

Cell growth

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The term **cell growth** is used in the contexts of biological <u>cell</u> development and <u>cell division</u> (reproduction). When used in the context of cell development, the term refers to increase in <u>cytoplasmic</u> and <u>organelle</u> volume (<u>G1 phase</u>), as well as increase in genetic material (<u>G2 phase</u>) following the replication during <u>S phase</u>. This is not to be confused with growth in the context of cell division, referred to as proliferation, where a cell, known as the "mother cell", grows and divides to produce two "daughter cells" (<u>M phase</u>).

Cell populations

Cell populations go through a particular type of <u>exponential growth</u> called doubling. Thus, each <u>generation</u> of cells should be twice as numerous as the previous generation. However, the number of generations only gives a maximum figure as not all cells survive in each generation.

Cell size

Cell size is highly variable among organisms, with some algae such as <u>Caulerpa taxifolia</u> being a single cell several meters in length. Plant cells are much larger than animal cells, and protists such as <u>Paramecium</u> can be 330 μ m long, while a typical human cell might be 10 μ m. How these cells "decide" how big they should be before dividing is an open question. Chemical gradients are known to be partly responsible, and it is hypothesized that mechanical stress detection by <u>cytoskeletal</u> structures is involved. Work on the topic generally requires an organism whose cell cycle is well-characterized.

Yeast cell size regulation

The relationship between cell size and <u>cell division</u> has been extensively studied in <u>yeast</u>. For some cells, there is a mechanism by which cell <u>division</u> is not initiated until a cell has reached a certain size. If the nutrient supply is restricted (after time t = 2 in the diagram, below), and the rate of increase in cell size is slowed, the time period between cell divisions is increased. [3] Yeast cell-size mutants were isolated that begin cell division before reaching a normal/regular size (*wee* mutants). [4]

Wee1 protein is a tyrosine kinase that normally phosphorylates the Cdc2 cell cycle regulatory protein (the homolog of <u>CDK1</u> in humans), a cyclin-dependent kinase, on a tyrosine residue. Cdc2 drives entry into mitosis by phosphorylating a wide range of targets. This <u>covalent</u> modification of the molecular structure of Cdc2 inhibits the enzymatic activity of Cdc2 and prevents cell division. Wee1 acts to keep Cdc2 inactive during early <u>G2</u> when cells are still small. When cells have reached sufficient size during

G2, the phosphatase Cdc25 removes the inhibitory phosphorylation, and thus activates Cdc2 to allow mitotic entry. A balance of Wee1 and Cdc25 activity with changes in cell size is coordinated by the mitotic entry control system. It has been shown in Wee1 mutants, cells with weakened Wee1 activity, that Cdc2 becomes active when the cell is smaller. Thus, mitosis occurs before the yeast reach their normal size. This suggests that cell division may be regulated in part by dilution of Wee1 protein in cells as they grow larger.

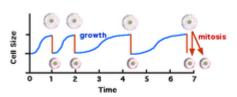


Figure 1:Cell cycle and growth

Linking Cdr2 to Wee1

The protein kinase <u>Cdr2</u> (which negatively regulates Wee1) and the Cdr2-related kinase <u>Cdr1</u> (which directly phosphorylates and inhibits Wee1 *in vitro*)^[5] are localized to a band of cortical nodes in the middle of interphase cells. After entry into mitosis, cytokinesis factors such as <u>myosin II</u> are recruited to similar nodes; these nodes eventually condense to form the <u>cytokinetic</u> ring. A previously uncharacterized protein, <u>Blt1</u>, was found to colocalize with Cdr2 in the medial interphase nodes. Blt1 knockout cells had increased length at division, which is consistent with a delay in mitotic entry. This finding connects a physical location, a band of cortical nodes, with factors that have been shown to directly regulate mitotic entry, namely Cdr1, Cdr2, and Blt1.

Further experimentation with <u>GFP</u>-tagged proteins and mutant proteins indicates that the medial cortical nodes are formed by the ordered, Cdr2-dependent assembly of multiple interacting proteins during interphase. Cdr2 is at the top of this hierarchy and works upstream of Cdr1 and Blt1. Mitosis is promoted by the negative regulation of Wee1 by Cdr2. It has also been shown that Cdr2 recruits Wee1 to the medial cortical node. The mechanism of this recruitment has yet to be discovered. A Cdr2 kinase mutant, which is able to localize properly despite a loss of function in phosphorylation, disrupts the recruitment of Wee1 to the medial cortex and delays entry into mitosis. Thus, Wee1 localizes with its inhibitory network, which demonstrates that mitosis is controlled through Cdr2-dependent negative regulation of Wee1 at the medial cortical nodes.

Cell polarity factors

Cell polarity factors positioned at the cell tips provide spatial cues to limit Cdr2 distribution to the cell middle. In fission yeast <u>Schizosaccharomyces pombe</u> (S. Pombe), cells divide at a defined, reproducible size during mitosis because of the regulated activity of Cdk1. The cell polarity protein kinase <u>Pom1</u>, a member of the dual-specificity tyrosine-phosphorylation regulated kinase (DYRK) family of kinases, localizes to cell ends. In Pom1 knockout cells, Cdr2 was no longer restricted to the cell middle, but was seen diffusely through half of the cell. From this data it becomes apparent that Pom1 provides inhibitory signals that confine Cdr2 to the middle of the cell. It has been further shown that Pom1-dependent signals lead to the phosphorylation of Cdr2. Pom1 knockout cells were also shown to divide at a smaller size than wild-type, which indicates a premature entry into mitosis. [7]

Pom1 forms polar gradients that peak at cell ends, which shows a direct link between size control factors and a specific physical location in the cell. [9] As a cell grows in size, a gradient in Pom1 grows. When cells are small, Pom1 is spread diffusely throughout the cell body. As the cell increases in size, Pom1 concentration decreases in the middle and becomes concentrated at cell ends. Small cells in early G2 which contain sufficient levels of Pom1 in the entirety of the cell have inactive Cdr2 and cannot enter mitosis. It is not until the cells grow into late G2, when Pom1 is confined to the cell ends

that Cdr2 in the medial cortical nodes is activated and able to start the inhibition of Wee1. This finding shows how cell size plays a direct role in regulating the start of mitosis. In this model, Pom1 acts as a molecular link between cell growth and mitotic entry through a Cdr2-Cdr1-Wee1-Cdk1 pathway. The Pom1 polar gradient successfully relays information about cell size and geometry to the Cdk1 regulatory system. Through this gradient, the cell ensures it has reached a defined, sufficient size to enter mitosis.

Cell cycle regulation in mammals

Many different types of eukaryotic cells undergo size-dependent transitions during the cell cycle. These transitions are controlled by the cyclin-dependent kinase Cdk1. Though the proteins that control Cdk1 are well understood, their connection to mechanisms monitoring cell size remains elusive. A postulated model for mammalian size control situates mass as the driving force of the cell cycle. A cell is unable to grow to an abnormally large size because at a certain cell size or cell mass, the S phase is initiated. The S phase starts the sequence of events leading to mitosis and cytokinesis. A cell is unable to get too small because the later cell cycle events, such as S, G2, and M, are delayed until mass increases sufficiently to begin S phase. [11]

Many of the signal molecules that convey information to cells during the control of cellular differentiation or growth are called growth factors. The protein mTOR is a serine/threonine kinase that regulates translation and cell division. [12] Nutrient availability influences mTOR so that when cells are not able to grow to normal size they will not undergo cell division. The details of the molecular mechanisms of mammalian cell size control are currently being investigated. The size of post-mitotic neurons depends on the size of the cell body, axon and dendrites. In vertebrates, neuron size is often a reflection of the number of synaptic contacts onto the neuron or from a neuron onto other cells. For example, the size of motoneurons usually reflects the size of the motor unit that is controlled by the motoneuron. Invertebrates often have giant neurons and axons that provide special functions such as rapid action potential propagation. Mammals also use this trick for increasing the speed of signals in the nervous system, but they can also use myelin to accomplish this, so most human neurons are relatively small cells.

Other experimental systems for the study of cell size regulation

One common means to produce very large cells is by cell fusion to form <u>syncytia</u>. For example, very long (several inches) <u>skeletal muscle</u> cells are formed by fusion of thousands of <u>myocytes</u>. Genetic studies of the fruit fly <u>Drosophila</u> have revealed several genes that are required for the formation of multinucleated muscle cells by fusion of <u>myoblasts</u>. Some of the key proteins are important for <u>cell adhesion</u> between myocytes and some are involved in adhesion-dependent cell-to-cell <u>signal transduction</u> that allows for a cascade of cell fusion events.

<u>Oocytes</u> can be unusually large cells in species for which embryonic development takes place away from the mother's body. Their large size can be achieved either by pumping in cytosolic components from adjacent cells through cytoplasmic bridges (*Drosophila*) or by internalization of nutrient storage granules (yolk granules) by endocytosis (frogs).

Increases in the size of <u>plant cells</u> are complicated by the fact that almost all plant cells are inside of a solid <u>cell wall</u>. Under the influence of certain plant hormones the cell wall can be remodeled, allowing for increases in cell size that are important for the growth of some plant tissues.

Most unicellular organisms are microscopic in size, but there are some giant bacteria and protozoa that are visible to the naked eye. See: Table of cell sizes (https://web.archive.org/web/200403281127 42/http://wikibooks.org/wiki/Biology_Cell_biology_Introduction_Cell_size) —Dense populations of a giant sulfur bacterium in Namibian shelf sediments [14]— Large protists of the genus *Chaos*, closely related to the genus *Amoeba* (https://web.archive.org/web/20040427144107/http://www.bms.ed.ac. uk/research/others/smaciver/chaos.htm)

In the rod-shaped bacteria E. coli, $Caulobacter\ crescentus$ and B. subtilis cell size is controlled by a simple mechanisms in which cell division occurs after a constant volume has been added since the previous division. [15][16] By always growing by the same amount, cells born smaller or larger than average naturally converge to an average size equivalent to the amount added during each generation.

Cell division

Cell reproduction is <u>asexual</u>. For most of the constituents of the cell, growth is a steady, continuous process, interrupted only briefly at M phase when the nucleus and then the cell divide in two.

The process of cell division, called <u>cell cycle</u>, has four major parts called phases. The first part, called **G₁ phase** is marked by synthesis of various <u>enzymes</u> that are required for DNA replication. The second part of the cell cycle is the **S phase**, where <u>DNA replication</u> produces two identical sets of <u>chromosomes</u>. The third part is the **G₂ phase** in which a significant protein synthesis occurs, mainly involving the production of <u>microtubules</u> that are required during the process of division, called <u>mitosis</u>. The fourth phase, <u>M phase</u>, consists of nuclear division (<u>karyokinesis</u>) and cytoplasmic division (<u>cytokinesis</u>), accompanied by the formation of a new <u>cell membrane</u>. This is the physical division of "mother" and "daughter" cells. The M phase has been broken down into several distinct phases, sequentially known as <u>prophase</u>, <u>prometaphase</u>, <u>metaphase</u>, <u>anaphase</u> and <u>telophase</u> leading to cytokinesis.

Cell division is more complex in <u>eukaryotes</u> than in other organisms. <u>Prokaryotic</u> cells such as <u>bacterial</u> cells reproduce by <u>binary fission</u>, a process that includes DNA replication, chromosome segregation, and cytokinesis. <u>Eukaryotic</u> cell division either involves <u>mitosis</u> or a more complex process called <u>meiosis</u>. Mitosis and meiosis are sometimes called the two "<u>nuclear</u> division" processes. Binary fission is similar to eukaryote cell reproduction that involves <u>mitosis</u>. Both lead to the production of two daughter cells with the same number of chromosomes as the parental cell. Meiosis is used for a special cell reproduction process of <u>diploid</u> organisms. It produces four special daughter cells (<u>gametes</u>) which have half the normal cellular amount of DNA. A <u>male</u> and a <u>female</u> gamete can then combine to produce a zygote, a cell which again has the normal amount of chromosomes.

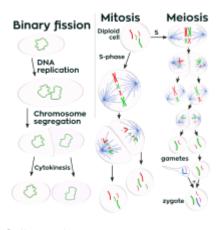
The rest of this article is a comparison of the main features of the three types of cell reproduction that either involve binary fission, mitosis, or meiosis. The diagram below depicts the similarities and differences of these three types of cell reproduction.

Comparison of the three types of cell division

The DNA content of a cell is duplicated at the start of the cell reproduction process. Prior to \overline{DNA} replication, the DNA content of a cell can be represented as the amount \mathbf{Z} (the cell has \mathbf{Z} chromosomes). After the DNA replication process, the amount of DNA in the cell is $\mathbf{2Z}$ (multiplication: 2 x Z = 2Z). During Binary fission and mitosis the duplicated DNA content of the reproducing parental cell is separated into two equal halves that are destined to end up in the two

daughter cells. The final part of the cell reproduction process is cell division, when daughter cells physically split apart from a parental cell. During meiosis, there are two cell division steps that together produce the four daughter cells.

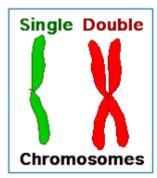
After the completion of binary fission or cell reproduction involving mitosis, each daughter cell has the same amount of DNA (**Z**) as what the parental cell had before it replicated its DNA. These two types of cell reproduction produced two daughter cells that have the same number of chromosomes as the parental cell. Chromosomes duplicate prior to cell division when forming new skin cells for reproduction. After meiotic cell reproduction the four daughter cells have half the number of chromosomes that the parental cell originally had. This is the <u>haploid</u> amount of DNA, often symbolized as **N**. Meiosis is used by diploid organisms to



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produce haploid gametes. In a diploid organism such as the human organism, most cells of the body have the diploid amount of DNA, 2N. Using this notation for counting chromosomes we say that human <u>somatic</u> cells have <u>46 chromosomes</u> (2N = 46) while human <u>sperm</u> and <u>eggs</u> have 23 chromosomes (N = 23). Humans have 23 distinct types of chromosomes, the <u>22 autosomes</u> and the special category of <u>sex chromosomes</u>. There are two distinct sex chromosomes, the X chromosome and the Y chromosome. A diploid human cell has 23 chromosomes from that person's father and 23 from the mother. That is, your body has two copies of human chromosome number 2, one from each of your parents.

Immediately after DNA replication a human cell will have 46 "double chromosomes". In each double chromosome there are two copies of that chromosome's DNA molecule. During mitosis the double chromosomes are split to produce 92 "single chromosomes", half of which go into each daughter cell. During meiosis, there are two chromosome separation steps which assure that each of the four daughter cells gets one copy of each of the 23 types of chromosome.



Chromosomes

Sexual reproduction

Though cell reproduction that uses mitosis can reproduce eukaryotic cells, eukaryotes bother with the more complicated process of meiosis because sexual reproduction such as meiosis confers a selective advantage. Notice that when meiosis starts, the two copies of sister chromatids number 2 are adjacent to each other. During this time, there can be genetic recombination events. Parts of the chromosome 2 DNA gained from one parent (red) will swap over to the chromosome 2 DNA molecule that received from the other parent (green). Notice that in mitosis the two copies of chromosome number 2 do not interact. It is these new combinations of parts of chromosomes that provide the major advantage for sexually reproducing organisms by allowing for new combinations of genes and more efficient evolution. However, in organisms with more than one set of chromosomes at the main life cycle stage, sex may also provide an advantage because, under random mating, it produces homozygotes and heterozygotes according to the Hardy-Weinberg ratio.

Disorders

A series of growth disorders can occur at the cellular level and these consequently underpin much of the subsequent course in <u>cancer</u>, in which a group of cells display uncontrolled growth and division beyond the normal limits, *invasion* (intrusion on and destruction of adjacent tissues), and sometimes *metastasis* (spread to other locations in the body via lymph or blood).

Measurement methods

The cell growth can be detected by a variety of methods. The **cell size growth** can be visualized by microscopy, using suitable stains. But the **increase of cells number** is usually more significant. It can be measured by manual counting of cells under microscopy observation, using the dye exclusion method (i.e. <u>trypan blue</u>) to count only viable cells. Less fastidious, scalable, methods include the use of <u>cytometers</u>, while <u>flow cytometry</u> allows combining cell counts ('events') with other specific parameters: fluorescent probes for membranes, cytoplasm or nuclei allow distinguishing dead/viable cells, cell types, cell differentiation, expression of a biomarker such as Ki67.

Beside the increasing number of cells, one can be assessed regarding the **metabolic activity growth**, that is, the <u>CFDA</u> and <u>calcein-AM</u> measure (fluorimetrically) not only the membrane functionality (dye retention), but also the functionality of cytoplasmic enzymes (esterases). The <u>MTT assays</u> (colorimetric) and the resazurin assay (fluorimetric) dose the mitochondrial redox potential.

All these assays may correlate well, or not, depending on cell growth conditions and desired aspects (activity, proliferation). The task is even more complicated with populations of different cells, furthermore when combining cell growth interferences or toxicity.

See also

- Bacterial growth
- Binary fission
- Cell cycle
- Clone (genetics)
- Developmental biology
- Meiosis
- Mitosis
- Stem cell

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Books

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External links

- A comparison of generational and exponential models of cell population growth (http://members.o ptusnet.com.au/exponentialist/Cells.htm)
- Local Growth in an Array of Disks (http://demonstrations.wolfram.com/LocalGrowthInAnArrayOfDisks/) Wolfram Demonstrations Project.

Image result for cell growth

Cell growth (or interphase) is shorthand for the idea of "growth in cell populations" by means of cell reproduction. It is the stage which cells are preparing for the next division, biochemical activities and reactions are taking place, however no obvious changes can be seen at this stage.

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