



# Cancer stem cells and brain tumors: uprooting the bad seeds

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**KEYWORDS:**  
glioma, medulloblastoma,  
progenitor cell, self-renewal, stem  
cell, stem cell niche, tumor  
microenvironment, tumor  
suppressor gene

The cancer stem cell (CSC) hypothesis is predicated on the idea that not all cells have equal proliferative potential and that, in brain tumors, the cells with the greatest ability to proliferate and contribute to tumorigenesis have phenotypic and functional properties similar to normal neural stem cells (NSCs). Over the past few years, multiple investigators have shown that CSCs isolated from human brain tumors (glioma and medulloblastoma) undergo self-renewal and multilineage cell differentiation, similar to normal NSCs. In addition, CSCs from these tumors, when implanted into rodent brains, generate tumors histologically identical to the parental tumors, suggesting that progenitor/stem cells can fully recapitulate the neoplastic phenotype *in vivo*. While these seminal studies clearly highlight the central role of stem cells in brain tumors, they also evoke important questions regarding the importance of these unique cells to tumor initiation, maintenance and treatment.

*Expert Rev. Anticancer Ther.* 7(11), 1581–1590 (2007)

## Brain tumors

Tumors of the nervous system represent the leading cause of cancer-related death in children and the fourth leading cause in adults [1]. Despite decades of research and clinical experience, the life expectancy of an adult diagnosed with a high-grade brain tumor is typically less than 1 year [2]. Our ability to develop new therapies for these deadly cancers is highly dependent on an improved understanding of the molecular and cellular changes that contribute to their development and continued growth.

Brain tumors are currently classified by the WHO according to the normal cell type in the brain that the tumor resembles most [3]. Based on this classification scheme, neuropathologists distinguish between glial cell tumors (e.g., astrocytomas, oligodendrogliomas and ependymomas) and neuronal tumors (e.g., medulloblastomas). While the WHO histologic classification system implies a cell of origin for brain tumors, in fact, the cell of origin has not been unequivocally identified for either glial or neuronal cell tumors. Recent studies have suggested that brain tumors are composed of numerous different

cell types, including cells resembling neural stem cells (NSCs), that each might contribute to tumorigenesis and continued tumor growth (maintenance). In this article, we will review what is known about brain tumor stem cells relevant to developing new approaches for treating these clinically challenging cancers.

## Heterogeneity of brain tumors

It has long been recognized that cancers are composed of heterogeneous populations of different cell types. In this regard, brain tumors comprise a complex microcosm of both neoplastic and non-neoplastic cells that each have distinct roles in dictating tumor formation and growth (FIGURE 1). The non-neoplastic (stromal) compartment contains immune system cells (microglia), entrapped normal cells (neurons, astrocytes, oligodendrocytes) and tumor-associated blood vessels. Taking a page from studies of other solid tumors, recent reports have shown that important cellular and molecular signals that emanate from the brain tumor microenvironment are critical for regulating neoplastic tumor cell growth. In this regard, high numbers of monocyte-like cells (microglia) are consistently found within

gliomas [4,5] and the number of these cells has been suggested to correlate with tumor growth [6]. Using both *in vitro* and *in vivo* experimental systems, microglia have been shown to provide critical growth signals that promote brain tumor growth [7]. Microglia are a rich source of growth factors and cytokines, including  $\text{TNF-}\alpha$ , EGF and chemokines, that increase brain tumor proliferation and migration [8,9]. Support for an important role for microglia in brain tumor proliferation derives from recent studies demonstrating that resident brain microglia contribute to brain tumor growth in a mouse model of neurofibromatosis-1 (NF1)-associated optic glioma [10].

In addition, microglia are important generators of chemokines, including stroma-derived factor-1 $\alpha$  (SDF-1 $\alpha$  or CXCL12). Studies in both medulloblastoma and glioma cell lines have shown that the chemokine, CXCL12, can regulate brain tumor growth in explant models *in vivo* [11,12]. Moreover, CXCL12 expression in the tumor microenvironment may also dictate where and when optic nerve gliomas grow in patients with NF1 [13]. In these studies, the restricted expression of CXCL12 concentrated along the optic nerve and chiasm in young mice parallels the growth pattern of

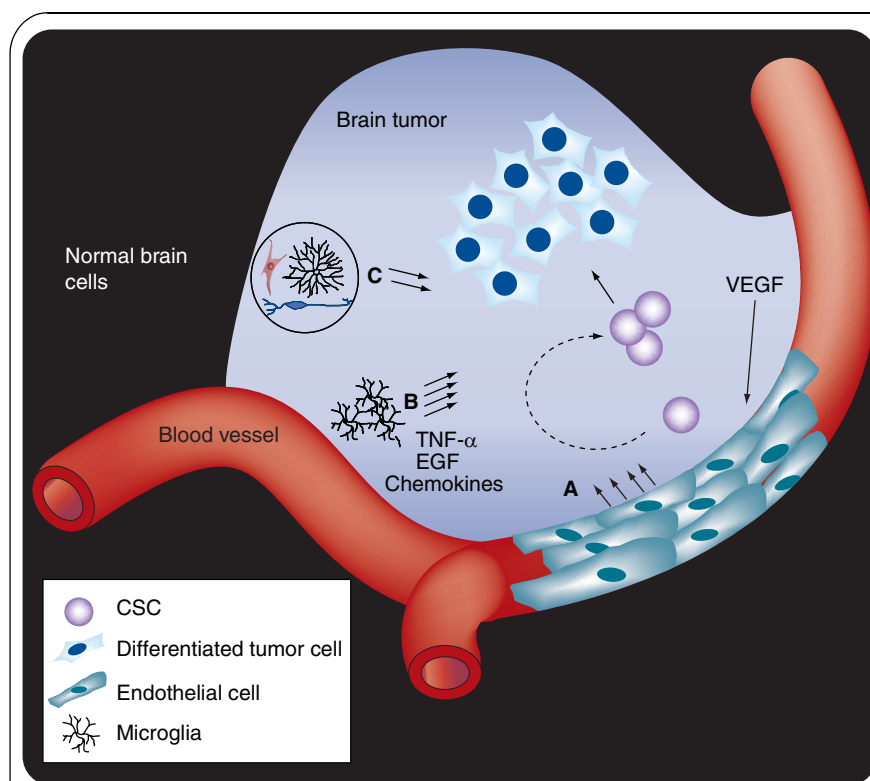
NF1-associated gliomas within the optic nerve and chiasm in children [14]. While no direct evidence currently exists to support a direct role for microglia in cancer stem cell (CSC) growth or function, CXCL12 promotes NSC proliferation [15] and may serve as a homing signal for stem cells during development and cancer [16].

Similar to microglia, tumor blood vessels have also been proposed to regulate brain tumor growth by providing essential nutrients and oxygen [17]. Recent studies have suggested that malignant glioma CSCs release VEGF and promote angiogenesis in the tumor microenvironment [18]. In addition, the tumor vasculature may create a unique stromal niche that maintains stem cells and progenitor cells [19–21]. The finding that endothelial cells produce novel growth factors, including the novel pigment epithelium-derived factor, which supports normal stem cell self-renewal, provides a molecular mechanism underlying the unique tropism of NSCs for the vascular niche [22].

### Cancer stem cells

The neoplastic compartment of brain tumors contains both progenitor (stem) cells and differentiated cells. Recent studies have focused on CSCs, based on experiments that suggest that these cells might be critical for generating or maintaining tumors. CSCs have been successfully identified and isolated in various cancers, including leukemia [23] and breast [24], pancreatic [25], head and neck [26], colon [27] and prostate cancers [28]. Several studies report that CSCs comprise a small percentage of the total cells in brain tumors, ranging from less than 1% in low-grade brain tumors to as many as a quarter of all cells in high-grade brain tumors using sphere formation and clonogenic assays [29–32]. However, only a small population of CSCs in solid cancers is clonogenic, and only one in 1000 to one in 5000 lung cancer, ovarian cancer or neuroblastoma CSCs can actually generate colonies in soft agar [33].

In brain tumors, CSCs, like their non-neoplastic NSC counterpart, are defined by their functional properties. Normal NSCs have the capacity for proliferation, self-renewal and the generation of multiple terminally differentiated cell types. CSCs isolated from human brain tumors can be maintained as neurospheres, similar to NSCs, and can generate new neurospheres from a single CSC (self-renewal) [30–32,34–36]. In addition, CSCs are also capable of differentiating into cellular lineages that arise from normal



**Figure 1. Tumor microcosm.** Brain tumors are composed of heterogeneous populations of both neoplastic cells and non-neoplastic cells. The non-neoplastic cell population contains microglia (monocyte-like cells), endothelial cells ('angiogenic niche') and normal brain cells. (A) CSCs in the angiogenic niche receive signals which induce CSC self-renewal (generation of new CSCs) or differentiation into more mature cancer cell types. (B) Microglia increase tumor growth and migration by releasing growth factors (e.g., EGF), cytokines (e.g.,  $\text{TNF-}\alpha$ ) and chemokines (e.g., CXCL12). (C) Normal brain cells (neurons, oligodendrocytes and astrocytes) in the tumor microenvironment also promote tumor growth. In addition, brain tumors express high levels of VEGF, which positively regulate endothelial cell proliferation. The dotted line denotes CSC proliferation and self-renewal. CSC: Cancer stem cell.

NSCs, including neurons, astrocytes and oligodendrocytes *in vitro* (multilineage differentiation), even though they are not functionally normal brain cells [29,30,34,35].

In addition to functional properties, immunocytochemical markers have also been used to identify NSCs and CSCs, including nestin [30,37,38], CD133 [29,30,32], Musashi-1 [30], Sox2 [30,39], brain lipid-binding protein (BLBP) [31,40] and maternal embryonic leucine zipper kinase (MELK) [30,41]. Of all these potential proteins, the most frequently used immunomarker is the cell surface protein, CD133.

Using CD133 to isolate brain tumor CSC populations, several investigators have shown that CD133<sup>+</sup> cells exhibit a greater ability than CD133<sup>-</sup> cells to form tumors in immunocompromised rodent brains *in vivo* [29,32,42]. CD133<sup>+</sup> CSCs were also capable of forming brain tumors at low cell concentrations, consistent with their high self-renewal capacity [36], as well as recapitulating the original tumor *in vivo* even after serial transplantations [30,32,34,43]. However, it is worth noting that a recent study demonstrated that secondary glioblastomas do not contain CD133-expressing cells, and that brain tumor stem cells in these tumors do not express CD133 [42].

Additional evidence for a direct relationship between NSCs and tumorigenesis in the brain derives from elegant studies by Ligon and colleagues [44]. They report that the CNS-restricted transcription factor, Olig-2, regulates the proliferation of neural progenitors as well as glioma-derived CSCs by modulating the function of p21<sup>WAF/CIP1</sup>. Moreover, they demonstrate that Olig-2 is required for glioma formation from NSCs. These exciting findings suggest that CSCs are critical for gliomagenesis.

NSCs live in specialized niches, typically in the subventricular zone, the lining of the ventricles and the hippocampal dentate gyrus [45–47]. These niches are important for maintaining NSCs and for providing extrinsic factors that control cell proliferation and cell fate [22,48,49]. Similar to NSCs, Calabrese and colleagues demonstrated that nestin<sup>+</sup>/CD133<sup>+</sup> brain tumor CSCs are maintained within vascular niches [19]. They also found that endothelial cell-derived factors in these specialized locations support human brain tumor xenograft growth in mice.

#### Growth control pathways

Some of the extracellular stromal signals that regulate cell proliferation and fate in normal NSCs are often co-opted during tumor formation to allow the tumor cell to escape normal growth regulation. The most frequently reported mutations involve three important signaling pathways specifying neuroglial cell specification and proliferation: the Sonic hedgehog (SHH), Wnt and receptor tyrosine kinase (RTK) signaling pathways (FIGURE 2).

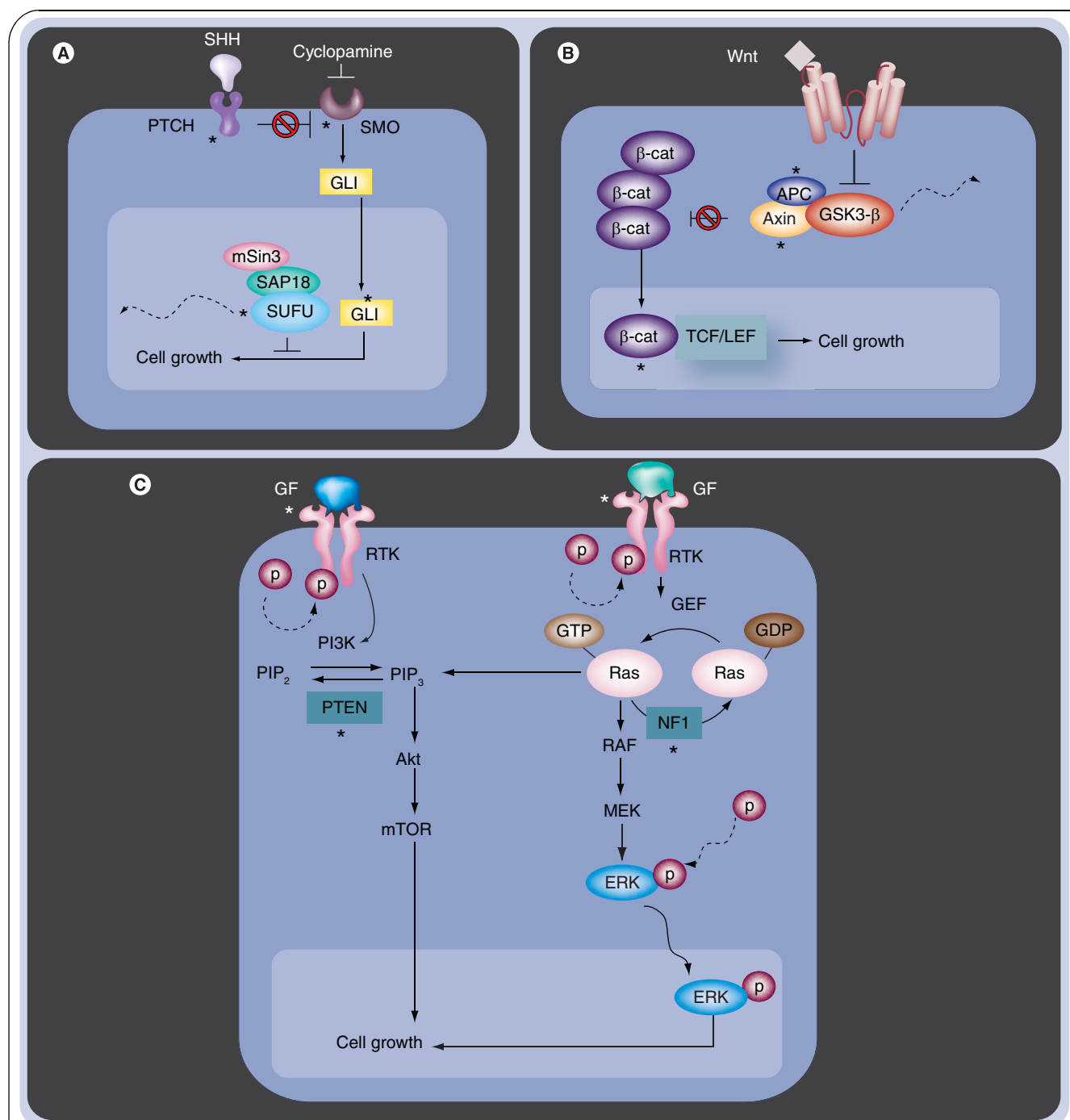
The identification of SHH pathway signaling abnormalities in brain tumors derives from initial studies of the inherited basal cell nevus cancer predisposition syndrome (Gorlin syndrome) in which a small proportion of affected patients develop medulloblastoma. Gorlin syndrome is caused by a germline

mutation in the SHH receptor, PATCHED1 (PTCH) gene. Analysis of sporadic medulloblastomas revealed *PTCH* inactivation in approximately 20% of tumors, and mice engineered to harbor a germline *Ptch* mutation develop medulloblastoma [50,51]. Other proteins involved in SHH signaling are also mutated or deregulated in brain tumors, including smoothened (SMO), the glioma-associated oncogene homologue 1–3 (*Gli1*, *Gli2*, *Gli3*) transcription factors and suppressor of fused (SUFU) [52–55]. During normal cerebellar development, Purkinje cell-derived SHH regulates the proliferation of granule precursor cells in the external granule layer (EGL) and stem cells in the developing neocortex [53,56,57]. The SHH–Gli pathway also influences foliation during cerebellum development by regulating the position and/or size of the lobes [58,59]. Increased *SHH–Gli* pathway signaling not only enhances NSC proliferation and neurogenesis, but is also necessary for brain tumorigenicity and growth [50,60,61].

Similarly, defects in Wnt/β-catenin pathway signaling in brain tumors were initially identified by studying patients with another inherited cancer predisposition syndrome, Turcot syndrome. Patients with Turcot syndrome develop medulloblastoma as a result of a germline mutation in the adenomatous polyposis coli (*APC*) gene [62]. Although mutations in the *APC* gene are rare in sporadic medulloblastoma, some tumors will harbor mutations in the β-catenin or Axin genes [63,64]. The Wnt/β-catenin axis is important for specifying the mid-brain–hindbrain boundary during development [65,66], and controls NSC proliferation and self-renewal [67,68].

RTK signaling through the EGF and PDGF receptors is a prime driver of NSC proliferation and differentiation. RTK activation leads to downstream activation of several key signaling molecules, including RAS and AKT. Although activating mutations in RAS are rare in human brain tumors, RAS can be activated by several mechanisms, including increased expression of PDGF receptors, mutational activation of the EGF receptor, and loss of the *NF1* tumor suppressor gene. Mutational activation and overexpression of the EGF receptor is observed in a third of high-grade gliomas, and cooperates with other genetic changes to promote gliomagenesis and malignant progression [69,70]. Similarly, PDGF receptor-expressing cells in the adult subventricular zone are NSCs that can form glioma-like growths in response to ectopic PDGF expression [71]. The importance of PDGF signaling in gliomagenesis is underscored by several additional observations. First, PDGF autocrine signaling regulates glioma survival [72]. Second, PDGF expression in nestin<sup>+</sup> progenitor cells results in glioma formation in mice *in vivo* [73]. Finally, a transforming PDGF receptor mutant has been identified in a high-grade glioma [74].

Children with the *NF1* inherited tumor predisposition syndrome are prone to the development of low-grade gliomas. The *NF1* tumor suppressor gene encodes a large cytoplasmic protein, termed neurofibromin, which negatively regulates RAS in astrocytes and other neural cell types. Moreover, neurofibromin controls NSC proliferation and self-renewal in a gene dose-dependent fashion *in vitro* [75]. Loss of neurofibromin expression in



**Figure 2. Signaling pathways involved in both neural stem cell proliferation and tumorigenesis.** Signaling pathways that involve SHH, WNT and RTKs activated by various GFs (e.g., EGF and PDGF) regulate normal neural stem cell (NSC) self-renewal, as well as brain tumor growth. **(A)** Mutations in the *PTCH*, *SMO* and *SUFU* genes are frequently observed in medulloblastoma (\*). *PTCH* mutations lead to hyperactivation of *SMO*, resulting in increased *GLI* expression. *SUFU* acts as a repressor of *GLI*, and increased *GLI* function promotes cell growth. The dotted line denotes dissociation of *SUFU* from the signaling complex. **(B)** WNT signaling regulates cell growth by controlling  $\beta$ -cat levels and  $\beta$ -cat nuclear translocation. The protein complex containing Axin, APC and GSK-3 $\beta$  acts as a negative regulator of  $\beta$ -cat by promoting  $\beta$ -cat degradation. Mutations in Axin, APC and  $\beta$ -cat have been reported in medulloblastoma (\*). The dotted line denotes dissociation of GSK-3 from the signaling complex. **(C)** RTK signaling initiated by the binding of mitogenic GFs (e.g., EGF and PDGF) increase proliferation in both normal NSCs and brain tumor cells. RTK binding by GFs leads to activation of several downstream signaling pathways that regulate cell survival and proliferation, including the Ras, Akt, mTOR and MAPK signaling intermediates. PTEN and NF1 function as tumor suppressors and inhibit signaling through these RTK pathways. Brain tumors (gliomas) have been reported to harbor mutations in the *PTEN*, *NF1* and *EGFR* genes or overexpress the PDGF receptor (\*). The dotted lines denote protein phosphorylation.  $\beta$ -cat:  $\beta$ -catenin; APC: Adenomatous polyposis coli; GEF: GTP exchange factor; GF: Growth factor; *GLI*: Glioma-associated oncogene homolog; GSK: Glycogen synthase kinase; LEF: Lymphoid enhancer-binding factor; NF: Nuclear factor; NF1: Neurofibromatosis 1 tumor suppressor, neurofibromin; P: Phosphate; PI3K: Phosphoinositide 3-kinase; PIP<sub>2</sub>: Phosphatidylinositol-4,5-bisphosphate; PIP<sub>3</sub>: Phosphatidylinositol-3,4,5-trisphosphate; RTK: Receptor tyrosine kinase; TCF: T-cell-specific transcription factor.

NSCs results in delayed maturation of both glial and neuronal cell lineages *in vivo* [40]. Moreover, glial progenitor *Nfl* inactivation results in low-grade glioma formation in the optic nerve and chiasm of genetically engineered mice *in vivo* [76,77].

Similarly, AKT activation does not result from mutational activation, but most often reflects loss of the phosphatase and tensin homolog (PTEN) tumor suppressor gene. PTEN regulates NSC proliferation *in vitro* and *in vivo* [78,79], but inactivation or loss of this gene accelerates high-grade glioma development *in vivo* [69,80].

Finally, many intracellular growth control pathways converge on cell-cycle growth regulators, including the p53 and cyclin-dependent protein kinase genes. High-grade gliomas frequently harbor mutations in the *TP53* (p53) tumor suppressor gene and *INK4a* (p16) cyclin-dependent kinase gene [81]. p53 functions as a negative regulator of self-renewal, proliferation and survival of NSCs *in vitro* [82]. Similarly, p16 is important for regulating NSC self-renewal and proliferation in the CNS [83,84]. Cooperativity between the *p53* and *Rb* pathway genes in promoting gliomagenesis has been demonstrated in genetically engineered mice in which loss of *Nfl* and *p53* or *Rb* and *p53* occurs [85–87].

#### Region-specific neural stem cells

A number of recent studies have suggested that NSCs in the brain are not homogeneous cellular populations, such that NSCs from different brain regions have different biological properties (self-renewal, proliferation and differentiation potential) [88–91]. In this regard, NSCs from the E14.5 mouse mid-brain–hindbrain exhibit greater self-renewal and lower neurogenic potential than identical-appearing NSCs isolated from the cortex. Forebrain-derived NSCs exhibit greater growth capacity than those isolated from the spinal cord or cerebellum, while optic nerve NSCs have the lowest capacity for self-renewal.

In addition to these differences in functional properties, NSCs from different brain regions also harbor brain region-specific gene expression patterns [88]. These brain region-specific genetic signatures are also retained in normal glia as well as glial tumors originating from these different brain regions [31,92]. These results suggest that NSCs from different brain regions are unique and give rise to differentiated progeny and tumors that harbor these distinct molecular profiles. These data also support the hypothesis that heterogeneity among pilocytic astrocytomas (PAs) based on brain region might be the result of innate functional differences in NSCs from different brain regions. It is tempting to speculate that these genetic signatures represent hard-wired gene expression programs that modify the ability of normal and preneoplastic cells in specific brain regions to respond to tumor-causing genetic changes and microenvironmental signals [93].

#### Expert commentary & five-year view

While it is not clear whether brain tumors originate from CSCs or are simply maintained by these progenitor cells, emerging evidence suggests that CSCs have unique properties that might

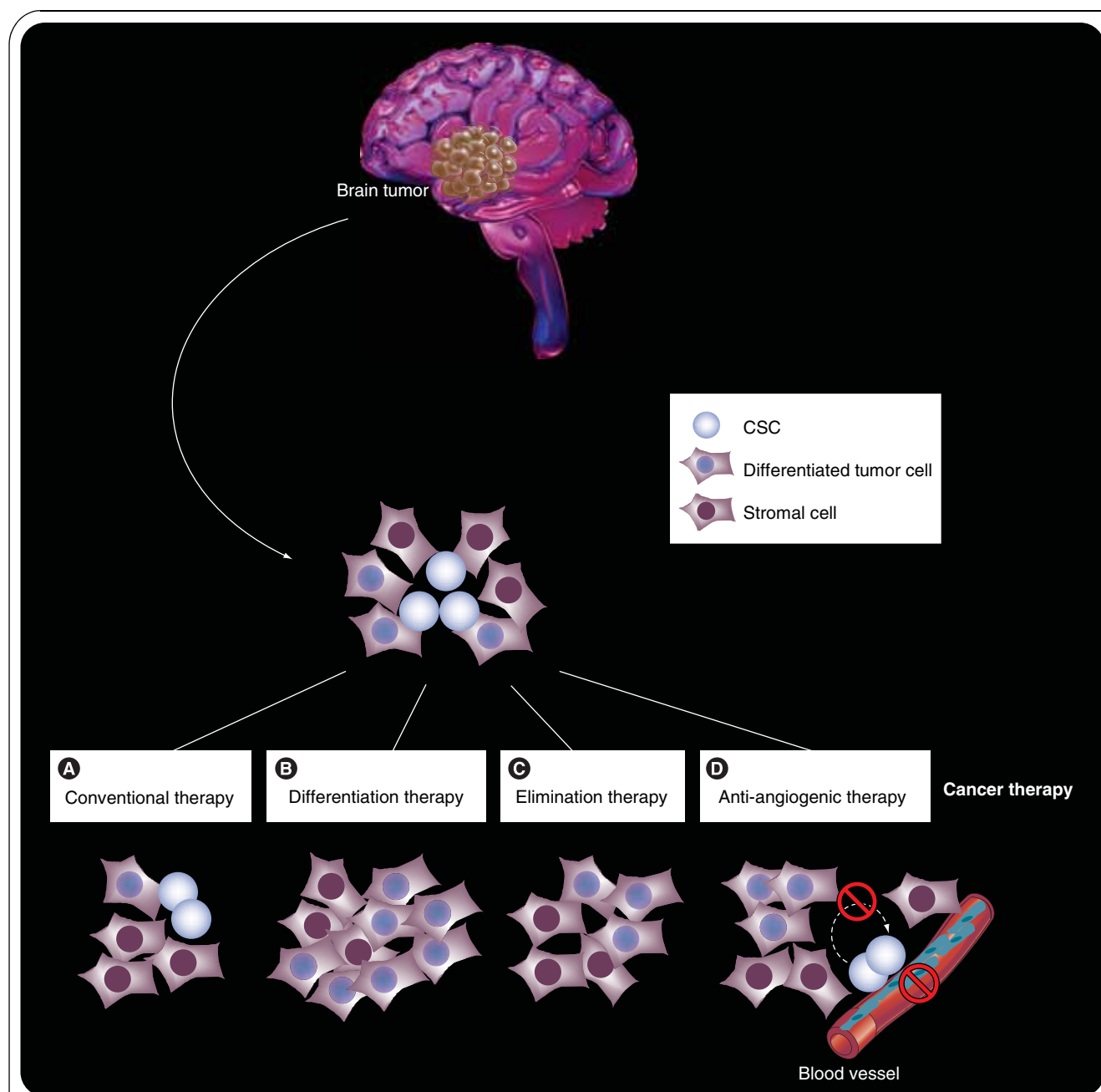
make them particularly resistant to conventional therapy. The current standard of care for many types of brain tumors includes radiation and chemotherapy. However, most high-grade brain tumors recur after treatment or fail to respond to initial therapy. Several groups have suggested that radiation-resistant stem cells in brain tumors may account for the observed resistance to conventional therapy.

One of the unique features of CSCs is an innate resistance to radiation and chemotherapy drugs [18,94–97]. In particular, the CD133<sup>+</sup> subpopulation is highly resistant to ionizing radiation in human glioma xenografts and can repair DNA damage more effectively than CD133<sup>−</sup> tumor cells [94]. In addition to relative radiation resistance, CSCs exhibit high levels of expression of multiple drug-resistance genes compared with the differentiated bulk tumor [18,96–98] and exhibit significant resistance to temozolomide, etoposide, carboplatin and paclitaxel [97].

As we learn more regarding the function of CSCs in tumorigenesis and maintenance, future therapeutic options may include specifically targeting the CSC by inducing CSC differentiation, eliminating CSCs and/or depriving the CSC of the normal regulatory cues that derive from the brain tumor microenvironment (FIGURE 3). Differentiation therapy could theoretically induce the loss of CSC self-renewal properties in brain tumors [99] and result in loss of the ‘tumor-maintaining’ cell. One of these anticancer drugs that can induce differentiation in tumors is retinoic acid (RA; vitamin A). All-*trans* RA is an effective drug for acute promyelocytic leukemia [100] and can induce differentiation in teratocarcinomas and melanomas [101]. Similarly, bone morphogenetic protein (BMP)-4 [102] and cannabinoids [103] also promote neural differentiation of stem-like, tumor-initiating precursors in human glioblastomas (GBMs). Another CSC-targeting therapy is to eliminate CSCs and/or target the key molecular signaling pathways that drive self-renewal, proliferation and cell-fate decisions in CSCs. Previous studies have shown that SHH pathway inhibitors can inhibit medulloblastoma growth in mice, whereas inhibition of the Notch pathway with  $\gamma$ -secretase inhibitors blocks CSC self-renewal and decreases mouse medulloblastoma tumor growth [104–106]. Future strategies might also take advantage of targets that derive from the analysis of gene expression profiles of tumorigenic CSCs relative to normal NSCs. Finally, strategies that target cellular and molecular signals that emanate from the tumor microenvironment and promote CSC self-renewal and proliferation might also be effective. In this regard, inhibiting VEGF and its receptor [107] or microglia function may deprive CSCs of the necessary trophic factors required for their maintenance. With continued advances in CSC biology, it is likely that additional treatments will be identified that specifically target the cell(s) most responsible for brain tumor growth.

#### Acknowledgement

We apologize to those authors whose studies we could not cite owing to space limitations.



**Figure 3. Potential cancer therapies.** Future therapies for brain tumors might target other cell types in the tumor microcosm, including CSCs and stromal cells (endothelial cells and microglia). **(A)** Conventional radiotherapy and chemotherapy result in a reduction in the overall tumor mass by killing cells with limited proliferative potential (e.g., differentiated tumor cells); however, this effect is transient and tumors typically recur when the surviving CSCs give rise to new differentiated tumor cells. **(B)** Differentiation therapy promotes CSC terminal differentiation resulting in a tumor composed predominantly of differentiated cancer cells that may be more sensitive to conventional therapy. **(C)** Elimination therapy is designed to kill the CSCs within the tumors. In the absence of these tumor-maintaining CSCs, conventional therapy may provide a more durable response. **(D)** Anti-angiogenic therapy targets the tumoral blood vessels, disrupting the angiogenic niche in the brain tumor. As a result, the CSCs lose their ability to undergo self-renewal and instead give rise to terminally differentiated tumor cells that are more likely to respond to conventional therapies. Similar strategies that eliminate tumor-promoting signals from microglia might also be effective as adjuvant agents in reducing tumor growth. The dotted line denotes CSC proliferation and self-renewal. CSC: Cancer stem cell.

#### Financial & competing interests disclosure

*This work was supported by grants from the National Institutes of Health (David H Gutmann) and Children's Tumor Foundation (Da Yong Lee). The authors have no other relevant affiliations or financial involvement with any organization or entity*

*with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.*

*No writing assistance was utilized in the production of this manuscript.*

## Key issues

- Brain tumors are composed of heterogeneous populations of neoplastic and non-neoplastic (stromal) cell types.
- Cells with stem cell-like properties can be isolated from human brain tumors and can recapitulate the original tumor when explanted into naive recipient rodent brains.
- Cancer stem cells, like normal neural stem cells, are capable of self-renewal, proliferation and multilineage differentiation.
- Cancer stem cells, like normal neural stem cells, reside in specialized niches that regulate their growth and differentiation.
- Signals that control normal neural stem/progenitor cell growth and differentiation are often mutated in human brain tumors.
- Neural stem/progenitor cells from different brain regions have unique biological properties that might dictate their response to cellular and molecular signals.
- A more complete understanding of the cellular components of brain tumors will provide unique opportunities to develop targeted therapies for these deadly cancers.

## References

Papers of special note have been highlighted as:

- of interest
- of considerable interest

- Stewart BW, Kleihues P. *World Cancer Report*. IARC Press, Lyon, France (2003).
- De Groot JF, Aldape KD, Colman H. High grade astrocytomas. In: *Principles of Neuro-Oncology*. Schiff D, O'Neill BP (Eds). McGraw Hill, NY, USA 259–288 (2005).
- Louis DN, Ohgaki H, Wiestler OD, Cavenee WK. *WHO Classification of Tumours of the Central Nervous System (4th Edition)*. IARC Press, Lyon, France (2007).
- Rossi ML, Hughes JT, Esiri MM, Coakham HB, Brownell DB. Immunohistological study of mononuclear cell infiltrate in malignant gliomas. *Acta Neuropathol. (Berl.)* 74(3), 269–277 (1987).
- Rossi ML, Jones NR, Candy E *et al*. The mononuclear cell infiltrate compared with survival in high-grade astrocytomas. *Acta Neuropathol. (Berl.)* 78(2), 189–193 (1989).
- Graeber MB, Scheithauer BW, Kreutzberg GW. Microglia in brain tumors. *Glia* 40(2), 252–259 (2002).
- Watters JJ, Schartner JM, Badie B. Microglia function in brain tumors. *J. Neurosci. Res* 81(3), 447–455 (2005).
- Platten M, Kretz A, Naumann U *et al*. Monocyte chemoattractant protein-1 increases microglial infiltration and aggressiveness of gliomas. *Ann. Neurol.* 54(3), 388–392 (2003).
- Sliwa M, Markovic D, Gabrusiewicz K *et al*. The invasion promoting effect of microglia on glioblastoma cells is inhibited by cyclosporin A. *Brain* 130(Pt 2), 476–489 (2007).
- Daginakatte GC, Gutmann DH. Neurofibromatosis-1 (NF1) heterozygous brain microglia elaborate paracrine factors that promote NF1-deficient astrocyte and glioma growth. *Hum. Mol. Genet.* 16(9), 1098–1112 (2007).
- Rubin JB, Kung AL, Klein RS *et al*. A small-molecule antagonist of CXCR4 inhibits intracranial growth of primary brain tumors. *Proc. Natl Acad. Sci. USA* 100(23), 13513–13518 (2003).
- Yang L, Jackson E, Woerner BM, Perry A, Piwnica-Worms D, Rubin JB. Blocking CXCR4-mediated cyclic AMP suppression inhibits brain tumor growth *in vivo*. *Cancer Res* 67(2), 651–658 (2007).
- Warrington NM, Woerner BM, Daginakatte GC *et al*. Spatiotemporal differences in CXCL12 expression and cyclic AMP underlie the unique pattern of optic glioma growth in neurofibromatosis type 1. *Cancer Res* 67(18), 8588–8595 (2007).
- Listernick R, Charrow J, Greenwald M, Mets M. Natural history of optic pathway tumors in children with neurofibromatosis type 1: a longitudinal study. *J. Pediatr.* 125(1), 63–66 (1994).
- Gong X, He X, Qi L, Zuo H, Xie Z. Stromal cell derived factor-1 acutely promotes neural progenitor cell proliferation *in vitro* by a mechanism involving the ERK1/2 and PI-3K signal pathways. *Cell Biol. Int.* 30(5), 466–471 (2006).
- Claps CM, Corcoran KE, Cho KJ, Rameshwar P. Stromal derived growth factor-1 $\alpha$  as a beacon for stem cell homing in development and injury. *Curr. Neurovasc. Res* 2(4), 319–329 (2005).
- Kaur B, Khwaja FW, Severson EA, Matheny SL, Brat DJ, Van Meir EG. Hypoxia and the hypoxia-inducible-factor pathway in glioma growth and angiogenesis. *Neuro. Oncol.* 7(2), 134–153 (2005).
- Bao S, Wu Q, Sathornsumetee S *et al*. Stem cell-like glioma cells promote tumor angiogenesis through vascular endothelial growth factor. *Cancer Res* 66(16), 7843–7848 (2006).
- Calabrese C, Poppleton H, Kocak M *et al*. A perivascular niche for brain tumor stem cells. *Cancer Cell* 11(1), 69–82 (2007).
- **Demonstrated that endothelial cells interact with cancer stem cells and secrete factors that maintain these cells in a stem cell-like state.**
- Farin A, Suzuki SO, Weiker M, Goldman JE, Bruce JN, Canoll P. Transplanted glioma cells migrate and proliferate on host brain vasculature: a dynamic analysis. *Glia* 53(8), 799–808 (2006).
- Folkins C, Man S, Xu P, Shaked Y, Hicklin DJ, Kerbel RS. Anticancer therapies combining antiangiogenic and tumor cell cytotoxic effects reduce the tumor stem-like cell fraction in glioma xenograft tumors. *Cancer Res* 67(8), 3560–3564 (2007).
- Ramírez-Castillejo C, Sánchez-Sánchez F, Andreu-Agulló C *et al*. Pigment epithelium-derived factor is a niche signal for neural stem cell renewal. *Nat. Neurosci.* 9(3), 331–339 (2006).
- Bonnet D, Dick JE. Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat. Med.* 3(7), 730–737 (1997).
- Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF. Prospective identification of tumorigenic breast cancer cells. *Proc. Natl Acad. Sci. USA* 100(7), 3983–3988 (2003).



- 25 Li C, Heidt DG, Dalerba P *et al*. Identification of pancreatic cancer stem cells. *Cancer Res* 67(3), 1030–1037 (2007).
- 26 Prince ME, Sivanandan R, Kaczorowski A *et al*. Identification of a subpopulation of cells with cancer stem cell properties in head and neck squamous cell carcinoma. *Proc. Natl Acad. Sci. USA* 104(3), 973–9789 (2007).
- 27 O'Brien CA, Pollett A, Gallinger S, Dick JE. A human colon cancer cell capable of initiating tumour growth in immunodeficient mice. *Nature* 445(7123), 106–110 (2007).
- 28 Collins AT, Berry PA, Hyde C, Stower MJ, Maitland NJ. Prospective identification of tumorigenic prostate cancer stem cells. *Cancer Res* 65(23), 10946–10951 (2005).
- 29 Singh SK, Clarke ID, Terasaki M *et al*. Identification of a cancer stem cell in human brain tumors. *Cancer Res* 63(18), 5821–5828 (2003).
- 30 Hemmati HD, Nakano I, Lazareff JA *et al*. Cancerous stem cells can arise from pediatric brain tumors. *Proc. Natl Acad. Sci. USA* 100(25), 15178–15183 (2003).
- 31 Taylor MD, Poppleton H, Fuller C *et al*. Radial glia cells are candidate stem cells of ependymoma. *Cancer Cell* 8(4), 323–335 (2005).
- **Demonstrated that restricted populations of radial glia cells are candidate ependymoma cancer stem cells, and suggested that subgroups of the same histologic tumor type are generated by different populations of progenitor cells in the tissues of origin.**
- 32 Singh SK, Hawkins C, Clarke ID *et al*. Identification of human brain tumour initiating cells. *Nature* 432(7015), 396–401 (2004).
- **This study was one of the first reports to describe the existence of stem-like cells in human brain tumors with the capacity for self-renewal, multilineage differentiation and recapitulation of the original tumor.**
- 33 Reya T, Morrison SJ, Clarke MF, Weissman IL. Stem cells, cancer, and cancer stem cells. *Nature* 414(6859), 105–111 (2001).
- 34 Galli R, Binda E, Orfanelli U *et al*. Isolation and characterization of tumorigenic, stem-like neural precursors from human glioblastoma. *Cancer Res* 64(19), 7011–7021 (2004).
- 35 Ignatova TN, Kukekov VG, Laywell ED, Suslov ON, Vrionis FD, Steindler DA. Human cortical glial tumors contain neural stem-like cells expressing astroglial and neuronal markers *in vitro*. *Glia* 39(3), 193–206 (2002).
- 36 Lee J, Kotliarova S, Kotliarov Y *et al*. Tumor stem cells derived from glioblastomas cultured in bFGF and EGF more closely mirror the phenotype and genotype of primary tumors than do serum-cultured cell lines. *Cancer Cell* 9(5), 391–403 (2006).
- 37 Dahlstrand J, Zimmerman LB, McKay RD, Lendahl U. Characterization of the human nestin gene reveals a close evolutionary relationship to neurofilaments. *J. Cell Sci.* 103(Pt 2), 589–597 (1992).
- 38 Tohyama T, Lee VM, Rorke LB, Marvin M, McKay RD, Trojanowski JQ. Nestin expression in embryonic human neuroepithelium and in human neuroepithelial tumor cells. *Lab. Invest.* 66(3), 303–313 (1992).
- 39 Zappone MV, Galli R, Catena R *et al*. Sox2 regulatory sequences direct expression of a (β)-geo transgene to telencephalic neural stem cells and precursors of the mouse embryo, revealing regionalization of gene expression in CNS stem cells. *Development* 127(11), 2367–2382 (2000).
- 40 Hegedus B, Dasgupta B, Shin JE *et al*. Neurofibromatosis-1 regulates neuronal and glial cell differentiation from neuroglial progenitors *in vivo* by both cAMP- and Ras-dependent mechanisms. *Cell Stem Cell* (2007) (In Press).
- 41 Nakano I, Paucar AA, Bajpai R *et al*. Maternal embryonic leucine zipper kinase (MELK) regulates multipotent neural progenitor proliferation. *J. Cell Biol.* 170(3), 413–427 (2005).
- 42 Beier D, Hau P, Proescholdt M *et al*. CD133<sup>+</sup> and CD133<sup>−</sup> glioblastoma-derived cancer stem cells show differential growth characteristics and molecular profiles. *Cancer Res* 67(9), 4010–4015 (2007).
- 43 Yuan X, Curtin J, Xiong Y *et al*. Isolation of cancer stem cells from adult glioblastoma multiforme. *Oncogene* 23(58), 9392–9400 (2004).
- 44 Ligon KL, Huillard E, Mehta S *et al*. Olig2-regulated lineage-restricted pathway controls replication competence in neural stem cells and malignant glioma. *Neuron* 53(4), 503–517 (2007).
- **Demonstrated that the CNS-restricted transcription factor, Olig-2, is required for neural progenitor proliferation and for glioma formation in mice. They show that Olig-2 functions to regulate gliomagenesis and stem cell proliferation by repressing p21 expression.**
- 45 Sanai N, Tramontin AD, Quiñones-Hinojosa A *et al*. Unique astrocyte ribbon in adult human brain contains neural stem cells but lacks chain migration. *Nature* 427(6976), 740–744 (2004).
- 46 Eriksson PS, Perfilieva E, Björk-Eriksson T *et al*. Neurogenesis in the adult human hippocampus. *Nat. Med.* 4(11), 1313–1317 (1998).
- 47 Alvarez-Buylla A, Lim DA. For the long run: maintaining germinal niches in the adult brain. *Neuron* 41(5), 683–686 (2004).
- 48 Palmer TD, Willhoite AR, Gage FH. Vascular niche for adult hippocampal neurogenesis. *J. Comp. Neurol.* 425(4), 479–494 (2000).
- 49 Shen Q, Goderie SK, Jin L *et al*. Endothelial cells stimulate self-renewal and expand neurogenesis of neural stem cells. *Science* 304(5675), 1338–1340 (2004).
- 50 Goodrich LV, Milenkovic L, Higgins KM, Scott MP. Altered neural cell fates and medulloblastoma in mouse patched mutants. *Science* 277(5329), 1109–1113 (1997).
- 51 Raffel C, Jenkins RB, Frederick L *et al*. Sporadic medulloblastomas contain PTCH mutations. *Cancer Res* 57(5), 842–845 (1997).
- 52 Reifemberger J, Wolter M, Weber RG *et al*. Missense mutations in SMOH in sporadic basal cell carcinomas of the skin and primitive neuroectodermal tumors of the central nervous system. *Cancer Res* 58(9), 1798–1803 (1998).
- 53 Palma V, Ruiz i Altaba A. Hedgehog-Gli signaling regulates the behavior of cells with stem cell properties in the developing neocortex. *Development* 131(2), 337–345 (2004).
- 54 Taylor MD, Liu L, Raffel C *et al*. Mutations in SUFU predispose to medulloblastoma. *Nat. Genet.* 31(3), 306–310 (2002).
- 55 Kinzler KW, Bigner SH, Bigner DD *et al*. Identification of an amplified, highly expressed gene in a human glioma. *Science* 236(4797), 70–73 (1987).
- 56 Wallace VA. Purkinje-cell-derived sonic hedgehog regulates granule neuron precursor cell proliferation in the developing mouse cerebellum. *Curr. Biol.* 9(8), 445–448 (1999).
- 57 Wechsler-Reya RJ, Scott MP. Control of neuronal precursor proliferation in the cerebellum by sonic hedgehog. *Neuron* 22(1), 103–114 (1999).
- 58 Corrales JD, Rocco GL, Blaess S, Guo Q, Joyner AL. Spatial pattern of sonic hedgehog signaling through Gli genes during cerebellum development. *Development* 131(22), 5581–5590 (2004).



- 59 Corrales JD, Blaess S, Mahoney EM, Joyner AL. The level of sonic hedgehog signaling regulates the complexity of cerebellar foliation. *Development* 133(9), 1811–1821 (2006).
- 60 Dahmane N, Sánchez P, Gitton Y *et al.* The sonic hedgehog–Gli pathway regulates dorsal brain growth and tumorigenesis. *Development* 128(24), 5201–5212 (2001).
- **Demonstrated that sonic hedgehog produced by Purkinje cells regulates the division of granule cell precursors, providing a link between normal brain development and tumorigenesis.**
- 61 Clement V, Sanchez P, de Tribolet N, Radovanovic I, Ruiz i Altaba A. Hedgehog–Gli1 signaling regulates human glioma growth, cancer stem cell self-renewal, and tumorigenicity. *Curr. Biol.* 17(2), 165–172 (2007).
- 62 Huang H, Mahler-Araujo BM, Sankila A *et al.* APC mutations in sporadic medulloblastomas. *Am. J. Pathol.* 156(2), 433–437 (2000).
- 63 Zurawel RH, Chiappa SA, Allen C, Raffel C. Sporadic medulloblastomas contain oncogenic  $\beta$ -catenin mutations. *Cancer Res.* 58(5), 896–899 (1998).
- 64 Dahmen RP, Koch A, Denkhäus D *et al.* Deletions of AXIN1, a component of the WNT/wingless pathway, in sporadic medulloblastomas. *Cancer Res.* 61(19), 7039–7043 (2001).
- 65 McMahon AP, Bradley A. The Wnt-1 (int-1) proto-oncogene is required for development of a large region of the mouse brain. *Cell* 62(6), 1073–1085 (1990).
- 66 Thomas KR, Musci TS, Neumann PE, Capocchi MR. Swaying is a mutant allele of the proto-oncogene *Wnt-1*. *Cell* 67(5), 969–976 (1991).
- 67 Lie DC, Colamarino SA, Song HJ *et al.* Wnt signaling regulates adult hippocampal neurogenesis. *Nature* 437(7063), 1370–1375 (2005).
- 68 Willert K, Brown JD, Danenberg E *et al.* Wnt proteins are lipid-modified and can act as stem cell growth factors. *Nature* 423(6938), 448–452 (2003).
- 69 Wei Q, Clarke L, Scheidenhelm DK *et al.* High-grade glioma formation results from postnatal pten loss or mutant epidermal growth factor receptor expression in a transgenic mouse glioma model. *Cancer Res.* 66(15), 7429–7437 (2006).
- 70 Holland EC, Hively WP, DePinho RA, Varmus HE. A constitutively active epidermal growth factor receptor cooperates with disruption of G1 cell-cycle arrest pathways to induce glioma-like lesions in mice. *Genes Dev.* 12(23), 3675–3685 (1998).
- 71 Jackson EL, Garcia-Verdugo JM, Gil-Perotin S *et al.* PDGFR  $\alpha$ -positive B cells are neural stem cells in the adult SVZ that form glioma-like growths in response to increased PDGF signaling. *Neuron* 51(2), 187–199 (2006).
- **Demonstrated that expression of PDGF in neural stem cells *in vivo* arrests neurogenesis and induces glial cell changes associated with the early stages of tumor formation.**
- 72 Lokker NA, Sullivan CM, Hollenbach SJ, Israel MA, Giese NA. Platelet-derived growth factor (PDGF) autocrine signaling regulates survival and mitogenic pathways in glioblastoma cells: evidence that the novel PDGF-C and PDGF-D ligands may play a role in the development of brain tumors. *Cancer Res.* 62(13), 3729–3735 (2002).
- 73 Dai C, Celestino JC, Okada Y, Louis DN, Fuller GN, Holland EC. PDGF autocrine stimulation dedifferentiates cultured astrocytes and induces oligodendrogliomas and oligoastrocytomas from neural progenitors and astrocytes *in vivo*. *Genes Dev.* 15(15), 1913–1925 (2001).
- **Demonstrated that autocrine stimulation of glial precursor cells by PDGF blocks glial cell differentiation and results in glioma formation.**
- 74 Clarke ID, Dirks PB. A human brain tumor-derived PDGFR- $\alpha$  deletion mutant is transforming. *Oncogene* 22(5), 722–733 (2003).
- 75 Dasgupta B, Gutmann DH. Neurofibromin regulates neural stem cell proliferation, survival, and astroglial differentiation *in vitro* and *in vivo*. *J. Neurosci.* 25(23), 5584–5594 (2005).
- 76 Bajenaru ML, Hernandez MR, Perry A *et al.* Optic nerve glioma in mice requires astrocyte *Nf1* gene inactivation and *Nf1* brain heterozygosity. *Cancer Res.* 63(24), 8573–8577 (2003).
- **Demonstrated that mouse optic gliomas in neurofibromatosis-1 (NF1) require a permissive brain environment composed of cells heterozygous for a mutation in the *Nf1* gene.**
- 77 Zhu Y, Harada T, Liu L *et al.* Inactivation of NF1 in CNS causes increased glial progenitor proliferation and optic glioma formation. *Development* 132(24), 5577–5588 (2005).
- 78 Groszer M, Erickson R, Scripture-Adams SD *et al.* Negative regulation of neural stem/progenitor cell proliferation by the Pten tumor suppressor gene *in vivo*. *Science* 294(5549), 2186–2189 (2001).
- 79 Groszer M, Erickson R, Scripture-Adams DD *et al.* PTEN negatively regulates neural stem cell self-renewal by modulating G0–G1 cell cycle entry. *Proc. Natl Acad. Sci. USA* 103(1), 111–116 (2006).
- 80 Xiao A, Yin C, Yang C, Di Cristofano A, Pandolfi PP, Van Dyke T. Somatic induction of Pten loss in a preclinical astrocytoma model reveals major roles in disease progression and avenues for target discovery and validation. *Cancer Res.* 65(12), 5172–5180 (2005).
- 81 Ohgaki H, Kleihues P. Genetic pathways to primary and secondary glioblastoma. *Am. J. Pathol.* 170(5), 1445–1453 (2007).
- 82 Gil-Perotin S, Marin-Husstege M, Li J *et al.* Loss of p53 induces changes in the behavior of subventricular zone cells: implication for the genesis of glial tumors. *J. Neurosci.* 26(4), 1107–1116 (2006).
- 83 Molofsky AV, He S, Bydon M, Morrison SJ, Pardoll R. Bmi-1 promotes neural stem cell self-renewal and neural development but not mouse growth and survival by repressing the p16<sup>Ink4a</sup> and p19<sup>Arf</sup> senescence pathways. *Genes Dev.* 19(12), 1432–1437 (2005).
- 84 Bruggeman SW, Valk-Lingbeek ME, van der Stoop PP *et al.* Ink4a and Arf differentially affect cell proliferation and neural stem cell self-renewal in Bmi1-deficient mice. *Genes Dev.* 19(12), 1438–1443 (2005).
- 85 Reilly KM, Loisel DA, Bronson RT, McLaughlin ME, Jacks T. Nf1; Trp53 mutant mice develop glioblastoma with evidence of strain-specific effects. *Nat. Genet.* 26(1), 109–113 (2000).
- 86 Marino S, Vooijs M, van Der Gulden H, Jonkers J, Berns A. Induction of medulloblastomas in p53-null mutant mice by somatic inactivation of Rb in the external granular layer cells of the cerebellum. *Genes Dev.* 14(8), 994–1004 (2000).
- 87 Zhu Y, Guignard F, Zhao D *et al.* Early inactivation of p53 tumor suppressor gene cooperating with NF1 loss induces malignant astrocytoma. *Cancer Cell* 8(2), 119–130 (2005).
- 88 Kim HT, Kim IS, Lee IS *et al.* Human neurospheres derived from the fetal central nervous system are regionally and temporally specified but are not committed. *Exp. Neurol.* 199(1), 222–235 (2006).

- 89 Hitoshi S, Tropepe V, Ekker M, van der Kooy D. Neural stem cell lineages are regionally specified, but not committed, within distinct compartments of the developing brain. *Development* 129(1), 233–244 (2002).
- 90 Fu SL, Ma ZW, Yin L, Iannotti C, Lu PH, Xu XM. Region-specific growth properties and trophic requirements of brain- and spinal cord-derived rat embryonic neural precursor cells. *Neuroscience* 135(3), 851–862 (2005).
- 91 Horiguchi S, Takahashi J, Kishi Y *et al.* Neural precursor cells derived from human embryonic brain retain regional specificity. *J. Neurosci. Res.* 75(6), 817–824 (2004).
- 92 Sharma MK, Mansur DB, Reifenger G *et al.* Distinct genetic signatures among pilocytic astrocytomas relate to their brain region origin. *Cancer Res.* 67(3), 890–900 (2007).
- **Demonstrated that glial tumors share an intrinsic, lineage-specific molecular signature that reflects the brain region in which their nonmalignant predecessors originated.**
- 93 Gilbertson RJ, Gutmann DH. Tumorigenesis in the brain: location, location, location. *Cancer Res.* 67(12), 5579–5582 (2007).
- 94 Bao S, Wu Q, McLendon RE *et al.* Glioma stem cells promote radioresistance by preferential activation of the DNA damage response. *Nature* 444(7120), 756–760 (2006).
- 95 Hambardzumyan D, Squatrito M, Holland EC. Radiation resistance and stem-like cells in brain tumors. *Cancer Cell* 10(6), 454–456 (2006).
- 96 Dean M, Fojo T, Bates S. Tumour stem cells and drug resistance. *Nat. Rev. Cancer* 5(4), 275–284 (2005).
- 97 Liu G, Yuan X, Zeng Z *et al.* Analysis of gene expression and chemoresistance of CD133<sup>+</sup> cancer stem cells in glioblastoma. *Mol. Cancer* 5, 67 (2006).
- 98 Eramo A, Ricci-Vitiani L, Zeuner A *et al.* Chemotherapy resistance of glioblastoma stem cells. *Cell Death Differ.* 13(7), 1238–1241 (2006).
- 99 Spira AI, Carducci MA. Differentiation therapy. *Curr. Opin. Pharmacol.* 3(4), 338–343 (2003).
- 100 Ohno R, Asou N, Ohnishi K. Treatment of acute promyelocytic leukemia: strategy toward further increase of cure rate. *Leukemia* 17(8), 1454–1463 (2003).
- 101 Sell S. Stem cell origin of cancer and differentiation therapy. *Crit. Rev. Oncol. Hematol.* 51(1), 1–28 (2004).
- 102 Piccirillo SG, Reynolds BA, Zanetti N *et al.* Bone morphogenetic proteins inhibit the tumorigenic potential of human brain tumour-initiating cells. *Nature* 444(7120), 761–765 (2006).
- 103 Aguado T, Carracedo A, Julien B *et al.* Cannabinoids induce glioma stem-like cell differentiation and inhibit gliomagenesis. *J. Biol. Chem.* 282(9), 6854–6862 (2007).
- 104 Cheng T. Cell cycle inhibitors in normal and tumor stem cells. *Oncogene* 23(43), 7256–7266 (2004).
- 105 Berman DM, Karhadkar SS, Hallahan AR *et al.* Medulloblastoma growth inhibition by hedgehog pathway blockade. *Science* 297(5586), 1559–1561 (2002).
- 106 Bar EE, Chaudhry A, Lin A *et al.* Cyclopamine-mediated hedgehog pathway inhibition depletes stem-like cancer cells in glioblastoma. *Stem Cells* 25(10), 2524–2533 (2007).
- 107 Jain RK, di Tomaso E, Duda DG, Loeffler JS, Sorensen AG, Batchelor TT. Angiogenesis in brain tumours. *Nat. Rev. Neurosci.* 8(8), 610–622 (2007).

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