# bacs\_hw5

110071010

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# **Backstory**

Verizon was an Incumbent Local Exchange Carrier (ILEC), responsible for maintaining land-line phone service in certain areas. Other competing providers, termed Competitive Local Exchange Carriers (CLEC), could also sell long-distance phone services in Verizon's areas. When something went wrong, Verizon would be responsible to respond and repair services as quickly for CLEC long-distance customers as for its own ILEC customers. The New York Public Utilities Commission (PUC) monitored fairness by comparing Verizon's response times for its ILEC customers versus CLEC customers. In each case, a hypothesis test was performed at the 1% significance level, to determine whether response times for CLEC customers were significantly slower than for Verizon's customers. If Verizon failed to provide fair treatment for CLEC customers, it would have to pay large penalties.

Verizon claims that mean response time for ILEC and CLEC customers are the same, but the PUC would like to test if CLEC customers were facing greater response times.

# Question 1

The Verizon dataset this week is provided as a "wide" data frame. Let's practice reshaping it to a "long" data frame

# **Getting data ready**

```
# Reading in the file
response_wide <- read.csv("verizon_wide.csv")
View(response_wide)</pre>
```

**1**a

#### Instruction

Pick a reshaping package – research them online and tell us why you picked it over others.

#### Answer

I chose tidyr over reshape2 for several reasons. For one thing, tidyr is more intuitive and integrated with tidyverse. For another, while reshape2 is powerful for more

complex reshaping tasks and works with a wider range of data types including matrices and arrays, this homework does not require such heavy data manipulation.

**1**b

## **Instruction**

Show the code to reshape the verizon\_wide.csv sample

```
# Reshaping the response_wide sample into response_long
library(tidyr)
response_long <- gather(response_wide, na.rm = TRUE, key = "host", valu
e = "response_time")</pre>
```

**1c** 

## **Instruction**

Show us the "head" and "tail" of the data to show that the reshaping worked

```
head(response_long)
     host response time
##
                  17.50
## 1 ILEC
## 2 ILEC
                   2.40
                   0.00
## 3 ILEC
## 4 ILEC
                   0.65
## 5 ILEC
                  22.23
## 6 ILEC
                   1.20
tail(response_long)
##
        host response time
                     24.20
## 1682 CLEC
## 1683 CLEC
                     22.13
## 1684 CLEC
                     18.57
                     20.00
## 1685 CLEC
## 1686 CLEC
                     14.13
## 1687 CLEC
                      5.80
```

**1d** 

## **Instruction**

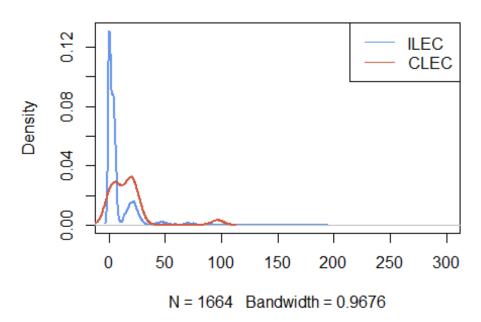
Visualize Verizon's response times for ILEC vs. CLEC customers

```
hosts <- split(x = response_long$response_time, f = response_long$host)

plot(density(hosts$ILEC), col = "cornflowerblue", lwd = 2, xlim= c(0,30
0), main = "Response Times of ILEC and CLEC")
lines(density(hosts$CLEC), col = "coral3", lwd = 2)</pre>
```

```
legend("topright", lty = 1, c("ILEC","CLEC"), col = c("cornflowerblue",
"coral3"))
```

# Response Times of ILEC and CLEC



# **Question 2**

Let's test if the mean of response times for CLEC customers is greater than for ILEC customers

## **2**a

# **Instruction**

State the appropriate null and alternative hypotheses (one-tailed)

### Answer

**Null Hypothesis H0:** Mean of CLEC customers' response times <u>is equal or less</u> compared to that of ILEC customers' response times

**Alternative Hypothesis H1:** Mean of CLEC customers' response times <u>is greater</u> than that of ILEC customers' response times

### 2b

## **Instruction**

Use the appropriate form of the t.test() function to test the difference between the mean of ILEC versus CLEC response times at 1% significance. For each of the following tests, show us the results and tell us whether you would reject the null hypothesis.

(i)

Conduct the test assuming variances of the two populations are equal

(ii)

Conduct the test assuming variances of the two populations are not equal

**2**c

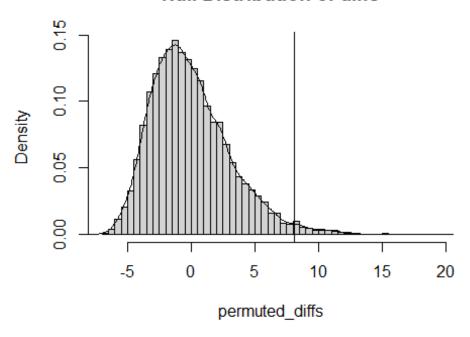
## Instruction

Use a permutation test to compare the means of ILEC vs. CLEC response times

Visualize the distribution of permuted differences, and indicate the observed difference as well.

```
# Setting seed
set.seed(123123)
# Observed difference
observed_diff <- mean(hosts$CLEC) - mean(hosts$ILEC)</pre>
# Getting the function ready for permutation
permute_diff <- function(values, groups){</pre>
  permuted <- sample(values, replace = FALSE)</pre>
 grouped <- split(permuted, groups)</pre>
 permute_diff <- mean(grouped$CLEC) - mean(grouped$ILEC)</pre>
}
# Visualize after 10000 permutations
nperms = 10000
permuted_diffs <- replicate(nperms, permute_diff(response_long$response)</pre>
_time, response_long$host))
hist(permuted_diffs, breaks = "fd", probability = TRUE, main = "Null Di
stribution of diffs")
lines(density(permuted diffs))
# Displaying observed difference
abline(v = observed diff)
```

# **Null Distribution of diffs**



(ii)

What are the one-tailed and two-tailed p-values of the permutation test?

```
# one-tailed p-value
p_1tailed <- sum(permuted_diffs > observed_diff)/ nperms
p_1tailed

## [1] 0.0172

# two-tailed p-value
p_2tailed <- sum(abs(permuted_diffs) > observed_diff)/ nperms
p_2tailed

## [1] 0.0172
```

# Question

(iii)

Would you reject the null hypothesis at 1% significance in a one-tailed test?

# <u>Answer</u>

No, we would not reject H0 since p-value > 0.01

# **Question 3**

Let's use the Wilcoxon test to see if the response times for CLEC are different than ILEC.

**3**a

## **Instruction**

Compute the W statistic comparing the values. You may use either the permutation approach (try the functional form) or the rank sum approach

```
# Define gt_eq() for comparing values from two groups
gt_eq <- function(a,b){
   ifelse(a > b, 1, 0) + ifelse(a ==b, 0.5, 0)
}

# W statistic
W <- sum(outer(hosts$CLEC , hosts$ILEC, FUN = gt_eq))
W
## [1] 26820</pre>
```

3b

### Instruction

Compute the one-tailed p-value for W

```
# Lengths of the CLEC and ILEC groups to be used in the p-value calcula
tion
n1 <- length(hosts$CLEC)
n2 <- length(hosts$ILEC)

wilcox_p_1tail<- 1 - pwilcox(W,n1,n2)
wilcox_p_2tail <- 2 * wilcox_p_1tail
wilcox_p_1tail
## [1] 0.0003688341</pre>
```

**3**c

## **Instruction**

Run the Wilcoxon Test again using the wilcox.test() function in R

```
wilcox.test(hosts$CLEC, hosts$ILEC, alternative = "greater")
## Wilcoxon rank sum test with continuity correction
##
## data: hosts$CLEC and hosts$ILEC
```

```
## W = 26820, p-value = 0.0004565
## alternative hypothesis: true location shift is greater than 0
```

**3d** 

## Question

At 1% significance, and one-tailed, would you reject the null hypothesis that the values of CLEC and ILEC are similar?

### Answer

Yes, we would reject H0 since p-value < 0.01

# **Question 4**

One of the assumptions of some classical statistical tests is that our population data should be roughly normal. Let's explore one way of visualizing whether a sample of data is normally distributed.

4a

### **Instruction**

Make a function called norm\_qq\_plot() that takes a set of values

```
norm_qq_plot <- function(values) {
    # (i) Create a sequence of probability numbers from 0 to 1, with ~100
0 probabilities in between
    probs1000 <- seq(0, 1, 0.001)
    # (ii) Calculate ~1000 quantiles of our values
    q_vals <- quantile(values, probs = probs1000)
    # (iii) Calculate ~1000 quantiles of a perfectly normal distribution
with the same mean and standard deviation as our values
    q_norm <- qnorm(p = probs1000, mean = mean(values), sd = sd(values))
    # (iv) Create a scatterplot comparing the quantiles of a normal distribution versus quantiles of values
    plot(q_norm, q_vals, xlab="normal quantiles", ylab="values quantiles")
    # (v) Draw a red line with intercept of 0 and slope of 1, comparing these two sets of quantiles
    abline(a = 0, b = 1, col="red", lwd=2) # a:intercept, b:slope
}</pre>
```

4b

## **Instruction**

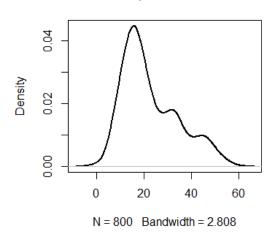
Confirm that your function works by running it against the values of our d123 distribution from week 3 and checking that it looks like the given plot

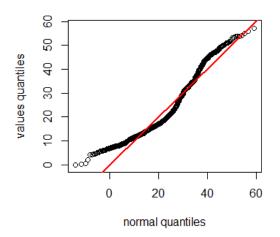
```
set.seed(123123)

d1 <- rnorm(n=500, mean=15, sd=5)
d2 <- rnorm(n=200, mean=30, sd=5)
d3 <- rnorm(n=100, mean=45, sd=5)
d123 <- c(d1, d2, d3)

par(mfrow = c(1,2))
plot(density(d123), main = "QQ plot of d123", lwd = 2)
norm qq plot(d123)</pre>
```

## QQ plot of d123





## **Comment**

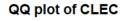
The QQplot of d123 suggests that it is not normally distributed. Rather, d123 appears to exhibit patterns of a bimodal distribution.

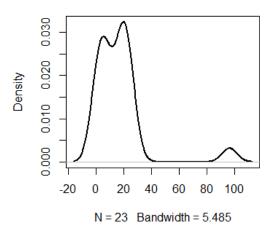
**4c** 

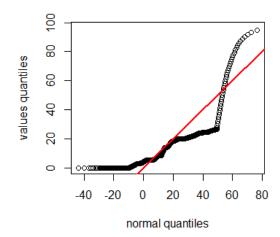
## **Instruction**

Traditional statistical t-tests to compare the means of two populations require that the two populations are normally distributed. Use your normal Q-Q plot function, norm\_qq\_plot, to check if the values from each of the CLEC and ILEC samples we compared in question 2 could be normally distributed. What's your conclusion?

```
# QQ-plotting for CLEC
par(mfrow = c(1,2))
response_wide <- response_wide[!is.na(response_wide$CLEC), ]
plot(density(response_wide$CLEC), main = "QQ plot of CLEC", lwd = 2)
norm_qq_plot(response_wide$CLEC)</pre>
```

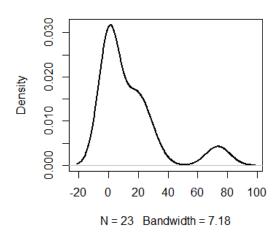


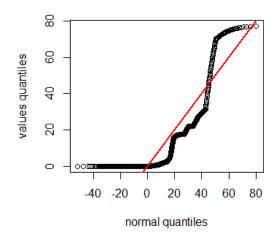




```
# QQ-plotting for ILEC
par(mfrow = c(1,2))
plot(density(response_wide$ILEC), main = "QQ plot of ILEC", lwd = 2)
norm_qq_plot(response_wide$ILEC)
```

# QQ plot of ILEC





# **Conclusion**

As observed in the graphs above, there are many unaligned data points in both CLEC and ILEC's QQplots, so one could easily conclude that both are not normally distributed.