

PIrANHA

Phylogenetics AND phylogeography



code quality **A** license GPL (≥ 2)

Scripts for file processing and analysis in phylogenomics & phylogeography

LICENSE

All code within the `PIrANHA` v0.1.7 repository is available "AS IS" under a generous GNU license. See the [LICENSE](#) file for more information.

CITATION

If you use scripts from this repository as part of your published research, I require that you cite the repository as follows (also see DOI information below):

- Bagley, J.C. 2019. PIrANHA v0.1.7. GitHub repository, Available at: <http://github.com/justincbagley/PIrANHA>.

Alternatively, please provide the following link to this software repository in your manuscript:

- <https://github.com/justincbagley/PIrANHA>

DOI

The DOI for `PIrANHA`, via [Zenodo](#), is as follows: `DOI 10.5281/zenodo.890815`. Here are some examples of citing PIrANHA using the DOI:

Bagley, J.C. 2019. PIrANHA v0.1.7. GitHub package, Available at: <http://doi.org/10.5281/zenodo.596766>.

Bagley, J.C. 2019. PIrANHA. Zenodo, Available at: <http://doi.org/10.5281/zenodo.596766>.

INTRODUCTION

Taking steps towards automating boring stuff during analyses of genetic data in phylogenomics & phylogeography...

`PIrANHA` v0.1.7 is a repository of shell scripts and R scripts written by the author, as well as additional code (R, Perl, and Python scripts) from other authors, that is designed to help automate processing and analysis of DNA sequence data in phylogenetics and phylogeography research projects (Avisé 2000; Felsenstein 2004). PIrANHA is fully command line-based and, rather than being structured as a single pipeline, it contains a series of scripts, some of which form pipelines, for aiding or completing tasks during evolutionary analyses of genetic data. Currently, PIrANHA scripts facilitate running or linking the following software programs:

- `pyRAD` (Eaton 2014) or `ipyrad` (Eaton and Overcast 2016)
- `PartitionFinder` (Lanfear et al. 2012, 2016)

- **BEAST** (Drummond et al. 2012; Bouckaert et al. 2014)
- **starBEAST** (Heled & Drummond 2010)
- **SNAPP** (Bryant et al. 2012)
- **MrBayes** (Ronquist et al. 2012)
- **ExaBayes** (Aberer et al. 2014)
- **RAxML** (Stamatakis 2014)
- **ðaði** (Gutenkunst et al. 2009)
- **fastSTRUCTURE** (Raj et al. 2014)
- **PhyloMapper** (Lemmon and Lemmon 2008)

The current code in **PIrANHA** has been written largely with a focus on 1) analyses of DNA sequence data and SNPs or SNP loci generated from massively parallel sequencing runs on ddRAD-seq genomic libraries (e.g. Peterson et al. 2012), and 2) automating running these software programs on the user's personal machine (e.g. **MAGNET** pipeline and **pyRAD2PartitionFinder** scripts) or a remote supercomputer machine, and then conducting post-processing of the results. In particular, a number of scripts have been written with sections allowing them to be run (or cause other software to be called) on a supercomputing cluster, using code suitable for **SLURM** or **TORQUE** (PBS; **Portable Batch System**) resource management systems (in some cases, this functionality is noted by adding "Super" in the script filename, as in **Super-pyRAD2PartitionFinder.sh**).

Distribution Structure and Pipelines

What's new in this release?

Some of the changes included in the current official release are detailed below. However, refer to [changeLog.md](#) for a full listing of updates to this repository since the last release (**PIrANHA** v0.1.6.1).

PIrANHA v0.1.7 (official minor version release) - February 19, 2019

- **February 19, 2019:** Improved **phylipSubsampler.sh** to check and account for differences in machine type. Now correctly accommodates UNIX (Mac) and LINUX systems.
- **February 2019:** Updated **MAGNET** script by adding a **getBipartTrees** function to the **MAGNET** pipeline, which organizes **RAxML** bipartitions trees for each locus (= best ML trees with bootstrap proportions along nodes the corresponding bootstrap searches search; resulting from **-f a -x** options, which are included in all **MAGNET** calls to **RAxML**). Edited header and script banner to be prepped for future official release of **MAGNET** with versioning 0.1.9.
- **December 2018:** Added new **MAGNET** script updated to include **--resume** option, and to set **raxml** executable name one of two ways after detecting machine type (**raxml** on Mac, **raxmlHPC-SS3** on Linux/supercomputer).
- **November 25, 2018:** Added to **MAGNET/shell** folder a new 'RAxMLRunChecker.sh' script v1.0, which counts the number of completed **RAxML** runs during the course of, or after, a **MAGNET** pipeline run, and also collates information on the dataset (e.g. number of patterns) and run (e.g. run time, optimum likelihood) for each locus/partition.
- **November 20, 2018 bug fix:** Updated **MAGNET** with edited 'MAGNET.sh' (now v0.1.7+) and 'NEXUS2gphocs.sh' (now v1.3+) scripts containing an important bug fix and some new code checking for whether the **NEXUS** to **fasta** file conversion succeeded.

What is possible with PIRANHA? Who cares?

How PIRANHA scripts work together

The goal of **PIrANHA** is to facilitate analysis pipelines that could be of interest to nearly anyone conducting evolutionary analyses of DNA sequence data using maximum-likelihood and Bayesian methods. **Figure 1** and **Figure 2** below demonstrate flow and interactions of the current partition scheme, population structure, and phylogenetics pipelines with **software** and **"file types"** used to generate input for PIrANHA in the left column, and the way these are processed within/using **PIrANHA** illustrated in the right column. External software programs are shown in balloons with names in black italic font, while **PIrANHA** scripts are given in blue. Arrows show the flow of files through different pipelines, which terminate in results (shown right of final arrows at far right of each diagram).

PIrANHA facilitates going from pyRAD “out-of-the-box” to evolutionary analyses

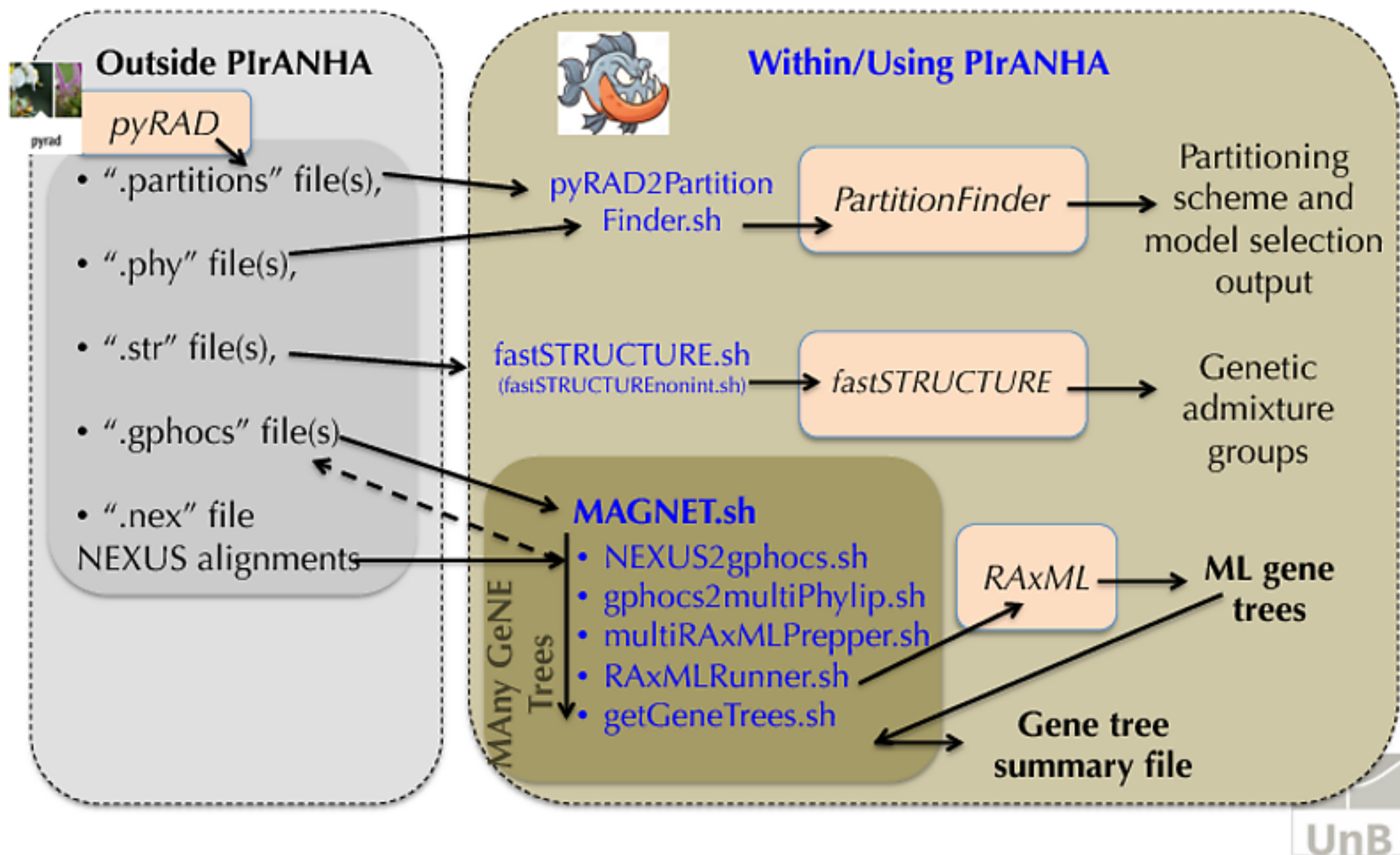


Figure 1

Bayesian phylogeny and divergence time analyses assisted with PIRANHA

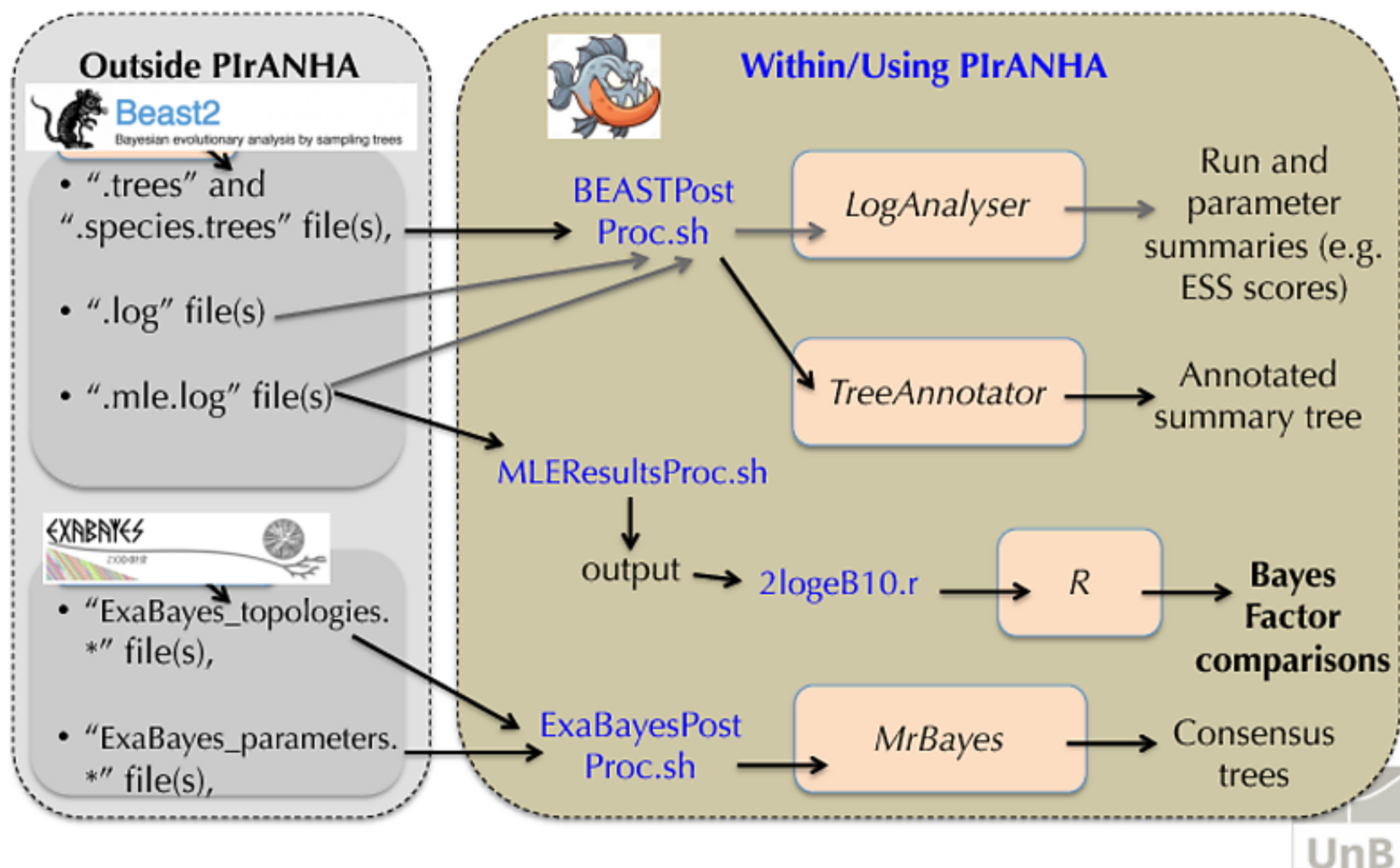


Figure 2

The following **Figure 3** illustrates new capacities of running and processing *daði* (Gutenkunst et al. 2009) files in PIRANHA (note: the post-processing script is still under development).

Whole-genome SNPs demographic modeling analyses assisted with PIRANHA

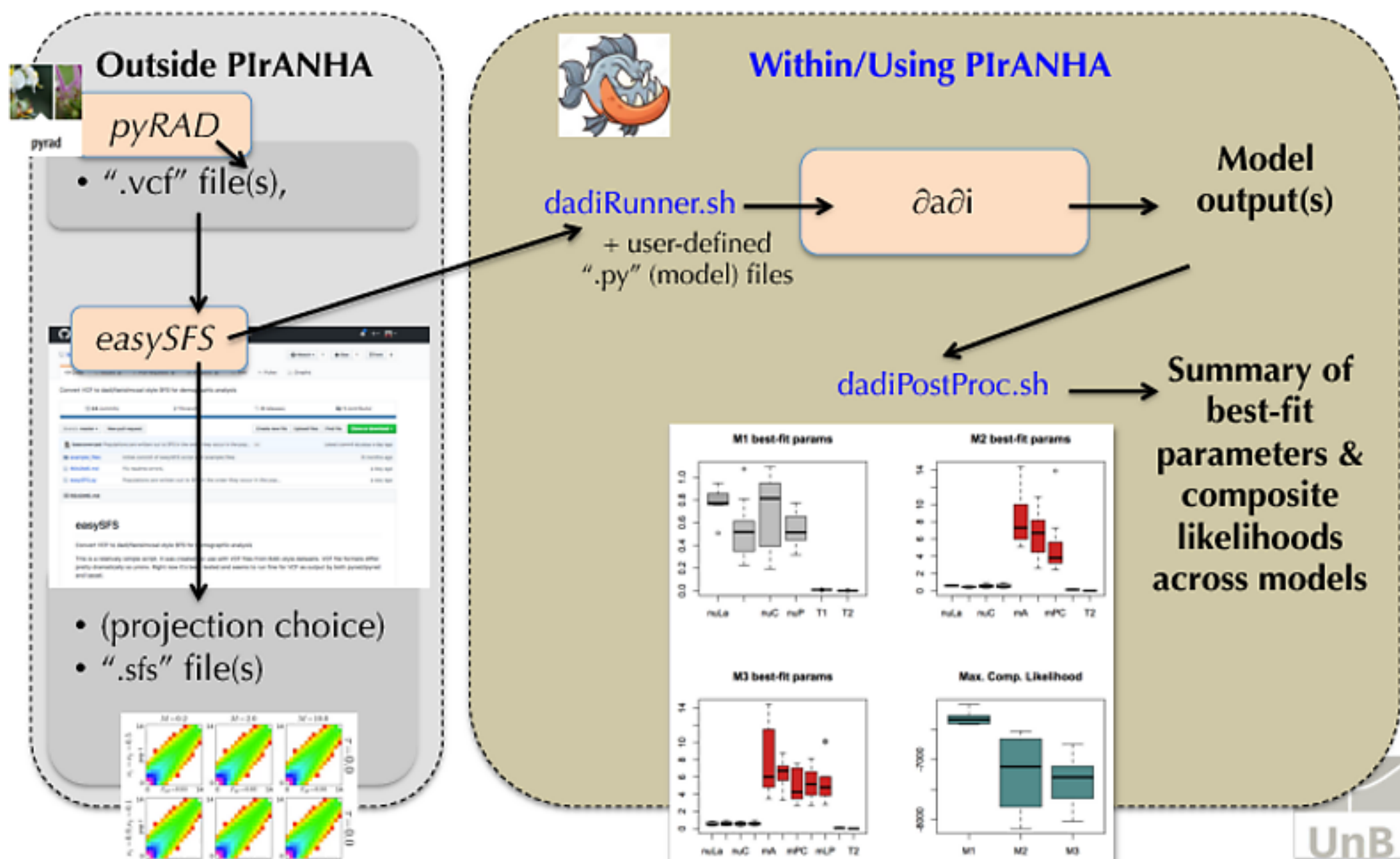


Figure 3

GETTING STARTED

Dependencies


PIRANHA, and especially the MAGNET package ([here](#) or [here](#)) within PIRANHA, relies on several software dependencies. These dependencies are described in some detail in README files for different scripts or packages; however, I provide a full list of them below, with asterisk marks preceding those already included in the "MAGNET-0.1.9" subdirectory of the current release. Of course, you can usually get away with not installing dependencies or software that are unrelated to the analysis you are conducting with PIRANHA, but it is recommended that you install all dependencies to take full advantage of PIRANHA's capabilities, or be prepared for any analysis!

- PartitionFinder
- BEAST v1.8.3++ and v2.4.2++ (or newer; available at: <http://beast.community> and <http://beast2.org>, respectively)
 - Updated Java, appropriate Java virtual machine / jdk required
 - BEAGLE in beagle-lib (libhmsbeagle * files) required
 - default BEAST packages required
 - SNAPP package addon required
- MrBayes v3.2++ (available at: <http://mrbayes.sourceforge.net/download.php>)
- ExaBayes (available at: <http://sco.h-its.org/exelixis/web/software/exabayes/>)
- RAXML (available at: <http://sco.h-its.org/exelixis/web/software/raxml/index.html>)
- Perl (available at: <https://www.perl.org/get.html>).

- *Nayoki Takebayashi's file conversion Perl scripts (available at: <http://raven.iab.alaska.edu/~ntakebay/teaching/programming/perl-scripts/perl-scripts.html>; note: some, but not all of these, come packaged within MAGNET)
- Python v2.7 and/or 3++ (available at: <https://www.python.org/downloads/>)
 - Numpy (available at: <http://www.numpy.org/>)
 - Scipy (available at: <http://www.scipy.org/>)
 - Cython (available at: <http://cython.org/>)
 - GNU Scientific Library (available at: <http://www.gnu.org/software/gsl/>)
 - bioscripts.convert v0.4 Python package (available at: <https://pypi.python.org/pypi/bioscripts.convert/0.4>; also see README for 'NEXUS2gphocs.sh')
- fastSTRUCTURE v1.0 (available at: <https://rajanil.github.io/fastStructure/>)
- dadi v1.7.0++ (or v1.6.3; available at: <https://bitbucket.org/gutenkunstlab/dadi/overview>)
- R v3++ (available at: <https://cran.r-project.org/>)

Users must install all software not included in PIRANHA , and ensure that it is available via the command line on their supercomputer and/or local machine (best practice is to simply install all software in both places). For more details, see the MAGNET README.

Installation

 As PIRANHA is primarily composed of UNIX shell scripts and customized R scripts, it is well suited for running on a variety of types of machines, especially UNIX/LINUX-like systems that are now commonplace in personal computing and dedicated supercomputer cluster facilities. The UNIX shell is common to all Linux systems and mac OS X. Using PIRANHA is thus very straightforward, because these systems come with the shell preinstalled. To install PIRANHA , simply clone this repository and modify permissions, as follows:

```
$ cd ~
$ git clone https://github.com/justincbagley/PIrANHA.git
$ cd ./PIrANHA/
$ chmod u+x ./*.sh; chmod u+x ./*/*.sh; chmod u+x ./*/*.pl
```

After doing this, PIRANHA should run "out-of-the-box" from most any folder on your machine. 🍌

IMPORTANT! - Passwordless SSH Access


PIrANHA largely focuses on allowing users with access to a remote supercomputing cluster to take advantage of that resource in an automated fashion. Thus, it is implicitly assumed in most scripts and documentation that the user has set up passwordless ssh access to a supercomputer account.

👉 If you have not done this, or are unsure about this, then you should set up passwordless acces by creating and organizing appropriate and secure public and private ssh keys on your machine and the remote supercomputer prior to using PIRANHA. By "secure," I mean that, during this process, you should have closed write privledges to authorized keys by typing "chmod u-w authorized keys" after setting things up using ssh-keygen.



❗ Setting up passwordless SSH access is **VERY IMPORTANT** as PIRANHA scripts and pipelines will not work without setting this up first. The following links provide a list of useful tutorials/discussions that can help users set up passwordless SSH access:

- <https://www.msi.umn.edu/support/faq/how-do-i-setup-ssh-keys>
- <https://coolestguidesontheplanet.com/make-passwordless-ssh-connection-osx-10-9-mavericks-linux/>
- <https://www.tecmint.com/ssh-passwordless-login-using-ssh-keygen-in-5-easy-steps/>

Input and Output File Formats

 PIRANHA scripts accept a number of different input file types, which are listed in Table 1 below. These can be generated by hand or are output by specific upstream software programs. As far as *output file types* go, PIRANHA outputs various text, PDF, and other kinds of graphical output from software that are linked through PIRANHA pipelines.

Input file types	Software (from)
.partitions	pyRAD / ipyrad
.phy	pyRAD / ipyrad / by hand
.str	pyRAD / ipyrad
.gphocs	pyRAD / ipyrad / MAGNET (NEXUS2gphocs.sh)
.loci	pyRAD / ipyrad
.nex	pyRAD / ipyrad / by hand
.trees	BEAST
.species.trees	BEAST
.log	BEAST
.mle.log	BEAST
.xml	BEAUti
.sfs	easySFS
Exabayes_topologies.*	ExaBayes
Exabayes_parameters.*	ExaBayes

 **NOTE: The following 'Getting Started' content is Under Construction! E-mail me about it, or check back soon for updates!** 

Phylogenetic Partitioning Scheme/Model Selection

pyRAD2PartitionFinder

Shell script for going directly from PHYLIP alignment (.phy) and partitions (.partisions) files output by pyRAD (Eaton 2014) or ipyrad (Eaton and Overcast 2016; for de novo assembly of reduced-representation sequence data from an NGS experiment) to inference of the optimal partitioning scheme and models of DNA sequence evolution for pyRAD -defined SNP loci in PartitionFinder (Lanfear et al. 2012, 2016). See current release of pyRAD2PartitionFinder [scripts](#) for more information (e.g. detailed comments located within the code itself; a README is coming soon).

Estimating Gene Trees for Species Tree Inference

MAGNET (MAny GeNE Trees)

Interactive shell pipeline for inferring maximum-likelihood gene trees in `RAXML` (Stamatakis 2014) for multilocus DNA sequence alignments (e.g. RAD loci from ddRAD-seq experiments, candidate genes, genomic contigs) to aid downstream summary-statistics species tree inference. Please see the [README](#) for the `MAGNET` Package, which is available as its own stand-alone repository so that it can be tracked and continually given its own updated doi and citation by Zenodo. Three starting input file formats are currently supported, including single NEXUS (.nex), single `G-PhoCS` (.gphocs; formatted for `G-PhoCS` software, Gronau et al. 2011), and multiple `PHYLIP` files.

Automating Bayesian evolutionary analyses in BEAST

BEASTRunner

[BEASTRunner](#) automates conducting multiple runs of `BEAST` v1 or v2 (Drummond et al. 2012; Bouckaert et al. 2014) XML input files on a remote supercomputing cluster that uses SLURM resource management with PBS wrappers, or a TORQUE/PBS resource management system. See the `BEASTRunner` [README](#) for more information.

BEAST_PathSampling

The `BEAST_PathSampling` directory is a new area of development within `PIrANHA` in which I am actively coding scripts to (1) edit `BEAST` v2++ XML files for path sampling analyses (Xie et al. 2011; Baele et al. 2012) and (2) automate moving/running the new path sampling XML files on a supercomputing cluster. Even as of August 2017, this is *very new stuff that is experimental and may still not be working*, so stay tuned for more updates soon.

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REFERENCES

- Aberer AJ, Kobert K, Stamatakis A (2014) ExaBayes: massively parallel Bayesian tree inference for the whole-genome era. *Molecular Biology and Evolution*, 31, 2553-2556.
- Avice JC (2000) *Phylogeography: the history and formation of species*. Cambridge, MA: Harvard University Press.
- Baele G, Lemey P, Bedford T, Rambaut A, Suchard MA, Alekseyenko AV (2012) Improving the accuracy of demographic and molecular clock model comparison while accommodating phylogenetic uncertainty. *Molecular Biology and Evolution*, 29, 2157-2167.
- Bouckaert R, Heled J, Kühnert D, Vaughan TG, Wu CH, Xie D, Suchard MA, Rambaut A, Drummond AJ (2014) BEAST2: a software platform for Bayesian evolutionary analysis. *PLoS Computational Biology*, 10, e1003537.
- Bryant D, Bouckaert R, Felsenstein J, Rosenberg NA, RoyChoudhury A (2012) Inferring species trees directly from biallelic genetic markers: bypassing gene trees in a full coalescent analysis. *Molecular Biology and Evolution*, 29, 1917–1932.
- Eaton DA (2014) PyRAD: assembly of de novo RADseq loci for phylogenetic analyses. *Bioinformatics*, 30, 1844-1849.
- Eaton DAR, Overcast I (2016) ipyrad: interactive assembly and analysis of RADseq data sets. Available at: <http://ipyrad.readthedocs.io/>.
- Drummond AJ, Suchard MA, Xie D, Rambaut A (2012) Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Molecular Biology and Evolution*, 29, 1969-1973.
- Felsenstein J (2004) *Inferring phylogenies*. Sunderland, MA: Sinauer Associates.
- Gronau I, Hubisz MJ, Gulko B, Danko CG, Siepel A (2011) Bayesian inference of ancient human demography from




individual genome sequences. Nature Genetics, 43, 1031-1034.

- Heled J, Drummond AJ (2010) Bayesian inference of species trees from multilocus data. Molecular Biology and Evolution, 27, 570–580.
- Lanfear R, Calcott B, Ho SYW, Guindon S (2012) PartitionFinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. Molecular Biology and Evolution, 29,1695-1701.
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B (2016) PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. Molecular Biology and Evolution.
- Lemmon AR, Lemmon E (2008) A likelihood framework for estimating phylogeographic history on a continuous landscape. Systematic Biology, 57, 544–561.
- Peterson BK, Weber JN, Kay EH, Fisher HS, Hoekstra HE (2012) Double digest RADseq: an inexpensive method for de novo SNP discovery and genotyping in model and non-model species. PLoS One, 7, e37135.
- Raj A, Stephens M, and Pritchard JK (2014) fastSTRUCTURE: Variational Inference of Population Structure in Large SNP Data Sets. Genetics, 197, 573-589.
- Ronquist F, Teslenko M, van der Mark P, Ayres D, Darling A, et al. (2012) MrBayes v. 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology, 61, 539-542.
- Stamatakis A (2014) RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics, 30, 1312-1313.
- Xie W, Lewis PO, Fan Y, Kuo L, Chen MH (2011) Improving marginal likelihood estimation for Bayesian phylogenetic model selection. Systematic Biology, 60, 150-160.

RECOMMENDED READING

- Unix shell background info [here](#), [here](#), [here](#), and [here](#).
- GNU [Bash Reference Manual](#)

TODO

- **Improve option and usage sections of `BEASTRunner.sh` script. DONE! **
- **Add `MrBayes` scripts DONE! **
- **Add options to `MrBayesPostProc` script, e.g. for burnin frac and stepping-stone MLE analysis. DONE! **
- Change new `phylipSubsampler` script so that it accounts for missing data (currently does not).
- Give supercomputer scripts options (header w/flags) that will work for both a) TORQUE/PBS and b) SLURM Workload Manager cluster management and job scheduling systems (need meticulous work on this in `Super-pyRAD2PartitionFinder.sh` , `BEASTRunner.sh` , `BEASTPostProc.sh` , and `RAxMLRunner.sh`)
- Add mrbayes and beast model options to `pyrad2PartitionFinder.sh` and supercomputer version of the same script.
- Make `pyrad` and `ipyrad` batch run scripts available
- Consider separate scripts to work with ipyrad
- Add capacity of adding or not adding path sampling/stepping-stone sampling to `BEAST` runs (use `BEASTRunner.sh` as springboard to develop tools in BEAST_PathSampling dir)

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