Table 1. Uptake Rate, Depuration Rate Constants (k_d) , Half-Life $(t_{1/2})$, Assimilation Efficiency (α) , Steady-State Bioaccumulation Factor (BAF), and Predicted Time to Achieve 85% Steady State $(t_{ss})^{\alpha}$

compound	exp. conc. (ng g^{-1})	tissue	uptake rate (pg $g^{-1} d^{-1}$)	$k_{\rm d} \times 10^{-2} \ ({\rm d}^{-1})$	$t_{1/2}$ (d)	α (%)	BAF	$t_{ m ss}$
P-CTX-1	19.1 ± 1.91	muscle	$33.7 \pm 2.53 (0.89)$	$2.52 \pm 0.215 (0.97)$	27.5 ± 2.34			
		skin	$66.3 \pm 3.49 (0.94)$	$0.966 \pm 0.475 (0.58)$	71.7 ± 35.3			
		brain	$48.5 \pm 2.77 (0.93)$	$5.92 \pm 1.40 (0.86)$	11.7 ± 2.78			
		gill	$45.5 \pm 3.03 (0.91)$	$1.67 \pm 0.501 (0.92)$	41.5 ± 12.4			
		liver	$95.0 \pm 6.12 (0.92)$	$16.5 \pm 1.27 (0.98)$	4.19 ± 0.323			
		intestine	$48.8 \pm 2.14 (0.96)$	$2.74 \pm 0.409 (0.96)$	25.3 ± 3.78			
		carcass	$25.7 \pm 1.38 (0.94)$	$2.91 \pm 0.287 (0.97)$	23.8 ± 2.35	6.01	0.0619	65.2
		whole fish	$25.2 \pm 1.30 \ (0.95)$	$3.23 \pm 0.32 \ (0.97)$	21.4 ± 2.12	6.13	0.0569	58.7
P-CTX-2	6.75 ± 1.06	muscle	$5.00 \pm 0.26 (0.94)$	$2.70 \pm 1.10 (0.67)$	25.6 ± 10.4			
		skin	$11.3 \pm 0.898 \ (0.88)$	$1.57 \pm 1.57 (0.33)$	44.1 ± 44.1			
		brain	$8.65 \pm 0.587 (0.91)$	$8.29 \pm 0.435 (0.99)$	8.36 ± 0.439			
		gill	$11.6 \pm 3.55 (0.33)$	$1.51 \pm 1.50 \ (0.25)$	45.9 ± 45.6			
		liver	$6.27 \pm 1.79 (0.53)$	$16.1 \pm 1.30 \ (0.98)$	4.31 ± 0.349			
		intestine	$10.8 \pm 0.817 (0.89)$	$2.00 \pm 0.840 \ (0.74)$	34.6 ± 14.5			
		carcass	$3.96 \pm 0.249 (0.92)$	$3.18 \pm 0.801 (0.84)$	21.8 ± 5.48	2.88	0.0271	59.7
		whole fish	$3.51 \pm 0.210 \ (0.93)$	$3.23 \pm 0.826 \ (0.84)$	21.5 ± 5.50	2.61	0.0243	58.7
P-CTX-3	4.00 ± 0.28	muscle	$1.37 \pm 0.0866 (0.92)$	$2.11 \pm 1.22 (0.60)$	32.9 ± 19.1			
		skin	$4.07 \pm 0.295 (0.90)$	$0.557 \pm 0.571 (0.24)$	125 ± 128			
		brain	$3.00 \pm 0.382 (0.74)$	$2.07 \pm 1.08 (0.55)$	33.5 ± 17.5			
		gill	$2.78 \pm 0.272 \ (0.83)$	$1.33 \pm 1.57 (0.42)$	52.0 ± 61.2			
		liver	$1.50 \pm 0.284 \ (0.56)$	$10.6 \pm 2.59 (0.94)$	6.53 ± 1.59			
		intestine	$2.54 \pm 0.165 (0.91)$	$1.66 \pm 1.07 (0.55)$	41.9 ± 26.9			
		carcass	$1.12 \pm 0.0786 \ (0.90)$	$1.82 \pm 1.57 (0.31)$	38.1 ± 32.8	1.28	0.0210	104
		whole fish	$0.956 \pm 0.0714 (0.89)$	$1.83 \pm 1.36 (0.37)$	38.0 ± 28.3	1.15	0.0189	104

[&]quot;Error represents \pm standard error, and values in parentheses represent the coefficient of determination (r^2) for the corresponding regression analysis.

$$\frac{C_{\text{fish}}}{C_{\text{feed}}} = \frac{\alpha F}{k_{\text{d}}} (1 - e^{-k_{\text{d}}t}) = BAF(1 - e^{-k_{\text{d}}t})$$
(3)

where F is the feeding rate (F = 0.03 g food g⁻¹ of fish d⁻¹). After long exposure times (i.e., $t \to \infty$), the equilibrium bioaccumulation factor (BAF) can be estimated through $\alpha F/k_{\rm d}$. Regarding the biotransformation between P-CTX-1, -2, and -3, a calculation³⁴ was conducted and the results indicated that the biotransformation of P-CTX-2 and -3 to P-CTX-1 in our exposed fish during the course of the experiment was negligible as shown in the Supporting Information.

■ RESULTS AND DISCUSSION

No orange-spotted groupers died during the experiment in either the control or treatment (exposed) groups. No detectable P-CTX-1, -2, and -3 were found in orange-spotted groupers in the control group at any time point during the entire accumulation-elimination phase (60 d). Significant differences in whole fish growth were observed between the exposed and control groups (p < 0.005). In addition, in the exposed group, the fish growth rate calculated at day 60 $(0.0144 \text{ g d}^{-1})$ was significantly lower than that at day 30 (i.e., the accumulation end day, 0.0211 g d^{-1}) (p < 0.001). To address the dilution effect of toxin concentrations due to fish growth during the entire experiment, tissue P-CTX concentrations were corrected by multiplying a factor of $(1 + g \times t)$.³⁵ Levels of P-CTX-1, -2, and -3 in various exposed grouper tissues (muscle, skin, gill, brain, liver, and intestine) were therefore growth-corrected using the two growth rates (i.e., day 30 and day 60), and the levels are shown in Figure 1. In the following discussion, the concentrations are all growthcorrected.

Uptake and Depuration Kinetics of P-CTXs. For P-CTX-1, during the uptake phase from 0 to 30 d, the detectable concentrations of P-CTX-1 in six tissues of exposed grouper followed a linear increase with linear coefficients (r^2) in the range of 0.89–0.96, as shown in Table 1. Except for liver (r^2 = 0.53) and gill ($r^2 = 0.33$), r^2 of the concentrations of P-CTX-2 for a linear increase in the other four tissues was in the range of 0.88-0.94. In regard to P-CTX-3, except for the liver $(r^2 =$ 0.56), r^2 of the concentrations for a linear increase in the other five tissues was in the range of 0.74-0.92. Furthermore, the concentrations of the three P-CTXs in all of the tissues did not reach a steady state during the 30 d exposure. An estimation of the time required to reach steady state (t_{ss}) was mathematically calculated for the whole fish and carcass (weight of carcass = weight of whole fish – weight of liver – weight of intestine).³⁵ According to eq 3, at steady state, $C_{\text{fish}}/C_{\text{feed}}$ approaches BAF asymptotically and, by the mathematical definition, never equals BAF (until $t \to \infty$).³³ To obtain $t_{\rm ss}$, we assumed 0.85 BAF (i.e., 85% to steady state) to replace $C_{\text{fish}}/C_{\text{feed}}$ and predict $t_{\rm ss}$ for P-CTXs during the uptake phase. As a result, the $t_{\rm ss}$ values of the three P-CTXs in the whole fish and carcass were in the range of 58.7–104 d, which exceeded 30 d (Table 1). Therefore, our experimental uptake data did not appear to level off during the 30 d exposure. 33,36 Similar results were reported in a previous study in which juvenile N. brevirostris dietarily exposed to ciguatoxic dinoflagellate daily achieved a steady state at approximately 8-16 weeks.²

During the elimination phase, P-CTX-1, -2, and -3 levels in muscle, brain, and liver of exposed orange-spotted groupers declined exponentially from day 32 to day 60. In the skin, intestine, and gill, however, the observed elimination curves indicated two stages of elimination kinetics. The concen-