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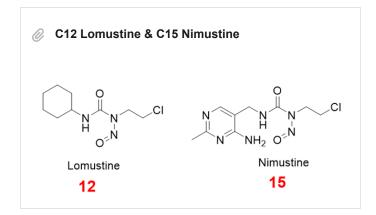
2025.6.2-4 C12 C15 C30 (on live/fixed cells); C12+C30 co-editing (on live cells)

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MONDAY, 6/2/2025

Plani	ned sa	mples					ı		
	#	Treate d on	Treatment	Incubation	RT ion	Index	i5	i7	RNA input (10ul RT rxn)
1	156	Live cells	Ctrl (5% DMSO)	80°C for 15min	Mn2+ (3mM)	i5-4 + i7-7	AGAGTAG A	GTAGAGA G	60ng
2	157	Live cells	Ctrl (5% DMSO)	80°C for 15min	Mn2+ (3mM)	i5-4 + i7-8	AGAGTAG A	CCTCTCT G	60ng
3	158	Live cells	C12 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-4 + i7-9	AGAGTAG A	AGCGTAG C	60ng
4	159	Live cells	C12 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-4 + i7-10	AGAGTAG A	CAGCCTC G	60ng
5	160	Live cells	C15 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-4 + i7-11	AGAGTAG A	TGCCTCT T	60ng
6	161	Live cells	C15 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-4 + i7-12	AGAGTAG A	TCCTCTA C	60ng
7	162	Live cells	C30 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-5 + i7-9	GTAAGGA G	AGCGTAG C	60ng
8	163	Live cells	C30 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-5 + i7-10	GTAAGGA G	CAGCCTC G	60ng
9	170	Fixed cells	Ctrl (5% DMSO)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-5	CTAAGCC T	AGGAGTC C	120ng
10	171	Fixed cells	Ctrl (5% DMSO)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-6	CTAAGCC T	CATGCCT A	120ng
11	172	Fixed cells	C12 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-7	CTAAGCC T	GTAGAGA G	120ng
12	173	Fixed cells	C12 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-8	CTAAGCC T	CCTCTCT G	120ng
13	174	Fixed cells	C15 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-9	CTAAGCC T	AGCGTAG C	120ng
14	175	Fixed cells	C15 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-10	CTAAGCC T	CAGCCTC G	120ng
15	176	Fixed cells	C30 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-11	CTAAGCC T	TGCCTCT T	120ng
16	177	Fixed cells	C30 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-8 + i7-12	CTAAGCC T	TCCTCTA C	120ng
17	178	Live cells	C12 (5mM final) + C30 (5mM final)	80°C for 15min	Mn2+ (3mM)	i5-5 + i7-8	GTAAGGA G	CCTCTCT G	60ng



TUESDAY, 6/3/2025

Cell treatment batch-3 Cell # **Treatment** Incubation number 1 150 2x10⁵ 37°C for 15min Ctrl (5% DMSO) 2 2x10^5 151 Ctrl (5% DMSO) 37°C for 15min 3 152 2x10^5 C30-FAI (1mM final) 37°C for 15min 4 153 2x10⁵ 37°C for 15min C30-FAI (1mM final) 5 2x10⁵ 37°C for 15min 154 FAI-N3 (1mM final) 6 155 2x10^5 FAI-N3 (1mM final) 37°C for 15min 7 156 2x10^5 80°C for 15min Ctrl (5% DMSO) 8 157 2x10⁵ Ctrl (5% DMSO) 80°C for 15min 9 158 4x10^5 C12 (5mM final) 80°C for 15min 10 159 4x10^5 C12 (5mM final) 80°C for 15min 11 4x10^5 160 C15 (5mM final) 80°C for 15min 12 161 4x10^5 C15 (5mM final) 80°C for 15min 13 162 8x10^5 C30 (5mM final) 80°C for 15min 14 163 8x10^5 C30 (5mM final) 80°C for 15min C12 (5mM final) + C30 (5mM 15 178 8x10^5 80°C for 15min final)

- For C12 (5mM final) + C30 (5mM final):
 - o 1x: 2.5ul 100mM C12, 0.25ul 1M C30, into 47.5ul cells in PBS
 - o 4x: 10ul 100mM C12, 1ul 1M C30, into 190ul cells in PBS

Cell treatment with RNA covalent modifiers:

- Treat "SARS-CoV2 5UTR"cells (SL5 HEK293) with TrypLE Express at room temp
- Collect and wash cells with 1x PBS
- Count cells: 3.65x10^6 cells/ml
- Prepare 2x10^5 cells/rxn x25 = 5M
- Resuspend cells:
 - o for 2x10^5 cells, resuspend in 47.5ul
 - o for 4x10^5 cells, resuspend in 95ul
 - o for 6x10^5 cells, resuspend in 142.5ul
 - o for 8x10^5 cells, resuspend in 190ul
- Add DMSO or covalent modifier:
 - o for 2x10⁵ cells, add 2.5ul
 - o for 4x10^5 cells, add 5ul
 - o for 6x10^5 cells, add 7.5ul
 - o for 8x10^5 cells, add 10ul

- Incubate at designated temp for 15min
- Chill on ice for 2min
- Add 300ul RLT Plus buffer (total volume=350ul) (for 8x10^5 cells, add 1200ul RLT Plus)
- Vortex for 30s to homogenize
- Store lysate in -80

Cell treatment batch-4						
	#	Cell number	Treatment	Incubation		
1	164	2.5x10^5	Ctrl (5% DMSO)	37°C for 15min		
2	165	2.5x10^5	Ctrl (5% DMSO)	37°C for 15min		
3	166	2.5x10^5	C30-FAI (1mM final)	37°C for 15min		
4	167	2.5x10^5	C30-FAI (1mM final)	37°C for 15min		
5	168	2.5x10^5	FAI-N3 (1mM final)	37°C for 15min		
6	169	2.5x10^5	FAI-N3 (1mM final)	37°C for 15min		
7	170	2.5x10^5	Ctrl (5% DMSO)	80°C for 15min		
8	171	2.5x10^5	Ctrl (5% DMSO)	80°C for 15min		
9	172	2.5x10^5	C12 (5mM final)	80°C for 15min		
10	173	2.5x10^5	C12 (5mM final)	80°C for 15min		
11	174	2.5x10^5	C15 (5mM final)	80°C for 15min		
12	175	2.5x10^5	C15 (5mM final)	80°C for 15min		
13	176	2.5x10^5	C30 (5mM final)	80°C for 15min		
14	177	2.5x10^5	C30 (5mM final)	80°C for 15min		

Making buffers right before the experiment:

1x PBS Plus (keep on ice)

• for cell resuspension and fixation

1x PBS Plus							
	Component	Vol (1ml)	Vol (15ml)	Final conc			
1	1x PBS	1ml	15ml	1x			
2	1M MgCl2	3ul	45ul	3mM			
3	10% (w/v) BSA/PBS	8ul	120ul	0.08%			
4	Total	1ml	15ml				

- · for cell fixation
- add at 4:1 ratio to cell suspension, e.g., 600ul fixing buffer to 150ul cells in washing buffer

Fixin	g buffer					^
	Component	Vol	Vol (16x)	Final conc	Note	
1	50x DSS	7.5ul	120ul	0.5mg/ml	Final conc if added to cell solution at 4:1 ratio	
2	Methanol (-20°C)	600ul	9.6ml	80%	Final conc if added to cell solution at 4:1 ratio	

Washing buffer (keep on ice)

• for both cell washing and later covalent modification reactions

Washing buffer							
	Component	Vol (1ml)	Vol (30ml)	Final conc	E		
1	1x PBS	940ul	28.2ml	1x			
2	1M MgCl2	3ul	90ul	3mM			
3	10% (w/v) BSA/PBS	50ul	1500ul	0.5%			
4	10% (v/v) Triton X100	10ul	300ul	0.1%			
5	RNaseIn	1ul	30ul	0.1%	Optional		
6	Total	1ml	30ml				

Cell collection:

- Treat "SARS-CoV2 5UTR"cells (SL5 HEK293) with TrypLE Express at room temp
- Collect and wash cells with 1x PBS Plus
- Count cells: 4.9x10^6 cells/ml
- Prepare <u>2.5x10^5 cells/rxn x16 = 4x10^6</u> cells in <u>2.4ml</u> 1x PBS Plus in a new 15ml tube (labeled as "fixed cell samples")

Cell fixation:

- Add 9.6ml fixing buffer to the 2.4ml "fixed cell samples"
- Incubate on ice for 15min, invert tube every 2min
- Add 3ml wash buffer, 300xg 3min
- Add 10ml wash buffer, 300xg 3min
- Add 10ml wash buffer, 300xg 3min
- Resuspend in 760ul wash buffer
- Aliquot 47.5ul to 14 tubes

Cell treatment with covalent modifiers:

- Add 2.5ul DMSO or covalent modifier in DMSO (total volume=50ul) to each sample at room temp, pipette to mix
- Incubate at designated temp for 15min
- · Chill on ice for 2min
- For fixed cell samples:
 - o Add 1.5ul proteinase K (NEB P8107S), incubate at 37°C for 10min, pipette to mix at 5min
 - Add 300ul RLT Plus buffer (total volume=350ul)
 - Vortex for 30s to homogenize
- Store lysate in -80

Note:

• Sample #174 and #175 had white precipitation after 15min incubation and chilling on ice, which is not observed on the corresponding live cell samples (#160 and #161).

WEDNESDAY, 6/4/2025

RNA extraction:

- Thaw cell lysate in RLT Plus buffer at RT, vortex to homogenize
- Transfer to DNA spin column, 8000xg 30s
- Add 350ul 70% EtOH to 350ul flow-through
- Transfer to Zymo Spin IC column, 8000xg 15s, discard the flow-through
- Add 700ul RW1 buffer, 8000xg 15s, discard the flow-through
- Add 500ul 70% EtOH instead of RPE buffer, 8000xg 15s, discard the flow-through
- Add 500ul 80% EtOH, 8000xg 15s, discard the flow-through
- Full speed 2min
- Elute with 15ul water
- Nanodrop:

RNA yield batch-3								
	#	RNA conc (ng/ul)	260/280	Add x ul water to dilute to 60ng/ul	60ng RNA (ul)	Water to 4.4ul (ul)	RT ion	
1	156	75.2	1.99	3.5	1.00	3.40	Mn2+ (3mM)	
2	157	65.9	1.98	1.4	1.00	3.40	Mn2+ (3mM)	
3	158	222.2	2.06	37.8	1.00	3.40	Mn2+ (3mM)	
4	159	274.2	2.01	50.0	1.00	3.40	Mn2+ (3mM)	
5	160	99.8	2.01	9.3	1.00	3.40	Mn2+ (3mM)	
6	161	84.5	1.96	5.7	1.00	3.40	Mn2+ (3mM)	
7	162	28.2	1.93	NA	2.13	2.27	Mn2+ (3mM)	
8	163	27.4	2.05	NA	2.19	2.21	Mn2+ (3mM)	
9	178	355.9	2.01	69.0	1.00	3.40	Mn2+ (3mM)	

RNA	RNA yield batch-4								
	#	RNA conc (ng/ul)	260/280	Add x ul water to dilute to 60ng/ul	120ng RNA (ul)	Water to 4.4ul (ul)	RT ion		
1	170	38.9	2.05	NA	3.08	1.32	Mn2+ (3mM)		
2	171	48.5	2.04	NA	2.47	1.93	Mn2+ (3mM)		
3	172	28.8	2.02	NA	4.17	0.23	Mn2+ (3mM)		
4	173	33.9	1.95	NA	3.54	0.86	Mn2+ (3mM)		
5	174	40.6	2.02	NA	2.96	1.44	Mn2+ (3mM)		
6	175	36.4	1.96	NA	3.30	1.10	Mn2+ (3mM)		
7	176	82.7	1.98	5.3	2.00	2.40	Mn2+ (3mM)		
8	177	69.0	1.99	2.1	2.00	2.40	Mn2+ (3mM)		

• Store RNA in -80

Reverse transcription:

• Input: 60ng total RNA for live cell samples, 150ng total RNA for fixed cell samples

Annealing							
	Component	vol (ul)	18x (ul)	Master mix (ul)			
1	60ng or 150ng RNA+water	4.4	4.4/EA				
2	CMV-5UTR-SHAPE-Rv (100uM)	0.05	0.9				
3	dNTP (10mM)	0.5	9				
4	Water	0.45	8.1	1/EA			
5	Total	5.4	5.4/EA				

• 65°C 5min, immediately put on ice

Exte	Extension							
	Component	vol (ul)	18x (ul)	Master mix (ul)				
1	Annealing product	5.4	5.4/EA					
2	375mM Tris/500mM KCI (5x buffer)	2	36					
3	100mM DTT (10x)	1	18					
4	Protoscript II	0.5	9					
5	RNaselN	0.1	1.8					
6	30mM MnCl2	1	18	4.6/EA				
7	Total	10	10/EA					

- 42°C 1h, 65°C 20min, 4°C hold
- Store cDNA in -20

Note:

- I will include these samples in the multiplexed library prep (2025.6.5 Amplicon library #2)
- Done in the same batches with "2025.6.4 C30-FAI (on live and fixed cells)"