

# Method

The method is only used as a reference for publication purpose.

Customers are responsible for the related risks of duplicate  
checking.

## **1 Experimental Procedure**

### **1.1 Sample Quality Control**

Please refer to QC report for methods of sample quality control.

### **1.2 Library Construction, Quality Control and Sequencing**

Sequencing libraries were generated and indexes were added to attribute sequences to each sample. Briefly, PCR amplification of targeted regions was performed by using specific primers connecting with barcodes. The PCR products with proper size were selected by 2% agarose gel electrophoresis. Same amount of PCR products from each sample was pooled, end-repaired, A-tailed and further ligated with Illumina adapters. Libraries were sequenced on a paired-end Illumina platform to generate 250bp paired-end raw reads. Subsequently, library quality was assessed and quantified by QPCR. Quantified libraries will be pooled and sequenced on Illumina platforms, according to effective library concentration and data amount required.

## **2 Bioinformatics Analysis Pipeline**

### **2.1 Paired-end reads assembly and quality control**

#### **2.1.1 Data split**

Paired-end reads was assigned to samples based on their unique barcode and truncated by cutting off the barcode and primer sequence.

#### **2.1.2 Sequence assembly**

Paired-end reads were merged using FLASH (<http://ccb.jhu.edu/software/FLASH/>) (Magoc T et al.,2011), a very fast and accurate analysis tool, which was designed to merge paired-end reads when at least some of the reads overlap the read generated from the opposite end of the same DNA fragment, and the splicing sequences were called raw tags.

#### **2.1.3 Data Filtration**

Quality filtering on the raw tags were performed using the fastp software to

obtain high-quality Clean Tags (Bokulich NA et al.,2012).

#### **2.1.4 Chimera removal**

The tags were compared with the reference database(Silva database (16S/18S), <https://www.arb-silva.de/>; Unite Database(ITS), <https://unite.ut.ee/>) using UCHIME Algorithm ([http://www.drive5.com/usearch/manual/uchime\\_algo.html](http://www.drive5.com/usearch/manual/uchime_algo.html))to detect chimera sequences, and then the chimera sequences were removed (Edgar RC et al.,2011). Then the Effective Tags finally obtained.

### 3 Reference

Bokulich NA, Subramanian S, Faith JJ, et al. Quality-filtering vastly improves diversity estimates from Illumina amplicon sequencing. *Nature Methods*. 2012;10(1):57-59. doi:10.1038/nmeth.2276.

Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R. UCHIME improves sensitivity and speed of chimera detection. *Bioinformatics*. 2011;27(16):2194-2200. doi:10.1093/bioinformatics/btr381.

Magoc T, Salzberg SL. FLASH: fast length adjustment of short reads to improve genome assemblies. *Bioinformatics*. 2011;27(21):2957-2963. doi:10.1093/bioinformatics/btr507.