Torrent Scout Light

V1.1, Chantal Roth March 22, 2012

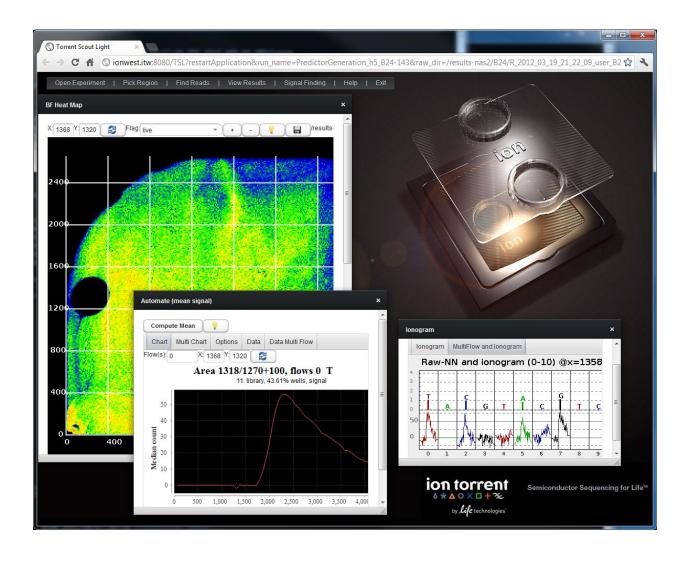




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Description

Torrent Scout Light is a pure web application written using Vaadin. The server side is written in Java and is the same code as used in the full Torrent Scout. Vaadin is a software framework that allows the developer to write 100% Java code, and then translates the gui for the web pages into Java Script, and uses the Google toolkit.

The goal of Torrent Scout Light is to provide quick access to data to get a first glance as to what the data looks like, with no installation overhead. As it is purely web based, the functionality is limited. To use the full functionality, you might want to try the full Torrent Scout client.

Starting

The most convenient way is to enable the TorrentScout plugin, and to view the results directly from the Default Report page. The plugin computes all the heat maps index files, so that you don't have to wait when you want to look at the data:



Just click on the View results in Torrent Scout Light link. This will automatically load this run and show you a heat map.

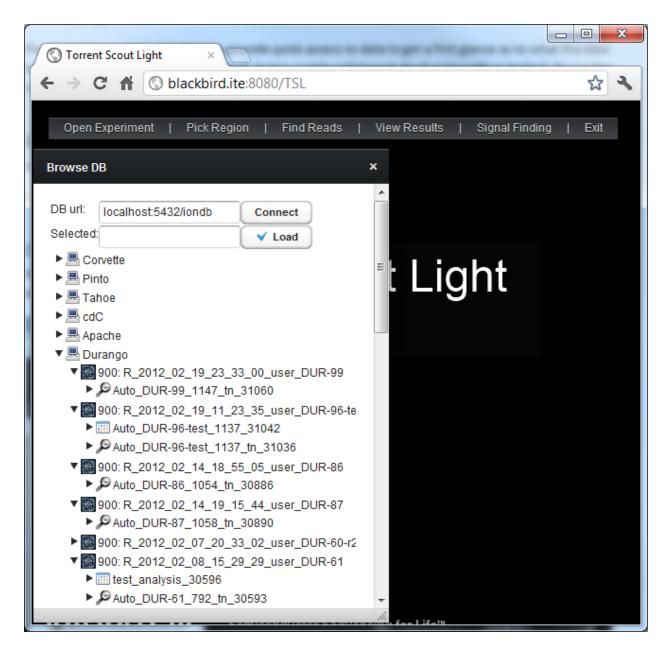
Alternatively, you can open a browser and type in the URL of your Torrent Server, followed by :8080/TSL. For instance:

http://myserver.com:8080/TSL

Opening an Experiment

You can select an experiment by browsing in the database. Open a PGM node, then an experiment node, and select an analysis result as shown in the image.

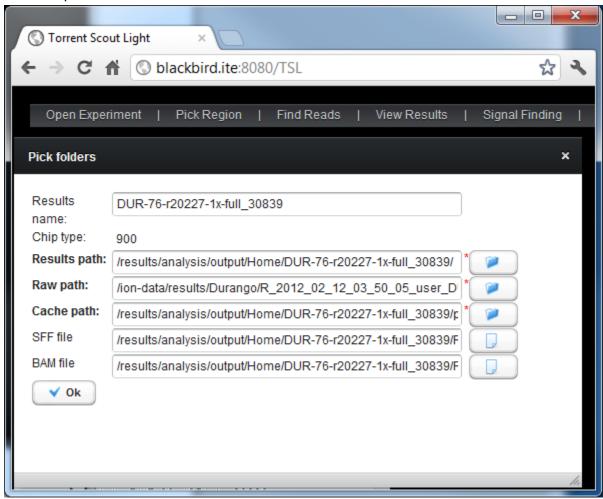




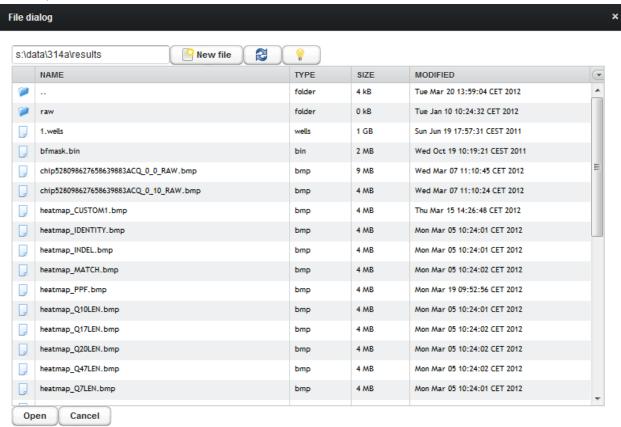
- Click on an experiment or run node to see a description popup.
- To load a run, click on a run node (such as Auto_xyz) and click on Load.
- You can specify a different db in the DB url but keep in mind that you can only view the data if the server has access to the files. One scenario that makes sense is if you have an rnd machine where you run TSL, but that machine is on the same network as the head node has access to the exact same files using the same paths as the head node.



After you click OK (the button is on top), it will show you the paths it has found from the database and will show you the details.



If the paths are not correct, you can change it here manually or by clicking the folder/document icon, which opens a file browser:



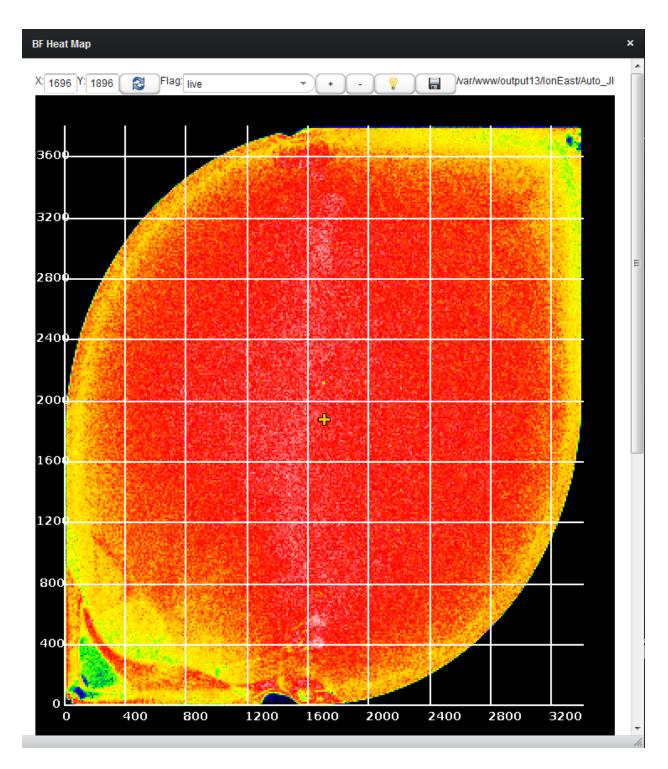
You can also open other experiments that way that are not in the database, by selecting "Open Experiment / Pick Folders".

Picking a region on the chip

BfMask Heat Map

If there is a bfmkask.bin file, it will open the BfMask Heat Map view that shows you the various flags such as bead, live, dud, ambiguous etc in a heat map:

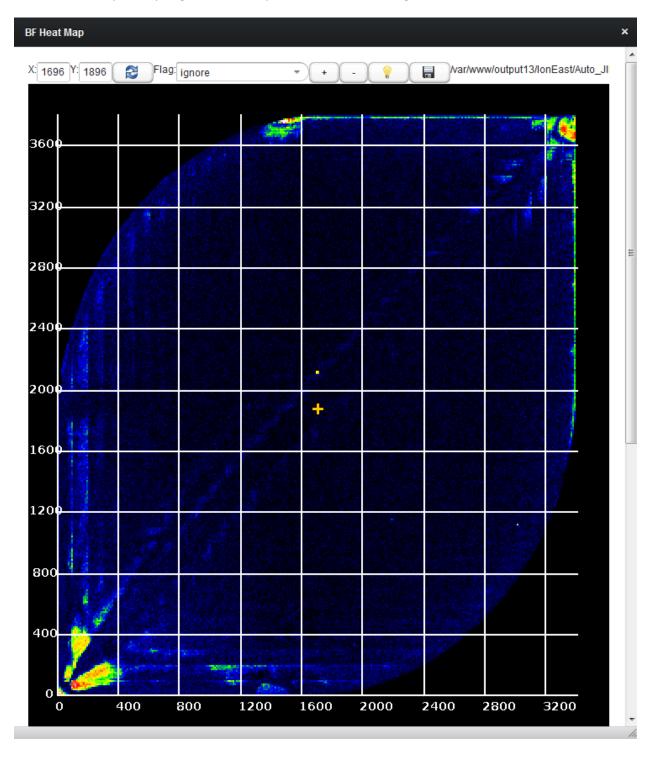




- You can also enter the coordinates manually (it won't move the orange cursor though unless you reopen the window)
- The +- buttons (in all components) allow you to zoom in and out
- The save icon lets you save the image
- The light bulb icon shows some info on what you can do (available in most components)



• You can pick any flag from the dropdown box – here the ignore mask:



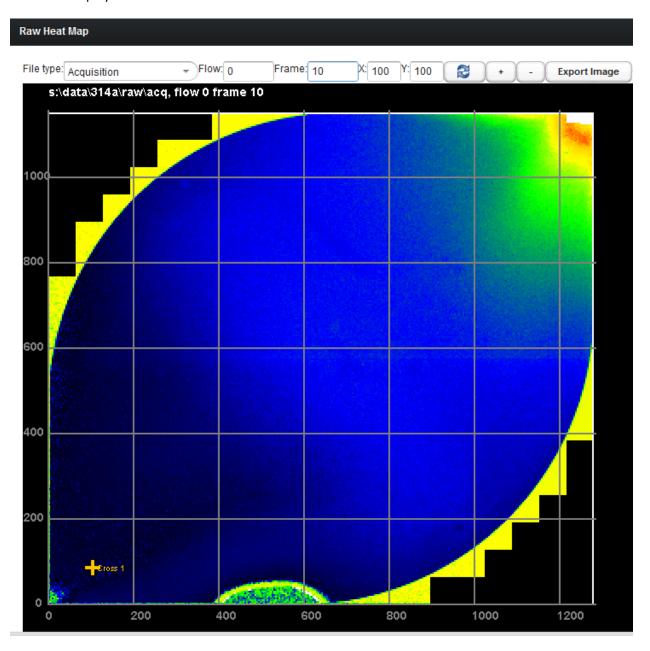
To pick a coordinate, move the orange cursor, or enter a coordinate in the X/Y text boxes.

In the flag drop down box you can chose different flags (such as bead, keypasse, dud etc) to view.



Raw Chip View

If there is no bfmask.bin file, or if you select "Pick region/ Raw heat map" it will open the WholeChip view that displays one frame of a raw .dat file:



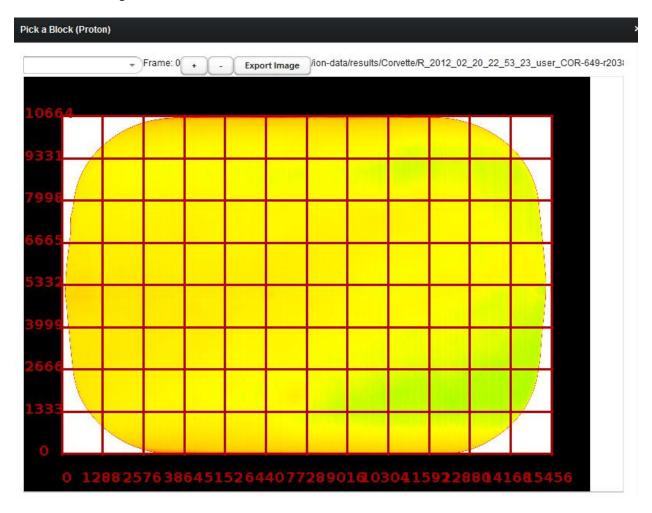
You can change the file type (acquisition, prerun bead find pre/post), the flow (any flow number starting from 0) and any frame (default is frame 10).

To pick a coordinate, move the orange cursor or enter the coordinates as in the bf mask view, and you can also zoom in and out with the +/- buttons.



Proton View

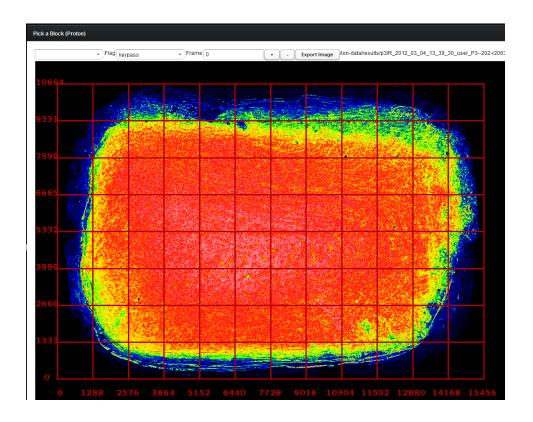
For proton chips, where the result is composed of multiple sub-blocks, you can pick a block for viewing with the "Pick Region/ Pick a block" menu:



The Proton view also lets you pick other heat maps as in the BF heat map component.

Note that if the heatmap has not been computed by the plugin, it will have to do it on the fly, which will take several minutes and use a lot of memory/CPU on the server...





Viewing results

Once you have chosen an experiment and a location (and a block in Proton experiments), you can now view results.

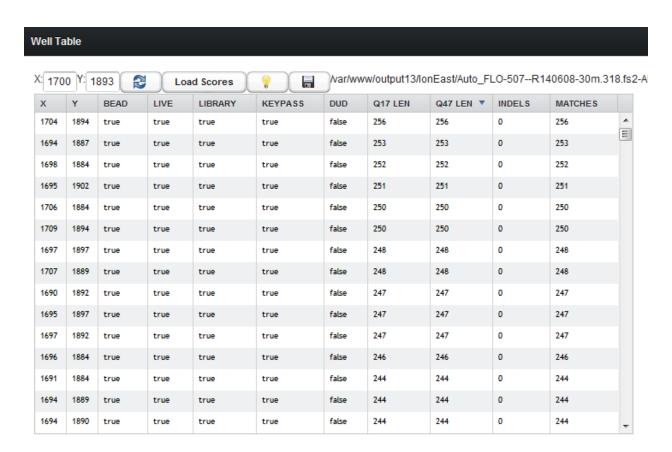
Well Table

The well table shows a few thousand wells surrounding the currently selected well. It shows a few flags and scores such as Q17 lengths, indels and so on. The scores might not always load if the index files have not been created yet. To force them to load, click on Load Scores (it might take a few minutes if there are no index files yet).

You can sort the table by clicking on the table header.

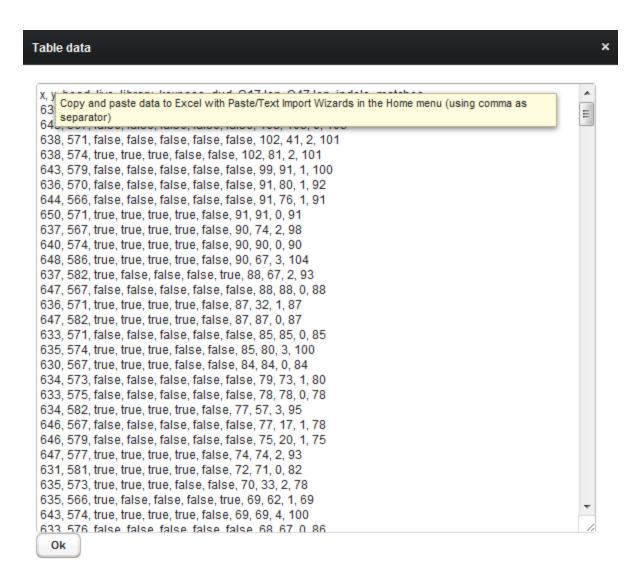
You can select a well by either clicking on a row, or by entering the coordinates on top. To view the results, select any of the viewers from the menu "View Results" (such as Ionogram, Alignment, Raw View etc)





The export button lets you export the table data (from all tables):

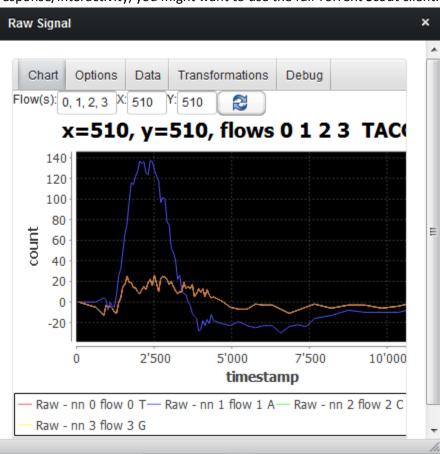




Raw Data View

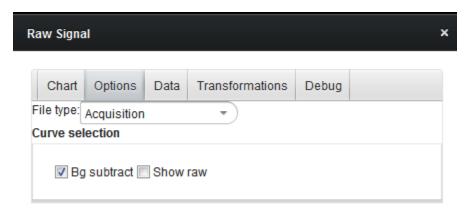
You can enter one of more flow numbers (separated by comma) into the flows text field. You can also enter a coordinate right there. Note that sometimes it takes time to load the raw data, in particular as this all goes to the torrent server first (which might be busy). For better performance and better





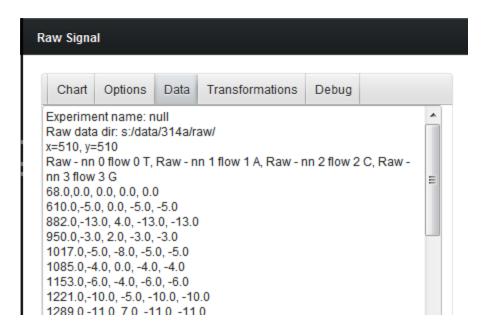
response/interactivity, you might want to use the full Torrent Scout client.

In the options tab you can pick to view the raw data and/or the bg subtracted data and the file type. The bg subtraction simply subtracts the average signal of the surrounding wells that are considered empty.



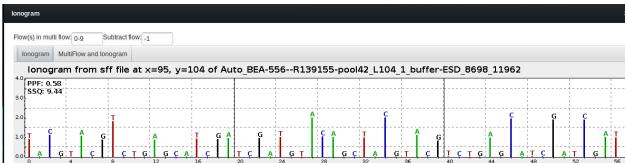
The data tab shows the currently loaded data:





You can copy and paste this to Excel (Use Paste/Text import wizard, comma as separator) to create your own charts.

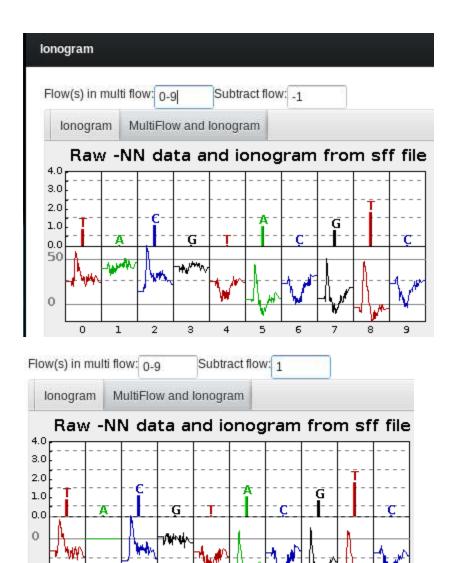
Ionogram



Viewing raw signals in ionogram view

The second tab shows any number of nearest neighbor subtracted raw signals for this well: just enter the flow numbers (starting at 0) into the text field and hit enter:



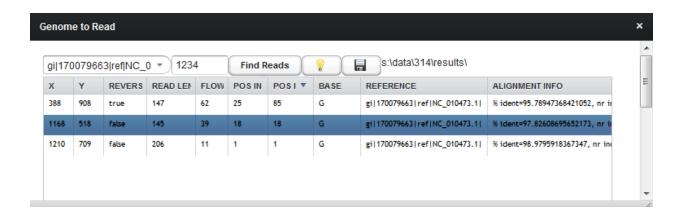


Genome To Read

In the component Find Reads/ Find reads by genome position you can locate any read that maps to a certain genome position.

Pick the reference sequence in the drop down box and enter a genome location in the text box. This will return a list of all reads (including the flow number) that map to that position. To view the ionogram or alignment of any read you can just open that component and select the read in the table.





Masks

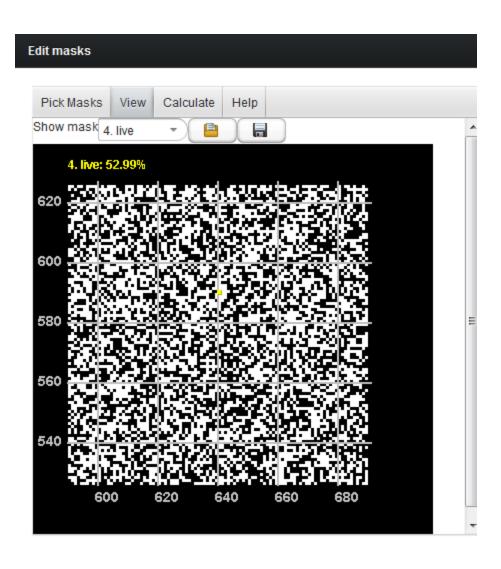
The Signal Finding/ Edit Masks component lets you view and do calculations with masks of a specific area on the chip.

The default is a 100x100 area, but you can zoom in and out of this with the Process component (upon request I can of course make this more flexible :-).

The View tab lets you view the masks and what % is flagged:

In the View tab you can save and load masks. It stores the coordinate in addition to all masks, so next time you load a masks file, it will automatically move the view to the coordinate of the mask. Note that you have to be in the correct experiment or block. So for Proton experiment currently you have to load the block for which you created the mask.



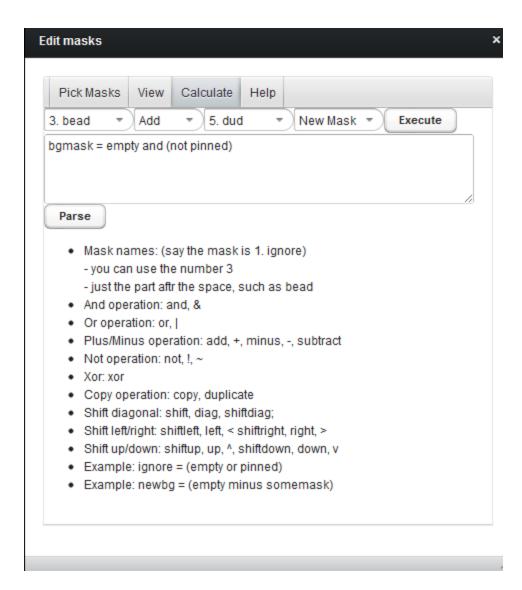


The calculate tab has 2 calculators:

- a) A simple drop down calculator
- b) A parser

The drop down calculator is really simple: pick mask A as first operand, an operation such as add, subtract, xor etc, and if it makes sense, a second operator mask B, and finally either an existing mask as result (Which will be overwritten) or else a new mask.





After the computation, it will print the command line string into the text box.

The parser allows you to copy/paste frequent operations directly into the text box, so you don't have to pick the masks in the drop down box :-)

The Pick masks tab is important for the Process and Automate component Shown later)



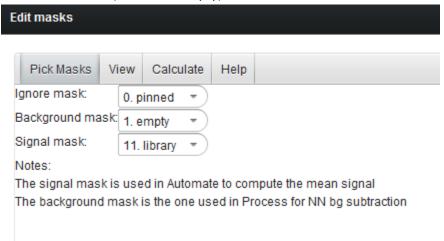
Pick Masks View Calculate Help Ignore mask: 0. pinned Background mask: 1. empty Signal mask: 11. library Notes: The signal mask is used in Automate to compute the mean signal The background mask is the one used in Process for NN bg subtraction

Process

Basically it lets you view an area of say 100x100 wells, pick a few wells in there at the same time (the cursors), and view NN subtracted signal. This allows you to compare the signal of different wells, maybe look at problems, and to see where approximately the signal appears.

You can snap the cursors to any of the masks (also your own!).

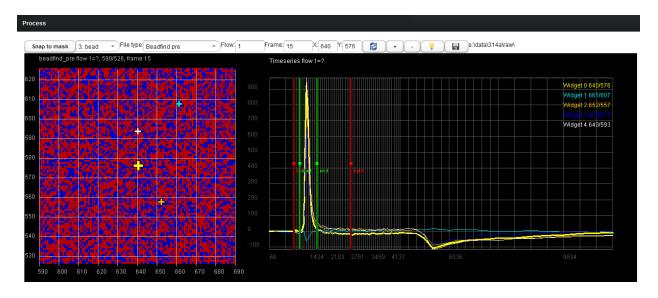
It automatically computes the NN bg subtraction, so make sure you have a reasonable mask selected in the Mask/Pick tab (default tis empty):



The zoom in magnifies the area, and reduces its size be a factor of 2 each time. **Note this currently reloads the data and masks**. So you have created your own masks, they would be lost (will think of a save features soon, but in the meantime, you might want to use the **full Torrent Scout client** to do more professional mask editing as it allows you to save one or all masks!)

You can pick the type of data and flow – here the beadfine pre flow 1 is shown with a very clear bead find signal:





Note the vertical red and green lines. They can be moved around with the mouse.

Those positions are used in the Fit component below:

Fit Component

The fit component is a bit difficult to understand at first I think: basically it helps you to classify wells. That way you can for instance create a mask such as "all live wells with a very high signal", or maybe "duds that don't look like duds", or "bead find with a low signal" etc.

The histogram is calculated using all wells from the selected histogram mask (or all wells if none is selected). For all those wells, it computes the values using the green (and red) frames.

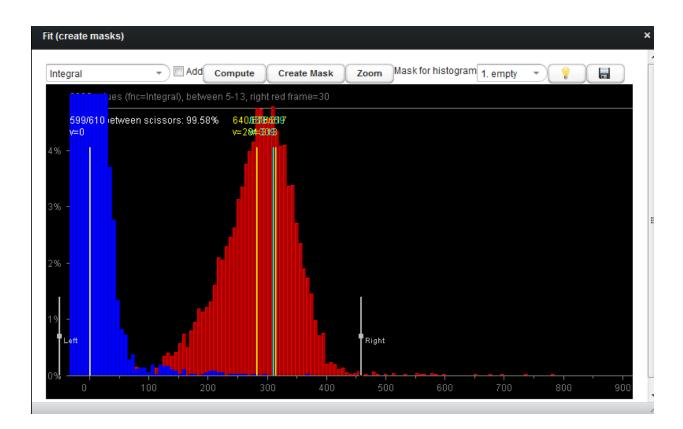
The integral computes the area under the curve between the green frames.

The max-end height computes the max between the green frames, and subtracts the value at the right red frame.

The add check box let's you add a second histogram the next time you compute it. So you might want to compare the data of the empty wells (blue) versus the bead wells (red) as in the example below.

The zoom button zooms into the area where the scissors are (it only shows the last data set currently...)

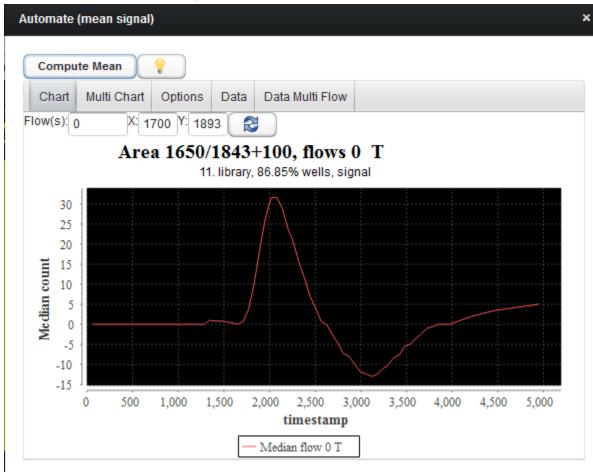






Automate (Compute Mean Signal)

Maybe you want to compare the signal shape of different areas the chip, maybe using different masks. This is where the automate component comes in:



It takes all wells from the Process component (such as a 100x100 area, but it can be larger/smaller depending on the zoom level), and computes the NN subtraction and mean signal for the entire area. Make sure you pick the signal mask you want in the Mask/Pick tab:

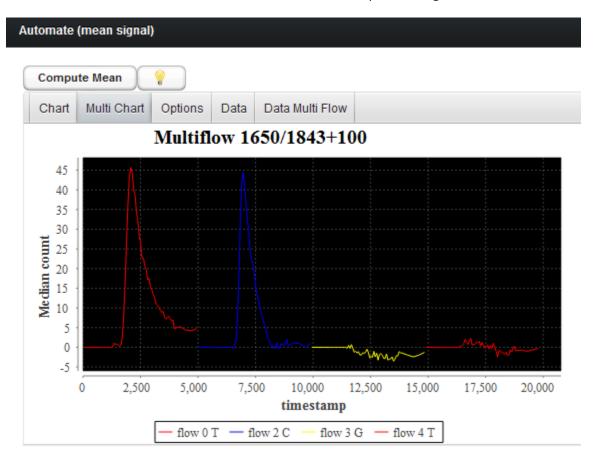


Edit masks										
Pi	ck Masks	Vie	ew	Calcula	te	Help				
Ignore mask:			0. pi	nned "	-)					
Background mask:			1. empty *							
Signal mask:		Ī	11. library 🔻							
Notes:										
The signal mask is used in Automate to compute the mean signal										
The background mask is the one used in Process for NN bg subtraction										

You can enter multiple flows in the text box (such as 1-4 or 1,2,3,4 or any other flow). Select the file type for beadfind in the Options tab (there you can also decide to subtract the result from another (empty) flow).

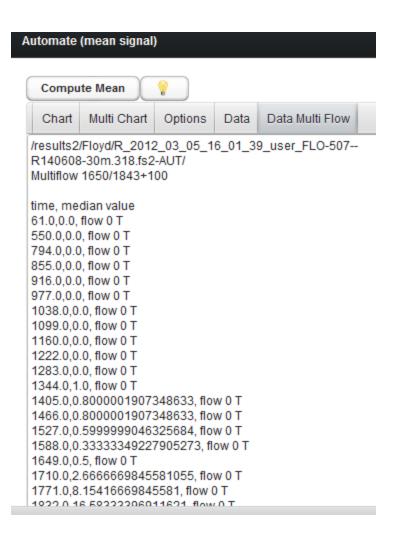


The Multi Chart tab shows the same data, but the flows are spread along the x axis:



You can export the data to excel with the Data tabs. In Excel, use Paste/Text Import Wizard/comma as separator.

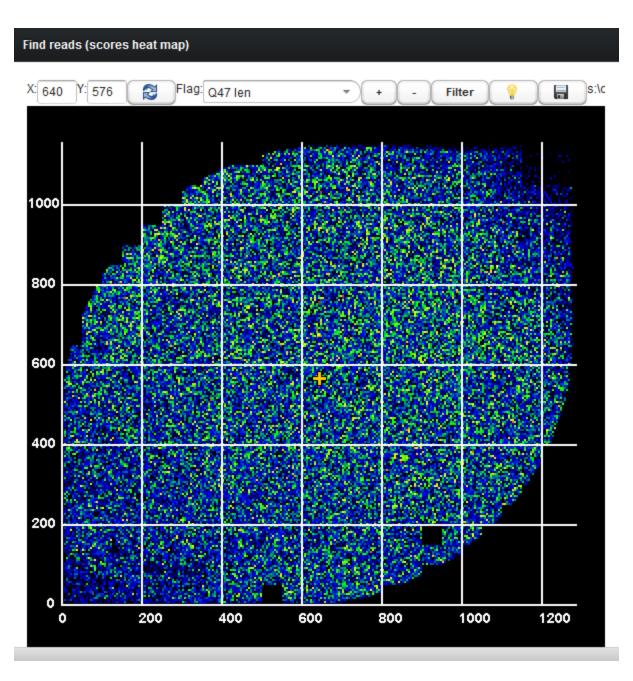




Find reads

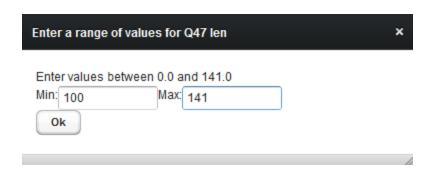
The find Reads component shows heat maps of scores such as Q17 length, and lets you filter the reads:

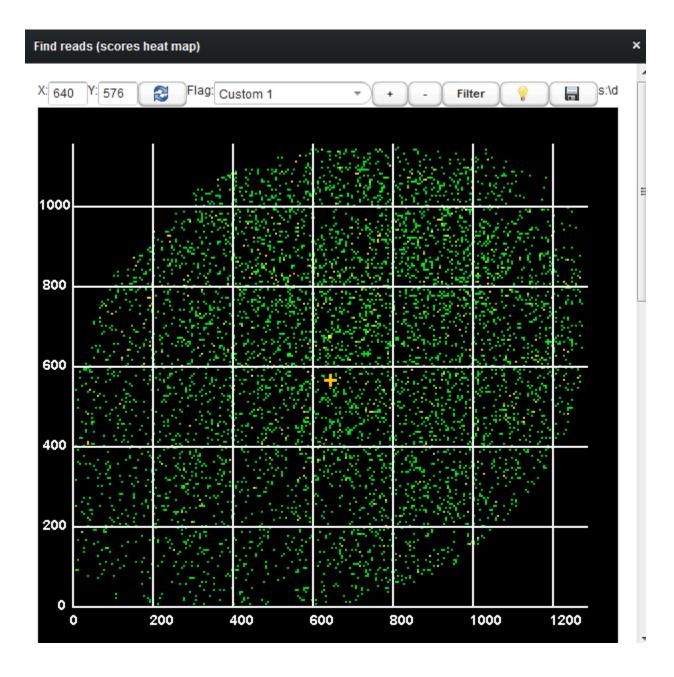




Select the filter button to only show wells that have certain values:

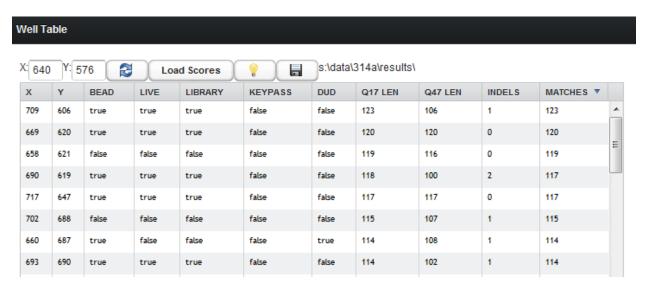








If you now select a well by moving the orange cursor, and open the well table, it will only select wells that fit the search criteria. In this case, only wells with a Q47 length of at least 100:

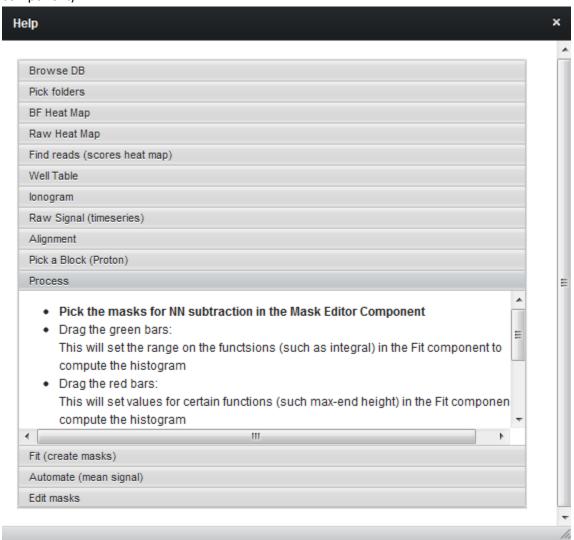


If there are fewer than 5000 wells in the entire chip that fit the result, it will load them all into the table.



Help

The help menu item shows a short description for each component (same as the light bulb icon in each component).





Troubleshooting and FAQ

I can't move the scroll bar

To move a scroll bar of any windows, first activate it by clicking on the title bar

I see a wait symbol and the GUI doesn't seem to respond

If your server is far away (network wise) then it might take a while to process clicks from the client. It is better to wait until the server response is back, otherwise it might make it worse:-). It could also indicate an error (is there a red exclamation point? If so move your mouse over it and you will see a stack trace)

I see a red exclamation point

This indicates an error. If you move your mouse over it, you will see the exception. If it is a memory error, see below on how to fix it. Otherwise, if you don't know what the issue is or it seems like a bug, let me know ©.

The application stopped working, and there is an error message saying "OutOfHeapSpace"

Since the application runs on the server, it can only handle 1-2 users at the same time as all the data is loaded in the **server** memory. You could increase the server memory (if your server has enough) by changing the setting in **/etc/default/tomcat6** and restarting tomcat. It is the line with:

JAVA_OPTS="-Djava.awt.headless=true -Xms2048m -Xmx6096m -XX:MaxPermSize=512M"

How can I check the log file for errors?

sudo tail -f /var/logs/tomcat6/catalina.out

How do I restart tomcat?

sudo /etc/init.d/tomcat6 restart

How do I deploy the app?

Copy the TSL.war file to /var/lib/tomcat6/webapps (you might have to restart with sudo /etc/init.d/tomcat6 restart)

How can Luse the Tomcat web admin interface?

Edit the file /etc/tomcat6/tomcat-users.xml:

```
<tomcat-users>
    <user username="tomcat" password="tomcat" roles="admin, manager, manager-gui"/>
</tomcat-users>
```

How do I access the Tomcat web admin interface?

http://yourserver.com:8080/manager/html (enter username/pw as specified in the file tomcatusers.xml)

