# Genetic Evaluation - Solution 4

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## Problem 1: Prediction Of Breeding Values Using A Sire Model

The dataset for this exercise is available from

https://charlotte-ngs.github.io/gelasmss2021/data/gel\_sire\_ex04\_p01.csv

to predict breeding values using a sire model. The sire model is a mixed linear effects model where the sire effects are random effects. This leads to the following model

$$y = Xb + Zs + e \tag{1}$$

where

- y vector of length n of observations
- b vector of length p of fixed effects
- s vector of length q of random sire effects
- e vector of length n of random errors

For the random effects, we have to specify the expected values and the variance-covariance matrices. Because the random errors e and the sire effects s are deviations from a common mean and hence their expected values are defined as

$$E[e] = 0$$

$$E[s] = 0$$

$$E[y] = Xb$$

The expected value E[y] is computed using the defined expected values for e and s and from the model (1).

The random error terms  $e_i$  are uncorrelated and hence the variance-covariance matrix var(e) is given by

$$var(e) = I * \sigma_e^2$$

In the case where the sires are unrelated, the sire effects are also uncorrelated and the variance-covariance matrix var(s) corresponds to

$$var(s) = I * \sigma_s^2$$

The values for  $\sigma_e^2$  and  $\sigma_s^2$  are taken from the results of the variance components estimation from last weeks exercise. The variance-covariance matrix of the observations y can be computed as

$$var(y) = ZZ^T * \sigma_s^2 + I * \sigma_e^2$$

### Hints

- Use the package pedigreemm to predict the breeding values for all the sires.
- The function ranef() can be used to extract realised values of random effects.

#### Solution

The solution is the same as in Exercise 2, but we will be interested in a different part of the solutions. We start by reading the data and with re-formatting the fixed effects as factors.

```
s_data_file <- "https://charlotte-ngs.github.io/gelasmss2021/data/gel_sire_ex04_p01.csv"</pre>
tbl_pb_sire <- readr::read_csv2(file = s_data_file)</pre>
## Using ',' as decimal and '.' as grouping mark. Use read_delim() for more control.
## Parsed with column specification:
## cols(
##
     Id = col_double(),
     slh = col_double(),
##
##
     hrd = col_double(),
     age = col_double(),
##
##
     cw = col_double(),
##
     sire = col_double()
## )
Reformat the fixed effects to factors
tbl_pb_sire$slh <- as.factor(tbl_pb_sire$slh)</pre>
tbl_pb_sire$hrd <- as.factor(tbl_pb_sire$hrd)</pre>
Setup the pedigree for this sire model
vec sire <- unique(tbl pb sire$sire)</pre>
```

Fitting the sire-model is done as follows. The summary function can be used to obtain the result for the variance components and for the fixed effects.

```
require(pedigreemm)
```

Data: tbl\_pb\_sire

## REML criterion at convergence: 12610.8

##

```
##
## Scaled residuals:
##
       Min
                1Q Median
                                        Max
  -3.2011 -0.6731 0.0137
                            0.6539
                                     3.4813
##
##
## Random effects:
##
    Groups
             Name
                         Variance Std.Dev.
##
    sire
             (Intercept)
                          6.257
                                   2.501
##
   Residual
                         90.181
                                   9.496
## Number of obs: 1716, groups:
                                 sire, 10
## Fixed effects:
##
                Estimate Std. Error t value
## (Intercept) -77.17270
                           16.67579
                                     -4.628
## slh2
                22.36751
                                     39.600
                            0.56484
## slh3
                 4.27798
                            0.56818
                                      7.529
## hrd2
                88.81545
                            0.73294 121.176
## hrd3
                 9.28428
                            0.72408
                                     12.822
                58.98147
                            0.71719
                                     82.239
## hrd4
## hrd5
                20.36389
                            0.72889
                                     27.938
                                     16.405
## age
                 0.68269
                            0.04161
##
## Correlation of Fixed Effects:
        (Intr) slh2
                      slh3
##
                             hrd2
                                     hrd3
                                            hrd4
                                                   hrd5
## slh2 -0.011
## slh3 -0.072 0.513
## hrd2 -0.018 0.014 -0.023
## hrd3 -0.012 -0.001 -0.003
                              0.493
## hrd4 0.009 -0.008 -0.025
                              0.497
                                      0.501
## hrd5 0.004 0.010 -0.028
                              0.490
                                     0.495 0.500
## age -0.998 -0.007 0.056 -0.004 -0.009 -0.031 -0.026
```

The realised values of the sire-effects correspond to the predicted breeding values for all sires. They can be obtained by

```
(pb_sire <- ranef(lmem_sire))</pre>
```

```
## $sire
##
      (Intercept)
## 1
        3.7549839
## 2
       -1.1362988
## 3
        1.3937495
## 4
       -2.7762615
## 5
       -2.1658113
## 6
        0.4890680
## 7
        0.9537987
## 8
        3.5275014
## 9
       -2.4808415
## 10 -1.5598885
```

### Problem 2: Compare Offspring Of Sires

For the purpose of livestockbreeding the realised values themselves are not so interesting. But for the selection decision require a ranking of the sires according to their breeding values. Find the ranking of the sires according to their breeding values.

According to the definition of breeding value, it corresponds to the deviation of the offspring from the population mean. Hence the offspring of the sire with the best breeding value should on average be better than the offspring of the sire with the worst breeding value. Verify the difference between the average phenotypic values of the offsprings of the sires with the best and the worst predicted breeding values.

### Solution

The ranking is obtained by

```
(vec_ranking_pb_sire <- order(pb_sire$sire$`(Intercept)`, decreasing = TRUE))
## [1] 1 8 3 7 6 2 10 5 9 4</pre>
```

The comparison of the average phenotypic values of the offsprings of the sires with the best and the worst predicted breeding values can be verified by

```
require(dplyr)
tbl_pb_sire %>%
  group_by(sire) %>%
  summarise(avg = mean(cw))

## # A tibble: 10 x 2
```

```
## # A tibble: 10 x 2
##
       sire
               avg
##
       <dbl> <dbl>
##
    1
           1 241.
##
    2
           2 238.
    3
           3 241.
##
##
    4
           4
              237.
    5
           5
##
             240.
##
    6
           6
             242.
##
    7
           7
              240.
##
    8
           8
              244.
##
    9
           9
              244.
## 10
          10
              238.
```

Comparing the average of the phenotypic values of the sire with the best and the worst predicted breeding value leads to

```
tbl_pb_sire %>%
  group by(sire) %>%
  summarise(avg = mean(cw)) %>%
  filter(sire == vec_ranking_pb_sire[1] |
           sire == vec_ranking_pb_sire[length(vec_ranking_pb_sire)])
## # A tibble: 2 x 2
##
      sire
             avg
##
     <dbl> <dbl>
## 1
         1
            241.
## 2
            237.
```

### Problem 3: Predict Breeding Values Using Animal Model

As in Exercise 2, we are using the full dataset to predicted breeding values with an animal model. The computations for the solution of this Problem will have a very long runtime. That is why the solution is only sketched and not explicitly computed.

The data is available from https://charlotte-ngs.github.io/gelasmss2021/data/gel\_bp\_ex04\_p03.csv.

### Solution

We first have to read the data

```
s data path gel ex2 <- "https://charlotte-ngs.github.io/GELASMSS2020/ex/w11/data bp w11.csv"
tbl_gel_ex2 <- readr::read_csv2(file = s_data_path_gel_ex2)</pre>
## Using ',' as decimal and '.' as grouping mark. Use read_delim() for more control.
## Parsed with column specification:
## cols(
##
     Id = col_double(),
##
     sex = col_double(),
     slh = col_double(),
##
##
     hrd = col_double(),
##
     age = col_double(),
##
     cw = col_double(),
     sire = col_double(),
     dam = col_double()
##
## )
colnames(tbl_gel_ex2);dim(tbl_gel_ex2)
## [1] "Id"
                      "slh"
                             "hrd" "age" "cw"
                                                   "sire" "dam"
## [1] 5325
               8
The fixed effects are converted into factors
tbl gel ex2$sex <- as.factor(tbl gel ex2$sex)
tbl_gel_ex2$slh <- as.factor(tbl_gel_ex2$slh)</pre>
tbl_gel_ex2$hrd <- as.factor(tbl_gel_ex2$hrd)</pre>
```

From the help file of pedigreemm, we can see that we first have to define a pedigree.

Now the model can be specified as for the other functions to fit linear mixed effects model, such as lmer.

```
# This takes more than one hour to run.
require(pedigreemm)
# according to https://stat.ethz.ch/pipermail/r-sig-mixed-models/2014g1/021609.html
options(lmerControl=list(check.nobs.vs.nlev="ignore",
     check.nobs.vs.rankZ = "ignore",
     check.nobs.vs.nRE="ignore"))
s_lmem_file <- "lmem_gel_ex2.rds"</pre>
if (file.exists(s_lmem_file)){
 load(file = s lmem file)
} else {
  lmem_gel_ex2 <- pedigreemm(cw ~ sex + slh + hrd + age + (1|Id),</pre>
                              data = tbl_gel_ex2,
                              pedigree = list(Id = ped))
  saveRDS(lmem_gel_ex2, file = s_lmem_file)
}
summary(lmem_gel_ex2)
```

The predicted breeding values are obtained by

ranef(lmem\_gel\_ex2)