

Cell-type-specific-GSA on test dataset

```
library(mremaR)
```

Here we will show a simple workflow for carrying out cell type-specific gene set analysis (CT-GSA). First let's load the CAR-seq package which is designed for carrying out cell type-specific differential expression analysis.

```
#devtools::install_github("chongjin/CARseq")  
library(CARseq)
```

##Simulation here is shown as we are running the test dataset. No need to do simulations on real data Now we will simulate gene expression for samples which are a mixture of six different cell types. This defaults to 1,000 genes in 100 cases and 100 controls. In each cell type 5% of genes are assigned a log-fold-change drawn from a normal distribution with a mean of plus/minus 4 and a standard deviation of 0.2.

```
sims <- CTexpSimulation(g = 1000, n = 100, p = 1, m_lfc = 0, sd_lfc = 1)  
sims$bulk.expression[c(1:5),c(1:5)] # bulk expression counts
```

```
##           sample.1 sample.2 sample.3 sample.4 sample.5  
## gene.1         162       162       142       144       117  
## gene.2          53        45        61        50        57  
## gene.3         467       452       378       498       552  
## gene.4         433       450       514       530       539  
## gene.5          33        45        41        50        33
```

```
sims$ols.mixture.estimate[c(1:5),] # cell type proportion estimates
```

```
##           cellType.1 cellType.2 cellType.3 cellType.4 cellType.5 cellType.6  
## sample.1 0.128638807 0.02636877 0.06552044 0.1713814 0.1404961 0.4675944  
## sample.2 0.007651266 0.10456251 0.10961245 0.1714795 0.1483554 0.4583389  
## sample.3 0.028606660 0.03348568 0.14458040 0.1218862 0.2342001 0.4372409  
## sample.4 0.067769561 0.01812786 0.03724474 0.2142052 0.2428260 0.4198267  
## sample.5 0.080075846 0.04148809 0.02638179 0.2954996 0.1986001 0.3579546
```

```
table(sims$trait) # sample traits
```

```
##  
## 0 1  
## 50 50
```

These outputs can then be passed to the CAR-seq package. The important results for us are the `shrunkened_lfc` and `shrunkened_lfcSE` matrices

```
# this will take a minute or two to run
res <- run_CARseq(
  count_matrix = sims$bulk.expression,
  cellular_proportions = sims$ols.mixture.estimate,
  groups = sims$trait,
  cores = 10 ##number of cores can be changed
)
```

As we are running on a test data, we are simulating a few gene sets. For original data, use open available datasets or the link provided below:

#kegg gene sets data

```
#kegg.gs <- "https://raw.githubusercontent.com/osedo/GSA-MREMA/main/real_data/kegg.RData"
```

```
#load(url(kegg.gs))
```

Since the gene set mentioned above contains genes in ENTREZ IDs, I recommend converting the gene names from Ensembl IDs to ENTREZ IDs. Alternatively, you can load a gene set dataset that uses Ensembl IDs or another dataset with your preferred ID format using `ensemblDb` or `gProfiler` R packages.

Let's simulate a few gene sets

```
# a few power gene sets, gs1 enriched for DE genes in cellType1, same for gs2 and so on
# we will give sets of 100 approx 20% DE gene
g <- 1000
gs.power <- lapply(c(1:6), function(x){
  ct_de <- which(abs(sims$simulated.lfc[,x]) > log2(2.5))
  ct_nonde <- c(1:g)[!c(1:g) %in% ct_de]
  ct_de <- paste0("gene.", ct_de)
  ct_nonde <- paste0("gene.", ct_nonde)
  genes_per_set <- 100
  p <- mean(abs(sims$simulated.lfc[,x]) > log2(2.5))
  enrichment <- 3
  c(sample(ct_de, genes_per_set*enrichment*p),
    sample(ct_nonde, (genes_per_set-genes_per_set*enrichment*p)))
})

# random gene sets
gs.null <- lapply(c(7:20), function(x){
  paste0("gene.", sample.int(1000, 100))
})

gs <- c(gs.power, gs.null)
names(gs) <- paste0("gs", 1:20)
```

Let's run `mrema()` on all cell-types

##parametric tests

```
mrema.res <- lapply(c(1:6), function(ct){
  postdata <- data.frame("Ensembl" = rownames(res$shrunk_lfcSE),
    "effect" = res$shrunk_lfc[,ct],
    "variance" = res$shrunk_lfcSE[,ct]^2,
    "pval" = res$p[,ct])
})
```

```

mrema_res <- mrema(postdata = postdata, raw.gs = gs, threshold = 2, DF = 4, ncores = 1)
mrema_res
})
names(mrema.res) <- colnames(res$p)
saveRDS(mrema.res, file = "samplepara.RDS")

```

##non-parametric tests – doesnt make any assumptions about the LFC distribution of a group of genes

```

full.gsa <- lapply(c(1:6), function(ct){
  data <- data.frame("genes" = rownames(res$shrunk_lfc),
                    "lfc" = res$shrunk_lfc[,ct],
                    "lfcSE" = res$shrunk_lfcSE[,ct])
  gsa.res <- REtest(data, log2(2), gs)
  gsa.res
})
names(full.gsa) <- colnames(res$p)[c(1:6)]
saveRDS(full.gsa, file = "samplenonpara.RDS")

```

We can now launch the shiny app to investigate our results

```

mremApp()    #parametric tests
REshine()    #non-parametric tests

```