

# Experimental datasets for benchmarking protein force fields

## [Article v0.1]

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**Abstract** 250 word limit

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## 1 Introduction

### • Background

- Role of molecular dynamics in understanding protein structure and function and in drug design
- Brief history of protein force fields

### • Gap in field

- Force fields are trained against different empirical targets and are expected to describe some behaviors well and others poorly
- Force fields for proteins often borrow parameters from more general force fields that aim to describe broader chemistry
- Need for a comprehensive collection of ex-

perimental datasets that interrogates a wide range of physical properties of proteins

### • Goals of current review

- Description of available datasets and not prescription of how comparisons should be made
- Focus on peptides and globular proteins without ligands or cofactors to narrow scope

### • Target audience

- Researchers involved in developing or assessing protein force fields
- Assume familiarity with molecular dynamics techniques, force field terms, and basics of protein structure

- Review format
  - Explanation of Perpetual Review format
  - Instructions for community involvement
- Outline of review sections

## 2 Goals of benchmark datasets

- Target observables
  - Target experimental observables instead of structural models or quantum chemistry data
- Accessibility
  - Identify datasets that are accessible without paywalls or restrictive licenses
- Multiple scales
  - Identify observables that interrogate physical properties at different length and time scales
  - Goal is to assess force fields rather than train parameters, so computational cost can be high
  - System size should range from small—small enough to sample an ensemble exhaustively—to medium—large enough to exhibit stable folding behaviors
- Discriminatory power
  - Identify systems that can discriminate between force fields
  - For example, most protein force fields can describe lysozyme well

## 3 Room-temperature (RT) crystallography

- Advantages of RT crystals
  - RT crystals are higher quality and exhibit lower mosaicity than low-temperature crystals
  - Proteins in RT crystals fluctuate more than those in low temperature crystals

- Observables are accessible in public databases in a common format
- Observables
  - Electron density
    - \* Electron density is independent of a structural model unless molecular replacement was used to solve phases
    - \* Electron density from solvent molecules can be included
    - \* Comparing simulations to experiments
      - Quality metrics for structural models, e.g. R-factors or correlation coefficients, are likely too sensitive to meaningfully discriminate between force fields
      - Differences can be visualized by an  $F_O - F_C$  map
      - A quantitative metric is a comparison between a structural model refined against simulated electron density and a structural model refined against experimental density, e.g. an RMSD
  - Reflections
    - \* Raw reflections are totally independent of a structural model
    - \* Reflections are available in PDB entries
    - \* Non-Bragg peaks from diffuse scattering inform on large-scale fluctuations
  - Debye-Waller (B) factors
    - \* B factors are available in PDB entries
    - \* B factors inform on local flexibility
    - \* A drawback is that B factors may reflect disorder in the crystal lattice rather than flexibility of the crystallized molecules
  - Populations of alternative conformations
    - \* Although alternative conformations rely on a structural model, this low resolution metric may discriminate between force fields that perform similarly on other observables
- Running crystal simulations
  - Simulations of single unit cells are less expensive but may miss fluctuations that are

- important for some observables
- Simulation of supercells are more realistic but may fail to maintain the correct symmetry
- May need to include co-solvents in mother liquor
- Systems
  - Criteria/desiderata
    - \* High resolution ( $\leq 1.2 \text{ \AA}$ ) crystals to ensure high quality target data and identify tautomers and protonation states
    - \* Protonation state can be determined unambiguously by neutron diffraction
    - \* Aim for diversity in secondary structure
    - \* Systems for which data from multiple crystals with different symmetry are available are useful
  - Systems
    - \* David Case
    - \* Julian Chen
    - \* James Fraser
    - \* Daniel Keedy
    - \* Michael Wall

## 4 Nuclear magnetic resonance spectroscopy

- Advantages of NMR
  - NMR experiments are performed in the desired ensemble for most applications
  - Comparison to NMR data may reveal native state bias that is difficult to diagnose with crystal simulations
  - Many NMR observables can be related to specific FF terms
- Observables
  - Chemical shift
    - \* Easily accessible for many systems in BMRB
    - \* Directly informs on local backbone conformation for unstructured peptides and disordered proteins
  - Scalar coupling
    - \* Difficult to interpret for larger, folded proteins due to aromatic ring currents, spin diffusion, etc.
    - \* Scalar coupling values for backbone amide proton inform on local backbone conformation
    - \* Requires Karplus parameters, which can be derived from QM
  - Helical propensities (merge with chemical shift section?)
    - \*  $^{13}\text{C}=\text{O}$  chemical shifts inform on helical propensities of amino acids
    - \* Benchmarks can target chemical shifts directly or Lifson-Roig helix extension parameters
  - Nuclear Overhauser effect (NOE) spectroscopy
    - \* NOEs inform on interactions between residues distant in primary sequence
    - \* NOE intensities are nonlinear averages that are difficult to converge, so they may serve better as ordinal (i.e. strong/medium/weak) rather than quantitative assessments
  - Residual dipolar coupling (RDC)
    - \* RDCs inform on large spatial motions
    - \* Calculating RDCs for large proteins requires computing an expensive alignment tensor
  - Spin relaxation
    - \* Spin relaxation rates inform on large spatial motions for folded proteins
    - \* Spin relaxation can discriminate between force fields that describe global conformations and those that describe only local conformations
    - \* There is error from zero point motion and difference between modeled and true bond lengths, but the necessary correction may be small enough to ignore
    - \* Spin relaxation rates will be difficult to converge for large, folded proteins
- Running NMR simulations

**Table 1.** Room-temperature crystallography datasets

Description	PDB ID	Experiments	Experimental references	Computational references
Endoglucanase	3X2P	X-ray diffraction Neutron diffraction		
Scorpion toxin II	1AHO	X-ray diffraction		

- Viscosity of water model is known to affect tumbling rates and thus spin relaxation rates

- Systems

- Kyle Beauchamp chemical shifts and scalar couplings
- Bernie Brooks spin relaxation dataset for lipids, good for methods
- Lillian Chong scalar couplings for protein mimetics, good for methods
- Kresten Lindorff-Larsen chemical shift and NOEs
- Samuli Ollila spin relaxation dataset for proteins
- Paul Robustelli chemical shifts, NOEs, and helical propensities
- Lars Schäfer c-Myb chemical shifts and NOEs

## 5 Hydrogen-deuterium exchange (HDX) experiments

- Advantages of HDX

- HDX informs on folding of small proteins with simple tertiary structures
- HDX discriminates between proteins with intermediate and high folding stability that have similar bulk properties or spin relaxation rates

- Observables

- Chemical shifts or HSQC measured by NMR
- Mass spectrometry

- Protection factor (exchange frequency relative to unfolded state) has an ambiguous relationship to computable quantities, e.g. free energies

- Systems

- Gabe Rocklin and Tobin Sosnick HDX dataset
- Vincent Shaw G proteins
- Vincent Voelz ubiquitin, BPTI, and myoglobin

## 6 List of potential figures

- Visualization of protein crystal supercell
- Visualization of differences in electron density with  $F_O - F_C$  map
- Solution protein structure with NMR observables labeled
  - Folded tertiary structure labeled with "RDC" and "Spin relaxation"
  - Long range contact labeled with "NOE"
  - Inset of  $\alpha$  helix labeled with "HDX" and "Helical propensity"
  - Inset of peptide backbone with "Chemical shift" and " $^3J$  coupling" labeled
- Histograms of observables in larger datasets (perhaps borrowed from original publications)
  - Distribution of spin relaxation rates in Ollila dataset
  - Distribution of HDX exchange rates in Rocklin/Sosnick dataset

## 7 Conclusions

- Summarize key points
- Additional type of experiments
  - Kirkwood-Buff integrals for co-solvents

**Table 2.** Nuclear magnetic resonance spectroscopy datasets

Description	PDB ID	Experiments	Experimental references	Computational references
c-Myb transactivation domain	1SB0	<sup>1</sup> H chemical shifts NOESY		
Short peptides		HDX exchange rates		

- Paramagnetic relaxation enhancement interactions
- Binding free energies
- Salt bridge dissociation rates
- Folding observables
  - \* Free energies
  - \* Kinetic rates
  - \* Melting temperatures
- Small angle x-ray scattering observables
  - \* Radii of gyration
  - \* Kratky plots
  - \* Pairwise distribution functions
- Additional protein systems
  - Membrane proteins (Benoit Roux)
  - Proteins with ligands or cofactors
  - Protein mimetics, e.g. peptoids or  $\beta$ -peptides (Lillian Chong)

## 8 Author Contributions

(Explain the contributions of the different authors here)

For a more detailed description of author contributions, see the GitHub issue tracking and changelog at <https://github.com/openforcefield/protein-benchmark-data>.

## 9 Other Contributions

(Explain the contributions of any non-author contributors here) For a more detailed description of contributions from the community and others, see the GitHub issue tracking and changelog at <https://github.com/openforcefield/protein-benchmark-data>.

## 10 Potentially Conflicting Interests

MKG has an equity interest in and is a cofounder and scientific advisor of VeraChem.

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