cloned in reverse orientation showed no CAT activity as expected. The ratio of CAT activity for promoters CTX:PLA₂:NTX was found to be 12:4:1 (Fig. 1).

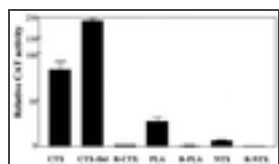


Figure 1

CAT assays on different toxin promoters. PLA, CTX and NTX represent PLA₂, CTX-2 and NTX-2 promoters; R-PLA, R-CTX and R-NTX represent the respective promoters in reverse orientation. CTX-Del denotes a selected deletion construct (Del 4; 11) of CTX-2 ...

Real-time PCR quantitation

The TaqMan assay utilizes the 5'→3' exonuclease activity of *Taq* DNA polymerase and a fluorogenic probe for automated quantification of DNA in a real-time manner. The C_T value refers to the threshold cycle at which a statistically significant increase in fluorescence is first detected by the sequence detection system. The increase in fluorescence is directly proportional to the exponential increase in PCR products and the measurement of signal is carried out in a real-time manner. Therefore, samples containing low concentration of target molecules would require more PCR cycles (hence higher C_T) to amplify enough copies to produce a significant fluorescent signal.

We have used the expression of the Neomycin gene present in the recombinant plasmid as an internal standard in these experiments. The optimal concentration of the total RNA for real-time detection was found to be 50 ng. A no-template control, in which sterile buffer was replaced for template RNA, was used in each experiment. This control was used to subtract any fluorescence that is not related to PCR amplification. The amount of target, normalized (ΔC_{TS}) to an endogenous reference (Neomycin gene) was calculated by subtracting the C_T value of the reference (C_{TR}) from the C_T value of the sample (C_{TS}; $\Delta C_{TS} = C_{TS} - C_{TR}$). The relative expression ($2^{-\Delta C_T}$) to an arbitrary calibrator (e.g. rCTX) is determined by subtracting the $\Delta C_{Tcalibrator}$ from the ΔC_{TS} value ($\Delta \Delta C_T = \Delta C_{TS} - \Delta C_{Tcalibrator}$). The ratio obtained from the results for the CAT mRNA produced by CTX:PLA₂:NTX was 2.01:1.5:1 (Table 1, column 6). These results corresponded to our previous study on the mRNA levels in the venom gland of *Naja naja sputatrix* determined by *in situ* hybridization, where CTX:PLA₂:NTX ratio was 2:1.3:1, respectively, (14). The venom protein profiles obtained by LC-MS (14) showed that the ratio of CTX:PLA:NTX in the venom was 13:3:1. Similarly, in a separate report, Tan (15) documented that CTX in *N.sputatrix* comprise almost 60% of venom dry weight, whereas PLA₂ and NTX comprise 14 and 4.5%, respectively, approximating to a similar ratio of 13:3:1. These differences between the mRNA and the protein content may be due to slow turnover rate of toxin transcripts and high translational efficiency in a manner similar to the observation made by other laboratories (16–19). These observations also imply that regulatory elements determining the mRNA stability and translational efficiency may be located within the toxin promoters. However, these speculations require further investigation.

Table 1.

Table 1 The threshold cycle (C _T) values of the promoter constructs									
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con
Con	Con	Con	Con	Con	Con	Con	Con	Con	Con

The threshold cycle (C_T) values of the promoter constructs

We obtain similar real-time results when the mRNA produced in each case has been normalized to either the pMAMneo promoter activity or to the rRNA in CHO cells (Table 1, columns 8 and 10, respectively). Hence, the results obtained by real-time PCR represent the actual amount of mRNA produced *in vivo*. From our CAT assays (based on the enzyme activity), it is clear that the values form an approximate estimation of the promotional activity of the promoters. We validated this by examining the expression of one of the toxin genes, CTX. Stable CHO clones containing the CTX promoter proximal to the CAT gene (in the pMAMneo-CAT plasmid) were prepared (11) and used in the measurement of CAT mRNA and CAT enzyme content. A ratio of CAT mRNA to CAT enzyme, 1:9, somewhat similar to the above results, was obtained.

In this study, we have also carried out an *in vitro* transcription of the CAT gene utilizing the CTX-2 promoter. Nuclear extracts from both the venom gland and CHO cells have been used. The results were found to be similar in both cases (Table 2) indicating that CHO cells form an alternative cell line for the analysis of toxin gene promoter activities. However, only a 2-fold increase in activity for the CTX-deletion construct (in contrast to the expected 4-fold increase) has been observed. We speculate that this may be due to the non-physiological environment present in *in vitro* assays. In our previous study (11), we demonstrated that the CTX promoter can initiate transcripts from the correct sites (similar to snake venom gland cells) when expressed in CHO cells. *In vitro* footprinting assays, using nuclear extracts from snake venom glands and CHO cells gave identical DNase I protection patterns, to demonstrate that CHO cells could be used as alternative cell lines to snake venom gland cells to investigate the expression toxin genes (11). Besides, Marchot *et al.* (20) have reported that CHO cells can be used to express and process toxin gene products.

Table 2 In vitro transcription on CTX-2 promoter				
In vitro transcription	C _T FAM (CAT)	ΔC _T rel to R-CTX	ΔΔC _T rel to R-CTX	2 ^{-ΔΔC_T}
Vin-CTX	19.3 ± 0.04	0	0	1.0
Vin-CTX	14.8 ± 0.09	-4.5	-4.5	22.63
Vin-CTXdel	13.7 ± 0.10	-5.6	-5.6	48.5
CHO-CTX	19.81 ± 0.06	0	0	1.0
CHO-CTX	15.40 ± 0.06	-4.41	-4.41	21.26

Table 2.
In vitro transcription on CTX-2 promoter

Real-time PCR has been widely used in the field of developmental studies (21), clinical diagnostics/prognosis (22–24) and the detection/screening of residual or very low copy numbers of gene or gene products (25,26). In this paper, we have demonstrated that real-time PCR could be used for quantifying the gene expression level. Compared to the traditional CAT assay, real-time PCR has provided a very rapid, sensitive and reproducible approach for quantitation of gene activity. The level of sensitivity (pg level) obtained by real-time PCR is rather difficult for assays such as northern hybridization and RNase protection. Real-time PCR also allows a large number of different RNA samples to be analysed and quantitated simultaneously. This method can also be used over a range of concentration of total RNA. Besides, the variations in results can be normalized using an internal control as, in this case, the expression of the neomycin gene. Hence, co-transfection experiments to determine the efficiency of transfections are not required. Thus, real-time PCR analysis forms the most direct and accurate method that is available to date for gene expression studies.

ACKNOWLEDGEMENTS

This work was supported by a Research Grant (R-183-000-031-112) from the National University of Singapore. D.M. received a University Scholarship.

NOTES

Go to: ☐ Go to: ☐

DDBJ/EMBL/GenBank accession nos⁺ To whom correspondence should be addressed. Tel: +65 874 3248; Fax: +65 779 1453; Email: AF097000, AF241224, AF101235

REFERENCES

Go to: ☐ Go to: ☐

1. Gorman C.M., Moffat,L.F. and Howard,B.H. (1982) Recombinant genomes which express chloramphenicol acetyltransferase in mammalian cells. *Mol. Cell Biol.*, 2, 1044–1051. [[PMC free article](#)] [[PubMed](#)]
2. Nguyen V.T., Morange,M. and Bensaude,O. (1988) Firefly luciferase luminescence assays using scintillation counters for quantitation in transfected mammalian cells. *Anal. Biochem.*, 171, 404–408. [[PubMed](#)]
3. Alam J. and Cook,J.L. (1990) Reporter genes: application to the study of mammalian gene transcription. *Anal. Biochem.*, 188, 245–254. [[PubMed](#)]
4. Kain S.R., Adams,M., Kondepudi,A., Yang,T.T., Ward,W.W. and Kitts,P. (1995) Green fluorescent protein as a reporter of gene expression and protein localization. *Biotechniques*, 19, 650–655. [[PubMed](#)]
5. Gibson U.E., Heid,C.A. and Williams,P.M. (1996) A novel method for real time quantitative RT–PCR. *Genome Res.*, 6, 995–1001. [[PubMed](#)]
6. Desjardin L.E., Chen,Y., Perkins,M.D., Teixeira,L., Cave,M.D. and Eisenach,K.D. (1998) Comparison of the ABI 7700 system (TaqMan) and competitive PCR for quantification of IS6110 DNA in sputum during treatment of tuberculosis. *J. Clin. Microbiol.*, 36, 1964–1968. [[PMC free article](#)] [[PubMed](#)]
7. Ishida K., Zhu,B. and Maeda,H. (2000) Novel approach to quantitative reverse transcription PCR assay of mRNA component in autopsy material using the TaqMan fluorogenic detection system: dynamics of pulmonary surfactant apoprotein A. *Forensic Sci. Int.*, 113, 127–131. [[PubMed](#)]
8. Stanners C.P., Eliceiri,G.L. and Green,H. (1971) Two types of ribosome in mouse–hamster hybrid cells. *Nat. New Biol.*, 230, 52–54. [[PubMed](#)]
9. Jordan M., Schallhorn,A. and Wurm,F.M. (1996) Transfecting mammalian cells: optimization of critical parameters affecting calcium-phosphate precipitate formation. *Nucleic Acids Res.*, 24, 596–601. [[PMC free article](#)] [[PubMed](#)]
10. Seed B. and Sheen,J.Y. (1988) A simple phase-extraction assay for chloramphenicol acyltransferase activity. *Gene*, 67, 271–277. [[PubMed](#)]
11. Ma D.H., Armugam,A. and Jeyaseelan,K. (2001) Expression of cardiotoxin-2 gene: cloning, characterization and deletion analysis of the promoter. *Eur. J. Biochem.*, 268, 1844–1850. [[PubMed](#)]
12. Afifiyan F., Armugam,A., Tan,C.H., Gopalakrishnakone,P. and Jeyaseelan,K. (1999) Postsynaptic α -neurotoxin gene of the spitting cobra, *Naja naja sputatrix*: structure, organization, and phylogenetic analysis. *Genome Res.*, 9, 259–266. [[PMC free article](#)] [[PubMed](#)]

13. Jeyaseelan K., Armugam,A., Donghui,M. and Tan,N.H. (2000) Structure and phylogeny of the venom group I phospholipase A(2) gene. *Mol. Biol. Evol.*, 17, 1010–1021. [[PubMed](#)]
14. Lachumanan R., Armugam,A., Durairaj,P., Gopalakrishnakone,P., Tan,C.H. and Jeyaseelan,K. (1999) *In situ* hybridization and immunohistochemical analysis of the expression of cardiotoxin and neurotoxin genes in *Naja naja sputatrix*. *J. Histochem. Cytochem.*, 47, 551–560. [[PubMed](#)]
15. Tan N.H. (1991) The biochemistry of venoms of some venomous snakes of Malaysia—a review. *Trop. Biomed.*, 8, 91–103.
16. Sato S., Yoshida,H., Abe,H. and Tamiya,N. (1969) Properties and biosynthesis of a neurotoxic protein of the venoms of sea snakes *Laticauda laticaudata* and *Laticauda colubrina*. *Biochem. J.*, 115, 85–90. [[PMC free article](#)] [[PubMed](#)]
17. Kochva E., Tonsing,L., Louw,A.I., Liebenberg,N.V. and Visser,L. (1982) Biosynthesis, secretion and *in vivo* isotopic labelling of venom of the Egyptian cobra, *Naja haje annulifera*. *Toxicon*, 20, 615–635. [[PubMed](#)]
18. Ogawa T., Onoue,H., Nakagawa,K., Nomura,S., Sueishi,K., Hattori,S., Kihara,H. and Ohno,M. (1995) Localization and expression of phospholipases A₂ in *Trimeresurus flavoviridis* (habu snake) venom gland. *Toxicon*, 33, 1645–1652. [[PubMed](#)]
19. Takeda M., Yoshida,H. and Tamiya,N. (1974) Biosynthesis of erabutoxins in the sea snake, *Laticauda semifasciata*. *Toxicon*, 12, 633–641. [[PubMed](#)]
20. Marchot P., Prowse,C.N., Kanter,J., Camp,S., Ackermann,E.J., Radic,Z., Bougis,P.E. and Taylor,P. (1997) Expression and activity of mutants of fasciculin, a peptidic acetylcholinesterase inhibitor from Mamba venom. *J. Biol. Chem.*, 272, 3502–3510. [[PubMed](#)]
21. Steuerwald N., Cohen,J., Herrera,R.J. and Brenner,C.A. (2000) Quantification of mRNA in single oocytes and embryos by real-time rapid cycle fluorescence monitored RT–PCR. *Mol. Hum. Reprod.*, 6, 448–453. [[PubMed](#)]
22. Fujimaki S., Funato,T., Harigae,H., Imaizumi,M., Suzuki,H., Kaneko,Y., Miura,Y. and Sasaki,T. (2000) A quantitative reverse transcriptase polymerase chain reaction method for the detection of leukaemic cells with t(8;21) in peripheral blood. *Eur. J Haematol.*, 64, 252–258. [[PubMed](#)]
23. Kreuzer K.A., Lass,U., Nagel,S., Ellerbrok,H., Pauli,G., Pawlaczyk-Peter,B., Siegert,W., Huhn,D. and Schmidt,C.A. (2000) Applicability of an absolute quantitative procedure to monitor intra-individual bcr/abl transcript kinetics in clinical samples from chronic myelogenous leukemia patients. *Int. J. Cancer*, 86, 741–746. [[PubMed](#)]
24. Pfitzner T., Engert,A., Wittor,H., Schinkothe,T., Oberhauser,F., Schulz,H., Diehl,V. and Barth,S. (2000) A real-time PCR assay for the quantification of residual malignant cells in B cell chronic lymphatic leukemia. *Leukemia*, 14, 754–766. [[PubMed](#)]
25. Zerr D.M., Huang,M.L., Corey,L., Erickson,M., Parker,H.L. and Frenkel,L.M. (2000) Sensitive method for detection of human herpesviruses 6 and 7 in saliva collected in field studies. *J. Clin. Microbiol.*, 38, 1981–1983. [[PMC free article](#)] [[PubMed](#)]
26. Miley W.J., Suryanarayana,K., Manns,A., Kubota,R., Jacobson,S., Lifson,J.D. and Waters,D. (2000) Real-time polymerase chain reaction assay for cell-associated HTLV type I DNA viral load.

Articles from Nucleic Acids Research are provided here courtesy of **Oxford University Press**