

Project Summary

The main goal of this project is to investigate bark thickness as a fire-adapted phenotype in four, economically-important, species of pines with a range along the eastern and south-eastern United States. The evolution of this complex trait will be studied in populations of slash pine (*Pinus elliottii*), pond pine (*P. serotina*), loblolly pine (*P. taeda*), and long leaf pine (*P. palustris*). Next generation sequencing (NGS) of 25 individuals from 20 natural populations for each species ($25 \times 20 \times 4 = 2000$ total individuals) will be used to genotype individuals with the goal of uncovering shared genetic architecture of these four closely related species. The results of this research, while providing new genomic resources for non-model species and augmenting existing resources, will address fundamental principles in evolutionary biology and serve to inform and improve breeding programs and land management initiatives.

Fire has played an important role in the evolutionary paths of many adaptive traits in plants, across a wide variety of species and habitats. Extant Pinaceae species occupy a variety of habitats across North and Central America, Asia, and Europe. These areas have been strongly influenced by fire, and as such, their plants have developed a range of fire-adapted traits. This study will investigate the underlying genetic architecture associated with the complex bark thickness phenotype, the degree to which this architecture is shared among four closely related species across their natural ranges and multiple evolutionary time scales, and how this variability can inform current economic interests. **Thus, this research directly applies to the overall goal of the NPGI by developing a basic knowledge of the structures and functions of economically important plant genomes.**

Intellectual Merit

By focusing on the dissection of an adaptive trait at both microevolutionary and macroevolutionary time scales, this study will address a fundamental principle in evolutionary biology: the mechanism associated with the vast majority of incomplete lineage sorting among populations of closely related species. The rise of shared phenotypes between related species has been a source of debate for many years, dividing into camps supporting either ancient inherited polymorphisms or multiple instances of convergent evolution. Very recently, Ségurel et al. (2012) showed that the ABO blood group antigens in humans are the result of vertical inheritance of maintained, ancient polymorphisms and Roux et al. (2012) demonstrated balancing selection around the S-locus in two species of *Arabidopsis*. In short, this study seeks to investigate the same phenomena in a quantitative, economically and environmentally-important, trait in four species of related plants at multiple evolutionary time scales. Additionally, the genomic resources developed in this project will augment and facilitate knowledge transfer of genomic data in non-model species which, until recently, have been technologically out of reach.

Broader Impacts

Data will be made available for download and exploration from VCU-hosted FTP and web sites. Also, the PI will develop and teach a new course, Applied Ecological Genomics, to introduce and expose students to the bioinformatic skills necessary to effectively manage terabases of data as well as how to exploit NGS data to infer population-level, evolutionary processes. Finally, this will give the PI additional teaching and research experience needed to jumpstart an academic career, focusing on ecologically-relevant problems in an evolutionary context.

Project Description

Fire has a been crucial influence in global ecosystem processes and on forested ecosystems, driving not only locally adapted phenotypes in plant populations (Lamont et al., 1991; Vega et al., 2008; Midgley and Bond, 2011; Keeley et al., 2011; He et al., 2012; Parchman et al., 2012), but also impacting carbon storage and climate (Bowman et al., 2009). Understanding the underlying genetic architecture associated with fire-associated traits is therefore critical to dealing with the environmental impacts of increased carbon inputs and changing climate. He et al. (2012) recently investigated five fire-adapted traits for 101 species of *Pinus* and found evidence, from the Cretaceous, for a strong influence of fire on trait evolution. However, they considered only presence or absence of the traits in their study. **There have been no studies to date which capture, as in this proposal, the quantitative nature of fire-associated trait evolution in these species.**

Plants display strong patterns of adaptation, especially local adaptation, when populations sizes are large (Leimu and Fischer, 2008). This characteristic is especially important in conifers, which display a lack of population structure and large effective population sizes (Neale and Savolainen, 2004); this is partly why association genetic studies have been so successful in these species (Gupta et al., 2005; González-Martínez et al., 2006, 2007; Eckert et al., 2009; Wegrzyn et al., 2010; Eckert et al., 2010, 2012).

The PI will investigate the genetic architecture of a complex phenotype in 2000 trees from four, economically-important species of pines with a range along the eastern and south-eastern United States (25 trees from 20 populations for each species). Bark thickness is a highly heritable trait for forest trees (e.g., $H^2 = 0.65$ for *P. taeda*; Pederick (1970)), and, as shown by He et al. (2012), is involved with adaptation to fire by species of *Pinus*, including the four focal species. The PI will study bark thickness, as a quantitative, fire-adapted phenotypic trait, in populations of slash pine (*Pinus elliottii*), pond pine (*P. serotina*), loblolly pine (*P. taeda*), and long leaf pine (*P. palustris*). **Using next-generation, high-throughput sequencing, the PI will dissect bark thickness as a complex and adaptive, quantitative trait across populations of these four closely related species with the goal of uncovering shared genetic architecture at multiple evolutionary time scales.** These findings have direct economic impact. For example, Marshall et al. (2006) demonstrated the negative financial impact that bark mass has on the forest industry. Linking model-based estimates of bark thickness with markers linked to the causative polymorphisms underlying variation for this trait can help to inform breeding plans to minimize this loss.

The application of next-generation sequencing technology to conifer genomes, as proposed

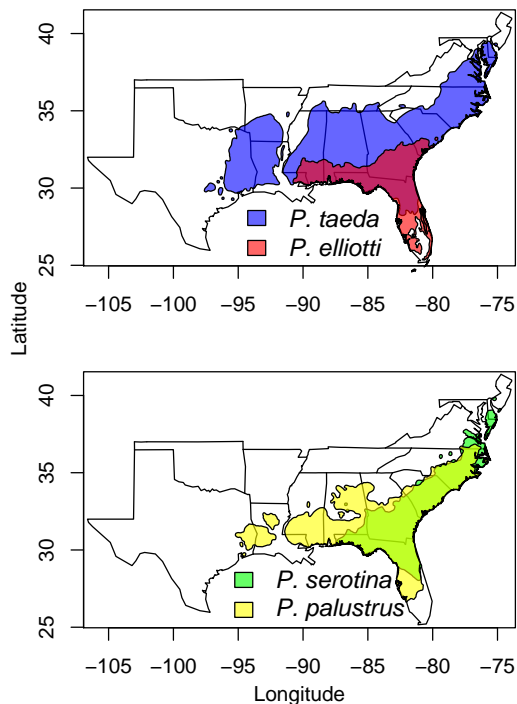


Figure 1: Natural ranges for four study species

in this study, holds the promise of being able to provide a way to dissect the genetic architecture of an adaptive trait. Until recently, techniques to consider adaptation on a genomic scale have been no match for the size and complexity of conifer genomes (Mackay et al., 2012). However, advances in genomic enrichment techniques such as those by Parchman et al. (2012) and Willing et al. (2011) coupled with the ever-decreasing cost of DNA sequencing have facilitated the application of new genotype-by-sequencing (GBS) approaches to answering fundamental questions in evolutionary biology in non-model species. **The PI is well-positioned to utilize the sophistication of the sequencing facilities at VCU, coupled with his backgrounds in evolutionary, molecular, and computational biology to study how bark thickness has evolved across *Pinus* and how this knowledge may serve to inform ecologically-relevant and economically-important decisions related this important genus.**

Dr. Andrew J. Eckert was chosen as a sponsor of this project due to his experience with association mapping and evolutionary genetics in conifers. He has published over 30 papers in this field and is recognized as an authority on complex trait dissection in forest tree populations. The PI has elected to stay at VCU for this research due to the availability of excellent faculty collaborators with extensive experience in phylogenetics and molecular evolution (Dr. Maria C. Rivera) and landscape genetics (Dr. Rodney J. Dyer). The research and teaching experience gained in this project will position the PI to competitively seek a future tenure-track position at a tier 1 research institution.

This study is laid out below in three general phases: (1) field sampling and DNA preparation, (2) DNA sequencing and bioinformatics, and (3) statistical analysis and hypothesis testing. The quarterly timeline for this project is shown in Figure 2.

Phase 1: Field sampling and DNA preparation

Field sampling The PI will sample four species in their natural ranges, with 25 individuals from 20 natural populations of each species (Figure 1). The locations of each population will be determined randomly. Within each population, a representative of the focal species will be chosen randomly. To avoid sampling close relatives, all other members of the population will be chosen to be at least 50 m from the first sampling location. Only trees with diameter at breast height (1.25 m above ground; DBH) of at least 15 cm will be sampled. All efforts will be made to sample adult trees with similar DBHs, so as to standardize ages within sites. Three to five needle fascicles will be collected from each tree for use in DNA extraction. Needles, with a desiccant, will be stored in 15 ml FalconTM tubes at ambient temperatures in the field and at -80°C long-term in the laboratory.

Bark thickness will be treated as a quantitative character in all downstream analyses, and as such, will be measured for each tree at five random locations around DBH. Measurements of bark thickness will be taken using a Haglöff[®] bark gauge, and the height of the tree will be measured using a clinometer. These values will also be used to obtain the best possible regression model for future studies in these species, using the regression equation from Cao and Pepper (1986); including

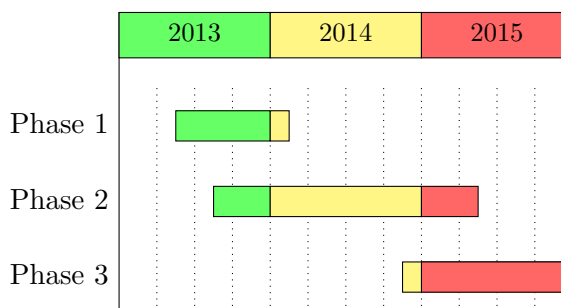


Figure 2: Project timeline

these covariates has been shown to significantly decrease estimation bias by up to 43% in some species (Li and Weiskittel, 2010). At the end of sampling, 2000 individual tubes of needles (20–25 needles/tube) will be banked along with GPS coordinates and measurements of bark thickness and height for all trees for all species.

DNA preparation Genomic DNA will be extracted from needle tissue using Qiagen DNEasy extraction kits in a 96-well format following an established tissue preparation protocol in the Eckert lab located at VCU. A single DNA extraction from multiple needles per individual will be performed for each sampled tree.

Criteria for completion Phase 1 will be designated complete when genomic DNA has been extracted for all 2000 individuals. DNA quantitation data as well as sample metadata will be stored in a relational database management system (RDBMS) that will also house sequencing and genotyping data from phase 2. It is estimated that phase 1 will be complete by June 2014.

Phase 2: DNA sequencing and bioinformatics

The PI will perform genotype-by-sequencing (GBS) utilizing highly-multiplexed libraries on the Illumina HiSeq 2000 platform. Genomic enrichment will be performed using the approach outlined in Parchman et al. (2012), in order to ensure high-coverage of the library given the complexity and size of conifer genomes (Mackay et al., 2012). Briefly, the genomic DNA from an individual is digested with two different restriction enzymes coupled with Illumina-specific adaptor ligation, including a computationally-correctable sequencing barcode (454 Life Sciences Corp., 2009) tied to an individual tree. The product is then amplified by PCR, size-selected on an agarose gel, purified, and pooled for multiplex sequencing. For 2000 trees, the amount of sequence data will approach 8 TB (terabases). All costs for sequencing will be absorbed by the Eckert lab.

The Nucleic Acids Research Facility (NARF) at VCU is currently producing paired-end reads of 150 bases in length. Using software that the PI has already written (Friedline, 2008–2012) coupled with existing libraries (e.g., BioPython (Cock et al., 2009), NGS QC Toolkit (Patel and Jain, 2012)), the processed sequencing data will be computationally divided into individuals and purged of low-quality reads. Additionally, reads that meet acceptance criteria globally, but possess bases (most often at the 3' end) that have low quality scores, will be trimmed and retained.

Once quality-controlled reads are obtained for each individual (and each population), they will be aligned (mapped) to the most current draft genome of *P. taeda*, available from Dendrome (dendrome.ucdavis.edu), using the Burroughs-Wheeler aligner (BWA) (Li and Durbin, 2009), and SNPs will be called. A SNP is called at particular alignment positions given enough high-quality reads to indicate a difference between the sample and the reference genome given a model of error. Sequence data from NGS technology requires the use of new SNP calling algorithms that properly model all the sources of errors (e.g, base calling errors, mapping errors, and sample preparation errors, particularly PCR errors which can have a significant impact on the analysis). Several tools have been developed to perform this task such as SAMtools (Li et al., 2009), GATK (McKenna et al., 2010), and, more recently, GeMS (You et al., 2012). The result of this phase will be a sample ($n = 2000$) by SNP matrix ($n = \text{thousands}$) for each species ($n = 4$).

Criteria for completion Phase 2 will be complete when all libraries have been sequenced and processed. Processing will include read demultiplexing, quality control, and storage as compressed files and in a relational database system. Additionally, mapping to the loblolly genome and SNP calling will result in a sample by SNP matrix. Sequence data will also be released at this time. This phase is the most computationally complex, and should be complete in early summer 2015.

Phase 3: Statistical analysis and hypothesis testing

This phase will focus on uncovering the underlying genetic architecture of bark thickness, summarized as four main questions. Each question will be answered by testing a null hypothesis, the rejection of which leads to the subsequent question.

Question 1: What is the genetic architecture of bark thickness for the four focal pine species that are adapted to fire?

Null hypothesis H_0^1 : There is no association between genotypic variation at surveyed SNPs and bark thickness in any of the four species.

Methods The PI will employ standard linear models that correct for kinship and population structure that are commonly used in genome-wide association studies (GWAS; Yu et al. (2006)). Association analysis has been widely used for dissecting the genetic basis of phenotypes for conifers (Neale and Kremer, 2011; Ingvarsson and Street, 2011). In addition, the PI will explore the use of the Bayesian models presented by Parchman et al. (2012), which was the first GWAS for a conifer, and the regression tree approaches presented by Holliday et al. (2012). Linear models will be used to explicitly test the effects of genotypic class of each discovered SNP on quantitative measures of bark thickness for each species. Multiple tests will be accounted for using a minimum P -value approach (Conneely and Boehnke, 2007).

Expected outcomes and relevance The null hypothesis will be rejected when at least one SNP in each of the four pine species is significantly associated to quantitative variation in bark thickness. These results will classify SNPs discovered in each species into two classes: those associated to bark thickness (which rejects the null hypothesis for this question) and those unassociated with bark thickness. The latter will be used as *controls* for questions two through four. The former establishes the genetic architecture of bark thickness for each focal species, and will be referred to as *candidates*.

Question 2: Are candidate SNPs shared across the four focal species, and if so, are they shared to a greater degree than randomly sampled SNPs from the controls?

Null hypotheses H_0^2 : There is no shared genetic architecture for bark thickness among the four focal species. H_0^3 : The degree of allele sharing is higher for candidate SNPs than randomly selected control SNPs.

Methods To evaluate H_0^2 , the PI will leverage the underlying, queryable data storage infrastructure to produce the intersection of SNPs associated with bark thickness in all four focal species (H_0^1). To evaluate H_0^3 , the union of all SNPs associated with bark thickness will be created. The fraction of this list that is shared across all four species will be used as a test statistic, the magnitude of which will be tested via a simple permutation analysis. The PI will construct a null distribution for the fraction of alleles shared across all four species for sets of SNPs randomly selected from those unassociated to bark thickness.

Allele sharing across species is common among conifer species and has been attributed largely to incomplete lineage sorting resulting from recent divergence times and large effective population sizes (Syring et al., 2007; Willyard et al., 2009), ancient admixture (Liston et al., 2007), and long-term gene flow (Zhou et al., 2010). For example, an analysis of levels of long-term gene flow between loblolly and slash pines, using multiple random sets of 50 nuclear genes and an isolation-with-migration model (Becquet and Przeworski, 2007), detected low but significant levels of gene flow

over the divergence history of these species (unpublished data, $4N_em = 2.5$ with a 95 % confidence interval of 0.52–5.14). The sets of loci randomly sampled from the unassociated loci will be assumed to largely reflect these processes, so that the enrichment of allele sharing at loci associated to a trait such as bark thickness, with clear adaptive relevance (He et al., 2012), would imply additional processes such as natural selection, as in Ségurel et al. (2012) and Roux et al. (2012), in promoting allele sharing among species.

Expected outcomes and relevance H_0^2 will be rejected when multiple SNPs associated with bark thickness are shared across species. The relevance of this result would be to establish the shared genetic architecture of an adaptive trait. H_0^3 will be rejected when the fraction of SNPs associated to bark thickness that are shared across the four focal species lies in the upper 1 % tail of the null distribution based upon random sets of unassociated SNPs. A significant result would support the existence of a process beyond incomplete lineage sorting, admixture, and gene flow that is contributing to the maintenance of shared alleles across the four focal species.

Question 3: Are reconstructed allele frequencies at shared candidate SNPs correlated more so with reconstructed bark thickness than shared control SNPs across the phylogeny for these four species?

Null hypothesis H_0^4 : Correlations between reconstructed allele frequencies at shared candidate SNPs and reconstructed values of bark thickness are no larger than those for reconstructed allele frequencies at randomly-chosen, shared control SNPs.

Methods Phylogenies and associated data matrices for genus *Pinus* will be obtained from TreeBASE (Morell, 1996) and GenBank (Benson et al., 2012). Most data matrices are constructed primarily from cpDNA coding sequences (Eckert and Hall, 2006; Gernandt et al., 2008) and cpDNA genome sequences (Parks et al., 2009). Chronograms will be obtained using the fossil calibration points from Willyard et al. (2007) as presented in He et al. (2012). Once a set of plausible chronograms has been generated, bark thickness and allele frequencies within lineages will be reconstructed as quantitative characters using a Bayesian method (Pagel et al., 2004). Specifically, we will utilize (1) a set of plausible chronograms ($n = 10\,000$) to account for uncertainty in the phylogenetic tree and (2) the character states, in combination with phylogenetically informed multiple regression analyses available in BayesTraits as BayesContinuous (Pagel et al., 2004), to test the association with reconstructed SNP allele frequencies and bark thickness for shared candidate and shared control SNPs. Allele frequencies and bark thickness will be specified for each species as averages and variances across populations.

Expected outcomes and relevance H_0^4 will be rejected when the multiple regression model for candidate SNPs better predicts bark thickness than for control SNPs. This suggests that the correlation structure of allele frequencies with bark thickness differs between the candidates and controls, as defined by answering questions 1 and 2. Since controls are matched in number and levels of heterozygosity with the candidates, this would suggest non-neutral processes driving this difference.

Question 4: To what extent is natural selection affecting rates of molecular evolution at candidate genes?

Null hypothesis H_0^5 : There is no evidence of selection in genes containing SNPs that are significantly associated with bark thickness.

Methods As the sequence resource, including annotations, for the loblolly pine genome continue to improve, establishing the genomic context of bark thickness-associated SNPs will enable the PI to test for evidence of selection in polymorphic, protein coding genes. From the results stemming from testing H_0^2 and H_0^3 , the PI will select 10 up to 20 SNPs from subsets of candidate and control loci from each population for each species exhibiting similar attributes (e.g., average heterozygosity). Using genomic position of SNPs and the loblolly genome annotations, the PI will design PCR primers to amplify the protein coding regions from each subsample ($n = 40$ trees), and each amplicon will be sent for Sanger sequencing. Once the sequences have been obtained, they will be aligned using standard tools such as MAFFT (Katoh et al., 2005) or MUSCLE (Edgar, 2004). From the alignment, d_N/d_S will be estimated, per locus and per site per locus, using established maximum likelihood methods (Yang, 2007).

Expected outcome and relevance Knowing whether or not genes containing SNPs are under positive ($d_N/d_S > 1$) or purifying selection ($d_N/d_S < 1$) addresses a fundamental question surrounding maintenance of these polymorphisms in natural populations. It is expected that d_N/d_S should be higher for candidates than controls.

Intellectual Merit and Broader Impacts

By answering these four questions, the PI will have broken down bark thickness as an adaptive trait into its genetic components, uncovering any shared genetic architecture in the focal species, at multiple evolutionary time scales. This project will benefit the local and global scientific communities, as well as help train the next generation of multi-discipline scientists in the world of big data. First, data will be made available for download and exploration from FTP and web sites hosted at VCU. Second, the PI will develop and teach a new three-credit course, BIOL 591 Applied Ecological Genomics, which will train advanced undergraduates and graduate students, using the data generated in this project, in skills necessary to both understand the technology available at VCU and to process, manage, and analyze the deluge of data generated from current genomics projects. This course will run in conjunction with an existing course, BIOL 693 Ecological Genomics taught by sponsor Eckert.

Given the diverse student body at VCU, the proposed research and teaching activities will increase research opportunities for underrepresented groups. VCU is among the top 20 universities in the nation for Biology degrees awarded to ethnic and racial minority students. In 2009, 18.3% of baccalaureate Biology degrees were conferred to African American students, exceeding the national average in Biology by 11.5% (NSF, 2010). A high percentage of VCU Biology (39.4%) baccalaureate degrees were also earned by Asian/Pacific Islanders, a value that far exceeds the national average in Biology of 6.2%. Caucasian students were awarded 34.9% of VCU baccalaureate Biology degrees as compared with the national average in Biology of 66.4% (NSF, 2010).

Results From Prior NSF Support

PI Friedline has yet to receive funding from the National Science Foundation.

References Cited

- 454 Life Sciences Corp. Using multiplex identifier (MID) adaptors for the GS FLX Titanium chemistry - extended MID set . *Technical Bulletin*, TCB 005-2009:1–7, Apr. 2009. URL http://www.liv.ac.uk/media/livacuk/centreforgenomicresearch/The_GS_FLX_Titanium_Chemistry_Extended_MID_Set.pdf.
- C. Becquet and M. Przeworski. A new approach to estimate parameters of speciation models with application to apes. *Genome Research*, 17(10):1505–1519, Oct. 2007. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=17712021&retmode=ref&cmd=prlinks>.
- D. A. Benson, I. Karsch-Mizrachi, K. Clark, D. J. Lipman, J. Ostell, and E. W. Sayers. GenBank. *Nucleic Acids Research*, 40(Database issue):D48–53, Jan. 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22144687&retmode=ref&cmd=prlinks>.
- D. M. J. S. Bowman, J. K. Balch, P. Artaxo, W. J. Bond, J. M. Carlson, M. A. Cochrane, C. M. D’Antonio, R. S. Defries, J. C. Doyle, S. P. Harrison, F. H. Johnston, J. E. Keeley, M. A. Krawchuk, C. A. Kull, J. B. Marston, M. A. Moritz, I. C. Prentice, C. I. Roos, A. C. Scott, T. W. Swetnam, G. R. van der Werf, and S. J. Pyne. Fire in the Earth system. *Science*, 324(5926):481–484, Apr. 2009. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19390038&retmode=ref&cmd=prlinks>.
- Q. V. Cao and W. D. Pepper. Predicting inside bark diameter for Shortleaf, Loblolly, and Longleaf pines. *Southern Journal of Applied Forestry*, 10:220–224, 1986. URL <http://www.ingentaconnect.com/content/saf/sjaf/1986/00000010/00000004/art00013>.
- P. J. A. Cock, T. Antao, J. T. Chang, B. A. Chapman, C. J. Cox, A. Dalke, I. Friedberg, T. Hamelryck, F. Kauff, B. Wilczynski, and M. J. L. de Hoon. Biopython: freely available Python tools for computational molecular biology and bioinformatics. *Bioinformatics*, 25(11):1422–1423, June 2009. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19304878&retmode=ref&cmd=prlinks>.
- K. N. Conneely and M. Boehnke. So many correlated tests, so little time! Rapid adjustment of P values for multiple correlated tests. *American Journal of Human Genetics*, 81(6):1158–1168, Dec. 2007. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=17966093&retmode=ref&cmd=prlinks>.
- A. J. Eckert and B. D. Hall. Phylogeny, historical biogeography, and patterns of diversification for Pinus (Pinaceae): phylogenetic tests of fossil-based hypotheses. *Molecular Phylogenetics and Evolution*, 40(1):166–182, July 2006. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=16621612&retmode=ref&cmd=prlinks>.
- A. J. Eckert, A. D. Bower, J. L. Wegrzyn, B. Pande, K. D. Jermstad, K. V. Krutovsky, J. B. St Clair, and D. B. Neale. Association genetics of coastal Douglas fir (*Pseudotsuga menziesii* var. *menziesii*, Pinaceae). I. Cold-hardiness related traits. *Genetics*, 182(4):1289–1302, Aug. 2009. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19487566&retmode=ref&cmd=prlinks>.

- A. J. Eckert, J. van Heerwaarden, J. L. Wegrzyn, C. D. Nelson, J. Ross-Ibarra, S. C. González-Martínez, and D. B. Neale. Patterns of population structure and environmental associations to aridity across the range of loblolly pine (*Pinus taeda* L., Pinaceae). *Genetics*, 185(3):969–982, July 2010. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=20439779&retmode=ref&cmd=prlinks>.
- A. J. Eckert, J. L. Wegrzyn, W. P. Cumbie, B. Goldfarb, D. A. Huber, V. Tolstikov, O. Fiehn, and D. B. Neale. Association genetics of the loblolly pine (*Pinus taeda*, Pinaceae) metabolome. *New Phytologist*, 193(4):890–902, Mar. 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22129444&retmode=ref&cmd=prlinks>.
- R. C. Edgar. MUSCLE: a multiple sequence alignment method with reduced time and space complexity. *BMC Bioinformatics*, 5:113, Aug. 2004. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=15318951&retmode=ref&cmd=prlinks>.
- C. J. Friedline. Code repository, 2008–2012. URL <http://code.friedline.net>.
- C. J. Friedline, R. B. Franklin, S. L. McCallister, and M. C. Rivera. Bacterial assemblages of the eastern Atlantic Ocean reveal both vertical and latitudinal biogeographic signatures. *Biogeosciences*, 9(6):2177–2193, 2012. URL <http://www.biogeosciences.net/9/2177/2012/>.
- D. S. Gernandt, S. Magallon, G. G. Lopez, O. Z. Flores, A. Willyard, and A. Liston. Use of simultaneous analyses to guide fossil-based calibrations of Pinaceae phylogeny. *International Journal Of Plant Sciences*, 169(8):1086–1099, 2008. URL <http://web.ebscohost.com/ehost/detail?sid=df0248f3-f8cc-4b19-9f27-6d88715a4381%40sessionmgr15&vid=1&hid=14&bdata=JkF1dGhUeXB1PWlwLHVybCxb29raWUzdWlkJnNpdGU9ZWVhc3QtbG12ZSZzY29wZT1zaXRl#db=a9h&AN=35434934>.
- S. C. González-Martínez, K. V. Krutovsky, and D. B. Neale. Forest-tree population genomics and adaptive evolution. *New Phytologist*, 170(2):227–238, 2006. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=16608450&retmode=ref&cmd=prlinks>.
- S. C. González-Martínez, N. C. Wheeler, E. Ersoz, C. D. Nelson, and D. B. Neale. Association genetics in *Pinus taeda* L. I. Wood property traits. *Genetics*, 175(1):399–409, Jan. 2007. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=17110498&retmode=ref&cmd=prlinks>.
- P. K. P. Gupta, S. S. Rustgi, and P. L. P. Kulwal. Linkage disequilibrium and association studies in higher plants: present status and future prospects. *Plant Molecular Biology*, 57(4):461–485, Mar. 2005. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=15821975&retmode=ref&cmd=prlinks>.
- T. He, J. G. Pausas, C. M. Belcher, D. W. Schwilk, and B. B. Lamont. Fire-adapted traits of *Pinus* arose in the fiery Cretaceous. *New Phytologist*, 194(3):751–759, May 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22348443&retmode=ref&cmd=prlinks>.

- J. A. J. Holliday, T. T. Wang, and S. S. Aitken. Predicting adaptive phenotypes from multilocus genotypes in Sitka Spruce (*Picea sitchensis*) using random forest. *G3: Genes, Genomes, Genetics*, 2(9):1085–1093, Sept. 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22973546&retmode=ref&cmd=prlinks>.
- P. K. Ingvarsson and N. R. Street. Association genetics of complex traits in plants. *New Phytologist*, 189(4):909–922, Mar. 2011. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=21182529&retmode=ref&cmd=prlinks>.
- K. Katoh, K.-i. Kuma, H. Toh, and T. Miyata. MAFFT version 5: improvement in accuracy of multiple sequence alignment. *Nucleic Acids Research*, 33(2):511–518, 2005. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=15661851&retmode=ref&cmd=prlinks>.
- J. E. Keeley, J. G. Pausas, P. W. Rundel, W. J. Bond, and R. A. Bradstock. Fire as an evolutionary pressure shaping plant traits. *Trends in Plant Science*, 16(8):406–411, Aug. 2011. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=21571573&retmode=ref&cmd=prlinks>.
- J. A. Lake and M. C. Rivera. Deriving the genomic tree of life in the presence of horizontal gene transfer: conditioned reconstruction. *Molecular Biology and Evolution*, 21(4):681–690, Apr. 2004. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=14739244&retmode=ref&cmd=prlinks>.
- B. B. Lamont, D. C. Maitre, R. M. Cowling, and N. J. Enright. Canopy seed storage in woody plants. *The Botanical Review*, 57(4):277–317, Oct. 1991. URL <http://www.springerlink.com/index/10.1007/BF02858770>.
- R. Leimu and M. Fischer. A meta-analysis of local adaptation in plants. *PLOS ONE*, 3(12):e4010, 2008. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19104660&retmode=ref&cmd=prlinks>.
- H. Li and R. Durbin. Fast and accurate short read alignment with Burrows-Wheeler transform. *Bioinformatics*, 25(14):1754–1760, July 2009. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19451168&retmode=ref&cmd=prlinks>.
- H. Li, B. Handsaker, . G. P. D. P. Subgroup, and 10. The Sequence Alignment/Map format and SAMtools. *Bioinformatics*, 25(16):2078–2079, Aug. 2009. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19505943&retmode=ref&cmd=prlinks>.
- R. Li and A. R. Weiskittel. Estimating and predicting bark thickness for seven conifer species in the Acadian Region of North America using a mixed-effects modeling approach: comparison of model forms and subsampling strategies. *European Journal of Forest Research*, 130(2):219–233, Aug. 2010. URL <http://www.springerlink.com/index/10.1007/s10342-010-0423-y>.
- A. A. Liston, M. M. Parker-Defeniks, J. V. J. Syring, A. A. Willyard, and R. R. Cronn. Interspecific phylogenetic analysis enhances intraspecific phylogeographical inference: a case study in *Pinus lambertiana*. *Molecular Ecology*, 16(18):3926–3937, Sept. 2007. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=17850554&retmode=ref&cmd=prlinks>.

- J. Mackay, J. F. D. Dean, C. Plomion, D. G. Peterson, F. M. Cánovas, N. Pavy, P. K. Ingvarsson, O. Savolainen, M. Á. Guevara, S. Fluch, B. Vinceti, D. Abarca, C. Díaz-Sala, and M.-T. Cervera. Towards decoding the conifer giga-genome. *Plant Molecular Biology*, Sept. 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22960864&retmode=ref&cmd=prlinks>.
- H. D. Marshall, G. E. Murphy, and B. Lachenbruch. Effects of bark thickness estimates on optimal log merchandising. *Forest Products Journal*, 56(11/12):87–92, 2006. URL <http://hdl.handle.net/1957/17956>.
- A. McKenna, M. Hanna, E. Banks, A. Sivachenko, K. Cibulskis, A. Kernytsky, K. Garimella, D. Altshuler, S. Gabriel, M. Daly, and M. A. DePristo. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Research*, 20(9):1297–1303, Sept. 2010. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=20644199&retmode=ref&cmd=prlinks>.
- J. Midgley and W. Bond. Pushing back in time: the role of fire in plant evolution. *New Phytologist*, 191(1):5–7, July 2011. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=21631504&retmode=ref&cmd=prlinks>.
- V. Morell. TreeBASE: the roots of phylogeny. *Science*, 273(5275):569, Aug. 1996. URL <http://www.treebase.org>.
- D. B. Neale and A. Kremer. Forest tree genomics: growing resources and applications. *Nature Reviews Genetics*, 12(2):111–122, Feb. 2011. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=21245829&retmode=ref&cmd=prlinks>.
- D. B. Neale and O. Savolainen. Association genetics of complex traits in conifers. *Trends in Plant Science*, 9(7):325–330, July 2004. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=15231277&retmode=ref&cmd=prlinks>.
- M. M. Pagel, A. A. Meade, and D. D. Barker. Bayesian estimation of ancestral character states on phylogenies. *Systematic Biology*, 53(5):673–684, Oct. 2004. URL <http://www.jstor.org/stable/4135357>.
- T. L. Parchman, Z. Z. Gompert, J. J. Mudge, F. D. Schilkey, C. W. Benkman, and C. A. Buerkle. Genome-wide association genetics of an adaptive trait in lodgepole pine. *Molecular Ecology*, 21(12):2991–3005, June 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22676074&retmode=ref&cmd=prlinks>.
- M. Parks, R. Cronn, and A. Liston. Increasing phylogenetic resolution at low taxonomic levels using massively parallel sequencing of chloroplast genomes. *Bmc Biology*, 7:84–84, 2009. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19954512&retmode=ref&cmd=prlinks>.
- R. K. Patel and M. Jain. NGS QC Toolkit: a toolkit for quality control of next generation sequencing data. *PLOS ONE*, 7(2):e30619–e30619, 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22312429&retmode=ref&cmd=prlinks>.

- L. A. Pederick. Variation and inheritance of stem form and bark thickness in young loblolly pine. *Technical Report.Sch.For.Resour.N.C.St.Univ.*, (41):44–44, 1970.
- M. C. Rivera and J. A. Lake. The ring of life provides evidence for a genome fusion origin of eukaryotes. *Nature*, 431(7005):152–155, Sept. 2004. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=15356622&retmode=ref&cmd=prlinks>.
- C. Roux, M. Pauwels, M. V. Ruggiero, D. Charlesworth, V. Castric, and X. Vekemans. Recent and ancient signature of balancing selection around the S-locus in *Arabidopsis halleri* and *A. lyrata*. *Molecular Biology and Evolution*, Oct. 2012. URL <http://mbe.oxfordjournals.org/content/early/2012/10/26/molbev.mss246.abstract>.
- L. Séguérel, E. E. Thompson, T. Flutre, J. Lovstad, A. Venkat, S. W. Margulis, J. Moyse, S. Ross, K. Gamble, G. Sella, C. Ober, and M. Przeworski. The ABO blood group is a trans-species polymorphism in primates. *Proceedings Of The National Academy Of Sciences Of The United States Of America*, Oct. 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=23091028&retmode=ref&cmd=prlinks>.
- J. Syring, K. Farrell, R. Businský, R. Cronn, and A. Liston. Widespread genealogical nonmonophyly in species of *Pinus* subgenus *Strobus*. *Systematic Biology*, 56(2):163–181, Apr. 2007. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=17454973&retmode=ref&cmd=prlinks>.
- J. A. Vega, C. Fernández, P. Pérez-Gorostiaga, and T. Fonturbel. The influence of fire severity, serotiny, and post-fire management on *Pinus pinaster* Ait. recruitment in three burnt areas in Galicia (NW Spain). *Forest Ecology and Management*, 256(9):1596–1603, 2008. URL <http://www.sciencedirect.com/science/article/pii/S0378112708005628>.
- J. L. Wegrzyn, A. J. Eckert, M. Choi, J. M. Lee, B. J. Stanton, R. Sykes, M. F. Davis, C.-J. Tsai, and D. B. Neale. Association genetics of traits controlling lignin and cellulose biosynthesis in black cottonwood (*Populus trichocarpa*, Salicaceae) secondary xylem. *New Phytologist*, 188(2): 515–532, Oct. 2010. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=20831625&retmode=ref&cmd=prlinks>.
- E.-M. Willing, M. Hoffmann, J. D. Klein, D. Weigel, and C. Dreyer. Paired-end RAD-seq for de novo assembly and marker design without available reference. *Bioinformatics*, 27(16): 2187–2193, Aug. 2011. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=21712251&retmode=ref&cmd=prlinks>.
- A. Willyard, W. Ann, J. Syring, D. S. Gernandt, A. Liston, and R. Cronn. Fossil calibration of molecular divergence infers a moderate mutation rate and recent radiations for *pinus*. *Molecular Biology and Evolution*, 24(1):90–101, Jan. 2007. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=16997907&retmode=ref&cmd=prlinks>.
- A. Willyard, R. Cronn, and A. Liston. Reticulate evolution and incomplete lineage sorting among the ponderosa pines. *Molecular Phylogenetics and Evolution*, 52(2):498–511, Aug. 2009. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=19249377&retmode=ref&cmd=prlinks>.

- Z. Z. Yang. PAML 4: phylogenetic analysis by maximum likelihood. *Molecular Biology and Evolution*, 24(8):1586–1591, Aug. 2007. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=17483113&retmode=ref&cmd=prlinks>.
- N. You, G. Murillo, X. Su, X. Zeng, J. Xu, K. Ning, S. Zhang, J. Zhu, and X. Cui. SNP calling using genotype model selection on high-throughput sequencing data. *Bioinformatics*, 28(5): 643–650, Mar. 2012. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=22253293&retmode=ref&cmd=prlinks>.
- J. Yu, G. Pressoir, W. H. Briggs, I. V. Bi, M. Yamasaki, J. F. Doebley, M. D. McMullen, B. S. Gaut, D. M. Nielsen, J. B. Holland, S. Kresovich, and E. S. Buckler. A unified mixed-model method for association mapping that accounts for multiple levels of relatedness. *Nature Genetics*, 38(2):203–208, Feb. 2006. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=16380716&retmode=ref&cmd=prlinks>.
- Y. F. Zhou, R. J. Abbott, Z. Y. Jiang, F. K. Du, R. I. Milne, and J. Q. Liu. Gene flow and species delimitation: a case study of two pine species with overlapping distributions in south-east China. *Evolution*, 64(8):2342–2352, Aug. 2010. URL <http://eutils.ncbi.nlm.nih.gov/entrez/eutils/elink.fcgi?dbfrom=pubmed&id=20298431&retmode=ref&cmd=prlinks>.

Biographical Sketch: Christopher J. Friedline

I Professional preparation

Gettysburg College	Biology	B.A.	1999
Virginia Commonwealth University	Bioinformatics	P.S.M.	2008
Virginia Commonwealth University	Integrative Life Sciences	Ph.D.	2013

II Appointments

- Teaching Assistant, Department of Biology, Virginia Commonwealth University, 2008, 2011
 - Biological Concepts Laboratory (BIOZ 101), Fundamentals of Molecular Genetics (BIOL 540), Plant and Animal Cell Biology (BIOL 676)
- Teaching Assistant, Department of Life Sciences, Virginia Commonwealth University, 2008
 - Introduction to Life Sciences (LFSC 101), Integrated Bioinformatics (BNFO 601)
- Senior Network Analyst, VCU Health System, Richmond, VA, 2006–2007
- IT Systems Manager, Infralogics, Inc., Richmond, VA, 2005–2006
- IT Systems Administrator, Infineon Technologies, Sandston, VA, 2005
- Senior Systems Administrator, Circuit City Stores, Inc., Richmond, VA, 2002–2005
- NT Systems Administrator, Virginia Interactive, Richmond, VA, 2002
- Systems Administrator/Senior Systems Analyst/Systems Analyst, Bently Systems, Inc., Exton, PA, 1999-2002

III Publications

- **Friedline, C. J.**, Franklin, R. B., McCallister, S. L. & Rivera, M. C. Bacterial assemblages of the eastern Atlantic Ocean reveal both vertical and latitudinal biogeographic signatures. *Biogeosciences* 9, 2177-2193 (2012).
- Fettweis, J. M., Alves, J. P., Borzelleca, J. F., **Friedline, C. J.**, et al. The Vaginal Microbiome: Disease, Genetics and the Environment. *Nature Precedings* (2011).
- **Friedline, C. J.**, Zhang, X., Zehner, Z. & Zhao, Z. FindSUMO: A PSSM-Based Method for Sumoylation Site Prediction. *Advanced Intelligent Computing Theories and Applications With Aspects of Artificial Intelligence* 1004–1011 (2008).

IV Synergistic activities

- Invited to participate in two 2011 MBL courses: Workshop on Molecular Evolution and Strategies and Techniques for Analyzing Microbial Population Structures.
- Designed and implemented the informatic pipeline and data acquisition system for the VCU Human Vaginal Microbiome Project, currently supporting 1962 samples of 53 million 454 sequencing reads (23 billion base pairs). Data are housed in a PostgreSQL database and made available via a Java-based web portal.
- Developed quality control metrics and assembled the genomes of three antibiotic resistant *S. aureus* strains using Illumina reads. Comparative genomic analyses identified several SNPs associated with the resistant phenotype.
- Created custom software to visually and statistically analyze microarray data.
<http://sledride.csbc.vcu.edu>
- Responsible for the design and implementation of multi-million dollar, enterprise-class systems involving integrated technology stacks, multi-terabyte, n-tier architecture, and high-availability across many industry sectors including academic, medical, manufacturing, and retail.

V Collaborators and other affiliations

Collaborators and Co-Editors

- Rima B. Franklin, Department of Biology, Virginia Commonwealth University
- S. Leigh McCallister, Department of Biology/Center for Environmental Studies, Virginia Commonwealth University

Graduate Advisors

- Maria C. Rivera, Ph.D. Advisor, Center for the Study of Biological Complexity/Department of Biology, Virginia Commonwealth University
- Paul M. Fawcett, P.S.M. Advisor, Center for the Study of Biological Complexity/Department of Internal Medicine, Virginia Commonwealth University

Dissertation Abstract

Using Molecular Sequence Data to Unravel the Relationships between Bacteria and their Environment

Microbial communities are recognized as major drivers of global biogeochemical processes. However, the genetic diversity and composition, as well as processes leading to the origin and diversification of these communities in space and time, are poorly understood. Characterization of microbial communities using high-throughput sequencing of 16S tags shows that OTU abundances can be approximated by a gamma distribution, which suggests structuring around small numbers of highly abundant OTUs and a large proportion of rare OTUs. The current methods used to characterize how communities are structured rely on multivariate statistics, which operate on pair-wise distance matrices. My analyses demonstrate that use of these methods, by reducing a highly-dimensional dataset (tens of samples, thousands of OTUs), result in a significant loss of information (Friedline et al., 2012). I demonstrate that, in some cases, up to 80 % of the most abundant OTUs may be removed while still recovering the same community relationships; this indicates these metrics are biased toward the highly abundant OTUs. I will demonstrate that the observed patterns of OTU abundance detected from microbial communities can be properly modeled using techniques similar to those used to model the presence and absence of genes in genome evolution (Lake and Rivera, 2004; Rivera and Lake, 2004). Using simulation studies, I demonstrate that general Markov models in a Bayesian inference framework outperform traditional, multivariate ecological methods in recovering true community structure. Applying this new methodology to Atlantic Ocean microbial communities uncovered a distance-decay effect which was not revealed by the traditional methods (Friedline et al., 2012). Although the ocean dataset operated on a much larger, continental scale, characterization of the sequence data generated from the nutrient-poor soil on Hog Island, a barrier island off the Virginia Coast, allows for a better characterization of the processes affecting these communities on a much smaller scale. Finally, using 16S data from the Human Vaginal Microbiome Project, generated here at VCU under the umbrella of the overall NIH HMP initiative, I show that quality filtering has a profound effect on the reliability of downstream analysis. In conclusion, my analyses of the metagenomic sequence data from three types of bacterial communities demonstrate that the proper identification of the biological process influencing these communities requires the development and implementation of new statistical and computational methodology that takes advantage of the extensive amount of information generated by high-throughput sequencing.

Data Management Plan

The PI is committed to fostering an open and transparent process of data generation, analysis, and publication. As required in the NSF PGRP guidelines, genome level sequence data will be made available as soon as data quality and integrity have been verified via simple FTP sites hosted by sponsor Eckert. In addition, we will deposit data and computer code in standard public repositories prior to, and in all cases regardless of, publication (e.g., GenBank, Dryad, TreeGenes).

Data Types

Genetic Genetic sequence data from the Illumina platform will be stored as compressed FASTA and/or FASTQ files on disk. The quality-filtered, working set of sequence data will be stored in a relational database management system (RDBMS) built on PostgreSQL following an existing Rivera lab data storage model currently in production for the Human Vaginal Microbiome Project. Reads will be tracked through a barcoding system relating sequence data uniquely with individuals, populations, and additional metadata, as appropriate (e.g., sampling location, SNP genotypes, annotation, etc.). A copy of the processed, quality-controlled data will be extracted automatically on a schedule and compressed for distribution via FTP. Additionally, a web interface will be provided for dynamic data exploration.

Spatial and phenotypic All metadata will be stored in the same relational database system. This includes latitude and longitude for all sampling locations as well as phenotypic measurements and observations (e.g., height, bark thickness, diameter at breast height (DBH)).

Data Standards

All data, including data generated in downstream analysis, will be made available in a plain-text format. Working sequence data will be stored as text in the RDBMS, as well as associated environmental and phenotypic data. These data will be made available in two ways. First, as downloadable, compressed files available via FTP. Second, as dynamically generated tables available from a web interface. Downloadable data will be in tab-delimited format, with informative headers as appropriate. This format was chosen as it conforms to a standard used in programs such as R, as well as to any current or new custom Java and Python code developed in this project.

Policies for Data Access and Sharing

Genomic data will be released following the Bermuda/Ft. Lauderdale agreement and will also be deposited in GenBank once data quality is sufficient for release. The remaining data will be released after publication in the text and web formats as described above, along with a copy of the publication, and will be posted on the Eckert laboratory web server. All relevant code and schemas used to process, store, and analyze these data will be made available under an open source license and distributed via the web (e.g., bitbucket.org). There will be no charge for these data.

Provisions for Appropriate Protection and Privacy

There are no known privacy or ethical issues associated with the data collected on this grant. There are no known copyright, licensing, or other intellectual property issues associated with these data.

Policies and Provisions for Re-use and Re-distribution

The data will not contain any permission restrictions beyond citation of our original publication(s) where the data were introduced. The genetic data will be of potential interest to the landscape genomic community (both professional and academic) for subsequent meta-analysis, statistical model development, and other summary uses. The PI does not foresee any reason to restrict the re-use of these data to any entity or group.

Plans for Archiving and Preservation of Access

Raw DNA template will be stored at VCU in the Eckert Laboratory in 1.5ml Eppendorf tubes at -80°C for least as long as it takes to publish all the materials associated with this project. Individual primers will not be made available, though published sequences can be used to create them de novo if necessary. Textual genetic data will be made available, via the web or some other publicly available medium as described above, for as long as the PI is engaged in academic research. Unprocessed leaf tissue samples will be similarly archived until publication, with raw physiological data (e.g. in the case of parameters derived from light-response curves) archived and available to the public following publication. For the foreseeable future, these data will be housed on the servers located within the Eckert Laboratory. All final data will be automatically backed up to VCU common spaces. There are no known issues associated with storing textual genetic data or anonymizing the identification numbers on these data sets.