

# Introduction to RNA-Seq – Sequence trimming

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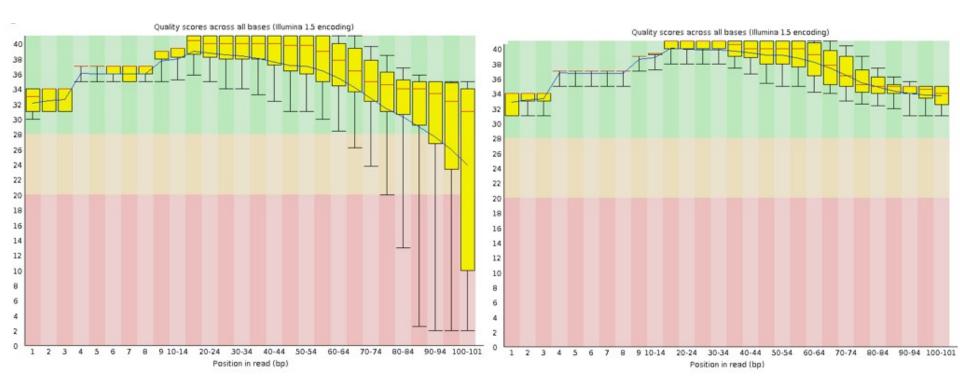




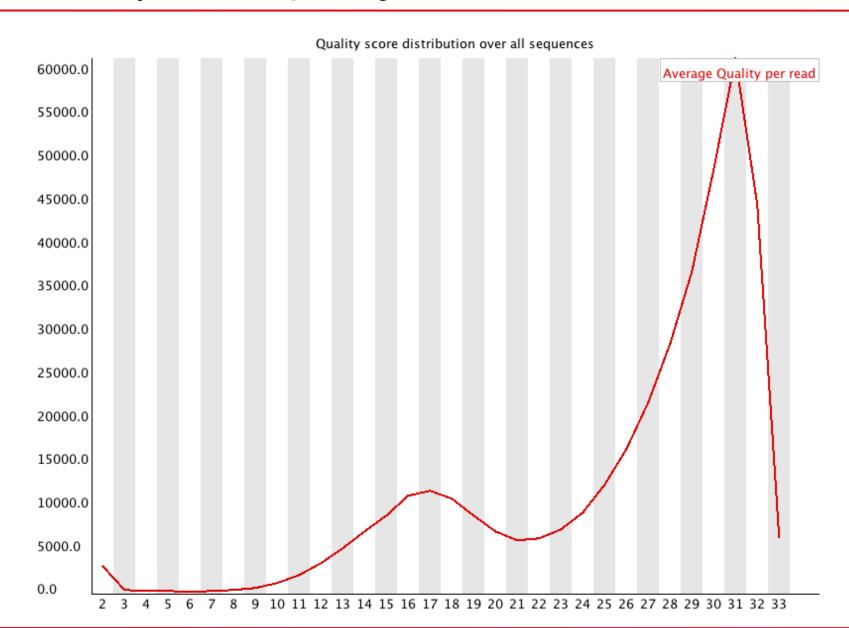


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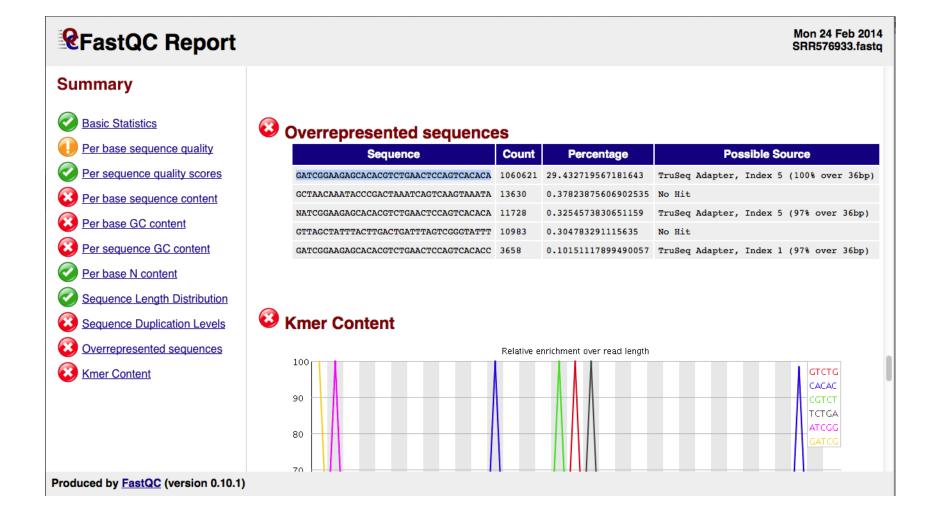
## Per Base Sequence Quality



## Per Sequence Quality Scores



### Overrepresented Sequences



## Trimming – Modifying with Trimmomatic

- Some dataset require modification prior to alignment, eg. trimming low quality base calls
- The decision to modify the dataset depends on the nature of the dataset and the question(s) asked
  - Transcriptome assembly, variant analysis, genome annotation → DO TRIMMING
  - Counting (eg. differential expression): most software can handle no trimming but still recommend some light trim. (continued)

## Trimming – Modifying with Trimmomatic

- Some dataset require modification prior to alignment, eg. trimming low quality base calls
- The decision to modify the dataset depends on the nature of the dataset and the question(s) asked
  - Trimming increases the rate of mapped reads
     BUT diminish the absolute numbers of reads

 Aggressive trimming (high quality threshold, low length filtering) can have negative impacts on expression quantification

see: William et al. (2016) Trimming of sequence reads alters RNA-Seq gene expression estimates. BMC bioinfo

## Trimmomatic – Clipping Adapter Sequences

- Generally, your sequencing facility will send you FASTQ files where all sequencing adapter sequences have been removed
- In practice, there can sometimes be unclipped adapter sequences. Why?
- Trimmomatic option:

ILLUMINACLIP:<fastaWithAdaptersEtc>:<seed mismatches>:<palindrome clip threshold>:<simple clip threshold>

ILLUMINACLIP:fastafilePATH:2:30:10

#### Trimmomatic Options

**ILLUMINACLIP**: Cut adapter and other illumina-specific sequences from the read.

**SLIDINGWINDOW**: Performs a sliding window trimming approach. It starts scanning at the 5" end and clips the read once the average quality within the window falls below a threshold.

**MAXINFO**: An adaptive quality trimmer which balances read length and error rate to maximise the value of each read

**LEADING**: Cut bases off the start of a read, if below a threshold quality

**TRAILING**: Cut bases off the end of a read, if below a threshold quality

CROP: Cut the read to a specified length by removing bases from the end

**HEADCROP**: Cut the specified number of bases from the start of the read

**MINLEN**: Drop the read if it is below a specified length

AVGQUAL: Drop the read if the average quality is below the specified level

**TOPHRED33**: Convert quality scores to Phred-33

TOPHRED64: Convert quality scores to Phred-64

#### **Trimmomatic**

#### eg Trim a paired-end dataset

```
trimmomatic PE \
reads_1.fq reads_2.fq \
./trimmed_data/paired_trimmed_1.fq. /trimmed_data/unpaired_trimmed_1.fq \
./trimmed_data/paired_trimmed_2.fq. /trimmed_data/unpaired_trimmed_2.fq \
SLIDINGWINDOW: 4:10
```

**Read the manual for more information.** This is good practice, as software is constantly evolving and you can't always rely on recipes provided by courses;-)

http://www.usadellab.org/cms/?page=trimmomatic

#### REFERENCES

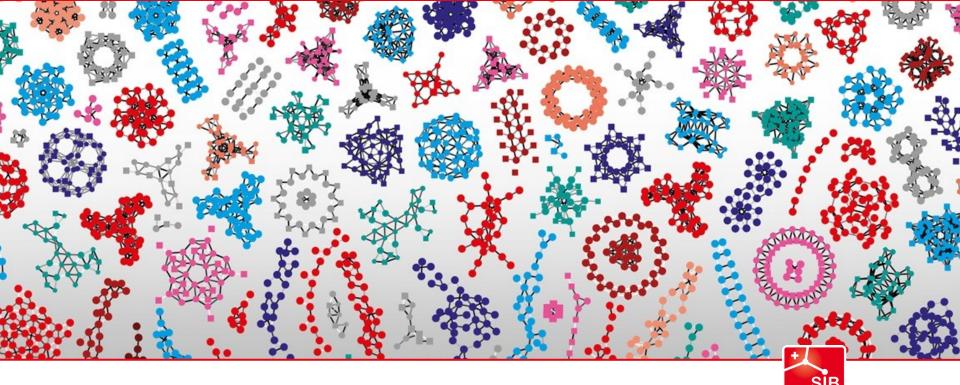
http://www.usadellab.org/cms/?page=trimmomatic

Bolger et al

(2014) "Trimmomatic: A flexible trimmer for Illumina sequen ce data" Bioinformatics 30(15): 2114-2120.

#### **Practical**

Go to the website and do the trimming practical



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