





## Draft Genome Sequence of *Micromonospora* sp. Strain WMMA1996, a Marine Sponge-Associated Bacterium

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ABSTRACT Micromonospora sp. strain WMMA1996 was isolated in 2013 off the coast of the Florida Keys, United States, from a marine sponge as part of bacterial coculture-based drug discovery initiatives. Analysis of the  $\sim$ 6.44-Mb genome reveals this microbe's potential role in the discovery of new drugs.

embers of the genus Micromonospora are important producers of antibiotics, having yielded more than 700 compounds of medical value over the years (1). Compounds such as the aminoglycoside antibiotics gentamicin (2, 3) and netilmicin (2), the antitumor antibiotics lomaiviticins (4–7) and tetrocarcins (8, 9), the anthracycline antibiotics (10–12), and the enediyne calicheamicin (13–15) highlight both the structural diversity and medicinal impact of Micromonospora-derived natural products. In addition to compounds with clear antimicrobial or anticancer activities, members of the Micromonospora have produced natural products such as juvenimicin C (16), an activator of phase II detoxifying enzymes with cancer chemopreventive activities, and diazepinomicin, a farnesylated dibenzodiazepine with antioxidant and antiproteolytic activities proposed to protect against an assortment of age-related diseases such as diabetes, atherosclerosis, and various cancers (17).

In addition to serving as important "stand-alone" producers of natural products beneficial to human health, the Micromonospora have important applications within coculture systems that have begun to emerge. Coculturing of microorganisms has proven an effective means of activating otherwise dormant biosynthetic gene clusters (BGCs) to generate otherwise unattainable natural products. For instance, we recently discovered the novel antimicrobial agent keyicin using a Micromonospora sp. WMMB285/Rhodococcus sp. WMMA185 coculture system; the Micromonospora sp. proved to be the keyicin producer (18). In a similar fashion, metabolomics analyses of Dietzia sp. WMMA184/Micromonospora sp. WMMA1996 cocultures revealed Dietziadependent production of a number of small polyketide natural products by the Micromonospora sp. Despite such advances, and the clear historical importance of Micromonospora in drug discovery, little genomic information is available for these microbes relative to other actinomycetes. In contrast, the life cycle traits and habitats of these organisms and their diverse applications (most recently focused on biofuel production) have been rigorously investigated (19).

Micromonospora sp. strain WMMA1996 was isolated in 2013 from a marineassociated sponge (Tedania sp.) collected off the coast of the Florida Keys, United States, and its complete genome was sequenced by the University of Washington PacBio Sequencing Service using PacBio RS II (Pacific Biosciences) technology. Reads were constructed into a total of 10 contigs using the Canu v. 1.4 assembler; associated contigs ranged in size from 8 kb to 4.1 Mb (20). Open reading frames were predicted by Prodigal (21) and annotated using HMMer models for the TIGRfam (22), KEGG (23), and PFAM (21, 22) databases. The genome has 73.76% GC content. The organism's

Received 23 January 2018 Accepted 1 February 2018 Published 22 February 2018

Citation Braun DR, Chevrette MG, Acharya DD, Currie CR, Rajski SR, Bugni TS. 2018. Draft genome sequence of Micromonospora sp. strain WMMA1996, a marine spongeassociated bacterium. Genome Announc 6:e00077-18. https://doi.org/10.1128/genomeA

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secondary metabolic content and potential were assessed on the basis of Anti-SMASH 4.0 (24) and PRISM (25). The Micromonospora sp. WMMA1996 genome was found to contain, but not be limited to, one type I polyketide (PKS), two type II PKS, one type III PKS, one nonribosomal peptide synthetase (NRPS) system, two type I PKS-NRPS hybrids, and four terpene biosynthetic gene clusters. The wealth of biosynthetic diversity housed within the Micromonospora WMMA1996 genome is unsurprising in light of our metabolomics analyses of Dietzia sp. WMMA184/Micromonospora sp. WMMA1996 cocultures, which revealed that the production of low-molecular weight (MW) polyketides is induced in coculture.

Accession number(s). The complete genome of Micromonospora sp. WMMA1996 has been deposited at the DDBJ/EMBL/GenBank under the project accession number PDHU00000000, which correlates to Bioproject number PRJNA407783.

## **ACKNOWLEDGMENTS**

This work was supported by the National Institutes of Health grants R01-GM104192 (T.S.B.) and U19-Al109673 (C.R.C.).

## **REFERENCES**

- 1. Talukdar M, Das D, Bora C, Bora TC, Deka Boruah HP, Singh AK. 2016. Complete genome sequencing and comparative analyses of broadspectrum antimicrobial-producing Micromonospora sp. HK10. Gene 594: 97-107. https://doi.org/10.1016/j.gene.2016.09.005.
- 2. Noone P. 1984. Sisomicin, netilmicin and dibekacin. A review of their antibacterial activity and therapeutic use. Drugs 27:548-578. https://doi .org/10.2165/00003495-198427060-00003.
- 3. Kumar CG, Himabindu M, Jetty A. 2008. Microbial biosynthesis and applications of gentamicin: a critical appraisal. Crit Rev Biotechnol 28: 173-212. https://doi.org/10.1080/07388550802262197.
- 4. He H, Ding WD, Bernan VS, Richardson AD, Ireland CM, Greenstein M, Ellestad GA, Carter GT. 2001. Lomaiviticins A and B, potent antitumor antibiotics from Micromonospora Iomaivitiensis. J Am Chem Soc 123: 5362-5363. https://doi.org/10.1021/ja010129o.
- 5. Woo CM, Beizer NE, Janso JE, Herzon SB. 2012. Isolation of Iomaiviticins C-E, transformation of Iomaiviticin C to Iomaiviticin A, complete structure elucidation of lomaiviticin A, and structure-activity analyses. J Am Chem Soc 134:15285-15288. https://doi.org/10.1021/ja3074984.
- 6. Woo CM, Gholap SL, Herzon SB. 2013. Insights into Iomaiviticin biosynthesis. Isolation and structure elucidation of (-)-homoseongomycin. J Nat Prod 76:1238-1241. https://doi.org/10.1021/np400355h.
- 7. Xue M, Herzon SB. 2016. Mechanism of nucleophilic activation of (-)lomaiviticin A. J Am Chem Soc 138:15559-15562. https://doi.org/10 .1021/jacs.6b09657.
- Fang J, Zhang Y, Huang L, Jia X, Zhang Q, Zhang X, Tang G, Liu W. 2008. Cloning and characterization of the tetrocarcin A gene cluster from Micromonospora chalcea NRRL 11289 reveals a highly conserved strategy for tetronate biosynthesis in spirotetronate antibiotics. J Bacteriol 190: 6014-6025. https://doi.org/10.1128/JB.00533-08.
- 9. Gui C, Zhang S, Zhu X, Ding W, Huang H, Gu YC, Duan Y, Ju J. 2017. Antimicrobial spirotetronate metabolites from marine-derived Micromonospora harpali SCSIO GJ089. J Nat Prod 80:1594-1603. https://doi.org/10 .1021/acs.jnatprod.7b00176.
- 10. Gewirtz DA. 1999. A critical evaluation of the mechanisms of action proposed for the antitumor effects of the anthracycline antibiotics adriamycin and daunorubicin. Biochem Pharmacol 57:727-741. https://doi .org/10.1016/S0006-2952(98)00307-4.
- 11. Kizek R, Adam V, Hrabeta J, Eckschlager T, Smutny S, Burda JV, Frei E, Stiborova M. 2012. Anthracyclines and ellipticines as DNA-damaging anticancer drugs: recent advances. Pharmacol Ther 133:26-39. https:// doi.org/10.1016/j.pharmthera.2011.07.006.
- 12. Sági JC, Kutszegi N, Kelemen A, Fodor LE, Gézsi A, Kovács GT, Erdélyi DJ, Szalai C, Semsei ÁF. 2016. Pharmacogenetics of anthracyclines. Pharmacogenomics 17:1075-1087. https://doi.org/10.2217/pgs-2016-0036.
- 13. Nicolaou KC, Pitsinos EN, Theodorakis EA, Saimoto H, Wrasidlo W. 1994. Synthetic calicheamicin mimics with novel initiation mechanisms: DNA cleavage, cytotoxicity, and apoptosis. Chem Biol 1:57-66. https://doi .org/10.1016/1074-5521(94)90041-8.

- 14. Nicolaou KC, Smith AL, Yue EW. 1993. Chemistry and biology of natural and designed enediynes. Proc Natl Acad Sci U S A 90:5881-5888.
- 15. Thorson JS, Sievers EL, Ahlert J, Shepard E, Whitwam RE, Onwueme KC, Ruppen M. 2000. Understanding and exploiting nature's chemical arsenal: the past, present and future of calicheamicin research. Curr Pharm Des 6:1841-1879. https://doi.org/10.2174/1381612003398564.
- 16. Carlson S, Marler L, Nam SJ, Santarsiero BD, Pezzuto JM, Murphy BT. 2013. Potential chemopreventive activity of a new macrolide antibiotic from a marine-derived Micromonospora sp. Mar Drugs 11:1152-1161. https://doi.org/10.3390/md11041152.
- 17. Abdelmohsen UR, Szesny M, Othman EM, Schirmeister T, Grond S, Stopper H, Hentschel U. 2012. Antioxidant and anti-protease activities of diazepinomicin from the sponge-associated Micromonospora strain RV115. Mar Drugs 10:2208-2221. https://doi.org/10.3390/md10102208
- 18. Adnani N, Chevrette MG, Adibhatla SN, Zhang F, Yu Q, Braun DR, Nelson J, Simpkins SW, McDonald BR, Myers CL, Piotrowski JS, Thompson CJ, Currie CR, Li L, Rajski SR, Bugni TS. 2017. Coculture of marine invertebrateassociated Bacteria and interdisciplinary technologies enable biosynthesis and discovery of a new antibiotic, keyicin. ACS Chem Biol 12:3093-3102. https://doi.org/10.1021/acschembio.7b00688.
- 19. Hirsch AM, Valdés M. 2010. Micromonospora: an important microbe for biomedicine and potentially for biocontrol and biofuels. Soil Biol Biochem 42:536-542. https://doi.org/10.1016/j.soilbio.2009.11.023.
- 20. Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722-736. https://doi .org/10.1101/gr.215087.116.
- 21. Hyatt D, Chen G-L, LoCascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. BMC Bioinformatics 11:119. https://doi.org/10.1186/1471 -2105-11-119
- 22. Haft DH, Selengut JD, Richter RA, Harkins D, Basu MK, Beck E. 2013. TIGRFAMs and genome properties in 2013. Nucleic Acids Res 41: D387-D395. https://doi.org/10.1093/nar/gks1234.
- 23. Kanehisa M, Goto S, Sato Y, Kawashima M, Furumichi M, Tanabe M. 2014. Data, information, knowledge and principle: back to metabolism in KEGG. Nucleic Acids Res 42:D199-D205. https://doi.org/10.1093/nar/
- 24. Blin K, Wolf T, Chevrette MG, Lu X, Schwalen CJ, Kautsar SA, Suarez Duran HG, de Los Santos ELC, Kim HU, Nave M, Dickschat JS, Mitchell DA, Shelest E, Breitling R, Takano E, Lee SY, Weber T, Medema MH. 2017. antiSMASH 4.0 —improvements in chemistry prediction and gene cluster boundary identification. Nucleic Acids Res 45:W36-W41. https://doi .org/10.1093/nar/gkx319.
- 25. Skinnider MA, Dejong CA, Rees PN, Johnston CW, Li H, Webster AL, Wyatt MA, Magarvey NA. 2015. Genomes to natural products PRediction Informatics for Secondary Metabolomes (PRISM). Nucleic Acids Res 43: 9645-9662. https://doi.org/10.1093/nar/gkv1012.