



# Practice: ChIP-seq data analysis

**USF Master Program in Bioinformatics** 

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## Steps

- Setting up R/Rstudio
- Data preparation
- Exploratory data analysis (QC)
- Peak detection







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#### R Project

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# The R Project for Statistical Computing

### **Getting Started**

R is a free software environment for statistical computing and graphics. It compiles and runs on a wide variety of UNIX platforms, Windows and MacOS. To **download R**, please choose your preferred CRAN mirror.

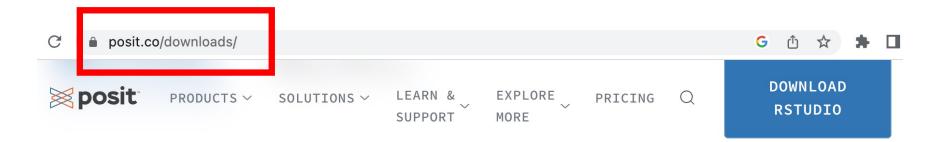
If you have questions about R like how to download and install the software, or what the license terms are, please read our answers to frequently asked questions before you send an email.

#### **News**

- R version 4.3.0 (Already Tomorrow) prerelease versions will appear starting
   Tuesday 2023-03-21. Final release is scheduled for Friday 2023-04-21.
- R version 4.2.3 (Shortstop Beagle) has been released on 2023-03-15.
- R version 4.1.3 (One Push-Up) was released on 2022-03-10.



### **Install Rstudio**



# RStudio Desktop

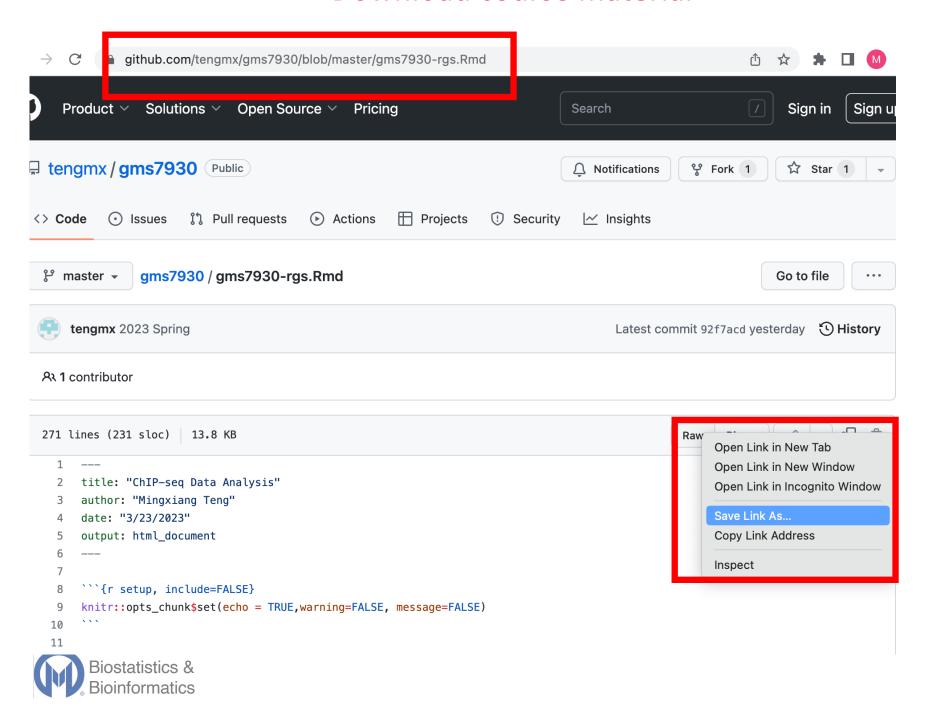
Find out more about RStudio Desktop and RStudio Desktop Pro below.

DOWNLOAD RSTUDIO

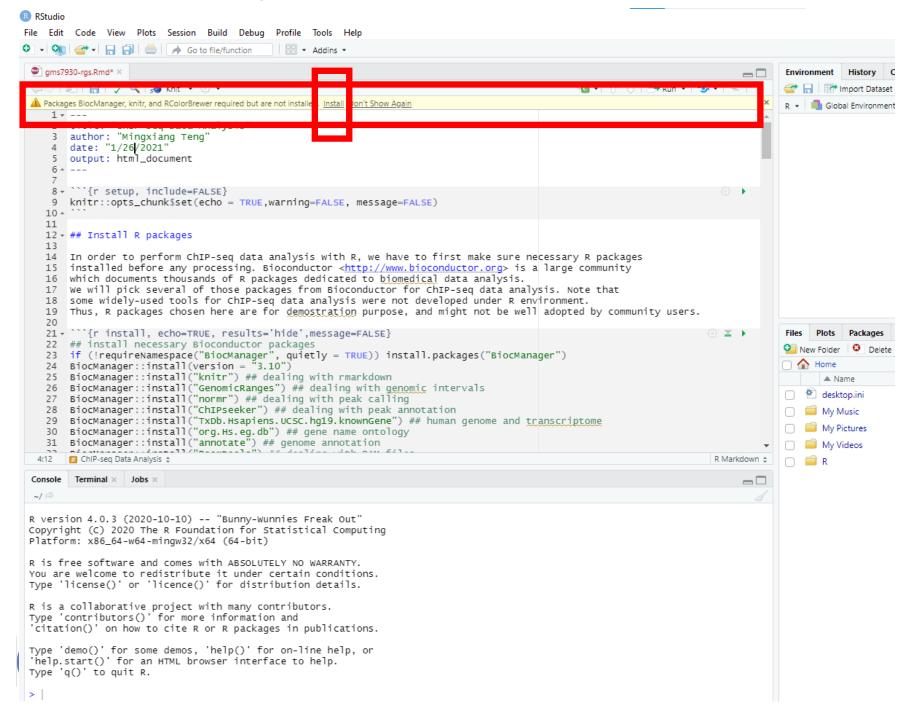




#### Download course material

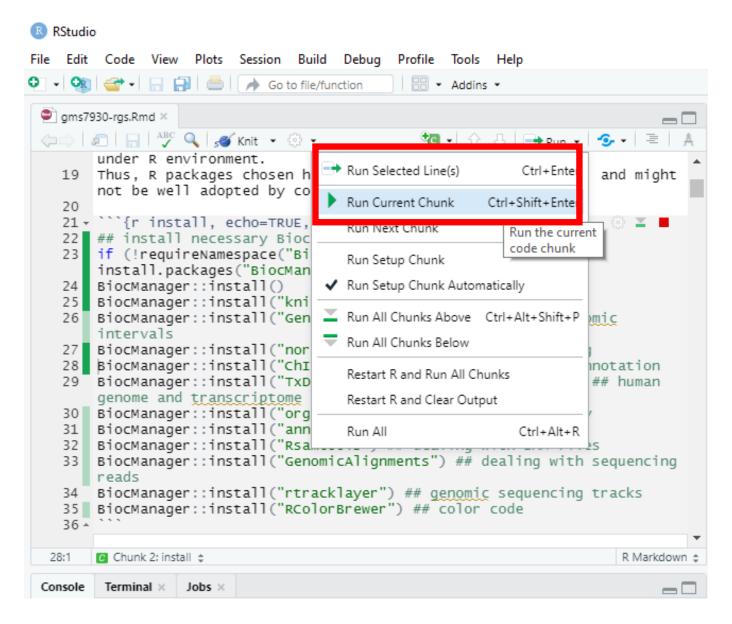


## Open .Rmd file in Rstudio



```
1
                  2 title: "ChIP-seq Data Analysis"
                      author: "Mingxiang Teng"
                      date: "3/23/2023"
                      output: html_document
                      ```{r setup, include=FALSE}
                  9
                      knitr::opts_chunk$set(echo = TRUE, warning=FALSE, message=FALSE)
                  10
                  11
   Non-code text
                  12
                      ## Install R packages
                     In order to perform ChIP-seg data analysis with R, we have to first make sure necessary R packages
                  14
                      installed before any processing. Bioconductor <a href="http://www.bioconductor.org">http://www.bioconductor.org</a> is a large community
                      which documents thousands of R packages dedicated to biomedical data analysis.
                      We will pick several of those packages from Bioconductor for ChIP-seq data analysis. Note that
                      some widely-used tools for ChIP-seq data analysis were not developed under R environment.
                  20
                  21
                      ```{r install, echo=TRUE, results='hide',message=FALSE}
                  23 r = getOption("repos")
                     r["CRAN"] = "http://cran.us.r-project.org" #https://cran.rstudio.com/
                  25
                      options(repos = r)
                      if (!requireNamespace("BiocManager", quietly = TRUE))
                        install.packages("BiocManager")
                  27
                                                                                                                           Install R packages
                      BiocManager::install()
                  28
                      install.packages('rmarkdown')
                  29
                      BiocManager::install("knitr") ## dealing with rmarkdown
                      BiocManager::install("GenomicRanges") ## dealing with genomic intervals
                  31
                      BiocManager::install("normr") ## dealing with peak calling
                      BiocManager::install("ChIPseeker") ## dealing with peak annotation
                  33
                      BiocManager::install("TxDb.Hsapiens.UCSC.hq38.knownGene") ## human genome and transcriptome
                  35
                      BiocManager::install("GenomeInfoDb")
                  36
                      BiocManager::install("BSgenome.Hsapiens.UCSC.hg38")
                  37
                      BiocManager::install("org.Hs.eg.db") ## gene name ontology
                  38
                      BiocManager::install("annotate") ## genome annotation
                      BiocManager::install("Rsamtools") ## dealing with BAM files
                  39
                      BiocManager::install("GenomicAlignments") ## dealing with sequencing reads
                      BiocManager::install("rtracklayer") ## genomic sequencing tracks
                  41
                      BiocManager::install("RColorBrewer") ## color code
                  44
                  45
                      After all required packages installed, we need to load these packages.
                  46
                         `{r load, echo=TRUE, results='hide',message=FALSE}
                  47
                      library(Rsamtools)
                      library(GenomicAlignments)
                      library(GenomicRanges)
                  50
                      library(normr)
                                                                                                                              Load R packages
                      library(TxDb.Hsapiens.UCSC.hg38.knownGene)
                  53
                      library(BSgenome.Hsapiens.UCSC.hg38)
                      library(org.Hs.eg.db)
                      library(ChIPseeker)
Biostatistic
                      library(annotate)
                      library(rtracklayer)
                      library(RColorBrewer)
```

#### How to run R code in .Rmd file





## Setting up R analysis environment

```
## install necessary Bioconductor packages
if (!requireNamespace("BiocManager", quietly = TRUE)) install.packages("BiocManager")
BiocManager::install(version = "3.10")
BiocManager::install("knitr") ## dealing with rmarkdown
BiocManager::install("GenomicRanges") ## dealing with genomic intervals
BiocManager::install("normr") ## dealing with peak calling
BiocManager::install("ChIPseeker") ## dealing with peak annotation
BiocManager::install("TxDb.Hsapiens.UCSC.hg19.knownGene") ## human genome and transcriptome
BiocManager::install("org.Hs.eg.db") ## gene name ontology
BiocManager::install("annotate") ## genome annotation
BiocManager::install("Rsamtools") ## dealing with BAM files
BiocManager::install("GenomicAlignments") ## dealing with sequencing reads
BiocManager::install("rtracklayer") ## genomic sequencing tracks
BiocManager::install("RColorBrewer") ## color code
```

Make sure to install these R packages before data analysis



## Questions you need to answer if installing packages one by one

```
** byte-compile and prepare package for lazy loading
** help
*** installing help indices
  converting help for package 'TxDb. Hsapiens. UCSC. hg19. knownGene'
    finding HTML links ... done
    package
                                             html
** building package indices
** testing if installed package can be loaded from temporary location
** testing if installed package can be loaded from final location
** testing if installed package keeps a record of temporary installation pat
* DONE (TxDb. Hsapiens. UCSC. hg19. knownGene)
The downloaded source packages are in
        'C:\Users\tengm\AppData\Local\Temp\Rtmpw50ztw\downloaded_packages'
Undate all/some/none? [a/s/n]:
n
```



# Data to be analyzed

ChIP: Gm12878Ctcf.bam

INPUT: Gm12878Control.bam

```
workdir = '~/' ##
download.file('https://github.com/tengmx/gms7930/raw/master/data/Gm12878Control.bam',paste0(workdir,'Gm12878Control.bam'))
download.file('https://github.com/tengmx/gms7930/raw/master/data/Gm12878Control.bam.bai',paste0(workdir,'Gm12878Control.bam.bai'))
download.file('https://github.com/tengmx/gms7930/raw/master/data/Gm12878Ctcf.bam',paste0(workdir,'Gm12878Ctcf.bam'))
download.file('https://github.com/tengmx/gms7930/raw/master/data/Gm12878Ctcf.bam.bai',paste0(workdir,'Gm12878Ctcf.bam.bai'))
```



## Data table generation

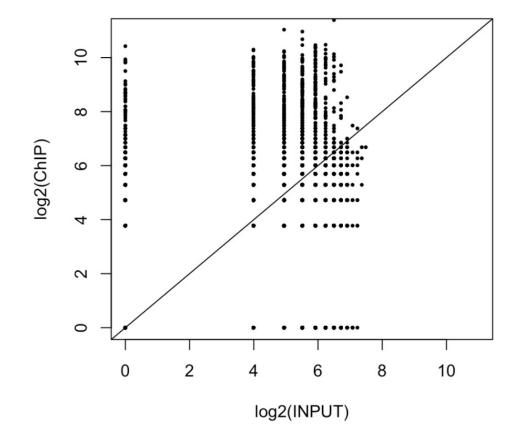
```
"``{r bam, echo=TRUE}
## sequencing files that only contain reads from chromosome 21
chip = file.path(workdir,'Gm12878Ctcf.bam')
input = file.path(workdir,'Gm12878Control.bam')
## chromosome length info from human genome build hg19
hg_chrs = getBSgenome("hg38")
seqlen_chr21 = seqlengths(hg_chrs)['chr21']
## create genomic 1000bp windows and store them using GenomicRanges
window_gr = unlist(tileGenome(seqlen_chr21, tilewidth=1000))
## count reads from both files for all windows
rc = summarizeOverlaps(window_gr, c(chip, input))
rc = assays(rc)[[1]]
## simple normalization based on library size
cpm_chr21 = t(t(rc)*(1000000/colSums(rc)))
head(cpm_chr21)
""
```

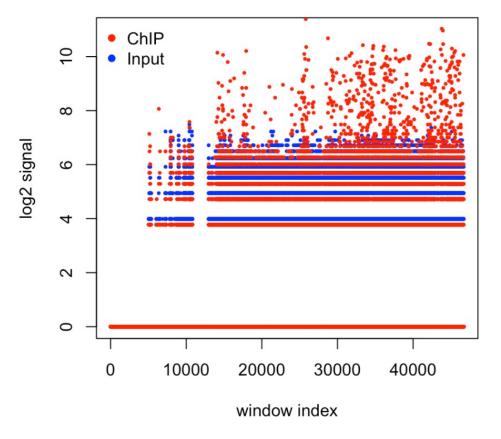
#### 



# Exploratory comparing ChIP to Input

```
par(mfrow=c(1,2))
## read counts in all windows
plot(log2(cpm_chr21[,2]+1),log2(cpm_chr21[,1]+1),pch=16,cex=0.5,xlab='log2(INPUT)',ylab='log2(ChIP)',ylim=c(0,11),xlim=c(0,11))
abline(a=0,b=1)
## read counts along the chromosome
plot(log2(cpm_chr21[,2]+1),col='blue',pch=16,cex=0.5,xlab='window index',ylab='log2 signal',ylim=c(0,11))
points(log2(cpm_chr21[,1]+1),col='red',pch=16,cex=0.5)
legend('topleft',c('ChIP','Input'),pch=16,col=c('red','blue'),bty='n')
```





## Peak detection at bin level

```
```{r range, echo=TRUE,message=FALSE}
## formalizing peak info with GRanges container
peaks = getRanges(peakfit)
peaks$sig = getEnrichment(peakfit) ## add enrichmend signal
peaks$pvalue = getPvalues(peakfit) ## add enrichment significance
peaks
```

GRanges object with 12353090 ranges and 3 metadata columns:

	seqnames	ranges	strand	1	component	sig	pvalue
	<rle></rle>	<iranges></iranges>	<rle></rle>	1	<integer></integer>	<numeric></numeric>	<numeric></numeric>
[1]	chr1	1-250	*	1	<na></na>	-8.27017e-18	1
[2]	chr1	251-500	*	Ι	<na></na>	-8.27017e-18	1
[3]	chr1	501-750	*	1	<na></na>	-8.27017e-18	1
[4]	chr1	751-1000	*	1	<na></na>	-8.27017e-18	1
[5]	chr1	1001-1250	*	1	<na></na>	-8.27017e-18	1
• • •	• • •	• • •		•			
[12353086]	chrY	57226251-57226500	*	I	<na></na>	-8.27017e-18	1
[12353087]	chrY	57226501-57226750	*	1	<na></na>	-8.27017e-18	1
[12353088]	chrY	57226751-57227000	*	1	<na></na>	-8.27017e-18	1
[12353089]	chrY	57227001-57227250	*	1	<na></na>	-8.27017e-18	1
[12353090]	chrY	57227251-57227415	*	I	<na></na>	-8.27017e-18	1

seqinfo: 24 sequences from an unspecified genome

# Select significant peaks

```
```{r peaksig, echo=TRUE, message=TRUE}
## only consider a peak as significantly enriched if its q-value is less than 0.01
peakssig = peaks[which(peaks$pvalue<0.01)]
peakssig = peakssig[order(peakssig$pvalue)]
peakssig
```</pre>
```

GRanges object with 706 ranges and 3 metadata columns:

```
ranges strand | component
  sia
   pvalue
      seanames
        <Rle>
                       <IRanges> <Rle> | <integer> <numeric>
  <numeric>
        chr21 25801501-25801750
                                      * |
  1
  1.24785 9.73874e-35
 [1]
 Γ27
        chr21 25801751-25802000
                                      * |
  1.18900 3.17057e-28
 Γ37
        chr21 43789251-43789500
  1 1.17497 7.37378e-27
  1 2.86317 9.60544e-27
 Γ47
        chr21 43939251-43939500
 Γ57
        chr21 43975501-43975750
  2.85912 2.15827e-26
[702]
        chr21 37582501-37582750
  1 0.683697 0.00837267
[703]
        chr21 42310001-42310250
   0.683697 0.00837267
[704]
        chr21 44298001-44298250
   0.683697
   0.00837267
[705]
        chr21 45308751-45309000
   0.683697 0.00837267
[706T
        chr21 45477751-45478000
  0.683697 0.00837267
```

-----

seqinfo: 24 sequences from an unspecified genome