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The Myxovirus Resistance A (MxA) Gene —88G>T Single Nucleotide Polymorphism Is Associated with Prostate Cancer

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Abstract

Background—Myxovirus (influenza virus) resistance A (MxA) is an interferon stimulated antiviral protein that is required for a complete antiviral response. MxA polymorphism (rs2071430) is located within an Interferon Stimulated Response Element (ISRE) at position –88 in the gene's promoter region, and it has been associated with increased susceptibility to infections and various diseases. In general, the low promoter activity genotype (GG) promotes susceptibility, whereas the high promoter activity genotype (TT) confers protection to Hepatitis C viral infection. MxA's role in prostate cancer is not fully understood. Previous literature has shown that MxA may be a mediator of the effect of IFN on normal and tumor cell motility. MxA may act as a tumor suppressor and the level of expression may be a predictor of metastatic potential. Based on this information, in this study we investigated the association of this functional polymorphism (rs2071430) in MxA with prostate cancer.

Methods—Sample size and power was calculated using the PGA software. Genomic DNA from a controls (n=140) and prostate cancer patients (n=164) were used for genotyping SNP rs2071430 on all samples. Statistical analysis was performed using logistic regression model.

Results—A significant association was observed between rs2071430 genotype GG and prostate cancer. Individuals harboring the GG genotype are at an increased risk of prostate cancer. Data stratification reveals that the mutant GT genotype offers either offers some protection against prostate cancer in Caucasians.

Conclusions—MxA SNP rs2071430 GG genotype is significantly associated with prostate cancer irrespective of race. However, data stratification also suggests that the GT genotype is under-represented in Caucasian subjects suggesting its role in protection against prostate cancer in Caucasians. Although MxA is primarily implicated in viral infection, but it may be also be associated with prostate cancer. Recent studies have implicated viral and bacterial infections with increased prostate cancer risk. Expression of the high promoter activity genotype may offer resistance to prostate cancer infection and possibly influence clinical outcomes.

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Keywords

MxA; Prostate; Cancer; polymorphism; rs2071430

1. Introduction

The human Myxovirus (influenza virus) resistance 1 gene encodes for the human MxA protein (76Kda) a cytoplasmic protein that is rapidly induced in response to acute viral infections (Roers et al., 1994). MxA belongs to the family of Mx proteins which are among the few effector proteins of the IFN a/b system with known antiviral activity. The Mx proteins are abundantly expressed in interferon-treated cells and play a crucial role in the early antiviral defense against certain RNA viruses which has been demonstrated in studies with transgenic mice also (Arnheiter et al., 1990; Hefti et al., 1999; Pavlovic et al., 1995). Human MxA is shown to inhibit multiplication of several ssRNA and dsRNA viruses including influenza, vesicular stomatitis virus (VSV) (Pavlovic et al., 1990), measles virus (Schnorr et al., 1993), Thogoto virus (Frese et al., 1995), Bunyavirus (Frese et al., 1996), and Semliki forest virus (Landis et al., 1998). MxA has also been shown to confer resistance to some DNA viral genomes (Gordien et al., 2001) (Netherton et al., 2009). In general, MxA appears to interfere with steps involving viral transcription although the exact mechanism of action is not known or fully understood.

MxA is a highly conserved large GTPase with additional homology to dynamin superfamily and have been found in all vertebrates species investigated so far, including mammals, birds, and fish (Staeheli et al., 1993). These proteins present a highly conserved N-terminal GTPase domain of ~300 amino acids, a central interactive domain of ~150 amino acids, and a GTPase effector domain (GED) of ~100 amino acids which encompasses two leucine zippers that have the capacity to form α-helices. The MxA GTPase domain has a low affinity for GTP but high GTP turnover rate which is indicative of efficient cycling achieved only within dense protein complexes typically assembled on the membrane surface (Goryachev and Pokhilko, 2006; Melen et al., 1994; Richter et al., 1995). Studies with human MxA protein have shown that the GED effector domain is able to specifically contact the middle domain, and that this interaction is critical to constitute a functional GTPase domain, as well as for oligomerization (Gao et al., 2010; Haller and Kochs, 2011) and antiviral functions (Di Paolo et al., 1999; Schumacher and Staeheli, 1998).

MxA has over 590 polymorphisms in its genome (NCBI dbSNP). It has previously been reported that a SNP at nucleotide position –88 (G or T) in the promoter region of the gene modulates MxA function at multiple levels including expression. MxA mRNA is significantly up-regulated with the mutant "T" (TT or GT) allele as compared to the wild type homozygous "G" allele (Fernandez-Arcas et al., 2004; Furuyama et al., 2006; Hijikata et al., 2001).

This functional polymorphic marker (rs2071430) is well studied and demonstrates an association with various diseases such as susceptibility to hepatitis C virus (Hijikata et al., 2000), SARS (Hamano et al., 2005), and sub-acute sclerosing panencephalitis (Torisu et al., 2004). Interestingly, an interferon stimulated response element (ISRE) is located around the single nucleotide polymorphism (rs2071430) in the promoter region and can also influence MX1 promoter activity. The T allele restores and the G allele attenuates ISRE binding due to sequence homology (Hijikata et al., 2000; Nakade et al., 1997). Functional promoter studies using MxA luciferase reporter assay has also demonstrated that promoter activity is highest in individuals with TT genotype and lowest in the GG genotype (Hijikata et al., 2001; Torisu et al., 2004).

The role of MxA in prostate cancer is not fully understood. A previous study (Mushinski et al., 2009) has implicated MxA as a potential metastasis suppressing gene in prostate cancer suggesting that its expression could be a key indicator of metastatic potential. We hypothesized that the functional polymorphism rs2071430 leading to attenuated MxA expression could be a genetic risk factor for prostate cancer. Our results demonstrate that rs2071430 polymorphism is associated with prostate cancer and that a specific rs2071430 genotype (GG) may confer an increased susceptibility to prostate cancer.

2. Materials and Methods

2.1 Samples

The genotyping protocol and use of human samples in the study were approved by IRB at Clark Atlanta University. Genotyping was performed on a total of 304 samples. The numbers of cancer and normal samples were approximately 50% each of the total sample set (Table I). Retrospectively collected Buffy coat samples were obtained from Bio-specimen Shared Resource, KU Cancer Center, University of Kansas Medical Centre and Cooperative Human Tissue Network (CHTN, Southern Division) following appropriate protocol review and approval. The purified genomic DNA samples were obtained commercially from BioServe Inc. (Beltsville, MD). All samples were stored at -80 until analysis. De-identified comprehensive clinical information regarding age, ethnicity and stage was available for all samples. However, family history of prostate cancer, PSA level and Gleason score was not available for all samples hence were not included in final statistical analysis.

2.2. DNA Isolation

Genomic DNA was isolated (and stored at -80C) from cultured cells or buffy coat using AquaPure total genomic DNA isolation kit (Bio-Rad). On an average approximately 30ug of DNA was routinely isolated from 300ul of buffy coat.

2.3. Genomic PCR

Genomic PCR was carried out in a 25ul PCR reaction that consisted of 12.5ul GoTaq Colorless Master Mix (Promega), 30ng genomic DNA and 400pm of 5' and 3' primer each. The PCR was carried out for 35 cycles with annealing temperatures 52.9C for rs2071430. The following primers were used rs2071430: Forward 5'-TGT ATA CCT GCA AGT CAC AGG, Reverse 5'-TGT TAG TTA CTA GCA GCC GAG, nested primer for rs2071430: 5'-GAG CAC CTT GAT CCT CAG AC.

2.4. SNP detection

The rs2071430 polymorphism was detected by sequencing the PCR amplicon spanning the above stated SNP using the primer pairs indicated above. An aliquot of the respective PCR reaction was first analyzed on 1.5% agarose gel to confirm specificity and quality of the reaction in terms of band size and absence of any background PCR product. Once confirmed, the remaining PCR product was cleaned using ExoSAP-IT (USB) before sequencing on the AB sequencer (DNA sequencing Lab, Morehouse School of Medicine, Atlanta, GA). The sequencing was performed using a nested primer within the rs2071430 PCR product. The sequences were scanned using ABI sequence scanner and the SNPs were manually detected. The amplicons showing low quality reads were re-sequenced. The single nucleotide polymorphisms were also detected by SNPdetctor (Zhang et al., 2005) to ensure that the SNPs were due to heterozygous allelic variations and not due to sequencing artifacts.

2.5. Statistical analysis

Each of the polymorphisms was tested for association with prostate cancer. Odds ratio and 95% confidence intervals were calculated for the genotype in association with prostate cancer using logistic regression analysis with adjustment for age. Relationship between genotype distribution with subjects stratified with race and age were also analyzed using the above model. NPSS version 2007, version 07.1.19 and SigmaStat v3.5 was used for statistical analysis. The sample size calculations and power were calculated using the "Power for Genetic Association Analyses" (PGA) package in Matlab (available on the National Cancer Institute website) (Menashe et al., 2008) using following parameters: 90% power, alpha=0.05, cancer prevalence of 250/100,000 men (actual for all races, white and black is 156.9, 150.4 and 234.6 per 100,000 men respectively as per SEER), minor and major allele frequencies shown in table III (excluding TT allele), case control ratio of 1 and relative risk of prostate cancer in African American men as 2.5 as compared to Caucasians. Using these calculations the sample size was approximately 110 samples (±23). This sample size was achieved in this study.

3. Results

3.1 Samples Demographics

A case-control study was performed to investigate rs2071430 genotype on 304 samples collected retrospectively. The samples were distributed equally (p=0.73) between cases (N=164, 53.9%) and controls (N=140, 46.1%). The mean age of cancer samples was 63.7 ± 0.746 years, and the mean age of normal samples was 60.2 ± 1.3 years. Both these groups were considered age matched (p = 0.273) (Table I).

Comprehensive statistical analysis based on Chi² analysis indicated the lack of any bias in the incidence of prostate cancer among the racial and age groups (Caucasians (53.3%) and African Americans sample sets (46.7%, Chi²=0.61).

3.2 MxA SNP rs2071430 in population

We first wanted to understand the population distribution of the rs2071430 genotype in the normal population published in NCBI dbSNP database. The results showed that Caucasian and Hispanic subjects lack the homozygous TT genotype (Table II). The TT genotype was observed at low frequency in African Americans (4.2%) whereas in the subjects with Pacific Rim heritage, the TT genotype frequency was highest at 16.7%. These results clearly suggested a strong racial distribution of the minor TT genotype (Table II). In the normal population, the GG genotype in NCBI database was 70% in African American (n=24) but 50% in our dataset (n-60). A recent study (Duc et al., 2012) the MxA GG genotype distribution in African population (consisting of subjects from Libya, Cameroun, Niger or Rwanda, none from African American background) was 80%. The genotype distribution in our data set and those reported elsewhere (e.g. Table II) could be due to sample size and ethnic background (Duc et al., 2012).

3.3 Sample set frequency of rs2071430 polymorphism

The frequency of rs2071430 genotype in our normal mixed race sample set (58.6% (GG), 35.7% (GT) and 5.7% (TT)) was nearly consistent with those reported for other heterogeneous control populations (NCBI dbSNP database, Table II, PI dataset: 72.5% (GG), 22.5% (GT) and 5% (TT). In our study, the rs2071430 allele frequency also conformed to Hardy-Weinberg equilibrium in the African American population (chi-square 2.23, df=2, p=0.11). There was a marked deviation from Hardy-Weinberg equilibrium in Caucasian sample set due to lack of TT genotype. The frequency distribution of rs2071430

in our complete sample set and samples stratified by race are listed in Tables III and IV respectively.

3.4. rs2071430 distribution in the sample and its association with prostate cancer

Each one of the genotypes was tested for its association with prostate cancer for all samples and in samples stratified by race. The combined cancer and control groups revealed that GG was a major (dominant) genotype (65.8%), whereas TT was a minor genotype (3.9%, Table III). The heterozygous genotype GT (frequency: 30.3%) was used as a reference to calculate the association of each genotype with prostate cancer. The odds of having cancer with GG genotype was found to be 71% higher (CI=1.041 – 2.818) than the odds with GT (reference) heterozygous genotype. On the other hand, the odds of the homozygous TT genotype decreased the risk of prostate cancer by 40% but was not statistically significant (CI= 0.167–2.116). Alternatively, the GG genotype was over represented in the cancer case group (72%) as compared to the control group (58.6%) while the TT genotype was overrepresented in the control group (5.7%) compared with the cancer group (2.4%, OR=0.6, Table III).

3.5. Association of rs2071430 with prostate cancer and Race

Sample stratification demonstrated that rs2071430 genotype distribution is associated with race. The MxA genotype frequencies in the Caucasian subpopulation were: 72%(GG), 27% (GT) and 1.2%(TT). The corresponding MxA genotype frequencies for the African American subpopulation were: 58.6%(GG), 34%(GT) and 7.1% (TT). The genotype frequencies in both these racial sub-populations tested showed statically significant difference (chi-square 14.3, p=0.0021). The results clearly demonstrated that GG genotype is over-represented in the Caucasian population whereas the heterozygous GT and homozygous TT genotype is over-represented in the African American population. These observation prompted us investigate the association of each genotype with cancer in African-American and Caucasian sub-populations. Although, GG genotype was associated with prostate cancer overall and over-represented in Caucasians as stated above but no significant race specific association was observed with cancer (OR 1.37, CI 0.769 – 2.442).

The distribution of GT genotype (used as reference to calculate overall risk, Table III) displayed strong association with African American as compared to Caucasians (OR 2.07, CI 0.897 – 4.780). The GT genotype was not associated with increased risk of cancer in African American (cancer 28.2% vs. normal 24%. Surprisingly, the GT genotype showed strong association with Normal Caucasians (30.4%) as compared to cancer Caucasians (17.4%). The odds ratio of cancer incidence in the Caucasian group with GT genotype was 3.1 (p=0.01) implying that the odds for cancer case is lower in Caucasians with a GT genotype. Unlike the African American subset, the TT genotype was absent in the Caucasian normal group but was observed in the cancer group in both racial sub-groups (16.6%). However, based on low frequency, the TT genotype showed no statistical significance in terms of its association with the disease and between racial sub-groups. No significant association with stage, Gleason grade and MxA rs2071430 was observed (data not shown).

4. Discussion

Together with other interferon inducible anti-viral genes such as oligoadenylate synthetase 1 (OAS1) and PKR, MxA (Sadler and Williams, 2008) polymorphism has also been shown to be associated with specific diseases for example multiple sclerosis and viral infections such as influenza and SARS. In mice, type I IFN is effective against influenza viruses only if the IFN-induced resistance factor Mx1 is present (Koerner et al., 2007).

Our previous study also demonstrated a strong association of OAS1 polymorphism with prostate cancer with racial disparities (Mandal et al., 2011). The OAS1 and prostate cancer association study prompted to ask the question whether polymorphisms in other classical IFN inducible anti-viral genes such as MxA are also common risk factors in prostate cancer. For this study we selected a single functional polymorphic marker at position –88 (rs2071430) in the promoter of MxA gene. In general the rs2071430 GG genotype promotes disease susceptibility whereas TT genotype confers protection towards Hepatitis C and B virus infection (Hijikata et al., 2001). At the mechanistic level, the functional rs2071430 GG, GT and TT genotype results in the lowest, intermediate and highest MX1 enzyme activity respectively (Fernandez-Arcas et al., 2004; Noguchi et al., 2012). In in vitro luciferase based reporter assays and in PBMC the allele TT/GT at –88 demonstrated higher reporter activity and MxA expression in response to IFN-alpha than GG allele suggesting a functional ISRE around rs2071430 (Fernandez-Arcas et al., 2004). Consistent with these observations, the rs2071430 genotype has been proposed to predict responsiveness to IFN-alpha therapy and susceptibility of Multiple sclerosis and Hepatitis C and B.

There is a general lack of studies associating MxA polymorphism with cancer. Our results suggest that the low activity GG genotype is a risk factor for prostate cancer whereas higher activity genotype TT protects against prostate cancer in general. These results are significant and suggest that low MxA expression due to GG genotype could be associated with prostate cancer especially in context of a functional study which demonstrated that MxA acts as tumor suppressor. MxA inhibits the motility and invasiveness of prostate cancer cells suggesting that its attenuated expression may promote aggressiveness of the disease (Mushinski et al., 2009). The GTPase activity of MxA appeared necessary and sufficient for blocking prostate cancer cell (PC3) invasiveness.

The low incidence of rs2071430 homozygous TT genotype in the Caucasian population is essentially consistent with population wide genotype distribution reported in NCBI dbSNP database (Table IV). According to previous literature, with the proximity of 35 base pairs apart the -88G>T and the -123C>A (rs17000900) SNPs have been assumed to be in linkage disequilibrium (LD) although not perfectly (Ching et al., 2010) . We also have genotyped this particular SNP collectively with rs2071430 (data not shown) but did not observe strong association with prostate cancer. These results further consolidate the association of rs2071430 with prostate cancer.

Collectively, the association studies and data stratification presented in this study suggests that rs2071430 is specifically associated with prostate cancer risk although the association between the two ethnic populations showed atypical results. Our present findings are clinically very useful for future studies to determine whether the –88G>T SNP may influence clinical outcome or is a genetic factor for increased risk or aggressiveness of prostate cancer. Furthermore, the association of two interferon inducible antiviral genes OAS1 (ref) and MxA, as shown in this study strongly suggests the possible viral etiology of prostate cancer. A larger study addressing the association between rs2071430 polymorphism and MxA expression in the prostate would be required to firmly establish whether MxA genotype is associated with altered expression. Alternatively, MxA rs2071430 genotype could lead to increased risk of prostate cancer due to complex gene-gene and/or gene-environment, (such increased susceptibility to infections) interactions.

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REFERENCES

- Arnheiter H, Skuntz S, Noteborn M, Chang S, Meier E. Transgenic mice with intracellular immunity to influenza virus. Cell. 1990; 62:51–61. [PubMed: 2194673]
- Ching JC, Chan KY, Lee EH, Xu MS, Ting CK, So TM, Sham PC, Leung GM, Peiris JS, Khoo US. Significance of the myxovirus resistance A (MxA) gene-123C>a single-nucleotide polymorphism in suppressed interferon beta induction of severe acute respiratory syndrome coronavirus infection. J Infect Dis. 2010; 201:1899–1908. [PubMed: 20462354]
- Di Paolo C, Hefti HP, Meli M, Landis H, Pavlovic J. Intramolecular backfolding of the carboxylterminal end of MxA protein is a prerequisite for its oligomerization. J Biol Chem. 1999; 274:32071–32078. [PubMed: 10542240]
- Duc TT, Farnir F, Michaux C, Desmecht D, Cornet A. Detection of new biallelic polymorphisms in the human MxA gene. Mol Biol Rep. 2012; 39:8533–8538. [PubMed: 22714910]
- Fernandez-Arcas N, Blanco A, Gaitan MJ, Nyqvist M, Alonso A, Reyes-Engel A. Differential transcriptional expression of the polymorphic myxovirus resistance protein A in response to interferonalpha treatment. Pharmacogenetics. 2004; 14:189–193. [PubMed: 15167707]
- Frese M, Kochs G, Feldmann H, Hertkorn C, Haller O. Inhibition of bunyaviruses, phleboviruses, and hantaviruses by human MxA protein. J Virol. 1996; 70:915–923. [PubMed: 8551631]
- Frese M, Kochs G, Meier-Dieter U, Siebler J, Haller O. Human MxA protein inhibits tick-borne Thogoto virus but not Dhori virus. J Virol. 1995; 69:3904–3909. [PubMed: 7745744]
- Furuyama H, Chiba S, Okabayashi T, Yokota S, Nonaka M, Imai T, Fujii N, Matsumoto H. Single nucleotide polymorphisms and functional analysis of MxA promoter region in multiple sclerosis. J Neurol Sci. 2006; 249:153–157. [PubMed: 16843495]
- Gao S, von der Malsburg A, Paeschke S, Behlke J, Haller O, Kochs G, Daumke O. Structural basis of oligomerization in the stalk region of dynamin-like MxA. Nature. 2010; 465:502–506. [PubMed: 20428112]
- Gordien E, Rosmorduc O, Peltekian C, Garreau F, Brechot C, Kremsdorf D. Inhibition of hepatitis B virus replication by the interferon-inducible MxA protein. J Virol. 2001; 75:2684–2691. [PubMed: 11222692]
- Goryachev AB, Pokhilko A. Computational Model Explains High Activity and Rapid Cycling of Rho GTPases within Protein Complexes. PLoS Comput Biol. 2006; 2
- Haller O, Kochs G. Human MxA protein: an interferon-induced dynamin-like GTPase with broad antiviral activity. J Interferon Cytokine Res. 2011; 31:79–87. [PubMed: 21166595]
- Hamano E, Hijikata M, Itoyama S, Quy T, Phi NC, Long HT, Ha LD, Ban VV, Matsushita I, Yanai H, Kirikae F, Kirikae T, Kuratsuji T, Sasazuki T, Keicho N. Polymorphisms of interferon inducible genes OAS-1 and MxA associated with SARS in the Vietnamese population. Biochem Biophys Res Commun. 2005; 329:1234–1239. [PubMed: 15766558]
- Hefti HP, Frese M, Landis H, Di Paolo C, Aguzzi A, Haller O, Pavlovic J. Human MxA protein protects mice lacking a functional alpha/beta interferon system against La crosse virus and other lethal viral infections. J Virol. 1999; 73:6984–6991. [PubMed: 10400797]
- Hijikata M, Mishiro S, Miyamoto C, Furuichi Y, Hashimoto M, Ohta Y. Genetic polymorphism of the MxA gene promoter and interferon responsiveness of hepatitis C patients: revisited by analyzing two SNP sites (-123 and -88) in vivo and in vitro. Intervirology. 2001; 44:379–382. [PubMed: 11805446]
- Hijikata M, Ohta Y, Mishiro S. Identification of a single nucleotide polymorphism in the MxA gene promoter (G/T at nt –88) correlated with the response of hepatitis C patients to interferon. Intervirology. 2000; 43:124–127. [PubMed: 10971132]
- Koerner I, Kochs G, Kalinke U, Weiss S, Staeheli P. Protective role of beta interferon in host defense against influenza A virus. J Virol. 2007; 81:2025–2030. [PubMed: 17151098]
- Landis H, Simon-Jodicke A, Kloti A, Di Paolo C, Schnorr JJ, Schneider-Schaulies S, Hefti HP, Pavlovic J. Human MxA protein confers resistance to Semliki Forest virus and inhibits the

amplification of a Semliki Forest virus-based replicon in the absence of viral structural proteins. J Virol. 1998; 72:1516–1522. [PubMed: 9445055]

- Mandal S, Abebe F, Chaudhary J. 2'-5' oligoadenylate synthetase 1 polymorphism is associated with prostate cancer. Cancer. 2011; 117:5509–5518. [PubMed: 21638280]
- Melen K, Ronni T, Lotta T, Julkunen I. Enzymatic characterization of interferon-induced antiviral GTPases murine Mx1 and human MxA proteins. J Biol Chem. 1994; 269:2009–2015. [PubMed: 7507489]
- Menashe I, Rosenberg PS, Chen BE. PGA: power calculator for case-control genetic association analyses. BMC Genet. 2008; 9:36. [PubMed: 18477402]
- Mushinski JF, Nguyen P, Stevens LM, Khanna C, Lee S, Chung EJ, Lee MJ, Kim YS, Linehan WM, Horisberger MA, Trepel JB. Inhibition of tumor cell motility by the interferon-inducible GTPase MxA. J Biol Chem. 2009; 284:15206–15214. [PubMed: 19297326]
- Nakade K, Handa H, Nagata K. Promoter structure of the MxA gene that confers resistance to influenza virus. FEBS Lett. 1997; 418:315–318. [PubMed: 9428735]
- Netherton CL, Simpson J, Haller O, Wileman TE, Takamatsu HH, Monaghan P, Taylor G. Inhibition of a large double-stranded DNA virus by MxA protein. J Virol. 2009; 83:2310–2320. [PubMed: 19109387]
- Noguchi S, Hijikata M, Hamano E, Matsushita I, Ito H, Ohashi J, Nagase T, Keicho N. MxA transcripts with distinct first exons and modulation of gene expression levels by single-nucleotide polymorphisms in human bronchial epithelial cells. Immunogenetics. 2012
- Pavlovic J, Arzet HA, Hefti HP, Frese M, Rost D, Ernst B, Kolb E, Staeheli P, Haller O. Enhanced virus resistance of transgenic mice expressing the human MxA protein. J Virol. 1995; 69:4506–4510. [PubMed: 7769712]
- Pavlovic J, Zurcher T, Haller O, Staeheli P. Resistance to influenza virus and vesicular stomatitis virus conferred by expression of human MxA protein. J Virol. 1990; 64:3370–3375. [PubMed: 2161946]
- Richter MF, Schwemmle M, Herrmann C, Wittinghofer A, Staeheli P. Interferon-induced MxA protein. GTP binding and GTP hydrolysis properties. J Biol Chem. 1995; 270:13512–13517. [PubMed: 7539429]
- Roers A, Hochkeppel HK, Horisberger MA, Hovanessian A, Haller O. MxA gene expression after live virus vaccination: a sensitive marker for endogenous type I interferon. J Infect Dis. 1994; 169:807–813. [PubMed: 7510764]
- Sadler AJ, Williams BR. Interferon-inducible antiviral effectors. Nature reviews. Immunology. 2008; 8:559–568. [PubMed: 18575461]
- Schnorr JJ, Schneider-Schaulies S, Simon-Jodicke A, Pavlovic J, Horisberger MA, ter Meulen V. MxA-dependent inhibition of measles virus glycoprotein synthesis in a stably transfected human monocytic cell line. J Virol. 1993; 67:4760–4768. [PubMed: 8392613]
- Schumacher B, Staeheli P. Domains mediating intramolecular folding and oligomerization of MxA GTPase. J Biol Chem. 1998; 273:28365–28370. [PubMed: 9774462]
- Staeheli P, Pitossi F, Pavlovic J. Mx proteins: GTPases with antiviral activity. Trends Cell Biol. 1993; 3:268–272. [PubMed: 14731745]
- Torisu H, Kusuhara K, Kira R, Bassuny WM, Sakai Y, Sanefuji M, Takemoto M, Hara T. Functional MxA promoter polymorphism associated with subacute sclerosing panencephalitis. Neurology. 2004; 62:457–460. [PubMed: 14872030]
- Zhang J, Wheeler DA, Yakub I, Wei S, Sood R, Rowe W, Liu PP, Gibbs RA, Buetow KH. SNPdetector: a software tool for sensitive and accurate SNP detection. PLoS Comput Biol. 2005; 1:e53. [PubMed: 16261194]

Highlights

In this study we investigated the association of the polymorphism (rs2071430) in MxA with prostate cancer. Our results suggest that rs2071430 GG genotype in Mx1 is significantly associated with prostate cancer. The results provide strong basis for the viral etiology of prostate cancer

Glymph et al.

Table I

Demographics of all samples used in the study

	Total no. of		AGE		
Sample type	Samples (N)	Mean ±SEM Median Range	Median	Range	P Value
Total no. of samples analyzed	304				
Cancers	164	63.707±0.746	63.5	43–86	27.0
Normal	140	60.214 ± 1.344	49	17–98	0.273
Caucasian					
Cancers	84	59.750±0.888	59.5	37–83	0
Normal	80	57.244±1.955	19	17–98	0.730
African American					
Cancers	80	67.862±1.028	89	43–86	0.072
Normal	09	63.952 ± 1.6	29	20-94	0.07

Page 10

Population Diversity and genotype of normal but ethnically diverse populations from NCBI dbSNP database for MX1 polymorphism rs2071430.

Table II

Glymph et al.

rs2071430 (-88G/F)	-88G/T)							
					Genotype		AII	Alleles
#ss	Population	Ethinicity	Sample Size	$\overline{9/9}$	$\overline{\mathrm{C}/\mathrm{T}}$	$\overline{\mathrm{L}/\mathrm{L}}$	$\overline{5}$	Ī
ss48295846	PI	anynomous samples	102	0.725	0.225	0.050	83.80%	16.20%
	CAUCI	Caucasian heritage	31	0.839	0.161	0.000	91.90%	8.10%
	AFR1	African/African American	24	0.708	0.250	0.042	83.30%	16.70%
	HISP1	Hispanic heritage	23	0.870	0.130	0.000	93.50%	6.50%
	PAC1	Pacific rim heritage	24	0.458	0.375	0.167	64.60%	35.40%

Page 11

Table III

Association of MX1 rs2071430 with prostate cancer. Genotype distribution for all samples, corresponding odds ratio, 95% confidence interval and p values are shown (OR= odds ratio, CI= confidence interval, N= number of samples and %= percent samples)

Glymph et al.

Genotype All Samples Cancer	All S	amples	Canc	er	Normal	lal	OR	95% CI Lower	95% CI Upper
	z	%	z	% N % N % N	z	%			
99	200	65.8	118	200 65.8 118 72 82 58.6 1.713	82	58.6		1.041	2.818
GT	92	30.3	42	25.6	50	35.7	92 30.3 42 25.6 50 35.7 Reference		
TT	12	3.9	4	12 3.9 4 2.4 8 5.7 0.60	8	5.7	09:0	0.167	2.116
Total	304	100.0	164	304 100.0 164 100.0 140 100.0	140	100.0			

Page 12

Table IV

Association of MX1 rs2071430 with prostate cancer among race. Genotype distribution for Caucasian samples and African-American samples and corresponding odds ratio and 95% confidence interval are shown (OR= odds ratio, CI= confidence interval)

Genotype	Race	Cancer	%	Cancer % Normal %	%	OR	95% CI 95% CI Lower Upper	95% CI Upper
99	African American	52	26	30	15	15 1.37	692.0	2.442
(n=200)	Caucasian	99	33	52	26	Reference		
GT	African American	26	28.2	22	24	2.07	0.897	4.780
(n=92)	Caucasian	16	17.4	28	30.4	Reference		
Ę	African American	2	16.6	8	9.99	66.6 0.0625	0.0019	1.998
(n=12)	Caucasian	2	16.6	0	0	0 Reference		